REVIEW

Linking the metabolic syndrome and obesity with vitamin D status: risks and opportunities for improving cardiometabolic health and well-being

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¹School of Arts and Sciences, American University in Dubai, Dubai, UAE; ²Sunlight, Nutrition, and Health Research Center, San Francisco, CA 94164-1603, USA **Abstract:** The global death toll from noncommunicable diseases is exceptionally high, reported to cause 71% of global deaths worldwide. Metabolic syndrome risk factors, especially excessive adiposity and obesity, are at the heart of the problem resulting in increased comorbidities such as cardiometabolic diseases and cancer, increased health costs, poorer quality of life, and shortened survival. Vitamin D₃ can positively reverse many of these adverse effects and outcomes through blocking signaling mechanisms that predispose to cardiometabolic and metastatic disease. As an affordable natural agent, vitamin D₃ can be used to counteract obesity-induced inflammation, block early adipogenesis, enhance glucose uptake, counteract hyperleptinemia, ameliorate insulin resistance, and reduce hypertension. This is supported by data from in vitro, in vivo and epidemiological studies and clinical trials. We propose that everyone in general and obese patients in particular consider raising 25-hydroxyvitamin D levels through UVB exposure and/or supplemental vitamin D₃ intake to reduce cardiometabolic and metastatic disease and increase longevity.

Keywords: vitamin D, metabolic syndrome, obesity, adipose tissue, cardiovascular disease, risk reduction

Introduction

The metabolic syndrome is characterized by a number of inter-dependent risk factors or diseases that collectively increase a person's risk for developing type 2 diabetes mellitus (T2DM) and cardiovascular diseases (CVDs). These risk factors include elevated blood glucose levels, hypertension, dyslipidemia (specifically increased low--density lipoprotein (LDL) cholesterol, reduced high-density lipoprotein (HDL) cholesterol, and elevated triglycerides (TG)), increased fat mass and adiposity with intraabdominal fat accumulation, a large waist circumference and high waist to hip ratio. All of the above abnormalities lead to obesity, which is characterized by high body mass index (BMI) >30 kg/m² and increased adiposity. Obesity is also an independent risk factor of the metabolic syndrome and cardiometabolic diseases.¹ Obesity is thought to double the risk of co-morbidity and mortality from noncommunicable diseases (NCD) and their complications, including T2DM, CVDs, and cancer.^{2,3} This indicates a complex relationship among obesity, cardiometabolic diseases, T2DM, and cancer,⁴ suggesting that many mechanistic regulatory pathways are common, perhaps intersecting, and may be collectively modulated in our search for therapeutic solutions. At the intersection of all these diseases is increased adiposity and inflammation associated with increasing obesity levels.⁵

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Global obesity rates remain unacceptably high. The WHO estimates of 2016 indicated that over 1.9 billion adults were overweight and over 650 million were obese. Unfortunately, childhood obesity is also on the rise globally, with over 41 million children under the age of 5 years characterized as overweight or obese and 18% of children aged 4–18 years as obese.⁶ The global NCD burden remains shockingly high and is responsible for 71% of deaths worldwide.⁷ The cost of co-morbidities, loss of productivity, increases in associated conditions such as mental health decline and poor quality-of-life associated with rising incidence of obesity and metabolic syndrome will greatly challenge global health services worldwide. The absolute global economic burden of these diseases is predicted to increase to \$2.1 trillion by 2030.^{8,9} Therefore, a global drive by all health stakeholders, including countries, global health institutions, health authorities, and private stakeholders, must be coordinated to reduce disease morbidity, mortality, and cost burdens. This can in part be achieved through implementing cost-effective interventions, such as education enabling basic exercise or nutrition interventions to prevent or reduce the collective disease burden.¹⁰ These can include increased exercise, encouragement of weight loss, or dietary management.¹ The latter could include supplementation with natural agents that can positively affect health, either through preventing risk factor occurrence, disease onset or pathological complications, via acting on several potentially intersecting metabolic pathways common to obesity, cardiometabolic diseases, and cancer. We propose below an argument for the use of vitamin D₃ for reduction of risk in obesity-associated NCD. How can we make this link? Epidemiological studies, which we review below, show that low circulating plasma levels of vitamin D₃ are present in obese individuals and are associated with poor morbidity and mortality outcomes from T2DM, CVD, and cancer. These have been shown to be ameliorated or reversed upon vitamin D₃ supplementation. Moreover, animal models indicate that vitamin D₃ is directly involved or implicated in adipogenesis and differentiation. Finally, mechanistic findings in vitro suggest that vitamin D₃ can simultaneously modulate several common intersecting hormone-regulated, obesity-induced, inflammation-potentiated signaling pathways common to T2DM, cardiometabolic diseases, and cancer.² As it is estimated that

Treatment of vitamin D deficiency on average costs less than 0.1% of the cost of investigations and treatment of worsening comorbidities and complications associated with hypovitaminosis D (ie, "vitamin D treatment cost of \$12/year versus, average cost for managing complications, of \$6,000 to 20,000/year per affected person").¹¹

Then, vitamin D_3 supplementation appears to be a potentially suitable cost-effective preventive and therapeutic agent that should be considered for all individuals, and especially in obese patients or those diagnosed with established metabolic syndrome risk factors to improve health outcomes and survival.

Vitamin D metabolism and genomic action in the human body

Vitamin D refers to a fat-soluble secosteroid which can be consumed in foods like supplemented milk, eggs and fish or synthesized in the body following exposure to UV light. Its effects were first identified by Mellanby and McCollum^{12,13} in cod liver oil and its biochemical metabolism pathways and physiological mechanisms of action for good health and well-being have been described in many tissue types and systems over the years.¹⁴⁻¹⁶ Vitamin D₃ is initially synthesized from 7-dehydrocholesterol upon UV exposure, then gets converted, respectively, to 25-hydroxyvitamin D [25(OH)D] in the liver, then the biologically active metabolite 1,25-dihydroxyvitamin D $[1,25(OH)_2D_3]$ in the kidneys. 25(OH)D is the circulating metabolite which gets measured routines as a marker of vitamin D sufficiency in the blood. Hydroxylation of vitamin D to 1,24,25(OH)₂D₃ by 25-hydroxyvitamin D 24hydroxylase (24-OHase), an enzyme found in liver, adipocytes and other cell types, results in the inactivation of vitamin D and its destination for excretion.¹⁷ The active metabolite of vitamin D, 1,25(OH)₂D₃ acts via binding to a member of the nuclear receptor superfamily, namely, the vitamin D receptor (VDR). Upon binding of 1,25(OH)₂D₃ ligand, VDR translocates to the nucleus, dimerizes with retinoid X receptor (RXR) (or other nuclear factors such as proliferator-activated peroxisome receptors-gamma (PPARy), retinoic acid receptor). The VDR/RXR dimer additionally recruits pioneer transcription factors, such as C/EBPa and SPI1 to activate transcription of several genes in several body organs and systems.^{14,18,19}

Vitamin D and obesity – epidemiological findings

Observational studies have often found an inverse relationship between obesity and serum 25(OH)D concentrations.^{20,21} The model that seems to best explain the observed correlations is volumetric dilution.²² A study of 686 adults taking <400 IU/d vitamin D found that the best fit to the data was a hyperbolic fit to weight, from 90 nmol/L (36 ng/mL) at 40 kg to 36 nmol/L (14 ng/mL) at 160 kg.²² It is assumed that 25(OH)D is stored in both adipose and non-adipose tissues. A review by Heaney found:

Mean serum 25(OH)D in all studies combined was 45 nmol/L. At the level of vitamin D repletion represented by this concentration, total body vitamin D would be 14,665 IU for a 70 kg adult woman. 65% of this total was present as native cholecalciferol and 35% as 25(OH) D. Nearly three-quarters of the cholecalciferol was in fat, while 25(OH)D was more evenly distributed throughout the body. (20% in muscle, 30% in serum, 35% in fat, and 15% in all other tissues)²³

The same group conducted a 10-week vitamin D supplementation study on 62 obese individuals with mean age ~45 years, weight ~107 \pm 20 kg, and BMI ~37 \pm 5 kg/m².²⁴ The changes in 25(OH)D after 21 weeks of supplementation were: for 1000 IU/d, 12 \pm 10 ng/mL; for 5000 IU/d, 28 \pm 10 ng/mL; and for 10,000 IU/d, 51 \pm 16 ng/mL. Note that the effect of vitamin D supplementation depended not only on baseline 25(OH)D, vitamin D₃ dose, weight, and BMI, but also genetic factors related to the absorption and conversion of vitamin D to 25(OH)D.²⁵

In the VITamin D and OmegA-3 (VITAL) trial, participants in the vitamin D treatment group were given 2000 IU/d vitamin D_3 .²⁶ Respective baseline and 1 year 25 (OH)D concentrations were 33.3 and 45.9 ng/mL for BMI<25 kg/m² (n=229), 29.5 and 41.4 ng/mL for 25 kg/m²<BMI<30 kg/m² (n=334), and 26.7 and 38.8 ng/mL for BMI>30 kg/m² (n=253). As a result, cancer incidence rates were significantly reduced only for those with BMI<25 kg/m².

Adipose tissues are active endocrine tissues that regulate various biochemical and physiological processes involved in body homeostasis, glucose and lipid metabolism, adipokine secretion, appetite control, vascular remodeling, and insulin action. In both humans and rodents, it has been shown that adipose tissues have a high concentration of 25(OH)D and its metabolizing enzymes.^{19,23,27} Obese individuals commonly store more vitamin D in adipose tissue than lean individuals.²⁸ Two rat model studies found support for effects of vitamin D in reducing the risk of metabolic syndrome, and by extension, risk of obesity. In one study, the combination of exercise and high-dose vitamin D reduced insulin, blood glucose, and homeostatic model assessment of insulin resistance (HOMA-IR).²⁹ In the other study, high-dose vitamin D supplementation significantly reduced weight, lipid profiles, visceral fat, and waist circumference of ovariectomized rats whether they were exercising or not.³⁰

It would be expected that during weight loss, stored vitamin D and its metabolites would be liberated and raise 25(OH)D concentrations. A meta-regression analysis of 25 (OH)D changes with respect to weight loss generally confirmed that expectation.³¹ However, smaller than expected increases in 25(OH)D were found, suggesting that conversion to other vitamin D metabolites also occurs.

It would be expected that various parameters associated with obesity would also be associated with 25 (OH)D concentrations. That is indeed the case. A crosssectional study of 20,360 US adults with lipid profiles and 25(OH)D concentrations from 2009 to 2011 found that vitamin D deficiency

was associated with significantly lower HDL-C (-5.1%) and higher total cholesterol (+9.4%), non-HDL-C (+15.4%), directly measured LDL-C (+13.5%), intermediate-density lipoprotein cholesterol (+23.7%), very low-density lipoprotein cholesterol (+19.0%), remnant lipoprotein cholesterol (+18.4%), and TG (+26.4%) when compared with the optimal group.³²

Vitamin D exerts a dose-dependent effect on adipocyte growth, differentiation, and survival

Adipocytes from white adipose tissue (WAT) are some of the most resilient cells in humans and other animals. This is partly an evolutionary survival mechanism that animals have adopted over millions of years to allow for prolonged storage given a limitation of available food sources in the wild. In humans, however, changing social and behavioral transitions have led to a life of abundance over the last century, and this protective adaptive mechanism and adipocyte resilience against destruction has become a curse with food overconsumption. In turn, storage of excessive fat has become a cause of increased metabolic disease risk. Therefore, any agents or mechanisms that can promote enhanced β-oxidation of stored fatty acids or triglycerides or promote adipocyte apoptosis without increasing inflammation can be considered as potential candidates for fat mass reduction, obesity management, and consequent reduction of metabolic risk. From a mechanistic perspective, the effects of $1,25(OH)_2D_3$ on adipose tissues are

conflicting and do not allow a clear conclusion to be drawn. The differences appear to change with several factors, including 1) absence or presence of VDR, 2) the concentration of $1,25(OH)_2D_3$ used in studies, 3) the species from which cells are derived, and (4) the stage of cell differentiation in which $1,25(OH)_2D_3$ is administered.^{33,34}

In vitro studies in mouse- and human-derived cells have shown opposite effects depending on the source of cells and their differentiation stages. Treatment with low concentrations of $1,25(OH)_2D_3$ was found to inhibit adipogenesis in mouse 3T3-L1 cells in a time-limited and dose-dependent manner.³⁵ This inhibition was ineffective 24–48 hrs after differentiation was initiated, suggesting that $1,25(OH)_2D_3$ inhibits only the early events of the adipogenic program. Transfection with hVDR in these cells blocked adipocyte differentiation and suppressed C/ EBP α and PPAR γ mRNA expression and protein activity. Similarly, in preadipocytes of brown adipose tissues (BAT), VDR expression and $1,25(OH)_2D_3$ treatment inhibited adipocyte differentiation and expression of C/EBP α as well as PPAR γ expression and activity.³⁶

Nobre et al confirmed that treatment of 3T3L1 preadipocytes, which are in the earlier stage of differentiation, with 1,25(OH)₂D₃ decreases C/EBP α , PPAR γ , and VDR protein levels, downregulates CYP7b1/1 α -hydroxylase and *VDR* mRNA expression, and decreases cell proliferation and adipocyte number. However, co-culture with leptin decreased the ability of 1,25(OH)₂D₃ to block adipogenesis in 3T3L1 preadipocytes co-cultured with both hormones.³⁷ Hyperleptinemia is known to be associated with increased adiposity and metabolic syndrome.

Findings in animal models support the notion that vitamin D is related to energy metabolism and specifically fat mass. The VDR knockout mouse (VDR-/mouse) presents a lean phenotype. Even upon high-fat feeding, these mice do not gain significant fat mass compared to wild-type animals. This VDR-/- phenotype is associated with increased levels of uncoupling proteins UCP-1, UCP-2, and UCP-3 in both BAT and WAT and an increased rate of β -oxidation in white adipose tissues. Both carnitine and carnitine palmitoyl transferase II responsible for transporting fatty acids into the mitochondrial matrix are increased in VDR-/- mice.³⁸ These data may suggest that VDR is an important factor for increasing adiposity and fat mass deposition. This increased adipose tissue formation sequesters plasma 25(OH)D and contributes to low circulating plasma levels and insufficiency.

Early research on $1,25(OH)_2D_3$ effects on apoptosis indicates that the hormone can exert a dose-dependent impact on adipocytes depending on their anatomical location and source by promoting glucocorticoid production and release.³⁹ In mature adipocytes, glucocorticoids may decrease lipid accumulation in differentiated adipocytes by promoting lipolysis and enhancing fatty acid mobilization, thus depleting the cell of its stored fats and sensitizing it to apoptosis. Additionally, treatment of mature mouse 3T3-L1 adipocytes induces Ca²⁺ mediated apoptosis via activation of calpain and caspase-12.⁴⁰

In a diet-induced mouse model of obesity, dietary supplementation with high levels of vitamin D_3 , high calcium or high calcium plus vitamin D_3 resulted in a significantly lower body weight and reduced fat mass gain in mice fed high-fat diets compared to controls. This was associated with Ca²⁺ dependent activation of apoptotic proteases in adipose tissues in mice given the supplementation.⁴¹ The above finding supports the suggestion that 1,25(OH)₂D₃ supplementation may potentially be an excellent agent to support weight loss and in turn reduce the risks associated with obesity.

Vitamin D, glucose metabolism, and insulin action – the case supporting a role for vitamin D in reducing risk of IR and T2DM

A prospective study conducted in southern California on 903 adults without T2DM or prediabetes at the time of enrollment in 1977–79 sought to determine the relationship between baseline 25(OH)D and incidence of T2DM.⁴² During the follow-up through 2009, there were 47 cases of T2DM and 337 cases of prediabetes. Those with baseline 25(OH)D >30 ng/mL compared with <30 ng/mL had a reduced risk of diabetes, with a HR of 0.31 (95% CI, 0.14–0.70) for those with 30 ng/mL <25(OH)D <39 ng/mL. There was a weak, non-significant inverse relationship between baseline 25(OH)D and prediabetes.

A high-dose vitamin D Randomized Controlled Trial (RCT) from Iran on vitamin D-deficient participants found a rate of progression from prediabetes to T2DM that was significantly lower in the intervention group (28% vs 3%; P=0.002).⁴³ A similar trial in the US with mean baseline 25(OH)D of 70 nmol/L did not find that supplementation with 4000 IU/d vitamin D₃ for 3.5 years significantly reduced progression from prediabetes to T2DM.⁴⁴ However, significant reductions in progression were

found for those with BMI <30 kg/m [HR=0.71 (95% CI, 0.53–0.95)] and those not taking calcium supplements [HR=0.81 (95% CI, 0.66–0.98)]. There was also a significant reduction in progression for those with baseline 25 (OH)D <30 nmol/L.

A cross-sectional analysis of 6868 adults without T2DM in the National Health and Nutrition Examination Survey examined the relationship between 25(OH)D and IR.⁴⁵ (IR was defined as the HOMA-IR; \geq 75th percentile, sex-specific: 3.9 in men or 3.6 in women). The analysis found that adding 25(OH)D to the model based on established risk factors could identify an additional 2–4 cases per 1000 people. The general 25(OH)D threshold was 60 nmol/L; however, it varied by ethnicity (Mexican-Americans: 54 nmol/L, non-Hispanic whites: 68 nmol/L, and non-Hispanic blacks: 41 nmol/L). This finding suggests that 25(OH)D is not an important determinant of IR.

A number of RCTs have been conducted to determine whether vitamin D supplementation could reduce IR. One such study was conducted in Melbourne, Australia on 54 overweight or obese adults with a mean age of 32 years who were vitamin D deficient (25(OH)D < 50 nmol/L).⁴⁶ Participants were given a bolus dose of 100,000 IU vitamin D₃ or placebo at outset, followed by 4000 IU/d or placebo thereafter for 16 weeks. In the treatment arm, mean 25(OH)D rose from 31 to 88 nmol/L. Differences between the treatment and control arms were not significant for insulin sensitivity or first-phase insulin secretion.

However, an RCT conducted in Iran with 162 participants aged 18-80 years with baseline 25(OH)D =12.5 ng/ mL (31 nmol/L) conducted over a 6-month period did find a reduction in the vitamin D treatment arm.⁴³ Those in the vitamin D treatment arm were given 50,000 IU vitamin D₃ pearls once weekly for 3 months, followed by one pearl per month for the next 3 months. 25(OH)D rose from 13 to 41 ng/mL in the treatment arm and from 13 to 16 ng/mL in the control arm. The mean HOMA-IR score dropped from 3.4 to 2.9 and 2.6 after 3 and 6 months, respectively, in the treatment arm and from 3.4 to 3.1 after 3 and 6 months in the control arm, with P=0.04 for the difference between the two arms. Glucose tolerance was not significantly different between the two arms after 3 months (P=0.68) but did reach significance after 6 months (P=0.002). The primary difference between the two studies was duration, but the achieved 25(OH)D was also higher in the Iranian study. These two examples illustrate the problems inherent in conducting vitamin D RCTs.

In vitro, treatment with 1,25(OH)₂ D₃ improves 2-deoxyglucose uptake in 3T3-L1 adipocytes, implying a positive effect on glucose metabolism. Incubation of cells with 1,25 (OH)₂D₃ for 24 hrs results in a fourfold increase in insulinstimulated uptake of 2-deoxyglucose in 3T3-L1 adipocytes compared to controls. Moreover, cells pre-treated with 1,25 $(OH)_2 D_3$ and then challenged with pro-inflammatory TNF- α had a 40% higher uptake of 2-deoxyglucose compared to those suppressed in their response to insulin stimulation following TNF- α treatment alone. Treatment with 1.25 $(OH)_2D_3$ counteracted the effect of TNF- α and restored glucose uptake response close to normal function (90%). The effect of 1,25(OH)₂D₃ was mediated by AKT phosphorylation.⁴⁷ These results have a huge implication for the recommended use of vitamin D₃ in diabetic, insulinresistant and obese patients to improve sensitization to insulin and promote glucose uptake as part of therapeutic protocols.

In an insulin-resistant cell model (IR BeWo model) exhibiting high inflammatory gene expression and elevated mTOR signaling, treatment with $1,25(OH)_2VitaminD_3$ reversed mTOR activation and reduced IR levels and lipid infiltration while decreasing leptin, TNF- α , and IL-6 levels.⁴⁸

In another diet-induced obesity model, mice fed high levels of vitamin D_3 , high calcium or high calcium plus vitamin D₃ all had lower plasma concentrations of glucose and insulin along with increased levels of adiponectin, the hormone that sensitizes adipocytes to insulin.⁴¹ Upon vitamin D_3 supplementation, with or without Ca,²⁺ the decrease in plasma insulin concentrations was almost completely normalized to concentrations measured in controls. Collectively, if animal model findings can be extrapolated to humans, these data may indicate that in vivo dietary supplementation of vitamin D₃ may attenuate diet-induced hyperglycemia, increase insulin sensitivity and action, and prevent postprandial hyperinsulinemia if administered to obese individuals. Hyperinsulinemia, present in insulin-resistant, diabetic and obese individuals can pose a secondary health risk by potentiating the incidence of disorders such as polycystic ovary syndrome and cancer, as well as increasing mortality.⁴⁹

Vitamin D's effect on obesityinduced inflammation

Obesity is associated with increased inflammation.⁵⁰ Adipocytes can secrete IL-6, which is a major inflammatory mediator in obesity. IL-6 also downregulates adiponectin, an anti-inflammatory adipokine. Decreased adiponectin is associated with increased pro-inflammatory effects associated with NF- κ B activation.⁵¹ Increased IL-6 concentrations and inflammation are associated with IR and a risk of T2DM and CVD.

In vitro studies show that treatment of lipopolysaccharide (LPS)-stimulated mouse 3T3-L1 adipocytes with 1,25 (OH)₂D₃ for 24 hrs significantly decreased IL-6 production but had no effect on IL-10 production. Similarly, a reduction in IL-6 production in epididymal WAT was observed in mice fed high-fat diets supplemented with vitamin D₃ compared to controls. The in vivo findings in mice are especially promising, as they indicate that vitamin D₃ can be considered as an adjuvant to reduce inflammation in obese subjects.⁵² Similarly, in human adipocytes from two different sources - bone marrow-derived human mesenchymal stromal cells (hMSCs) differentiated into adipocytes and adipocytes isolated from biopsies stimulated with LPS – 24-hr treatment with $1,25(OH)_2$ D₃ resulted in a significant reduction in IL-6 secretion. This was associated with a reduction in nuclear factor-kB (NF-кВ) nuclear translocation and activity.⁵³

Treatment with $1,25(OH)_2$ D₃ can reduce pro-inflammatory effects induced by TNF- α in 3T3-L1 adipocytes and adipocyte-macrophage co-cultures. This attenuation of the inflammatory response is mediated by dephosphorylation of mitogen activated protein kinase p38, downregulation of NF- κ B, and induction of dual-specificity phosphatases (DUSP) DUSP 1, 10, 11 and 16.⁴⁷

Human adipocytes pre-sensitized with IL-1 β and treated with 1,25(OH)₂D₃ showed a significant reduction in mRNA levels of pro-inflammatory interleukins IL-6, IL-8 and monocyte chemoattractant protein-1 (MCP-1), along with reduced IL-8 secretion. However, a similar reduction was not observed in adipose tissue biopsies removed from obese subjects receiving an oral treatment of vitamin D₃ (7000 IU).⁵⁴

Observational studies do not lend compelling evidence to support an effect of vitamin D on inflammatory markers in metabolic syndrome patients. There is only weak evidence from observational studies to show that vitamin D reduces biomarkers of inflammation. For example, an RCT involving overweight/obese adults given high-dose vitamin D for 16 weeks did not find any significant reductions in inflammatory biomarkers.⁴⁶ However, RCTs involving participants with various diseases have found beneficial effects, such as for T2DM^{55,56} and polycystic ovary syndrome.⁵⁷

A meta-analysis investigated the effects of vitamin D supplementation on biomarkers of inflammation and oxidative stress in patients with different forms of DM.55 A total of 33 studies conducted between 2012 and 2017 involving 1053 randomized patients were included in the analyses. Twenty of the studies were conducted in Iran, which tended to have patients with low baseline 25(OH)D. Trial durations were from 6 weeks to 12 months. Vitamin D₃ doses ranged from 400 IU/d to 50,000 IU every 2 weeks, and some trials included calcium supplements. The effect of vitamin D supplementation on serum high-sensitivity C-reactive protein(hs-CRP) for T2DM was -0.73 (95% CI, -1.07 to -0.39). Most of the other included studies also found significant reductions in hs-CRP, except for studies on pregnant women with gestational DM in People's Republic of China. Other analyses found significant increases in nitric oxide, antioxidant concentrations, and reduction in malondialdehyde concentration, all in a direction that improves biochemical status (Table 1). However, there was an indication of publication bias for hs-CRP and malondialdehyde concentration.

Vitamin D regulates circulating lipids, blood pressure and improves CVD outcomes

The endothelium plays an important role in the maintenance of blood fluidity, vessel wall tone and permeability. Endothelial damage via oxidative stress and metabolic

Table I Findings reported in a meta-analysis of the effects of vitamin D supplementation on biomarkers of inflammation and oxidative stress in patients with T2DM⁵⁵

Outcome	N	Condition	GDM included?	WMD (95% CI)	Þ
Hs-CRP	14	T2DM	No	-0.73 (-1.07, -0.39)	0.000
Hs-CRP	28	All	Yes	-0.27 (-0.35, -0.20)	<0.001
Nitric oxide concentration	7	T2DM	No	4.3 (1.0, 7.7)	
Total serum antioxidant capacity	11	All	Yes	57 (33, 81)	0.28
Glutathione concentrations	12	All	Yes	83 (44, 121)	0.43
Malondialdehyde concentration	11	T2DM	No	-0.43 (-0.62, -0.25)	0.04

Notes: p-value from Egger's test (if <0.05, indication of publication bias).

Abbreviations: N, number of studies; NA, not applicable; T2DM, type 2 diabetes mellitus; GDM, gestational diabetes mellitus; WMD, weighted mean difference; CI, confidence interval.

disorders contributes to vascular endothelial dysfunction, impaired vascular relaxation, vascular hypertrophy fibrosis, plaque deposition, and arterial stiffness, all of which contribute to hypertension, atherosclerosis and CVDs.58,59 Obesity-associated hyperleptinemia has been shown to cause endothelial cell dysfunction and increase inflammation and oxidative stress in vitro and in vivo.^{60,61} Obese individuals have been reported to have significantly higher plasma leptin levels compared to eutrophic individuals.⁶² Most recently, plasma leptin level has been reported to be positively associated with increased blood pressure independent of gender and BMI.⁶³ Recent findings demonstrate that 1,25 (OH)₂D₃ can counteract leptin-induced oxidative stress and inflammation in human umbilical vein endothelial cells. Treatment of these cells with leptin alone significantly increased reactive oxygen species production, NF-KB translocation to the nucleus, and upregulated the mRNA expression of MCP1, TGF β , and vascular adhesion molecule 1. Pretreatment with 1,25(OH)₂D₃ at physiological concentrations (10^{-10} M) and supraphysiological concentrations (10^{-7} m) M) prevented leptin-induced superoxide anion production, significantly increased nuclear factor (erythroid-derived 2)like 2 (NRF2) translocation to the nucleus, and upregulated expression of the antioxidant defense genes superoxide dismutase 2, glutathione peroxidase, NADPH hydrogenase (quinone) 1 (NQO1), and heme oxygenase 1.64 These findings reinforce the idea that 1,25(OH)₂D₃ may have potent protective effects against oxidative stress-induced damage and hence prevent metabolic complications of obesity such as hypertension or other inflammation-induced complications mediated by increased NFkB activity.

VDR-null mice display less body fat mass and lower plasma triglyceride and cholesterol levels compared with wild-type male mice.³⁸ VDR-/- mice also have higher blood pressure than wild-type mice. These effects are associated with increases in renin (RER) mRNA expression and plasma angiotensin II. These mice remain responsive to challenges in salt variation, though not as effectively as wild-type controls, indicating that VDR plays a role in the renin-angiotensin system and ultimately in blood pressure control. At the gene level, the renin gene RER contains a transcription response element that can be repressed by VDR binding.^{65,66} Mice with a deletion in VDR also exhibit a reduction in end-diastolic and end-systolic volume, impaired ventricular function and cardiac hypertrophy due to increased calcineurin inhibitory protein 1 expression (MCIP1) and impairments in the calcineurin/nuclear factor of activated T cell (NFATc1) signaling pathways.⁶⁷

Additionally, studies by Cheng et al indicate that 1,25 $(OH)_2D_3$ ameliorates hepatic abnormalities in T2DM. In a mouse model of T2DM, 8-week supplementation of type 2 diabetes db/db mice with 1,25(OH)₂D₃ ameliorates hepatic triglyceride accumulation and glucose production via the activation of Ca2+ mediated CaMKKB/AMPK signaling pathway.⁶⁸ Imga et al have shown that vitamin D administration in overweight or obese vitamin D-deficient premenopausal women, can result in a significant reduction in LDL-C levels and improvement of IR.⁶⁹ Effects on vitamin D on postprandial lipid profiles in obese women may be dose-dependent and favors a reduction in LDL and vLDL at lower vitamin D levels, and an elevation of lipids at higher vitamin D doses.⁷⁰ Perhaps the latter may be due to the effects of vitamin D on increased lipolysis in adipocytes and β-oxidation associated with concentrated vitamin action on WAT. However, further studies need to be conducted in this respect and should include male subjects to confirm the mechanisms explaining the dose-dependent effects.

As for observational studies, there is limited evidence that vitamin D reduces blood pressure and hypertension. In a 6-month vitamin D supplementation study involving middle-aged African Americans, each 1 ng/mL increase in 25(OH)D resulted in a significant 0.2 mm Hg drop in systolic blood pressure (SBP) but no effect on diastolic blood pressure (DBP).⁷¹ In a community-based open-label study in Canada where participants were provided vitamin D₃ supplements and encouraged to achieve >40 ng/mL 25(OH)D, those who were hypertensive at baseline (SBP ~155 mm Hg, DBP ~95 mm Hg) reduced their SBP by ~14–18 mm Hg and DBP by ~12 mm Hg.⁷² However, participants who were prehypertensive.

However, in CVD outcomes, there is reasonable evidence that vitamin D reduces the risk of CVD. A meta-analysis of 34 prospective observational studies involving 180,667 participants found that the pooled risk ratio for the incidence of a 10-ng/mL increase was 0.90 (95% CI, 0.86–0.94), while the risk ratio for mortality was 0.88 (95% CI, 0.80–0.96).⁷³

A meta-analysis of the effects of vitamin D supplementation on CVD risk factors based on 81 studies found significant beneficial effects for blood pressure, lipid profile, parathyroid hormone, and serum high-sensitivity Creactive protein (hs-CRP) but not for peak wave velocity or augmentation index.⁷⁴

Additional evidence that vitamin D deficiency plays an important role in the risk of CVD is that CVD rates are 15–25% higher in winter than in summer,⁷⁵ even in warm countries such as Kuwait.⁷⁶

In general, a majority of the evidence points to a beneficial effect of vitamin D in reducing the risk of CVD, although RCTs are not available to support these associations.^{77,78} For example, the VITAL study found a HR for major CVD from 2000 IU/d vitamin D₃ vs placebo over 5 years =0.97 (95% CI, 0.85-1.12).²⁶ However, VITAL was based on vitamin D dose, while proper RCTs should be based on 25(OH)D.⁷⁹ A recent meta-analysis of observational studies of CVD events vs 25 (OH)D found few effects above 15 ng/mL.⁷³ The mean baseline 25(OH)D in VITAL was 27.8 ng/mL among 395 males and 31.7 ng/mL in 441 females.²⁶ Thus, a significant beneficial effect would not be expected.

Vitamin D reduces risk of and improves survival from cancer

There is very strong evidence that vitamin D reduces the risk of incidence and death for many types of cancer. Geographical ecological studies from mid-latitude countries generally find inverse correlations between indices of solar UVB doses and mortality rates for 15–20 types of cancer.¹⁴ Prospective observational studies have found significant inverse correlations between serum 25(OH)D at the time of enrollment and incidence of colorectal cancer.⁸⁰ Case-control studies have identified a significant inverse correlation between 25(OH)D measured near time of diagnosis, but this correlation was not observed in prospective studies due to the fact that breast cancer can develop rapidly.⁸¹ Vitamin D is more effective at reducing cancer mortality rates than incidence rates. For example, a meta-analysis of the effects of vitamin D supplementation on risk of cancer found no significant effect on cancer incidence but a significant 15%

reduction in cancer mortality rates.⁸² The mechanisms whereby vitamin D reduces the risk of cancer are well known. The most important of these affect cellular differentiation, proliferation and apoptosis, angiogenesis, and metastasis.^{14,83,84} Recently, results of the VITAL trial were released.²⁶ This RCT included over 25,000 participants, including more than 5000 African Americans, who were followed for 5 years. The treatment arm was given 2000 IU/d vitamin D₃, and a placebo was given to the control arm. The results revealed three important findings. First, participants with BMI <25 kg/m² had a HR for cancer incidence =0.76 (95% CI, 0.61-0.90). Second, death from cancer was significantly reduced if results from the first 1 or 2 years were omitted: HR=0.79 (95% CI, 0.63-0.99) for 1 year and 0.75 (95% CI, 0.59-0.96) for 2 years. It is reasonable to perform this type of analysis, as 25(OH)D can affect cancer risk for some time. The third finding was that cancer incidence for black Americans was marginally non-significantly reduced [HR=0.77 (95% CI, 0.59-1.01)]. Other findings regarding vitamin D were not significant. Significant effects on incidence were found among the groups that had lower baseline and/or achieved higher 25(OH)D levels. The main problem with VITAL was that the outcomes were based on vitamin D dose, whereas 25(OH)D should have been used.⁷⁹

Conclusion and recommendations for improved metabolic health and wellbeing

We propose the use of vitamin D_3 for all individuals to protect against cardiometabolic and metastatic disease (Table 2). In vitro studies, animal models, observational studies' findings, and limited number of existing RCTs

Table 2 Beneficial actions of vitamin D_3 bioactive metabolites of	n various risk factors	s influencing cardiometabo	lic health outcomes
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Factor or marker	Beneficial actions of vitamin D ₃ bioactive metabolites
Circulating glucose levels	Improves glucose uptake from the blood stream
Insulin sensitivity	Improves insulin sensitivity hence reduces insulin resistance
Leptin	Reduces circulating obesity-induced hyperleptinemia hence decreases its negative effects associated with
	inflammation, insulin resistance, hypertension, and oxidative stress
Hypertension	Regulates blood pressure and critical for the renin-angiotensin system
BMI and weight loss	Mild reduction effects on body mass observed primarily in women; more significant effects observed with concurrent calcium supplementation.
Adipocyte function, growth and storage	Depends on state of differentiation of pre-adipocyte or mature adipocyte; enhances β -oxidation of fatty acids and triglycerides; prevents against high-fat diet-induced adipogenesis
Dyslipidemia	Improved lipid profiles especially downregulation of plasma triglycerides and reduction in triglyceride accumulation in the liver.
Inflammation	Significant anti-inflammatory effect resulting in the reduction of CRP, interleukins and pro-inflammatory cytokines which would otherwise increase noncommunicable disease risk and comorbidities such as cancer.

support the proposal that vitamin D₃ will have therapeutic power/benefits to reduce the burden of disease in obese patients by counteracting the effects of many metabolic risk factors. This will greatly improve patient health outcomes, quality of life, and survival. This is expected to reduce health costs of treating NCDs, related comorbidities, and associated indirect loss of productivity costs. Overall, the cost of a maintenance dose of 1000-5000 IU vitamin D₃ daily needed to achieve plasma circulating levels in the range of 30-50 ng/mL is cost-effective compared to costs of treatment of NCDs. In addition, there are many additional health benefits of vitamin D supplementation and very limited risk of adverse effects. Given that increased adiposity has the effect of depleting much of the 25(OH)D precursor out of the plasma,⁸⁵ the higher daily dose of vitamin D₃ is advised in obese individuals. More RCTs need to be conducted in obese patients to determine the correct daily dose-titration needed in patients with increasing number of metabolic risk factors or increasing disease severity.

Disclosure

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1447