

Hemolytic uremic syndrome: pathogenesis and update of interventions

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Marina S Palermo[†],
Ramón A Exeni and
Gabriela C Fernández

[†]Author for correspondence
Lab Inmunología, Instituto de
Investigaciones Hematológicas,
Academia Nacional de Medicina
P. de Melo 3081 (C1425AUM),
Ciudad de Buenos Aires,
Argentina
Tel.: +54 114 805 5695
Fax: +54 114 803 9475
mspalamero@hematologia.anm.edu.ar

The typical form of hemolytic uremic syndrome (HUS) is the major complication of Shiga toxin-producing *Escherichia coli* infections. HUS is a critical health problem in Argentina since it is the main cause of acute renal failure in children and the second cause of chronic renal failure, accounting for 20% of renal transplants in children and adolescents in Argentina. Despite extensive research in the field, the mainstay of treatment for patients with HUS is supportive therapy, and there are no specific therapies preventing or ameliorating the disease course. In this review, we present the current knowledge about pathogenic mechanisms and discuss traditional and innovative therapeutic approaches, with special focus in Argentinean contribution. The hope that a better understanding of transmission dynamics and pathogenesis of this disease will produce better therapies to prevent the acute mortality and the long-term morbidity of HUS is the driving force for intensified research.

KEYWORDS: *Escherichia coli* O157:H7 • hemolytic uremic syndrome • inflammation • Shiga toxin • treatment

General concepts

The typical form of hemolytic uremic syndrome (HUS) is caused by particular serotypes of enterohemorrhagic *Escherichia coli* (EHEC) that produce Shiga toxin (Stx), and are known as STEC (Stx-producing *E. coli*). Full development of HUS is characterized by thrombocytopenia, hemolytic anemia and acute renal failure, which appear a few days after the infection. Wagner may have been the first to report a case of HUS [1], and Gasser *et al.* described the features of HUS in 1955 [2]. In Argentina, the group of Gianantonio in 1964 published the first complete description of the clinical features and evolution in more than 50 cases of HUS [3] and most importantly, they proposed peritoneal dialysis as treatment in the acute period [4]. Later, in 1983, Karmali *et al.* described the association between *E. coli*, Stx and HUS [5].

Hemolytic uremic syndrome is the main cause of acute renal failure in children in Argentina, and a high percentage of children with HUS (20–30%) have long-term permanent renal sequelae, making this disease the second cause of chronic renal failure and accounting for 20% of renal transplants in children and adolescents in Argentina [6,7]. HUS is not exclusively restricted

to the kidney. Many patients during the acute period experience neurological symptoms (20%, including lethargy, seizures, headache, convulsions and coma), hypertension (30–50%), pancreatic insufficiency (8–10%) and/or gastrointestinal complications (severe colitis and rectal prolapse) [6,8]. In this sense, severe cases of HUS, and usually death, are associated with central nervous system involvement. Other extra renal long-term complications can also be observed, mainly hypertension, but also neurological sequelae (moderate and severe disability) and insulin-dependent diabetes mellitus have been reported [7,9,10].

Infection begins with ingestion of contaminated food or water. Acid-resistance mechanisms of STEC facilitate their survival through the low pH of the stomach [11]. The bacteria pass through the small intestine, and virulence genes are turned on by environmental signals in the colon [12]. Adherence to enterocytes of the colon is produced by a characteristic mechanism called attaching and effacing (A/E) lesions, in which bacteria intimately attach to the intestinal epithelial cells and rearrange the cytoskeletal actin underneath, causing diarrhea and intestinal inflammation [13]. Local damage in colonic blood vessels induced by Stx results

in bloody diarrhea. Recent evidence indicates that macropinocytosis might be responsible for toxin uptake and transcytosis by globotriosylceramide (Gb₃)-free intestinal epithelial cells [14]. If sufficient Stx is absorbed into the circulation, vascular endothelial sites rich in the toxin receptor are damaged, leading to impaired function.

Pathophysiology

Gianantonio described in 1973: 'The pathologic renal findings in the acute stage of HUS show that most of the lesions are expressions of intravascular clotting' [15]. Renal histology reveals thickening of the capillary wall and swelling of endothelial cells within the glomerulus, with reduction of glomerular capillaries [16]. Nowadays, it is well known that the microvascular endothelial damage is a central pathogenic process underlying the development of HUS. However, human glomerular endothelial cells *in vitro* are not very susceptible to Stx toxic effects unless pretreated with inflammatory mediators [17]. Histology also shows that renal tubular cells can be necrotic. It has been demonstrated that Stx directly affects renal tubular cells [18] and triggers the activation of the coagulation and inflammatory systems [19,20], which, in turn, sensitize endothelial glomerular cells to Stx toxic effects [17]. Regardless of which cell type is the first target of Stx, the extent of glomerular injury will be determinant in long-term outcome. In fact, affected glomeruli will become fibrotic reducing the number of functional nephrons.

Shiga toxin

Stx is the main virulence factor in STEC infections. The *stx* gene is encoded in lysogenic bacteriophages contained by the bacteria. The production of Stx is linked to the replication cycle of the Stx phage, and the bacteria must be lysed in order to release the toxin [21]. Two major subfamilies of Stx exist: Stx1 and Stx2. In addition, each subfamily consists of different variants. Members included in the Stx1 subfamily are Stx1, Stx1c [22] and Stx1d [23], whereas Stx2, Stx2c [24], Stx2d [24], Stx2d_{activable} [25], Stx2e [26] and Stx2f [27] belong to the Stx2 group. Although all these variants have been isolated from patients, not all of them are associated with severe disease. In this sense, it has been reported that bacteria producing only Stx2 are more pathogenic than those producing Stx1 alone and those producing Stx1 with Stx2. Those producing Stx2, Stx2c and Stx2d_{activable} have been associated with more severe disease, such as hemorrhagic colitis and HUS, whereas the other members of Stx1 or Stx2 subfamilies were associated with uncomplicated diarrhea and asymptomatic infections [28]. All Stx members possess one catalytically active A subunit and five B subunits that mediate the binding to Stx receptor, Gb₃. Stx induces different receptor-mediated biological effects depending on the cell type, which is related to the organization of Gb₃ in the membrane and directs the fate of the toxin inside the cell. Stx cytotoxicity is dependent on Gb₃ presentation within detergent-resistant membrane (DRM) domains or 'lipid rafts' [29,30]. Internalization of Stx through Gb₃ within plasma membrane DRM results in a retrograde transport ending in the cytosol, and the interaction of the A subunit with the 28S

rRNA leads to inhibition of protein synthesis and, eventually, cell death [31]. By contrast, non-DRM plasma membrane Gb₃ mediates binding and subsequent traffic to lysosomes for Stx degradation [32,33]. In addition, the dose is also a determinant in Stx-mediated effects. In fact, sublethal amounts of Stx have only minor effects on overall protein synthesis but have dramatic effects on gene regulation, causing increased expression of hallmark proadhesive, prothrombotic and inflammatory genes in, for example, endothelial cells [34–37].

Additional factors

Hemolytic uremic syndrome is multifactorial in etiology involving complex interactions between bacterial and host factors. Although in the USA and Europe, 95% of HUS cases are associated with a STEC infection [38], in Argentina this association is confirmed only in the 60% of HUS cases, by at least one of the diagnosis criteria used [39]. This discrepant result may be attributed to deficiencies in local systems of diarrhea surveillance. On the other hand, not all STEC infections result in HUS. In fact, it may result in no disease, watery diarrhea, bloody diarrhea or HUS. Only 5–10% of infected children develop HUS [40]. The initial bacterial inoculum, the amount and type of Stx produced by the bacteria ingested, additional bacterial virulence factors, and the specific characteristic of activation of the thrombotic and inflammatory responses of the host are some of the factors that may determine the outcome of a STEC infection.

Additional factors from the bacteria

Besides Stx, putative virulence factors of STEC including adhesins, other toxins and proteases [39,41,42], are required to develop disease. The precise role of some of these factors in STEC disease remains to be fully established. It is possible that the degree of adhesion is correlated with the ability to cause disease. Whereas adhesins are probably implicated in the first contact to cells, intimate adhesion is mediated by the proteins encoded in the locus of enterocyte effacement (LEE), which participate in the formation of the A/E lesion. These include a type III secretion system, the *E. coli*-secreted proteins (Esp) and the surface protein intimin [43]. It is important to point out that several LEE-positive STEC strains are not associated with HUS, and that the type III secretion system also secretes many other effector molecules encoded outside the LEE, which are referred to as non-LEE-encoded effectors [44]. At least three of these non-LEE-encoded effectors have been linked to non-O157 STEC that cause HUS [45].

The intimin gene *eae* has been identified as a risk factor for HUS, as the vast majority of strains implicated in HUS are intimin positive [46]. However, recently it has been shown that highly pathogenic strains producing Stx2d_{activable} are intimin negative [47].

Among non-Stx toxins, the plasmid-encoded enterohemolysin (Ehx) is a pore-forming cytotoxin produced by both *eae*-positive and *eae*-negative STEC, but the frequency is higher among *eae*-positive STEC [48]. Ehx could contribute to disease by its membrane damaging effect on a wide variety of cell types, including

erythrocytes, its ability to induce production of proinflammatory cytokines, or both [49]. Although Ehx has been found in many STEC serotypes that are commonly associated with diarrheal disease and HUS [50–52], it is not clear how this toxin could enter the circulation to cause systemic damage.

Subtilase cytotoxin is a toxin with an A1:B5 structure, detected in a high percentage of STEC strains, mainly in LEE-negative strains [53]. In contrast to Stx, it was recently described that glycosphingolipids are not pivotal receptors for this toxin *in vivo* [54]. It has been shown to transiently inhibit protein synthesis and cause apoptosis of the Stx-sensitive Vero cell line [55,56]. In addition, intraperitoneal injection of subtilase cytotoxin to mice causes death, microangiopathic hemolytic anemia, thrombocytopenia and renal impairment, characteristic features of Stx-induced HUS [53,57].

Bielaszewska *et al.* described another novel toxin, the cytolethal distending toxin (CDT-V), present in 87 and 6% of O157:H⁻ and O157:H7 strains, respectively [58]. CDT-V has also been found in non-O157 STEC strains, all of which were *eae* negative [59]. This toxin directly injures endothelial cells leading to their death, and may thus contribute to the pathogenesis of HUS [59].

Several proteases (EspA, EspP, EspJ, EspI and StcE) are produced by STEC, and their activities suggest a putative role in the pathogenesis of the disease [43,60,61].

Lipopolysaccharide (LPS) is a major product of the Gram-negative bacteria that synergizes Stx-induced toxicity. Although Louise *et al.* suggested that Stx may enhance the procoagulant effect of LPS [62], *in vivo* studies have demonstrated that toxicity of Stx is potentiated by LPS [63–65]. In particular, LPS, chemokines and cytokines released by inflammatory cells or injured cells upregulate the expression of Stx receptors and sensitize microvascular endothelial cells (glomerular and others) to Stx-induced injury [17,66–68].

In summary, although Stx-induced endothelial injury is the primary pathogenic event, multiple bacterial components may contribute to the whole pathogenic mechanism.

Factors from the host

Several authors have reported the importance of the inflammatory and thrombotic responses in the development of HUS. Initially, STEC colonization induces acute colonic inflammation. In this regard, the infiltration of the gut and the presence of leukocytes in feces are seen in many STEC-infected patients. Several pathogenic factors of STEC have been demonstrated to induce the expression of proinflammatory chemokines in epithelial cells, which was accompanied by an influx of polymorphonuclear leukocytes (PMNs) [18,69]. Transmigration of PMNs from the basolateral to apical area of an intestinal epithelial cell line significantly increased the movement of Stx1 and Stx2 in the opposite direction [70]. In addition, PMN recruitment in the intestine may also increase the risk of HUS by inducing the Stx2 prophage *in vivo* and augmenting Stx2 production, mainly through the production of H₂O₂ [71].

Patients also evidence a marked inflammatory response as demonstrated by systemically (blood) and locally (urine) increased levels of various inflammatory mediators, including interleukins,

chemokines, soluble adhesion molecules, growth factors and acute-phase response proteins. In addition, they also show markers of endothelial injury, activation of the coagulation cascade and inhibition of fibrinolysis [72]. It has been suggested that the degree of the prothrombotic activation early in infection could be decisive in the course of the disease [73].

The activation of PMNs is evidenced by a high peripheral blood PMN count at presentation, which has been correlated with a poor prognosis, and increased levels of serum elastase and IL-8. In addition, PMNs from HUS patients show increased adhesive capacity *in vitro* and reduction in their granule content as demonstrated by ultrastructural, phenotypic and functional studies [74–76]. In this sense, the severity of renal impairment has been correlated with the degree of PMN degranulation [77]. Platelet activation has been suggested by several evidences. Thrombocytopenia is a main feature of HUS, and is caused by consumption of platelets, probably after activation and aggregation. Platelets from HUS patients showed impaired aggregating responses and reduced β -thromboglobulin (β -TG) content [78]. Platelet-derived products such as β -TG, platelet factor 4 and soluble P-selectin were found to be elevated during acute HUS [79,80]. Furthermore, changes in platelet ultramorphology and increased platelet-derived microvesicles were found in these patients, indicating platelet activation [81]. The activation of both PMNs and platelets will potentiate the inflammatory process and may enhance the primary Stx-induced endothelial damage.

Activated monocytes may also contribute to Stx toxic effects by the secretion of several chemokines and cytokines (TNF- α , IL-1 β , IL-8, RANTES and tissue factor), which increase endothelial susceptibility to Stx. An increased percentage of CD16⁺ monocytes have been reported in peripheral monocytes from HUS patients [82]. These CD16⁺ monocytes are considered to represent an activated and more mature subset with characteristics that resemble macrophages and dendritic cells [83,84]. Fractalkine (FKN; CX₃CL1) is a transmembrane chemokine present on endothelial and epithelial cells, whereas the FKN receptor (CX₃CR1) is expressed on monocytes, among other cells. Under conditions of physiologic flow, FKN mediates adhesion of monocytes, and is upregulated upon inflammation. A disappearance of CX₃CR1⁺ monocytes has been reported in HUS patients, and this correlated with the severity of renal failure [85]. The fact that CX₃CR1⁺ leukocytes were observed in renal biopsies from patients with HUS suggest that the interaction of CX₃CR1⁺ cells with FKN present on activated endothelial cells may contribute to renal injury in HUS.

Epidemiology

Globally, 70–80% of HUS cases have been attributed to epidemic outbreaks with O157:H7/H⁻ strains. However, other serotypes are also capable of causing human disease [86]. In Argentina, where HUS is endemic, the dominant STEC serotype isolated from HUS patients is O157:H7, followed by O145:NM, O26:H11, O103:H21, O174:H21, O8:H19 and O145:H25 serotypes, in descending order [87]. It is important to point out that non-O157 STEC is much harder to detect microbiologically, and for this reason the proportion of HUS cases caused by O157 STEC may be an overestimate.

The incidence of HUS varies according to the country. Argentina shows the highest incidence worldwide with 12–14 cases per 100,000 children under 5 years of age per year, and more than 400 new cases per year [87]. This rate is ten-times higher than in other industrialized countries [88].

In Argentina, as in the rest of the world, healthy cattle are the principal and natural reservoir of *E. coli* O157:H7 [89]. STEC strains have been recovered from fecal samples of 39% of healthy animals in a recent Argentine survey [90]. STEC is transmitted to humans through contaminated food and water [91–93]. Other routes of transmission are associated to prolonged fecal shedding of STEC among children attending daycare centers [94] and direct contact with infected individuals [95] or animals [96].

Examples of reported sources include undercooked ground beef, private or municipal water sources and other food products, such as unpasteurized apple cider or milk, fresh vegetables, sprouts and salami [97]. Visits to petting zoos, dairy farms, camping grounds where cattle have previously grazed, and recreational water sources have all been shown as risk factors for infection. In Argentina, person-to-person transmission has been proposed as an important route of infection supporting endemic behavior and family outbreaks [98]. A very low infectious dose (100–500 organisms) of this microorganism is required to become infected, accounting for the ability to cause severe and epidemic disease [44].

Treatment

Although there is extensive research in the field, the mainstay of treatment for patients with HUS is supportive therapy that generally includes control of fluid and electrolyte imbalance, use of dialysis if required, control of hypertension and blood transfusion as required.

Treatments during pre-HUS interval

Since vascular occlusion could partly underlie renal insufficiency during HUS, Ake *et al.* recommended parenteral volume expansion of children with bloody diarrhea associated to *E. coli* O157:H7 infection and before HUS develops [99]. Volume expansion might attenuate renal failure and also mitigate the nephrotoxicities of filtered urate and hemoglobin. They have encouraged hospitalization and intravenous administration of isotonic fluids, based on the fact that salt loading protects against nonthrombotic nephrotoxicity and hyponatremia, a potential complication of hypotonic maintenance fluids. However, more randomized studies evaluating this point would be necessary to generalize this early treatment, and most important, children who receive intravenous volume expansion need hospitalization and careful monitoring. Moreover, this recommendation makes essential a very rapid assessment of stools for *E. coli* O157:H7 detection by microbiologists and the immediate report of presumptive positives, since this treatment must be given not later than 2 days after the beginning of *E. coli* O157:H7-associated diarrhea.

The use of antimotility drugs or opioid narcotics have been discouraged in children with acute diarrhea, because of their association with complications of *E. coli* O157:H7 infection and

with the prolongation of symptoms. Thus, antidiarrheal agents are usually avoided, as it is thought that this contributes to retention of Stx within the colon, which could enhance absorption of the toxin [40,100–102].

Antibiotic treatment & potential preventive agents

There is a long history of the discussion of antibiotic treatment for EHEC-induced diarrhea. An extensive analysis of the *E. coli* O157:H7 outbreak in Sakai City, Japan, suggested that treatment with fosfomycin was associated with a significantly decreased risk of HUS. However, this study has several drawbacks. Fosfomycin was compared only with other antibiotics, not with the absence of antibiotic-treatment [103,104], the benefit for antibiotic treatment was only restricted to patients that received the drug on day 2 of their illness, and this study was limited to children infected with the same strain, as is the case in analyses of outbreaks. On the other hand, most occidental researchers currently believe that antibiotics should not be administered to patients with definite or possible enteric STEC infection [105]. Several studies demonstrated that children with hemorrhagic colitis associated with EHEC who received antibiotic therapy were more likely to develop HUS compared with children who did not receive antibiotic therapy [102,106,107]. Antibiotics increase the risk of the HUS by enhancing phage induction and subsequent *stx* gene expression, and by increasing Stx release after induced bacteria lysis [108].

A recent prospective cohort study conducted by a network of 47 laboratories from different states in the USA confirmed that administering sulfa-containing antibiotics to children infected with *E. coli* O157:H7 increases their risk of developing HUS [107], and indicate that β -lactam antibiotics are associated with a similar degree of risk.

We retrospectively reviewed the outcome of 54 patients who received trimetoprim/sulphamethoxazole out of 641 HUS patients admitted to our nephrology unit from 1970 to 1990, and compared their long-term follow-up with those who had not received antibiotics. There was no significant difference between the two groups [RAMÓN A EXENI, PERSONAL OBSERVATION].

In conclusion, during the diarrhea phase, antibiotic treatment should be avoided, as beneficial effects regarding initiation of HUS cannot be deduced from recent studies.

Michael *et al.* have recently reviewed seven randomized, controlled trials for interventions to evaluate their effectiveness for relevant clinical outcomes [109]. They reported that, since the first randomized trial conducted in Argentina [110], several anticoagulation therapies were assayed including heparin alone, heparin and urokinase, or heparin and dipyridamole [109]. In all these trials, the treatment and control groups received supportive therapy. Although there was significant heterogeneity between studies, there was no significant difference between groups for any of the primary or secondary outcomes, including mortality, neurological events and proteinuria or hypertension at the last follow-up. However, the incidence of bleeding (adverse effect) was significantly greater in the group that received anticoagulation therapy compared with supportive therapy alone.

Another Argentinean group conducted a trial (94 patients) comparing steroids with placebo, and there was no significant difference between the two groups for any of the outcome measures of interest [111].

Based on a previous report showing inadequate erythropoietin synthesis in children with HUS [112], it has been recently proposed early administration of erythropoietin. This new treatment reduced the need for red blood cell transfusion in HUS children [113]. The results of this pilot study will have to be confirmed in a larger multicenter trial.

Some trials have used fresh frozen plasma infusion, given no additional benefits compared with supportive therapy [114,115].

A nonconventional treatment for a very severe group of HUS patients, with clinical hemodynamic parameters of septic shock and neurological dysfunction at onset, has been proposed by Valles *et al.* [116]. The protocol includes fresh frozen plasma infusions, methylprednisolone pulses (10 mg/kg/day) for three consecutive days and plasma exchange for 5 days, starting after admission to the intensive care unit. Since nine out of 12 patients survived, compared with five deaths among a historical group of six children with the same severe form, the authors suggest that early introduction of this protocol could benefit HUS patients with hemodynamic instability and neurological dysfunction at onset.

Dialysis

The majority of children with HUS develop some degree of renal insufficiency. Gianantonio innovatively established peritoneal dialysis to manage the acute renal failure in HUS, reducing the mortality from 50 to 5% [4]. Approximately two-thirds of children with HUS will require dialysis therapy, and approximately a third will have milder renal involvement without the need for dialysis therapy [117]. Peritoneal dialysis and hemodialysis modes have been used. In most centers in Argentina, as well as in other countries, peritoneal dialysis is the preferential choice especially when patients are below 1–2 years of age [118,119]. It has been argued that peritoneal dialysis may have a higher risk of peritonitis in patients with bloody diarrhea. However, this has not yet been reported, and there are no randomized, control trials comparing the effectiveness of different types of dialysis (peritoneal dialysis, hemodialysis or continuous venous hemofiltration) in patients with acute renal failure caused by HUS.

Experimental strategies in patients with HUS

One of the first specific therapeutic approaches that raise expectation among pediatric nephrologists was the idea of binding the released Stx in the gut via amorphous compounds. In this regard, a diatomaceous silicon diamide compound linked to an oligosaccharide chain (Synsorb® Pk) was shown to avidly bind and neutralize Stx. However, the Synsorb Pk was not found to be beneficial in preventing extrarenal complications or decreasing the duration of dialysis in children with new-onset HUS [120]. Thereafter, Gb₃ polymers have been developed and showed to entrap Stx in the gut and prevent EHEC toxicity in mice [121]. Under a similar reasoning, Paton *et al.* engineered a recombinant

bacterium displaying a Stx receptor mimic on its surface, which neutralized Stx with very high efficiency, and completely protected STEC-challenged mice when administered three-times daily [122]. This approach is still under investigation. The major drawback of these proposals is that barely traces of Stx that reach circulation would be enough to induce HUS, although these compounds showed a high affinity for Stx and can neutralize significant amounts of Stx in the intestine.

Taking this consideration into account, analogs of the Gb₃ receptor for administration systemically are being developed. Among them, Starfish® is a new compound shown to bind Stx 1000-times more efficiently than Synsorb Pk and has the potential to be administered intravenously. Starfish has been shown to protect mice against a lethal dose of Stx1 but not Stx2, whereas a modified version of Starfish, called Daisy®, protected mice against lethal doses of Stx1 and Stx2 [123]. Alternatively, Nishikawa *et al.* identified a structure with potent Stx-neutralizing activity in the circulation. This compound, named SUPER TWIGS, is formed by 18 trisaccharides of Gb₃, with the capacity of forming complexes with Stx in circulation that enables efficient uptake and degradation of Stx by macrophages [124]. The same authors have described a peptide-based Stx2 inhibitor that has remarkable therapeutic potency and appears to function by inducing aberrant cellular transport and degradation of Stx2 [125]. In addition, there are new therapeutic strategies, including pharmacologic inhibitors of Gb₃ [126], and several anti-Stx antibodies against the different subunits of Stx, from both Stx1 or Stx2 variants, and humanized monoclonal antibodies against Stx. The antibodies are intended to neutralize circulating Stx1 and Stx2, thereby treating the pathogenic agent of the disease and preventing serious complications such as bloody diarrhea, destruction of red blood cells and platelets, and HUS.

The production, characterization and evaluation of a panel of human monoclonal antibodies in transgenic mice were shown to effectively protect mice and piglets against the corresponding toxin challenge [127]. However, it is important to highlight that the absence of antigenic cross-reactivity between B subunits of Stx1 and Stx2 encourage the development of Stx2 antibodies, since Stx2 or its derivatives are the most frequently HUS-associated toxins.

Moreover, monoclonal antibodies directed to the Stx2 A subunit have been proved to be equal or more protective than those directed to the B subunit [128,129]. Although the mechanisms involved are still a matter of discussion, it has been recently reported that anti-A subunit antibodies interfere with retrograde transport of the toxin, preventing toxin-mediated cell death, and may interact with Stx2 when still bound to membrane receptors [130]. In addition, antibodies directed to the Stx2 A subunit as opposed to those directed against the B subunit, have broad-spectrum activity that includes other Stx2 variants, such as Stx2c [128]. Some of these antibodies have recently been approved with the orphan drug status by the US FDA and by the EMEA, for preventing HUS in a dose-escalating, Phase I clinical trial of STEC-infected pediatric patients.

Although therapeutic strategies generally attempt to neutralize bacterial virulence factors, some new proposals have challenged the blocking of host factors that contribute to the pathogenic process. In particular, it has recently been postulated that during typical HUS, the local activation of complement via the alternative pathway by Stx may also play a pathogenic role and then, inhibition of complement cascade may be beneficial [131]. On the other hand, since the inflammatory reaction is important in HUS evolution, TNF- α and cytokines might also be candidates as targets for therapeutic inhibition [28].

STEC diagnosis

One of the limitations for all these new therapeutic approaches is that the window for application is very small. In particular, the diagnosis of STEC infection should be made within 2 days after the initiation of diarrhea, a challenge that cannot be achieved routinely, at least in Argentina. Different reasons may contribute to the inability to identify STEC-infected children as early and accurate as should be done. STEC bacteria not always can be isolated and characterized from stools of diarrheic children, even in children where bloody diarrhea progressed to typical HUS [39]. In addition, the huge number of childhood diarrheas during warm seasons makes critical a sustained community education regarding the need to seek immediate medical attention in the event of bloody diarrhea and highlight the importance of microbiologic evaluation of all of them. An additional limitation is that not all healthy centers from undeveloped regions have the capability to characterize the isolated bacteria by molecular techniques such as PCR. Other criterion for STEC infection is the presence of Stx in stools. However, it has recently been reported that free fecal Stx is detected only in a low percentage of STEC-infected children and, noticeably, the lowest percentage of detection was in the group of children who evolved to HUS [132]. These findings raise the possibility that Stx was produced in greater quantities earlier in the illness and absorbed from the small bowel and not from colonic contents. The third criterion for STEC infection has been the detection of anti-Stx1/Stx2 antibodies in serum. However, the kinetic of antibody production not always allow the detection of them early in the course of infection. Therefore, community education, improvement of local surveillance systems, development of simple and economic diagnostic tools, and identification of predictors for bad evolution are imperative to allow application of new therapeutic tools in those children at high risk of HUS development.

Interventions to avoid STEC transmission

Since cattle and their products are associated with the majority of cases of *E. coli* O157:H7 infection in humans, they represent an attractive target for preslaughter intervention to reduce the risk in humans. A number of approaches are being studied to reduce levels of the organism in cattle, including animal management practices such as chlorination of water [133], modifications in animal feeding [134], the use of probiotics [135] and bacteriophage therapy [136]. Vaccinations using different

immunogens are also being investigated [137]. One big potential barrier is that ranchers and feedlots may have little incentive to pay for such treatments.

Efforts to develop vaccines for people also face barriers. A Phase II trial of O157:H7 serotype LPS vaccine was conducted in 49 children between 2–5 years of age in the USA. An increase in serum IgG LPS antibodies was reported up to 6 months after vaccination [138]. However, since this vaccine does not protect against non-O157 STEC strains and outbreaks in industrialized countries are sporadic, it is difficult to test it in clinical trials. Another concern is that any proposed vaccine should be very safe, because it would be given to children and because the majority of STEC-infected children improve without any intervention.

Expert commentary & five-year view

Currently, there are no specific therapies preventing or ameliorating the disease course. Although there are new therapeutic modalities in the horizon for typical HUS, present recommended therapy is merely symptomatic and supportive. Parenteral volume expansion may counteract the effect of thrombotic processes before development of HUS and attenuate renal injury. A promising erythropoietin treatment has been proposed to reduce the need for red blood cell transfusion in HUS children. Use of antibiotics, antimotility agents, antithrombotic agents, narcotics and nonsteroidal anti-inflammatory drugs should be avoided during the acute phase. Immunization of cattle and humans, and new therapeutic protocols are under investigation, although none are ready for use in the near future. Thus, our weapons to cure HUS are still limited, and prevention is best done by preventing primary STEC infection. For this aim, more strict controls at all points of the food industry and a sustained educational campaign is mandatory, specially in Argentina, where there are over 400 new cases of HUS each year.

The hope that a better understanding of the pathogenesis and transmission routes of this disease will produce better therapies to prevent the acute mortality and long-term morbidity of HUS is the driving force for intensified research. Accumulating evidence from experimental therapies in animal models, and those from the few protocols assayed in humans suggest that the window for application of these new approaches is very small. In particular, the diagnosis of STEC should be made within 2 days after initiation of diarrhea, a challenge that cannot be achieved routinely, at least in Argentina. Therefore, improvement of community awareness of bloody diarrheas, new diagnostic tools and predictors for bad evolution are imperative to allow application of new therapeutic tools in those children at high risk of HUS development.

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The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

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Key issues

- The typical or epidemic form of hemolytic uremic syndrome (HUS) is the major complication of gastrointestinal infections with seropathotypes of Shiga toxin (Stx)-producing *Escherichia coli* (STEC).
- HUS is a public-health problem in Argentina, since it is the second cause of chronic renal failure and accounts for 20% of renal transplantations in children and adolescents.
- Although Stx is the main virulence factor in STEC infections, additional factors from both the bacteria and the host would be necessary to lead to HUS in 10% of STEC infections. Among host factors, there is a great consensus about the importance of the inflammatory and thrombotic responses in the development of HUS.
- Volume expansion has been recently proposed during pre-HUS interval, as a new possibility to attenuate renal injury. On the contrary, the use of antimotility drugs or opioid narcotics has been discouraged as it is thought that these agents contribute to retention of Stx within the colon.
- Similarly, antibiotic treatment should be avoided, as beneficial effects regarding initiation of HUS cannot be deduced from recent studies.
- The early administration of erythropoietin has been recently proposed as a novel approach to reduce red blood cell transfusions in HUS children.
- In addition, there are experimental therapeutic strategies, including analogs of the globotriosylceramide receptor to be administered systemically and several anti-Stx antibodies that are being developed.
- Interventions in cattle are an interesting alternative to reduce the risk of human disease. They include modifications in animal feeding and management, use of probiotics and vaccination.
- Early diagnosis of STEC infection is essential for effective future therapy. Therefore, new diagnostic tools and predictors for bad evolution are imperative to allow application of new therapeutic tools in those children at high risk of HUS development.

References

Papers of special note have been highlighted as:

- of interest

- Wagner J. Acute tubular necrosis with anaemia. *Great Ormond St. J.* 7, 66–72 (1954).
- Gasser C. Atypical, constitutional, non-spherocytic hemolytic anemia. *Schweiz Med Wochenschr.* 88(41), 1014–1018 (1958).
- Gianantonio C, Vitacco M, Mendilaharzu F, Rutty A, Mendilaharzu J. The hemolytic–uremic syndrome. *J. Pediatr.* 64, 478–491 (1964).
- Gianantonio CA, Vitacco M, Mendilaharzu J, Mendilaharzu F, Rutty A. Acute renal failure in infancy and childhood. Clinical course and treatment of 41 patients. *J. Pediatr.* 61, 660–678 (1962).
- Karmali MA, Steele BT, Petric M, Lim C. Sporadic cases of haemolytic–uraemic syndrome associated with faecal cytotoxin and cytotoxin-producing *Escherichia coli* in stools. *Lancet* 1(8325), 619–620 (1983).
- Exeni RA. Hemolytic uremic syndrome. Clinical manifestations. Treatment. *Medicina (B Aires).* 66(Suppl. 3), 6–10 (2006).
- Repetto HA. Long-term course and mechanisms of progression of renal disease in hemolytic uremic syndrome. *Kidney Int. Suppl.* 97, S102–S106 (2005).
- Lopez EL, Devoto S, Fayad A, Canepa C, Morrow AL, Cleary TG. Association between severity of gastrointestinal prodrome and long-term prognosis in classic hemolytic–uremic syndrome. *J. Pediatr.* 120(2 Pt 1), 210–215 (1992).
- Nesmith JD, Ellis E. Childhood hemolytic uremic syndrome is associated with adolescent-onset diabetes mellitus. *Pediatr. Nephrol.* 22(2), 294–297 (2007).
- Suri RS, Mahon JL, Clark WF, Moist LM, Salvadori M, Garg AX. Relationship between *Escherichia coli* O157:H7 and diabetes mellitus. *Kidney Int. Suppl.* 112, S44–S46 (2009).
- Large TM, Walk ST, Whittam TS. Variation in acid resistance among Shiga toxin-producing clones of pathogenic *Escherichia coli*. *Appl. Environ. Microbiol.* 71(5), 2493–2500 (2005).
- Spears KJ, Roe AJ, Gally DL. A comparison of enteropathogenic and enterohaemorrhagic *Escherichia coli* pathogenesis. *FEMS Microbiol. Lett.* 255(2), 187–202 (2006).
- Kaper JB, Nataro JP, Mobley HL. Pathogenic *Escherichia coli*. *Nat. Rev. Microbiol.* 2(2), 123–140 (2004).
- Malyukova I, Murray KF, Zhu C *et al.* Macropinocytosis in Shiga toxin 1 uptake by human intestinal epithelial cells and transcellular transcytosis. *Am. J. Physiol. Gastrointest. Liver Physiol.* 296(1), G78–G92 (2009).
- Gianantonio CA, Vitacco M, Mendilaharzu F, Gallo GE, Sojo ET. The hemolytic–uremic syndrome. *Nephron* 11(2), 174–192 (1973).
- Remuzzi G, Ruggenenti P. The hemolytic uremic syndrome. *Kidney Int.* 48(1), 2–19 (1995).
- van Setten PA, van Hinsbergh VW, van der Velden TJ *et al.* Effects of TNF α on verocytotoxin cytotoxicity in purified human glomerular microvascular endothelial cells. *Kidney Int.* 51(4), 1245–1256 (1997).
- Creydt VP, Silberstein C, Zotta E, Ibarra C. Cytotoxic effect of Shiga toxin-2 holotoxin and its B subunit on human renal tubular epithelial cells. *Microbes Infect.* 8(2), 410–419 (2006).
- Lee JE, Kim JS, Choi IH, Tagawa M, Kohsaka T, Jin DK. Cytokine expression in the renal tubular epithelial cells stimulated by Shiga toxin 2 of *Escherichia coli* O157:H7. *Ren. Fail.* 24(5), 567–575 (2002).
- Nestoridi E, Kushak RI, Duguerre D, Grabowski EF, Ingelfinger JR. Up-regulation of tissue factor activity on human proximal tubular epithelial cells in response to Shiga toxin. *Kidney Int.* 67(6), 2254–2266 (2005).
- Herold S, Karch H, Schmidt H. Shiga toxin-encoding bacteriophages – genomes in motion. *Int. J. Med. Microbiol.* 294(2–3), 115–121 (2004).

- 22 Zhang W, Bielaszewska M, Kuczius T, Karch H. Identification, characterization, and distribution of a Shiga toxin 1 gene variant (*stx(1c)*) in *Escherichia coli* strains isolated from humans. *J. Clin. Microbiol.* 40(4), 1441–1446 (2002).
- 23 Burk C, Dietrich R, Acar G, Moravek M, Bulte M, Martlbauer E. Identification and characterization of a new variant of Shiga toxin 1 in *Escherichia coli* O157:H19 of bovine origin. *J. Clin. Microbiol.* 41(5), 2106–2112 (2003).
- 24 Friedrich AW, Bielaszewska M, Zhang WL *et al.* *Escherichia coli* harboring Shiga toxin 2 gene variants: frequency and association with clinical symptoms. *J. Infect. Dis.* 185(1), 74–84 (2002).
- 25 Kokai-Kun JF, Melton-Celsa AR, O'Brien AD. Elastase in intestinal mucus enhances the cytotoxicity of Shiga toxin type 2d. *J. Biol. Chem.* 275(5), 3713–3721 (2000).
- 26 Sonntag AK, Bielaszewska M, Mellmann A *et al.* Shiga toxin 2c-producing *Escherichia coli* isolates from humans and pigs differ in their virulence profiles and interactions with intestinal epithelial cells. *Appl. Environ. Microbiol.* 71(12), 8855–8863 (2005).
- 27 Sonntag AK, Zenner E, Karch H, Bielaszewska M. Pigeons as a possible reservoir of Shiga toxin 2f-producing *Escherichia coli* pathogenic to humans. *Berl. Munch. Tierarztl. Wochenschr.* 118(11–12), 464–470 (2005).
- 28 Scheiring J, Andreoli SP, Zimmerhackl LB. Treatment and outcome of Shiga-toxin-associated hemolytic uremic syndrome (HUS). *Pediatr. Nephrol.* 23(10), 1749–1760 (2008).
- 29 Tam P, Mahfoud R, Nutikka A *et al.* Differential intracellular transport and binding of verotoxin 1 and verotoxin 2 to globotriaosylceramide-containing lipid assemblies. *J. Cell Physiol.* 216(3), 750–763 (2008).
- 30 Smith DC, Sillence DJ, Falguieres T *et al.* The association of Shiga-like toxin with detergent-resistant membranes is modulated by glucosylceramide and is an essential requirement in the endoplasmic reticulum for a cytotoxic effect. *Mol. Biol. Cell* 17(3), 1375–1387 (2006).
- 31 Tam PJ, Lingwood CA. Membrane cytosolic translocation of verotoxin A1 subunit in target cells. *Microbiology* 153(Pt 8), 2700–2710 (2007).
- 32 Falguieres T, Mallard F, Baron C *et al.* Targeting of Shiga toxin B-subunit to retrograde transport route in association with detergent-resistant membranes. *Mol. Biol. Cell* 12(8), 2453–2468 (2001).
- 33 Hoey DE, Sharp L, Currie C, Lingwood CA, Gally DL, Smith DG. Verotoxin 1 binding to intestinal crypt epithelial cells results in localization to lysosomes and abrogation of toxicity. *Cell Microbiol.* 5(2), 85–97 (2003).
- 34 Zoja C, Angioletti S, Donadelli R *et al.* Shiga toxin-2 triggers endothelial leukocyte adhesion and transmigration via NF- κ B dependent up-regulation of IL-8 and MCP-1. *Kidney Int.* 62(3), 846–856 (2002).
- 35 Morigi M, Galbusera M, Binda E *et al.* Verotoxin-1-induced up-regulation of adhesive molecules renders microvascular endothelial cells thrombogenic at high shear stress. *Blood* 98(6), 1828–1835 (2001).
- 36 Petruzzello TN, Mawji IA, Khan M, Marsden PA. Verotoxin biology: molecular events in vascular endothelial injury. *Kidney Int. Suppl.* 112, S17–S19 (2009).
- 37 Warnier M, Romer W, Geelen J *et al.* Trafficking of Shiga toxin/Shiga-like toxin-1 in human glomerular microvascular endothelial cells and human mesangial cells. *Kidney Int.* 70(12), 2085–2091 (2006).
- 38 Schmidt H, Scheef J, Huppertz HI, Frosch M, Karch H. *Escherichia coli* O157:H7 and O157:H(-) strains that do not produce Shiga toxin: phenotypic and genetic characterization of isolates associated with diarrhea and hemolytic-uremic syndrome. *J. Clin. Microbiol.* 37(11), 3491–3496 (1999).
- 39 Rivas M, Miliwebsky E, Chinen I *et al.* Characterization and epidemiologic subtyping of Shiga toxin-producing *Escherichia coli* strains isolated from hemolytic uremic syndrome and diarrhea cases in Argentina. *Foodborne Pathog. Dis.* 3(1), 88–96 (2006).
- **A total of 99 isolates from Argentine children were investigated to establish the phenotypic and genotypic characteristics of the Shiga toxin-producing *Escherichia coli* (STEC) strains and the clonal relatedness of STEC O157:H7 strains using subtyping techniques.**
- 40 Tarr PI, Gordon CA, Chandler WL. Shiga-toxin-producing *Escherichia coli* and haemolytic uraemic syndrome. *Lancet* 365(9464), 1073–1086 (2005).
- 41 Nielsen EM, Scheutz F, Torpdahl M. Continuous surveillance of Shiga toxin-producing *Escherichia coli* infections by pulsed-field gel electrophoresis shows that most infections are sporadic. *Foodborne Pathog. Dis.* 3(1), 81–87 (2006).
- 42 Roldgaard BB, Scheutz F, Boel J *et al.* VTEC O157 subtypes associated with the most severe clinical symptoms in humans constitute a minor part of VTEC O157 isolates from Danish cattle. *Int. J. Med. Microbiol.* 294(4), 255–259 (2004).
- 43 Gyles CL. Shiga toxin-producing *Escherichia coli*: an overview. *J. Anim. Sci.* 85(13 Suppl.), E45–62 (2007).
- **Excellent review highlighting the importance of cattle in STEC infections, focusing on bacterial virulence factors.**
- 44 Karmali MA. Host and pathogen determinants of verocytotoxin-producing *Escherichia coli*-associated hemolytic uremic syndrome. *Kidney Int. Suppl.* 112, S4–S7 (2009).
- 45 Coombes BK, Wickham ME, Mascarenhas M, Gruenheid S, Finlay BB, Karmali MA. Molecular analysis as an aid to assess the public health risk of non-O157 Shiga toxin-producing *Escherichia coli* strains. *Appl. Environ. Microbiol.* 74(7), 2153–2160 (2008).
- 46 Ethelberg S, Olsen KE, Scheutz F *et al.* Virulence factors for hemolytic uremic syndrome, Denmark. *Emerg. Infect. Dis.* 10(5), 842–847 (2004).
- 47 Bielaszewska M, Friedrich AW, Aldick T, Schurk-Bulgrin R, Karch H. Shiga toxin activatable by intestinal mucus in *Escherichia coli* isolated from humans: predictor for a severe clinical outcome. *Clin Infect Dis.* 43(9), 1160–1167 (2006).
- 48 Eklund M, Leino K, Siitonen A. Clinical *Escherichia coli* strains carrying *stx* genes: *stx* variants and *stx*-positive virulence profiles. *J. Clin. Microbiol.* 40(12), 4585–4593 (2002).
- 49 Taneike I, Zhang HM, Wakisaka-Saito N, Yamamoto T. Enterohemolysin operon of Shiga toxin-producing *Escherichia coli*: a virulence function of inflammatory cytokine production from human monocytes. *FEBS Lett.* 524(1–3), 219–224 (2002).
- 50 Schmidt H, Karch H. Enterohemolytic phenotypes and genotypes of shiga toxin-producing *Escherichia coli* O111 strains from patients with diarrhea and hemolytic uremic syndrome. *J. Clin. Microbiol.* 34(10), 2364–2367 (1996).
- 51 Zhang WL, Bielaszewska M, Liesegang A *et al.* Molecular characteristics and epidemiological significance of Shiga toxin-producing *Escherichia coli* O26 strains. *J. Clin. Microbiol.* 38(6), 2134–2140 (2000).

- 52 Cookson AL, Croucher D, Pope C, Bennett J, Thomson-Carter F, Attwood GT. Isolation, characterization, and epidemiological assessment of Shiga toxin-producing *Escherichia coli* O84 isolates from New Zealand. *J. Clin. Microbiol.* 44(5), 1863–1866 (2006).
- 53 Paton AW, Srimanote P, Talbot UM, Wang H, Paton JC. A new family of potent AB(5) cytotoxins produced by Shiga toxigenic *Escherichia coli*. *J. Exp. Med.* 200(1), 35–46 (2004).
- 54 Kondo Y, Tokuda N, Fan X *et al.* Glycosphingolipids are not pivotal receptors for subtilase cytotoxin *in vivo*: sensitivity analysis with glycosylation-defective mutant mice. *Biochem. Biophys. Res. Commun.* 378(2), 179–181 (2009).
- 55 Matsuura G, Morinaga N, Yahiro K *et al.* Novel subtilase cytotoxin produced by Shiga-toxigenic *Escherichia coli* induces apoptosis in Vero cells via mitochondrial membrane damage. *Infect. Immun.* 77(7), 2919–2924 (2009).
- 56 Morinaga N, Yahiro K, Matsuura G, Moss J, Noda M. Subtilase cytotoxin, produced by Shiga-toxigenic *Escherichia coli*, transiently inhibits protein synthesis of Vero cells via degradation of BiP and induces cell cycle arrest at G1 by downregulation of cyclin D1. *Cell Microbiol.* 10(4), 921–929 (2008).
- 57 Wang H, Paton JC, Paton AW. Pathologic changes in mice induced by subtilase cytotoxin, a potent new *Escherichia coli* AB5 toxin that targets the endoplasmic reticulum. *J. Infect. Dis.* 196(7), 1093–1101 (2007).
- 58 Janka A, Bielaszewska M, Dobrindt U, Greune L, Schmidt MA, Karch H. Cytolethal distending toxin gene cluster in enterohemorrhagic *Escherichia coli* O157:H and O157:H7: characterization and evolutionary considerations. *Infect. Immun.* 71(6), 3634–3638 (2003).
- 59 Bielaszewska M, Fell M, Greune L *et al.* Characterization of cytolethal distending toxin genes and expression in shiga toxin-producing *Escherichia coli* strains of non-O157 serogroups. *Infect. Immun.* 72(3), 1812–1816 (2004).
- 60 Grys TE, Siegel MB, Latham WW, Welch RA. The StcE protease contributes to intimate adherence of enterohemorrhagic *Escherichia coli* O157:H7 to host cells. *Infect. Immun.* 73(3), 1295–1303 (2005).
- 61 Dahan S, Wiles S, La Ragione RM *et al.* EspJ is a prophage-carried type III effector protein of attaching and effacing pathogens that modulates infection dynamics. *Infect. Immun.* 73(2), 679–686 (2005).
- 62 Louise CB, Obrig TG. Shiga toxin-associated hemolytic uremic syndrome: combined cytotoxic effects of Shiga toxin and lipopolysaccharide (endotoxin) on human vascular endothelial cells *in vitro*. *Infect. Immun.* 60(4), 1536–1543 (1992).
- 63 Palermo M, Alves-Rosa F, Rubel C *et al.* Pretreatment of mice with lipopolysaccharide (LPS) or IL-1 β exerts dose-dependent opposite effects on Shiga toxin-2 lethality. *Clin. Exp. Immunol.* 119(1), 77–83 (2000).
- 64 Keepers TR, Psotka MA, Gross LK, Obrig TG. A murine model of HUS: Shiga toxin with lipopolysaccharide mimics the renal damage and physiologic response of human disease. *J. Am. Soc. Nephrol.* 17(12), 3404–3414 (2006).
- 65 Sauter KA, Melton-Celsa AR, Larkin K, Troxell ML, O'Brien AD, Magun BE. Mouse model of hemolytic-uremic syndrome caused by endotoxin-free Shiga toxin 2 (Stx2) and protection from lethal outcome by anti-Stx2 antibody. *Infect. Immun.* 76(10), 4469–4478 (2008).
- 66 van Setten PA, Monnens LA, Verstraten RG, van den Heuvel LP, vanHinsbergh VW. Effects of verocytotoxin-1 on nonadherent human monocytes: binding characteristics, protein synthesis, and induction of cytokine release. *Blood* 88(1), 174–183 (1996).
- 67 Thorpe CM, Hurley BP, Lincicome LL, Jacewicz MS, Keusch GT, Acheson DW. Shiga toxins stimulate secretion of interleukin-8 from intestinal epithelial cells. *Infect. Immun.* 67(11), 5985–5993 (1999).
- 68 Tesh VL, Samuel JE, Perera LP, Sharefkin JB, O'Brien AD. Evaluation of the role of Shiga and Shiga-like toxins in mediating direct damage to human vascular endothelial cells. *J. Infect. Dis.* 164(2), 344–352 (1991).
- 69 Miyamoto Y, Iimura M, Kaper JB, Torres AG, Kagnoff MF. Role of Shiga toxin versus H7 flagellin in enterohemorrhagic *Escherichia coli* signalling of human colon epithelium *in vivo*. *Cell Microbiol.* 8(5), 869–879 (2006).
- 70 Hurley BP, Thorpe CM, Acheson DW. Shiga toxin translocation across intestinal epithelial cells is enhanced by neutrophil transmigration. *Infect. Immun.* 69(10), 6148–6155 (2001).
- 71 Wagner PL, Acheson DW, Waldor MK. Human neutrophils and their products induce Shiga toxin production by enterohemorrhagic *Escherichia coli*. *Infect. Immun.* 69(3), 1934–1937 (2001).
- 72 Bielaszewska M, Karch H. Consequences of enterohaemorrhagic *Escherichia coli* infection for the vascular endothelium. *Thromb. Haemost.* 94(2), 312–318 (2005).
- 73 Chandler WL, Jelacic S, Boster DR *et al.* Prothrombotic coagulation abnormalities preceding the hemolytic-uremic syndrome. *N. Engl. J. Med.* 346(1), 23–32 (2002).
- 74 Fernandez GC, Gomez SA, Rubel CJ *et al.* Impaired neutrophils in children with the typical form of hemolytic uremic syndrome. *Pediatr. Nephrol.* 20(9), 1306–1314 (2005).
- 75 Fernandez GC, Rubel C, Barrionuevo P *et al.* Phenotype markers and function of neutrophils in children with hemolytic uremic syndrome. *Pediatr. Nephrol.* 17(5), 337–344 (2002).
- 76 Exeni RA, Fernandez GC, Palermo MS. Role of polymorphonuclear leukocytes in the pathophysiology of typical hemolytic uremic syndrome. *Scientific World Journal* 7, 1155–1164 (2007).
- Summarizes the current knowledge regarding the contribution of neutrophils in the pathogenic mechanisms of hemolytic uremic syndrome (HUS), arising from findings in both experimental models and direct studies on neutrophils from HUS patients.
- 77 Fernandez GC, Gomez SA, Ramos MV *et al.* The functional state of neutrophils correlates with the severity of renal dysfunction in children with hemolytic uremic syndrome. *Pediatr. Res.* 61(1), 123–128 (2007).
- 78 Sasseti B, Vizcarguenaga MI, Zanaro NL *et al.* Hemolytic uremic syndrome in children: platelet aggregation and membrane glycoproteins. *J. Pediatr. Hematol. Oncol.* 21(2), 123–128 (1999).
- 79 Appiani AC, Edefonti A, Bettinelli A, Cossu MM, Paracchini ML, Rossi E. The relationship between plasma levels of the factor VIII complex and platelet release products (β -thromboglobulin and platelet factor 4) in children with the hemolytic-uremic syndrome. *Clin. Nephrol.* 17(4), 195–199 (1982).
- 80 Katayama M, Handa M, Araki Y *et al.* Soluble P-selectin is present in normal circulation and its plasma level is elevated in patients with thrombotic thrombocytopenic purpura and haemolytic uraemic syndrome. *Br. J. Haematol.* 84(4), 702–710 (1993).

- 81 Galli M, Grassi A, Barbui T. Platelet-derived microvesicles in thrombotic thrombocytopenic purpura and hemolytic uremic syndrome. *Thromb. Haemost.* 75(3), 427–431 (1996).
- 82 Fernandez GC, Ramos MV, Gomez SA *et al.* Differential expression of function-related antigens on blood monocytes in children with hemolytic uremic syndrome. *J. Leukoc. Biol.* 78(4), 853–861 (2005).
- 83 Ziegler-Heitbrock HW, Fingerle G, Strobel M *et al.* The novel subset of CD14⁺/CD16⁺ blood monocytes exhibits features of tissue macrophages. *Eur. J. Immunol.* 23(9), 2053–2058 (1993).
- 84 Ancuta P, Weiss L, Haeffner-Cavaillon N. CD14⁺CD16⁺ cells derived *in vitro* from peripheral blood monocytes exhibit phenotypic and functional dendritic cell-like characteristics. *Eur. J. Immunol.* 30(7), 1872–1883 (2000).
- 85 Ramos MV, Fernandez GC, Patey N *et al.* Involvement of the fractalkine pathway in the pathogenesis of childhood hemolytic uremic syndrome. *Blood* 109(6), 2438–2445 (2007).
- 86 Brooks JT, Sowers EG, Wells JG *et al.* Non-O157 Shiga toxin-producing *Escherichia coli* infections in the United States, 1983–2002. *J. Infect. Dis.* 192(8), 1422–1429 (2005).
- 87 Rivas M, Miliwsky E, Chinen I, Deza N, Leotta GA. The epidemiology of hemolytic uremic syndrome in Argentina. Diagnosis of the etiologic agent, reservoirs and routes of transmission. *Medicina (B Aires)* 66(Suppl. 3), 27–32 (2006).
- 88 Mead PS, Griffin PM. *Escherichia coli* O157:H7. *Lancet* 352(9135), 1207–1212 (1998).
- 89 Parma AE, Sanz ME, Blanco JE *et al.* Virulence genotypes and serotypes of verotoxigenic *Escherichia coli* isolated from cattle and foods in Argentina. Importance in public health. *Eur. J. Epidemiol.* 16(8), 757–762 (2000).
- 90 Meichtri L, Miliwsky E, Gioffre A *et al.* Shiga toxin-producing *Escherichia coli* in healthy young beef steers from Argentina: prevalence and virulence properties. *Int. J. Food Microbiol.* 96(2), 189–198 (2004).
- 91 Rivas M, Caletti MG, Chinen I *et al.* Home-prepared hamburger and sporadic hemolytic uremic syndrome, Argentina. *Emerg. Infect. Dis.* 9(9), 1184–1186 (2003).
- 92 Oteiza JM, Chinen I, Miliwsky E, Rivas M. Isolation and characterization of Shiga toxin-producing *Escherichia coli* from precooked sausages (morcillas). *Food Microbiol.* 23(3), 283–288 (2006).
- 93 Chinen I, Epszteyn S, Melamed CL *et al.* Shiga toxin-producing *Escherichia coli* O157 in beef and chicken burgers, and chicken carcasses in Buenos Aires, Argentina. *Int. J. Food Microbiol.* 132(2–3), 167–171 (2009).
- 94 Miliwsky E, Deza N, Chinen I *et al.* Prolonged fecal shedding of Shiga toxin-producing *Escherichia coli* among children attending day-care centers in Argentina. *Rev. Argent. Microbiol.* 39(2), 90–92 (2007).
- 95 Belongia EA, Osterholm MT, Soler JT, Ammend DA, Braun JE, MacDonald KL. Transmission of *Escherichia coli* O157:H7 infection in Minnesota child day-care facilities. *JAMA* 269(7), 883–888 (1993).
- 96 Crump JA, Sulka AC, Langer AJ *et al.* An outbreak of *Escherichia coli* O157:H7 infections among visitors to a dairy farm. *N. Engl. J. Med.* 347(8), 555–560 (2002).
- 97 Yoon JW, Hovde CJ. All blood, no stool: enterohemorrhagic *Escherichia coli* O157:H7 infection. *J. Vet. Sci.* 9(3), 219–231 (2008).
- 98 Gonzalez G, Pianciola L, Russo VD *et al.* *Escherichia coli* outbreaks, San martin de los Andes, Neuquén, Argentina. Presented at: VTEC 2009. Buenos Aires, Argentina, 10–13 May (2009).
- 99 Ake JA, Jelacic S, Ciol MA *et al.* Relative nephroprotection during *Escherichia coli* O157:H7 infections: association with intravenous volume expansion. *Pediatrics* 115(6), e673–e680 (2005).
- 100 Andreoli SP, Trachtman H, Acheson DW, Siegler RL, Obrig TG. Hemolytic uremic syndrome: epidemiology, pathophysiology, and therapy. *Pediatr. Nephrol.* 17(4), 293–298 (2002).
- 101 Siegler R, Oakes R. Hemolytic uremic syndrome; pathogenesis, treatment, and outcome. *Curr. Opin. Pediatr.* 17(2), 200–204 (2005).
- 102 Zimmerhackl LB. *E. coli*, antibiotics, and the hemolytic–uremic syndrome. *N. Engl. J. Med.* 342(26), 1990–1991 (2000).
- 103 Ikeda K, Ida O, Kimoto K, Takatorige T, Nakanishi N, Tataru K. Effect of early fosfomicin treatment on prevention of hemolytic uremic syndrome accompanying *Escherichia coli* O157:H7 infection. *Clin. Nephrol.* 52(6), 357–362 (1999).
- 104 Kawamura N, Yamazaki T, Tamai H. Risk factors for the development of *Escherichia coli* O157:H7 associated with hemolytic uremic syndrome. *Pediatr. Int.* 41(2), 218–222 (1999).
- 105 Iijima K, Kamioka I, Nozu K. Management of diarrhea-associated hemolytic uremic syndrome in children. *Clin. Exp. Nephrol.* 12(1), 16–19 (2008).
- 106 Safdar N, Said A, Gangnon RE, Maki DG. Risk of hemolytic uremic syndrome after antibiotic treatment of *Escherichia coli* O157:H7 enteritis: a meta-analysis. *JAMA* 288(8), 996–1001 (2002).
- 107 Wong CS, Jelacic S, Habeeb RL, Watkins SL, Tarr PI. The risk of the hemolytic–uremic syndrome after antibiotic treatment of *Escherichia coli* O157:H7 infections. *N. Engl. J. Med.* 342(26), 1930–1936 (2000).
- **Important revision about antibiotic treatment: prospective, randomized and with a significant result.**
- 108 Karch H, Friedrich AW, Gerber A, Zimmerhackl LB, Schmidt MA, Bielaszewska M. New aspects in the pathogenesis of enteropathic hemolytic uremic syndrome. *Semin. Thromb. Hemost.* 32(2), 105–112 (2006).
- 109 Michael M, Elliott EJ, Craig JC, Ridley G, Hodson EM. Interventions for hemolytic uremic syndrome and thrombotic thrombocytopenic purpura: a systematic review of randomized controlled trials. *Am. J. Kidney Dis.* 53(2), 259–272 (2009).
- **Very complete and prospective revision about different interventions.**
- 110 Vitacco M, Sanchez Avalos J, Gianantonio CA. Heparin therapy in the hemolytic–uremic syndrome. *J. Pediatr.* 83(2), 271–275 (1973).
- 111 Perez N, Spizzirri F, Rahman R, Suarez A, Larrubia C, Lasarte P. Steroids in the hemolytic uremic syndrome. *Pediatr. Nephrol.* 12(2), 101–104 (1998).
- 112 Exeni R, Donato H, Rendo P *et al.* Low levels of serum erythropoietin in children with endemic hemolytic uremic syndrome. *Pediatr. Nephrol.* 12(3), 226–230 (1998).
- 113 Pape L, Ahlenstiel T, Kreuzer M *et al.* Early erythropoietin reduced the need for red blood cell transfusion in childhood hemolytic uremic syndrome – a randomized prospective pilot trial. *Pediatr. Nephrol.* 24(5), 1061–1064 (2008).
- **Refers to an interesting and promising approach to reduce the number of blood transfusions in HUS patients.**
- 114 Loirat C, Sonsino E, Hinglais N, Jais JP, Landais P, Fermanian J. Treatment of the childhood haemolytic uraemic syndrome with plasma. A multicentre randomized controlled trial. The French Society of Paediatric Nephrology. *Pediatr. Nephrol.* 2(3), 279–285 (1988).

- 115 Rizzoni G, Claris-Appiani A, Edefonti A *et al.* Plasma infusion for hemolytic-uremic syndrome in children: results of a multicenter controlled trial. *J. Pediatr.* 112(2), 284–290 (1988).
- 116 Valles PG, Pesle S, Piovano L *et al.* Postdiarrheal Shiga toxin-mediated hemolytic uremic syndrome similar to septic shock. *Medicina (B Aires)* 65(5), 395–401 (2005).
- 117 Serna At, Boedeker EC. Pathogenesis and treatment of Shiga toxin-producing *Escherichia coli* infections. *Curr. Opin. Gastroenterol.* 24(1), 38–47 (2008).
- 118 Cobenas CJ, Alconcher LF, Spizzirri AP, Rahman RC. Long-term follow-up of Argentinean patients with hemolytic uremic syndrome who had not undergone dialysis. *Pediatr. Nephrol.* 22(9), 1343–1347 (2007).
- 119 Zambrano OP, Delucchi BA, Cavagnaro SF *et al.* Hemolytic-uremic syndrome in Chile: clinical features, evolution and prognostic factors. *Rev. Med. Chil.* 136(10), 1240–1246 (2008).
- 120 Trachtman H, Nnaan A, Christen E *et al.* Effect of an oral Shiga toxin-binding agent on diarrhea-associated hemolytic uremic syndrome in children: a randomized controlled trial. *JAMA* 290(10), 1337–1344 (2003).
- 121 Watanabe M, Matsuoka K, Kita E *et al.* Oral therapeutic agents with highly clustered globotriose for treatment of Shiga toxin-producing *Escherichia coli* infections. *J. Infect. Dis.* 189(3), 360–368 (2004).
- 122 Paton AW, Morona R, Paton JC. Neutralization of Shiga toxins Stx1, Stx2c, and Stx2e by recombinant bacteria expressing mimics of globotriose and globotetraose. *Infect. Immun.* 69(3), 1967–1970 (2001).
- 123 Mulvey GL, Marcato P, Kitov PI, Sadowska J, Bundle DR, Armstrong GD. Assessment in mice of the therapeutic potential of tailored, multivalent Shiga toxin carbohydrate ligands. *J. Infect. Dis.* 187(4), 640–649 (2003).
- 124 Nishikawa K, Matsuoka K, Watanabe M *et al.* Identification of the optimal structure required for a Shiga toxin neutralizer with oriented carbohydrates to function in the circulation. *J. Infect. Dis.* 191(12), 2097–2105 (2005).
- 125 Nishikawa K, Watanabe M, Kita E *et al.* A multivalent peptide library approach identifies a novel Shiga toxin inhibitor that induces aberrant cellular transport of the toxin. *FASEB J.* 20(14), 2597–2599 (2006).
- 126 Silberstein C, Copeland DP, Chiang W, Repetto HA, Ibarra C. A glucosylceramide synthase inhibitor prevents the cytotoxic effects of Shiga toxin-2 on human renal tubular epithelial cells. *J. Epithel. Biol. Pharmacol.* 1(5), 71–75 (2009).
- 127 Mukherjee J, Chios K, Fishwild D *et al.* Human Stx2-specific monoclonal antibodies prevent systemic complications of *Escherichia coli* O157:H7 infection. *Infect. Immun.* 70(2), 612–619 (2002).
- 128 Sheoran AS, Chapman S, Singh P, Donohue-Rolfe A, Tzipori S. Stx2-specific human monoclonal antibodies protect mice against lethal infection with *Escherichia coli* expressing Stx2 variants. *Infect. Immun.* 71(6), 3125–3130 (2003).
- 129 Smith MJ, Carvalho HM, Melton-Celsa AR, O'Brien AD. The 13C4 monoclonal antibody that neutralizes Shiga toxin type 1 (Stx1) recognizes three regions on the Stx1 B subunit and prevents Stx1 from binding to its eukaryotic receptor globotriaosylceramide. *Infect. Immun.* 74(12), 6992–6998 (2006).
- 130 Krautz-Peterson G, Chapman-Bonofiglio S, Boisvert K *et al.* Intracellular neutralization of shiga toxin 2 by an a subunit-specific human monoclonal antibody. *Infect. Immun.* 76(5), 1931–1939 (2008).
- **Highlights the importance of including antibodies against the A subunit in therapeutic formulations with Stx2-neutralizing perspectives.**
- 131 Orth D, Khan AB, Naim A *et al.* Shiga toxin activates complement and binds factor H: evidence for an active role of complement in hemolytic uremic syndrome. *J. Immunol.* 182(10), 6394–6400 (2009).
- 132 Cornick NA, Jelacic S, Ciol MA, Tarr PI. *Escherichia coli* O157:H7 infections: discordance between filterable fecal Shiga toxin and disease outcome. *J. Infect. Dis.* 186(1), 57–63 (2002).
- 133 Rice EW, Clark RM, Johnson CH. Chlorine inactivation of *Escherichia coli* O157:H7. *Emerg. Infect. Dis.* 5(3), 461–463 (1999).
- 134 Buchko SJ, Holley RA, Olson WO, Gannon VP, Veira DM. The effect of different grain diets on fecal shedding of *Escherichia coli* O157:H7 by steers. *J. Food Prot.* 63(11), 1467–1474 (2000).
- 135 Stephens TP, Loneragan GH, Karunasena E, Brashears MM. Reduction of *Escherichia coli* O157 and *Salmonella* in feces and on hides of feedlot cattle using various doses of a direct-fed microbial. *J. Food Prot.* 70(10), 2386–2391 (2007).
- **Demonstrates of valuable data on reducing the level of bacteria in the main reservoir of *E. coli* O157.**
- 136 Sheng H, Knecht HJ, Kudva IT, Hovde CJ. Application of bacteriophages to control intestinal *Escherichia coli* O157:H7 levels in ruminants. *Appl. Environ. Microbiol.* 72(8), 5359–5366 (2006).
- 137 Horne C, Vallance BA, Deng W, Finlay BB. Current progress in enteropathogenic and enterohemorrhagic *Escherichia coli* vaccines. *Expert Rev. Vaccines* 1(4), 483–493 (2002).
- 138 Ahmed A, Li J, Shiloach Y, Robbins JB, Szu SC. Safety and immunogenicity of *Escherichia coli* O157 O-specific polysaccharide conjugate vaccine in 2–5-year-old children. *J. Infect. Dis.* 193(4), 515–521 (2006).

Affiliations

- Marina S Palermo, PhD
Lab Inmunología, Instituto de Investigaciones Hematológicas, Academia Nacional de Medicina P. de Melo 3081 (C1425AUM), Ciudad de Buenos Aires, Argentina
Tel.: +54 114 805 5695
Fax: +54 114 803 9475
mspalermo@hematologia.anm.edu.ar
- Ramón A Exeni, MD
Departamento de Nefrología, Hospital Municipal del Niño, San Justo, Provincia de Buenos Aires, Argentina
Tel.: +54 114 441 9371
rexeni@pccp.com.ar
- Gabriela C Fernández, PhD
Lab Inmunología, Instituto de Investigaciones Hematológicas, Academia Nacional de Medicina P. de Melo 3081 (C1425AUM), Ciudad de Buenos Aires, Argentina
Tel.: +54 114 805 5695
Fax: +54 114 803 9475
gfernandez@hematologia.anm.edu.ar