

RESEARCH ARTICLE

Postprandial PYY increase by resistant starch supplementation is independent of net portal appearance of short-chain fatty acids in pigs

Anne Krog Ingerslev¹, Shivaprakash Jagalur Mutt², Helle Nygaard Lærke¹, Mette Skou Hedemann¹, Peter Kappel Theil¹, Kirstine Lykke Nielsen^{1a}, Henry Jørgensen¹, Karl-Heinz Herzig^{2,3,4}, Knud Erik Bach Knudsen^{1*}

1 Department of Animal Science, Aarhus University, Tjele, Denmark, **2** Research Unit of Biomedicine and Biocenter of Oulu, Department of Physiology, University of Oulu, Oulu, Finland, **3** Department of Gastroenterology and Metabolism, Poznan University of Medical Sciences, Poznan, Poland, **4** Medical Research Center (MRC) and University Hospital, Oulu, Finland

^a Current Address: Department of Forensic Medicine, Forensic Chemistry, Aarhus University, Aarhus N, Denmark

* KnudErik.Bach.Knudsen@anis.au.dk



OPEN ACCESS

Citation: Ingerslev AK, Mutt SJ, Lærke HN, Hedemann MS, Theil PK, Nielsen KL, et al. (2017) Postprandial PYY increase by resistant starch supplementation is independent of net portal appearance of short-chain fatty acids in pigs. *PLoS ONE* 12(10): e0185927. <https://doi.org/10.1371/journal.pone.0185927>

Editor: François Blachier, National Institute for Agronomic Research, FRANCE

Received: June 14, 2017

Accepted: September 21, 2017

Published: October 5, 2017

Copyright: © 2017 Ingerslev et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: The present research projects were financially supported by The Danish Strategic Research Council, project “Concepts for enhanced butyrate production to improve colonic health and insulin sensitivity – ButColns” (Project No. 10-093526), and by the European Commission in the Communities Sixth Framework Programme,

Abstract

Increased dietary fiber (DF) fermentation and short-chain fatty acid (SCFA) production may stimulate peptide tyrosine-tyrosine (PYY) secretion. In this study, the effects of hindgut SCFA production on postprandial PYY plasma levels were assessed using different experimental diets in a porto-arterial catheterized pig model. The pigs were fed experimental diets varying in source and levels of DF for one week in 3×3 Latin square designs. The DF sources were whole-wheat grain, wheat aleurone, rye aleurone-rich flour, rye flakes, and resistant starch. Postprandial blood samples were collected from the catheters and analyzed for PYY levels and net portal appearance (NPA) of PYY was correlated to NPA of SCFA. No significant effects of diets on NPA of PYY were observed ($P > 0.05$), however, resistant starch supplementation increased postprandial NPA of PYY levels by 37 to 54% compared with rye-based and Western-style control diets ($P = 0.19$). This increase was caused by higher mesenteric artery and portal vein PYY plasma levels ($P < 0.001$) and was independent of SCFA absorption ($P > 0.05$). The PYY levels were higher in response to the second daily meal compared with the first daily meal ($P < 0.001$), but similar among diets ($P > 0.10$). In conclusion, the increased postprandial PYY responses in pigs fed with different levels and sources of DF are not caused by an increased SCFA absorption and suggest that other mechanisms such as neural reflexes and possibly an increased flow of digesta in the small intestine may be involved. The content of DF and SCFA production did not affect PYY levels.

Project HEALTHGRAIN (FOOD-CT-2005-514008) to KEBK. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Introduction

The gastrointestinal tract is a large endocrine organ releasing regulatory peptides in response to nutrient content [1]. Peptide tyrosine-tyrosine (PYY) is a hormone synthesized and released in response to food intake from the endocrine L-cells mainly in the distal part of the gastrointestinal tract, such as ileum and colon, and has several gut functions that contribute to post-prandial satiety and decreased food intake [2]. These functions mediate, among others, ileal and colonic breaks to slow gastric emptying and promote digestive activities including regulation of insulin secretion and glucose homeostasis [2, 3].

Dietary fiber (DF) is a nutritional component associated with increased satiety feelings [4]. This is due to properties of adding bulk and producing intraluminal viscosity in the gastrointestinal tract that delay gastric emptying, and inhibit the rate of digestion and absorption of macronutrients [5, 6] with beneficial effects on glycaemia and insulinemia [7]. Furthermore, Karhunen *et al.* [8] showed that in humans a soluble psyllium-fiber enriched meal induced a prolonged increase in PYY concentration. However, Juvonen *et al.* [9] found no short-term PYY effects of oat bran supplemented puddings in healthy subjects, suggesting that the discrepancies might be fiber dependent. Fermentation of DF by microbes in the distal part of the small intestine and colon additionally produces short-chain fatty acid (SCFA), mainly acetate, propionate, and butyrate, that are known to affect PYY release. Previous studies have shown that rectal and ileal infusions of SCFA in rodents, pigs, and humans stimulate PYY release [10–12]. Particularly butyrate has received considerable attention due to its multiple effects on colonic health [13–15]. A recent *in vivo* study in mice showed that oral propionate or butyrate administration increased PYY secretion and reduced feed intake [16]. However, appetite regulation by SCFA is a more long-term regulation since nutrients need to reach the colon before DF fermentation and SCFA absorption can take place.

Based on the importance of SCFA in the release of PYY and its suppressive effect on appetite, we determined the effect of absorbed SCFA on PYY levels by analyzing the plasma PYY concentrations and correlated them with previously studied varying SCFA levels from feed containing different levels of DF. Previously, we studied the effects of resistant starch and rye-based diets on portal absorption of SCFA and insulin sensitivity, demonstrating that a high portal SCFA absorption was associated with lower insulin secretion [17, 18]. In addition, SCFA absorption was higher and insulin levels lower after the second meal in the same day compared to the first meal. Therefore, in this study we investigated if PYY levels are affected by DF supplementations and how these changes correlate with the net portal appearance (NPA) of SCFA. Therefore, plasma samples from two separate and previously conducted studies [17, 18] were analyzed for PYY concentrations. The experimental diets used differed in their fermentative capacity, causing different absorption of SCFA. We hypothesized that an increased intake of DF and hence an augmented SCFA absorption would increase PYY concentrations.

Materials and methods

Experimental diets

The experimental diets provided all the nutrients, vitamins and minerals required by growing pigs (S1 and S2 Tables). These diets contained varying amounts of DF to either have high or low effects on colonic fermentation and SCFA absorption. The experimental details and the composition of the diets are summarized in the flowchart (S1 Fig).

The diets consisted of a low-DF Western-style diet (WSD) based on white wheat flour, and two high-DF diets, an arabinoxylan-rich diet (AXD) and a resistant-starch rich diet (RSD) [18]

or wheat whole grain (WWG), wheat aleurone-rich flour (WAF) and rye aleurone-rich flour (RAF) [17]. Diets were formulated to be balanced with regard to protein, fat, and metabolizable energy, but to significantly differ in DF content (Table 1). This was successfully achieved in the different diets with the same energy density of the diet (19.24–20.43 MJ), except for the protein content in AXD. This difference was the result of a higher than estimated protein content of the wheat flour incorporated in the RSD and WSD and thus a lower protein content in the AXD [19]. Consumption of AXD resulted in the highest NPA of total SCFA (102 mmol/h), with intermediate levels after RSD consumption (66 mmol/h), and lowest levels in WSD-fed pigs (37 mmol/h, $P_{Diet} = <0.001$) [18] (Reprinted with permission). Consumption of the RAF tended to increase the NPA of SCFA (46 mmol/h), compared to intermediate levels in the WAF-fed pigs (39 mmol/h), and the lowest levels in the WWG-fed pigs (33 mmol/h, $P_{Diet} = 0.07$) [17] (Reprinted with permission). In addition, a parallel study with intact pigs fed the same experimental diets as in experiment 1 of the present study, Nielsen *et al.* [19] found a higher proportion of digesta residues, particularly as starch degradation products, in the distal small intestine 1.5 hours after feeding in response to the RSD compared to the WSD and AXD diets.

Ethics statement

The animal experiments were conducted according to licenses obtained from the Danish Animal Experiments Inspectorate, Ministry of Food, Agriculture and Fisheries, Danish Veterinary

Table 1. Chemical composition of the experimental diets and postprandial net portal appearance of total SCFA after consumption of the experimental diets.

	Experimental diets					
	Experiment 1			Experiment 2		
	WSD	AXD	RSD	WWG	WAF	RAF
DM ^a (g/kg as-fed basis)	915	891	903	654	704	691
Chemical composition (units/kg DM)						
Gross energy, MJ	19.68	19.24	20.30	20.35	20.35	20.43
Ash	37	51	34	43	58	41
Protein (N × 6.26), g	207	154	191	173	177	173
Fat, g	152	135	150	137	134	146
Digestible carbohydrates, g	535	442	473	525	484	522
Sugars, g	113	22	3	19	14	17
Starch, g	422	420	470	506	470	505
Non-digestible carbohydrates, g ^b	72	169	183	116	115	122
Resistant starch, g	6	8	113	11	10	18
Total NSP (soluble), g	58 (11)	144 (33)	55 (8)	105 (26)	105 (15)	104 (39)
Fructans, g	0	22	3	3	2	10
AX (soluble)	18 (6)	72 (22)	15 (4)	61 (17)	62 (9)	50 (19)
Klason lignin, g	6	15	13	22	23	26
Total dietary fiber, g ^c	70	167	181	138	138	148
NPA of SCFA (mmol/h) ^d	37	102	66	33	39	46

Diets were analyzed as described in the Materials and Method section. WSD, Western-style diet; AXD, arabinoxylan-rich diet; RSD, resistant starch-rich diet; WWG, whole-wheat grain; WAF, wheat aleurone flour; RAF, rye aleurone flour.

^a Abbreviations: DM, dry matter; N, nitrogen; NSP, non-starch polysaccharides.

^b Calculated as fructans + resistant starch + total NSP + lignin.

^c Calculated as non-digestible carbohydrates + Klason lignin

^d The statistical significance of diet was $P_{Diet} < 0.001$ in experiment 1, and $P_{Diet} = 0.07$ in experiment 2. Reprinted with permission [17, 18].

<https://doi.org/10.1371/journal.pone.0185927.t001>

and Food Administration. The studies were in compliance with the guidelines concerning animal experiments and care of animals under study according to the Danish Ministry of Justice, Act. 726 of September 9, 1993, and as amended in Act 1306 of November 23, 2007. The health of the animals was monitored throughout the experimental period, and no serious illnesses were observed. All surgery was performed under general anesthesia, and all efforts were made to minimize suffering by providing postsurgical analgesia. After the end of the experiments, the pigs were euthanized with an overdose of pentobarbital, followed by bleeding for postmortem autopsy. Plasma samples were collected from previously conducted animal experiments and evaluated in a novel context [17, 18].

Animals and experimental design

The pigs were obtained from Department of Animal Science, Aarhus University Foulum, Tjele, Denmark. The animals were surgically equipped with catheters in the portal vein and mesenteric artery [17, 18]. The experimental period lasted for three consecutive weeks and was designed as a repeated 3×3 Latin square design. In each experimental week, the pigs were adapted to the experimental diets for 4–6 days before blood samples were collected on days 7, 14, and 21 as described by others [20, 21].

In each experiment six female Danish landrace × Yorkshire pigs (58.8 (SEM 1.6) kg) (experiment 1) or (56.5 (SEM 1.8) kg) (experiment 2) were included.

Experiment 1: Feeding WSD, AXD, and RSD. The WSD supplied 70 g DF per day, and the RSD and AXD supplied 192 and 189 g DF per day, respectively, for a period of 1 week on each diet. Meal portions were weighed from day-to-day and divided into three meals per day, given at 09:00, 14:00, and 19:00 hours supplying 33.3% to mimic a typical human meal pattern. Consecutive blood samples were collected on days 7, 14, and 21 at -15 min (first daily meal fed at 0 min), 15, 30, 45, 60, 90, 120, 180, 240, and 300 min relative to feeding the first daily meal. Plasma SCFA was analyzed at time points 0, 60, 180, and 300 min relative to the first daily meal. Blood samples were collected in Na-heparin vacutainers, and plasma was collected by centrifugation (12 min, 2000g, 4°C).

Experiment 2: Feeding WWG, WAF, and RAF. The animals received one of the three experimental diets on days 4–7, after having been fed with a washout diet for an initial 3 days of the experimental week. The daily ration supplied 210 g DF per day and was weighed from day-to-day. The daily ration was divided into three meals given at 09:00, 14:00, and 19:00 hours providing 40, 40 and 20% of the daily feed allowance to mimic the diurnal variation in the human intake of DF. Consecutive blood samples were collected at days 7, 14, and 21 at 0 min (first daily meal), 30, 60, 120, 180, 240, 300 min (second daily meal), 330, 360, 420, 480, 540, and 600 min relative to feeding the first daily meal. Blood samples were collected in Na-heparin vacutainers, and plasma was collected by centrifugation (12 min, 2000g, 4°C). Plasma samples for arterial and portal SCFA analyses were pooled in 4 pools (pool 1–4) on each sampling day to represent the mean SCFA content in the blood within each pig. One and a half mL arterial and 1.5 mL portal plasma were pooled for arterial and portal SCFA analyses, respectively, from time points 15, 60, and 120 min (pool 1), 180, 240, and 300 min (pool 2), 315, 360, and 420 min (pool 3), and 480, 540, and 600 min (pool 4).

Analytical methods

Portal venous and arterial plasma samples were analyzed for PYY (total) using the MILLIPLEX MAP Mouse Metabolic Hormone Magnetic Bead Panel, Antibody-Immobilized Magnetic PYY beads having 3–36 amino-acid sequences with 100% specificity to the rat, mouse, canine and porcine species (Millipore, Billerica, MA, USA) with an inter-assay CV of 5.6–6.6% and

5–17% and intra-assay CV of 6.0–8.4% and 4–13% in experiments 1 and 2, respectively. Plasma SCFA was determined as previously described [17, 18].

Calculations and statistics

Net portal appearance (NPA) was calculated as described by Rerat *et al.* [22]:

$$NPA = PPF \times (c_{(PV)} - c_{(MA)}) \tag{1}$$

where PPF is the portal plasma flow, $c_{(PV)}$ is the concentration in the portal vein, and $c_{(MA)}$ is the concentration in the mesenteric artery. For the correlations between NPA of PYY and SCFA in experiment 2, an average NPA of PYY was calculated using mean PYY concentrations and mean blood flows at the following time intervals after feeding: time points 30, 60, and 120 min (pool 1); 180, 240, and 300 min (pool 2); 330, 360, and 420 min (pool 3); 480, 540, and 600 min (pool 4).

Effects of diet, time, and their two-factor interactions in experiment 1 were analyzed as repeated measurements using the MIXED procedure of Statistical Analysis Software (SAS, version 9.3, SAS Institute Inc., Cary, NC, USA). Plasma variables were analyzed as a linear mixed model

$$Y_{ijkl} = \mu + \alpha_i + \beta_j + \alpha\beta_{ij} + \gamma_k + \delta_{ikl} + \rho_{ijkl} + \epsilon_{ijkl} \tag{2}$$

where Y_{ijkl} is the dependent variable; μ is the overall mean; α_i is the effect of diet ($i = \text{WSD, RSD, AXD}$); β_j is the time after the first daily meal on day 7 ($j = -15, 15, 30, 45, 60, 120, 180, 240, \text{ and } 300 \text{ min}$); and $\alpha\beta_{ij}$ is the interaction term. The three terms γ_k ($k = \text{pig } 1, 2, 3, 4, 5, 6$), δ_{ikl} ($l = \text{week } 1, 2, 3$), and ρ_{ijkl} accounted for repeated measurements being performed on the same pig (γ_k) each week (δ_{ikl}) and on the same pig within a sampling period (ρ_{ijkl}), while ϵ_{ijkl} describes the residual error component. Fasting concentrations were calculated without effects of time and diet-time interactions. The covariance structure of ρ_{ijkl} was modelled using the spatial power option, which takes into account the different intervals between repeated measurements. The random effects and residuals are assumed to be normally distributed and independent and their expectations were assumed to be zero.

Effects of the first and second meal in experiment 2 were analyzed as repeated measurements using the MIXED procedure of Statistical Analysis Software (version 9.3, SAS Institute Inc., Cary, NC, USA). Plasma variables were analyzed as a linear mixed model

$$Y_{ijklmn} = \mu + \alpha_i + \beta_j + \gamma_k + \alpha\beta_{ij} + \alpha\gamma_{ik} + \beta\gamma_{jk} + \alpha\beta\gamma_{ijm} + \delta_l + \rho_m + \vartheta_{ilm} + \tau_{ilmn} + \epsilon_{ijklm} \tag{3}$$

where Y_{ijklmn} is the dependent variable; μ is the overall mean; α_i is the effect of diet ($i = \text{WWG, WAF, or RAF}$); β_j is the effect of meal ($j = 1 \text{ or } 2$), and γ_k is the time after a meal ($k = 0, 30, 60, 120, 180, 240 \text{ and } 300 \text{ min}$), and δ_l is the effect of week ($l = 1, 2, \text{ or } 3$). The four terms $\alpha\beta_{ij}$, $\alpha\gamma_{ik}$, $\beta\gamma_{jk}$, and $\alpha\beta\gamma_{ijk}$ accounted for the interactions between the main effects. The covariance structure ρ_m ($m = \text{pig}$) accounted for repeated measurements being performed on the same pig fed different diets each week (ϑ_{ilm}) and on the same pig within a sampling period (τ_{ilmn}), and was modelled using the spatial power option, which takes into account the adverse time intervals between repeated measurements. The ϵ_{ijklm} is the residual error component under the assumptions that the random effects and residuals are normally distributed and independent, and their expectations were assumed to be zero.

The hormone concentrations were transformed to $\ln(x)$ before statistical analysis to obtain variance homogeneity and presented as least square means with 95% confidence intervals. The NPA was calculated on original scale and presented as least square means with standard error (SE). Pearson correlations were evaluated using the PROC CORR procedure of SAS (version

9.3, SAS Institute Inc., Cary, NC, USA). The significance level was set at $P < 0.05$ while $P \leq 0.10$ was reported being a tendency.

Results

Net portal appearance of PYY and plasma concentrations

Experiment 1: Effect of WSD, AXD, and RSD. The 5 h postprandial NPA of PYY was 37–54% higher in response to RSD consumption compared to AXD and WSD intake, although this difference was not statistically significant ($P_{Diet} = 0.19$; Table 2). Pigs fed RSD secreted 2.07 nmol/h, compared to 1.34 nmol/h and 1.51 nmol/h in WSD and AXD fed pigs, respectively (Table 2). The PYY secretion increased postprandially on all three diets ($P_{Time} = 0.03$; Fig 1) followed by a decrease. The NPA of PYY peaked at 3.4–3.5 nmol/h 15 and 45 min after feeding in RSD and WSD-fed pigs, respectively, and at 3.2 nmol/h after 60 min in response to AXD intake (Fig 1). Fasting and postprandial mesenteric artery and portal vein PYY concentrations were higher in response to RSD compared to WSD and AXD ($P_{Diet} < 0.01$; Table 2; S2 Fig). All experimental diets resulted in a rapid increase in mesenteric arterial and portal vein PYY concentrations in response to a meal, followed by a gradual decrease over the next 4 hours ($P_{Time} < 0.001$; S2 Fig). Arterial PYY concentrations tended to be higher in response to RSD compared to WSD and AXD throughout the sampling period ($P_{Diet \times Time} = 0.053$). Mesenteric arterial PYY concentrations peaked at 220 and 183 pM 60 min after feeding RSD and WSD, respectively, as compared with 156 pM 45 min after feeding AXD. Portal vein PYY concentrations in response to RSD and AXD peaked 60 min after feeding at 250 and 169 pM, respectively, compared to 203 pM 30 min after feeding WSD.

Experiment 2: Effect of WWG, WAF, and RAF. Overall, the NPA of PYY was not affected by diet, meal, time, or any interactions hereof ($P > 0.05$; Table 3). The NPA of PYY displayed an inclination towards a postprandial increase after the first daily meal (Fig 2A), although not statistically significant ($P_{Diet \times Time \times Meal} = 0.34$; Table 3). The NPA peaked at 1.1–1.5 nmol/h 60–120 min after feeding, followed by a decrease. The second daily meal caused a secondary increase, although less prominent as compared to the first response (Fig 2A). The average NPA of PYY for all three diets suggested a more distinct postprandial increase followed by a decrease in response to the first meal compared to the second meal which appeared more fluctuating (Fig 2B), although not statistically significant ($P_{Meal} = 0.33$). Postprandial PYY concentrations in the mesenteric artery and portal vein were not different between the three experimental diets ($P_{Diet} > 0.1$; Table 3). Mesenteric artery and portal vein PYY concentrations at day 7 following the first daily meal showed a fast increase the first 60 min postprandial after which a plateau phase was reached until the second daily meal 5 hours/300 min later (S3 Fig). The second daily meal caused a drop in PYY concentrations at 330 min with a subsequent rise at 360 min ($P_{Meal \times Time} < 0.001$). Plasma PYY concentrations reached a higher postprandial plateau after the second daily meal (420 min) as compared with the plateau after the first daily meal (120 min). Overall, PYY concentrations were higher after the second meal compared with the first meal ($P_{Meal} > 0.001$; Table 3). Postprandial mesenteric arterial concentrations increased by 12–34% (from 129–137 pM to 154–163 pM) after the first daily meal to the second meal, and portal vein concentrations increased by 9–25% (from 135–142 pM to 155–169 pM) after the first to second meal.

Correlations between SCFA absorption and PYY secretion

Pearson correlations were used to assess whether there were correlations between NPA of PYY and NPA of total SCFA (Fig 3) for the individual time points. No significant correlations ($P > 0.05$) were found between NPA of PYY and NPA of total or individual SCFA (acetate,

Table 2. Postprandial net portal PYY appearance and mesenteric artery and portal vein PYY concentrations of pigs fed WSD, AXD, and RSD.

		Experimental diets			SE	P-values		
		WSD	AXD	RSD		Diet	Time	Diet×Time
Net portal appearance (nmol/h)		1.34	1.51	2.07	0.38	0.19	0.03	0.14
Arterial PYY (pM)	Fasting	105 ^b	120 ^b	146 ^a	14	0.004	-	-
	Postprandial	140 [114; 172] ^b	135 [110; 165] ^b	182 [148; 223] ^a	-	< 0.001	< 0.001	0.053
Portal PYY (pM)	Fasting	124 ^b	113 ^b	173 ^a	12	0.001	-	-
	Postprandial	157 [130; 189] ^b	147 [122; 177] ^b	203 [169; 245] ^a	-	< 0.001	< 0.001	0.55

Postprandial net portal appearance of PYY and PYY concentrations in the mesenteric artery and portal vein of pigs fed the experimental diets in response to first the daily meal. Fasting concentrations and net portal appearance are means of time points relative to the first daily feeding (-15, 15, 30, 45, 60, 120, 180, 240, and 300 min.) with standard error (SE), and postprandial concentrations are least square means [95% cis], n = 6.

^{a,b}Means within a row without common superscript differ ($P < 0.05$).

WSD, Western-style diet; AXD, arabinoxylan-rich whole-grain diet; RSD, resistant starch-rich diet.

<https://doi.org/10.1371/journal.pone.0185927.t002>

propionate, butyrate; $P > 0.05$), as shown in Fig 3 and S3 Table. A Pearson correlation comparing the NPA of SCFA against the NPA of PYY for RSD and WSD fed pigs from experiment 1 specifically, did not reveal significant correlations ($P = 0.14$; S4 Fig).

Discussion

Effects of SCFA on PYY levels and secretion

In the present study, we found no significant correlation between SCFA absorption to the portal vein and PYY secretion in pigs in spite of a significant higher SCFA absorption obtained with AXD and RSD in comparison with WSD in experiment 1. Consumption of RSD resulted in 35% and 38% higher arterial and portal vein PYY concentrations, respectively, compared to PYY levels found in AXD fed pigs. In addition, we found similar PYY levels in portal and

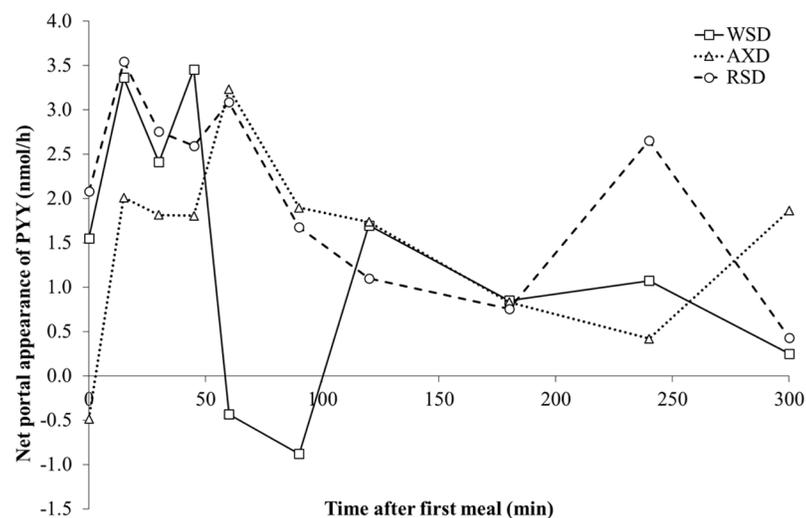


Fig 1. Net portal appearance of PYY in pigs fed WSD, AXD, and RSD. Net portal appearance of PYY following the first daily meal (0 min). Values are means, n = 6; $P_{Diet} = 0.19$, $P_{Time} = 0.03$, $P_{Diet \times Time} = 0.14$). The solid lines represent means from pigs fed the Western-style diet (WSD); dotted lines represent means from pigs fed the arabinoxylan-rich whole-grain diet (AXD); the dashed line represent means from pigs fed the resistant starch-rich diet (RSD).

<https://doi.org/10.1371/journal.pone.0185927.g001>

Table 3. Postprandial net portal appearance of PYY and PYY concentrations in the mesenteric artery and portal vein of pigs fed of pigs fed WWG, WAF, and RAF¹.

	Experimental diets			P-values						
	WWG	WAF	RAF	Diet	Time	Meal	Meal×Time	Diet×Meal	Diet×Time	Diet×Time×Meal
Net portal appearance (nmol/h)	184	561	440	0.21	0.11	0.33	0.17	0.96	0.87	0.34
1. meal	298	641	494							
2. meal	70	480	387							
Arterial PYY (pM)				0.87	< 0.001	< 0.001	< 0.001	0.99	0.97	0.97
1. meal	129 [107;154]	130 [108;157]	137 [114;164]							
2. meal	154 [128;184]	155 [129;187]	163 [136;196]							
Portal PYY (pM)				0.76	< 0.001	< 0.001	< 0.001	0.89	0.91	0.84
1. meal	135 [115;157]	136 [116;160]	142 [122;167]							
2. meal	155 [133;181]	159 [135;187]	169 [144;198]							

¹ Postprandial net portal appearance and concentrations are means of time points relative to the first daily feeding (meal 1; 0, 30, 60, 120, 180, 240, 300 min (meal 2) 330, 360, 420, 480, 540, and 600 min), n = 6. Arterial and portal concentrations are given with 95% cis, and net portal appearance with a standard error of 157. WWG, whole-wheat grain; WAF, wheat aleurone flour; RAF, rye aleurone flour.

<https://doi.org/10.1371/journal.pone.0185927.t003>

arterial plasma, indicating a low hepatic clearance of PYY. Our findings are in accordance with previous observations using *in situ* perfusion of rat liver [23], demonstrating that from portal infusion of up to 500 pM, only up to 10% was extracted by the liver. Increasing the intake of fermentable carbohydrates was previously linked to increased colonic SCFA production and increased endogenous PYY secretions in murine models [24, 25]. The DF level in the RSD diet was increased using high-amylose maize and raw potato starch, which was shown to increase PYY concentrations in rodents [25, 26]. In addition, a 3-day human intervention study by Nilsson *et al.* [27] found that ingestion of barley-kernel based bread increased serum SCFA and significantly elevated the plasma PYY levels. The same results were reported by Sandberg *et al.* [28] who demonstrated that whole grain rye kernel bread given as a late evening meal increased both fasting and postprandial plasma concentrations of PYY, glucagon-like peptide 1 (GLP-1), and fasting SCFA the following morning compared to white wheat flour based bread. Brown *et al.* [29] suggested that the PYY increase is mediated by increased colonic SCFA, activating the G-protein-coupled free fatty acid receptors FFAR3 and FFAR2. In the present study, however, postprandial PYY concentrations did not correlate with SCFA levels, as total and individual SCFA concentrations and absorption were lowest in response to WSD, intermediate with RSD, and highest when AXD was fed [18]. We previously found that the changes in SCFA absorption correlated to changes in the fecal microbial composition, particularly in response to the AXD, while the microbial composition did not differ significantly in the RSD compared to either the WSD or the AXD diets [19]. Similarly, despite comparable levels of DF in the WWG, WAF, and RAF based diets, SCFA plasma concentration and absorption were lowest in pigs fed WWG, intermediate in WAF, and highest in response to the rye-based RAF diets [17]. In line with this, we previously found no correlation between the increased SCFA production and colonic FFAR2/FFAR3 gene expression, but discovered an inverse correlation between FFAR3 and FFAR2 gene expression and cecal SCFA/butyrate pool sizes [30]. This apparent discrepancy between increased PYY secretion and lack of effect on FFAR3 gene expression was also recently reported in rats [31]. Adam *et al.* [31] suggested that

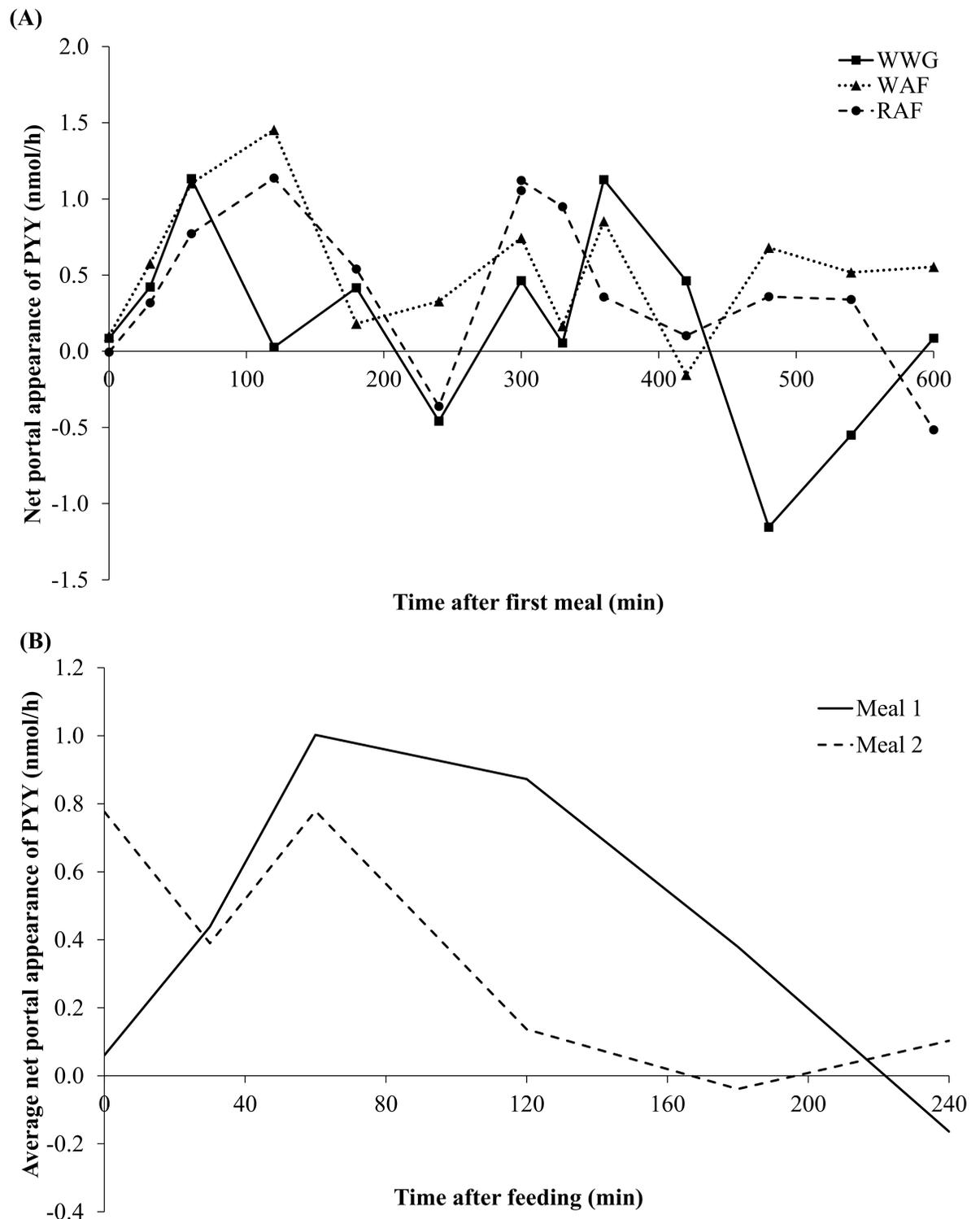


Fig 2. Net portal appearance of PYY following the first (0 min) and second (300 min) daily meals in pigs fed WWG, WAF, and RAF. (A) Net portal appearance of PYY after the first (0 min) and second (300 min) daily meal ($P_{Diet} = 0.21$, $P_{Meal} = 0.33$, $P_{Time} = 0.11$, $P_{Meal \times Time} = 0.87$, $P_{Diet \times Meal} = 0.96$, $P_{Diet \times Time} = 0.87$, $P_{Diet \times Time \times Meal} = 0.34$). Values are means, $n = 6$. The solid lines represent means from pigs fed the whole-wheat grain diet (WWG); dotted lines represent means from pigs fed the wheat aleurone flour diet (WAF); dashed lines represent means from pigs fed the rye aleurone flour diet (RAF). (B) Average net portal appearance of PYY for the three experimental diets following the first (solid line) and second daily meal (dashed line).

<https://doi.org/10.1371/journal.pone.0185927.g002>

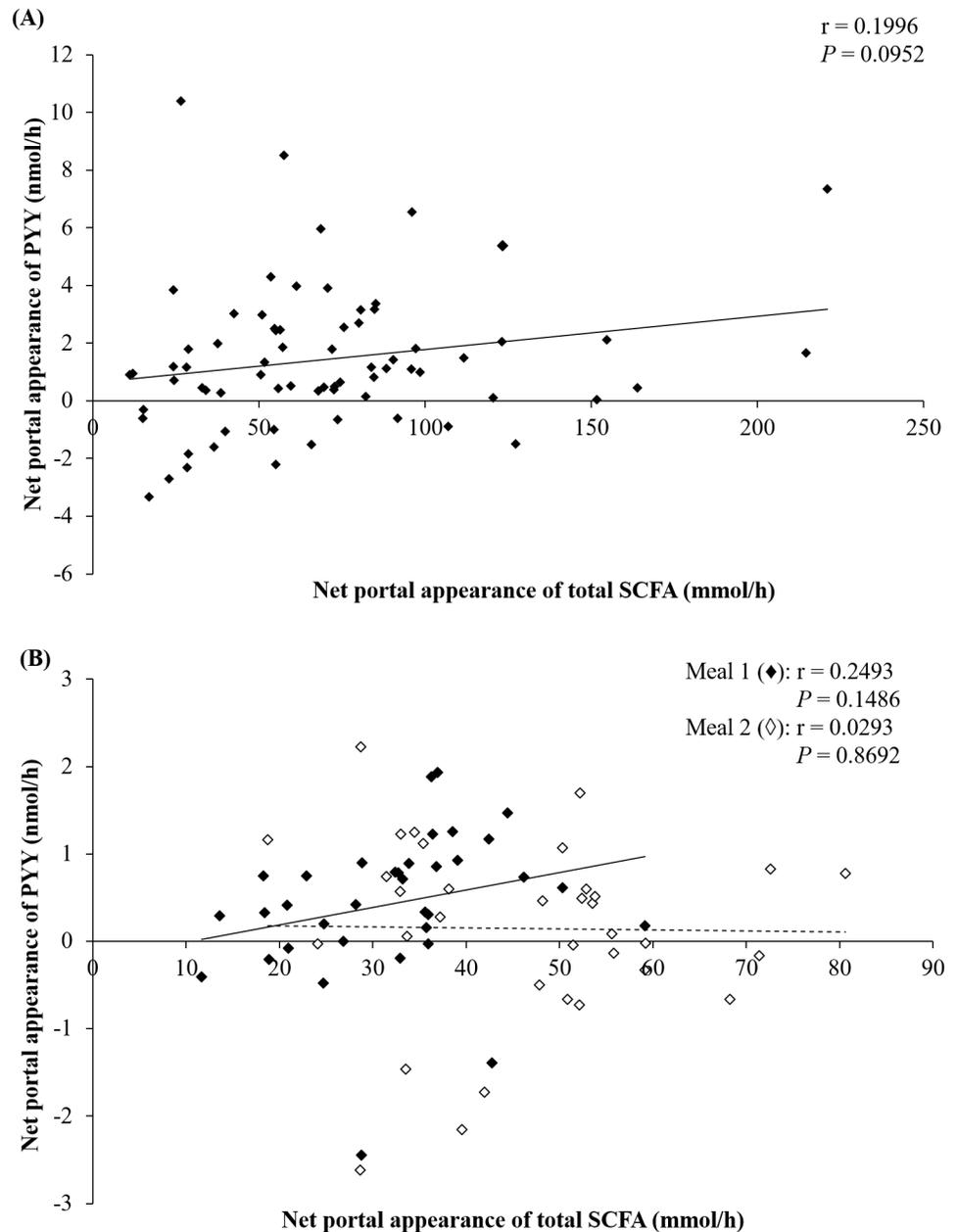


Fig 3. Correlations between net portal appearance of PYY and NPA of total SCFA in pigs fed WSD, AXD, and RSD. (A) Correlation between net portal appearance of PYY and SCFA for each pig after consumption of WSD, AXD, or RSD, $n = 6$. Each point represents a correlation between net portal appearance of PYY and SCFA for a given pig on a specific diet (time points 0, 60, 180, and 300 min relative to first daily meal). (B) Correlation with first (\blacklozenge) and second (\diamond) meals after consumption of WWG, WAF, or RAF, $n = 6$. Each point represents the average net portal appearance of PYY (pools 1–4 as described in the Materials and Methods) correlated to the net portal appearance of SCFA (pools 1–4) for each pig on each experimental diet. Inserts show the Pearson's r correlation and the corresponding P -value. Solid line represent means for meal 1, and the dashed line represent means for meal 2.

<https://doi.org/10.1371/journal.pone.0185927.g003>

an increased secretory capacity or an increased number of colonic L-cells was responsible for the increased PYY secretion [31]. Indeed, others have reported increased L-cell number and differentiation in rodents in response to standard diets supplemented with fermentable carbohydrates (oligofructose and fructo-oligosaccharides) [32, 33]. The results presented here thus support previous findings suggesting that increased levels of circulating PYY may be a result of a FFAR3/FFAR2 independent stimulation [16].

The *in vivo* effects of DF and SCFA supplementation on PYY secretion have generated ambiguous results and appear to be related to species differences and animal models used. Production and secretion of PYY are commonly considered to be mediated by L-cells in the distal part of the small intestine and the colon, along with GLP-1 [34]. Recent studies in mice and rats have shown a differential expression of these hormones in the different segments of the intestine with expression of GLP-1 secretion from L-cells dominating in the distal small intestine while both GLP-1 and PYY are expressed in the colon [35–37]. The anatomical distribution of PYY secreting L-cells along the intestinal tract differs between mice, pigs, and humans, especially in the proximal intestine [38, 39]. Interestingly, Cho *et al.* [37] showed that the small intestinal-colon gradient in the expression of PYY was less pronounced in pigs compared to mice, showing that significant inter-species differences exist which may account for the observed differences in PYY responses between rodent and porcine models, and thus possibly humans. Indeed, several rodent studies have shown increased PYY levels after DF consumption [25, 40–42], whereas porcine studies are limited and have produced equivocal results so far [11, 43, 44].

Effects of digesta residues on PYY absorption

The release of PYY into circulation occurs in response to food intake, and reaches plateau levels within 1–2 hours postprandially followed by a plateau for up to 5 hours in healthy human subjects [45]. This initial release occurs before food enters the distal parts of the gut and it has therefore been suggested that the initial release is neurally mediated in dogs and pigs [46, 47]. The continuous PYY production after an initial increase in plasma PYY levels suggests, however, that the PYY production is related to the presence of digesta residues in the distal small intestine and proximal large intestine acting on ileal and colonic endocrine L-cells. Furthermore, the observation described in the parallel study by Nielsen *et al.* [19], showing a higher proportion of small intestinal digesta residues and starch degradation products present in the small intestines 1.5 hours after feeding in response to the RSD compared to the WSD and AXD, indicate that the effects on postprandial PYY responses reported here are primarily due to neural regulation and ileal digesta residues, rather than SCFA production and absorption. This assumption is supported by the results from experiment 2 where the average second meal PYY increments were significantly higher compared to the first meal. In addition, a parallel study to experiment 2 using ileal cannulated pigs showed an increased flow of digesta in response to the first daily meal compared to the second [48], suggesting that after an overnight fast lasting 14 hours, the small intestine is relatively empty because the small previous evening meal (only 20% of the daily load), has passed through the gastrointestinal tract. Thus, the higher PYY concentrations in response to the second meal could be a result of a cumulative effect of food present in the small intestine.

In conclusion, the results of the present study suggest that in pigs fed diets varying in DF with comparable digestibility in the small intestine and variable SCFA and butyrate NPA total DF intake and SCFA production *per se* were not responsible for the increased PYY secretion. The increased flow of intestinal digesta residues in combination with neural regulation may be responsible for the increased PYY response in pigs.

Supporting information

S1 Fig. Flowchart of experimental design and diets. Experimental design and diets. Flowchart showing the example of repeated 3×3 Latin square design in experiments. Six pigs completed each experiment. WSD, Western-style diet; AXD, arabinoxylan-rich diet; RSD, resistant starch-rich diet; WFL, white wheat flour (washout diet); WWG, whole-wheat grain; WAF, wheat aleurone flour; RAF, rye aleurone flour.
(TIF)

S2 Fig. PYY concentrations following the first daily meal in pigs fed WSD, AXD, and RSD. PYY concentrations following the first daily meal (0 min) in pigs fed WSD, AXD, and RSD. A) Mesenteric artery PYY concentrations ($P_{Diet} < 0.001$, $P_{Time} < 0.001$, $P_{Diet \times Time} = 0.0531$). B) Portal vein PYY concentrations ($P_{Diet} < 0.001$, $P_{Time} < 0.001$, $P_{Diet \times Time} = 0.55$). Data was ln-transformed before statistical analysis to obtain variance homogeneity, and back transformed to original scale after statistical analyses. Values are means, $n = 6$. WSD, Western-style diet; AXD, arabinoxylan-rich whole-grain diet; RSD, resistant starch-rich diet.
(TIFF)

S3 Fig. PYY concentrations following the first daily meal in pigs fed WWG, WAF, and RAF. PYY concentrations following the first daily meal (0 min) in pigs fed WWG, WAF, and RAF. Data was ln-transformed before statistical analysis to obtain variance homogeneity, and back transformed to original scale after statistical analyses. A) Mesenteric arterial PYY concentrations at day 7 following the first daily meal (0 min) and second daily meal (300 min). Values are means, $n = 6$. $P_{Diet} = 0.87$, $P_{Time} < 0.001$, $P_{Meal} < 0.001$, $P_{Meal \times Time} < 0.001$, $P_{Diet \times Meal} = 0.99$, $P_{Diet \times Time} = 0.97$, $P_{Diet \times Time \times Meal} = 0.97$. B) Portal vein PYY concentrations at day 7 following the first daily meal (0 min) and second daily meal (300 min). $P_{Diet} = 0.76$, $P_{Time} < 0.001$, $P_{Meal} < 0.001$, $P_{Meal \times Time} < 0.001$, $P_{Diet \times Meal} = 0.89$, $P_{Diet \times Time} = 0.91$, $P_{Diet \times Time \times Meal} = 0.84$. WWG, whole-wheat grain; WAF, wheat aleurone flour; RAF, rye aleurone flour.
(TIFF)

S4 Fig. Correlations between net portal appearance of PYY and net portal appearance of total SCFA in pigs fed the RSD and WSD diets. Correlations from experiment 1 between net portal appearance of PYY and net portal appearance of total SCFA in pigs fed the RSD and WSD diets, $n = 6$. Each point represents a correlation between net portal appearance of PYY and SCFA for a given pig on a specific diet (time points 0, 60, 180, and 300 min relative to first daily meal).
(TIF)

S1 Table. Ingredients list of experimental diets used in experiment 1.
(DOCX)

S2 Table. Ingredients list of experimental diets used in experiment 2.
(DOCX)

S3 Table. Correlations between net portal appearance of PYY and total or individual short-chain fatty acids in experiments 1 and 2. Correlations between net portal appearance of PYY (nmol/h) and total or individual short-chain fatty acids (SCFA; mmol/h) in experiments 1 and 2.
(DOCX)

Acknowledgments

The authors thank the staff at the animal facility of the Department of Animal Science for valuable technical assistance.

Author Contributions

Conceptualization: Anne Krog Ingerslev, Knud Erik Bach Knudsen.

Data curation: Anne Krog Ingerslev, Kirstine Lykke Nielsen.

Formal analysis: Anne Krog Ingerslev.

Funding acquisition: Knud Erik Bach Knudsen.

Investigation: Anne Krog Ingerslev, Shivaprakash Jagalur Mutt, Helle Nygaard Lærke, Mette Skou Hedemann, Peter Kappel Theil, Kirstine Lykke Nielsen.

Methodology: Helle Nygaard Lærke, Mette Skou Hedemann, Peter Kappel Theil, Kirstine Lykke Nielsen, Henry Jørgensen, Knud Erik Bach Knudsen.

Project administration: Anne Krog Ingerslev, Helle Nygaard Lærke, Mette Skou Hedemann, Peter Kappel Theil, Kirstine Lykke Nielsen, Henry Jørgensen.

Resources: Anne Krog Ingerslev, Shivaprakash Jagalur Mutt, Kirstine Lykke Nielsen, Karl-Heinz Herzig, Knud Erik Bach Knudsen.

Software: Anne Krog Ingerslev.

Supervision: Knud Erik Bach Knudsen.

Visualization: Anne Krog Ingerslev.

Writing – original draft: Anne Krog Ingerslev, Kirstine Lykke Nielsen.

Writing – review & editing: Anne Krog Ingerslev, Karl-Heinz Herzig, Knud Erik Bach Knudsen.

References

1. Murphy KG, Bloom SR. Gut hormones and the regulation of energy homeostasis. *Nature*. 2006; 444(7121):854–9. Epub 2006/12/15. <https://doi.org/10.1038/nature05484> PMID: 17167473.
2. Karhunen LJ, Juvonen KR, Huotari A, Purhonen AK, Herzig KH. Effect of protein, fat, carbohydrate and fibre on gastrointestinal peptide release in humans. *Regul Pept*. 2008; 149(1–3):70–8. <https://doi.org/10.1016/j.regpep.2007.10.008> PMID: 18456350
3. Boey D, Sainsbury A, Herzog H. The role of peptide YY in regulating glucose homeostasis. *Peptides*. 2007; 28(2):390–5. Epub 2007/01/11. <https://doi.org/10.1016/j.peptides.2006.07.031> PMID: 17210210.
4. Slavin J, Green H. Dietary fibre and satiety. *Nutrition Bulletin*. 2007; 32:32–42. <https://doi.org/10.1111/j.1467-3010.2007.00603.x>
5. Juvonen KR, Purhonen AK, Salmenkallio-Marttila M, Lahteenmaki L, Laaksonen DE, Herzig KH, et al. Viscosity of oat bran-enriched beverages influences gastrointestinal hormonal responses in healthy humans. *J Nutr*. 2009; 139(3):461–6. Epub 2009/01/30. <https://doi.org/10.3945/jn.108.099945> PMID: 19176745.
6. Smith CE, Tucker KL. Health benefits of cereal fibre: a review of clinical trials. *Nutr Res Rev*. 2011; 24(1):118–31. <https://doi.org/10.1017/S0954422411000023> PMID: 21320383
7. Gemen R, de Vries JF, Slavin JL. Relationship between molecular structure of cereal dietary fiber and health effects: focus on glucose/insulin response and gut health. *Nutr Rev*. 2011; 69(1):22–33. <https://doi.org/10.1111/j.1753-4887.2010.00357.x> PMID: 21198632
8. Karhunen LJ, Juvonen KR, Flander SM, Liukkonen KH, Lahteenmaki L, Siloaho M, et al. A psyllium fiber-enriched meal strongly attenuates postprandial gastrointestinal peptide release in healthy young adults. *J Nutr*. 2010; 140(4):737–44. Epub 2010/02/12. <https://doi.org/10.3945/jn.109.115436> PMID: 20147463.

9. Juvonen KR, Salmenkallio-Marttila M, Lyly M, Liukkonen KH, Lahteenmaki L, Laaksonen DE, et al. Semisolid meal enriched in oat bran decreases plasma glucose and insulin levels, but does not change gastrointestinal peptide responses or short-term appetite in healthy subjects. *Nutr Metab Cardiovasc Dis.* 2011; 21(9):748–56. Epub 2010/07/08. <https://doi.org/10.1016/j.numecd.2010.02.002> PMID: 20605427.
10. Cherbut C, Ferrier L, Roze C, Anini Y, Blottiere H, Lecannu G, et al. Short-chain fatty acids modify colonic motility through nerves and polypeptide YY release in the rat. *Am J Physiol.* 1998; 275(6 Pt 1): G1415–22. Epub 1998/12/09. PMID: 9843779.
11. Cuhe G, Cuber JC, Malbert CH. Ileal short-chain fatty acids inhibit gastric motility by a humoral pathway. *Am J Physiol Gastrointest Liver Physiol.* 2000; 279(5):G925–30. Epub 2000/10/29. PMID: 11052989.
12. Freeland KR, Wolever TM. Acute effects of intravenous and rectal acetate on glucagon-like peptide-1, peptide YY, ghrelin, adiponectin and tumour necrosis factor-alpha. *Br J Nutr.* 2010; 103(3):460–6. Epub 2009/10/13. <https://doi.org/10.1017/S0007114509991863> PMID: 19818198.
13. Brouns F, Kettlitz B, Arrighoni E. Resistant starch and "the butyrate revolution". *Trends Food Sci Technol.* 2002; 13(8):251–61. [https://doi.org/10.1016/s0924-2244\(02\)00131-0](https://doi.org/10.1016/s0924-2244(02)00131-0)
14. Hamer HM, Jonkers D, Venema K, Vanhoutvin S, Troost FJ, Brummer RJ. Review article: the role of butyrate on colonic function. *Aliment Pharmacol Ther.* 2008; 27(2):104–19. <https://doi.org/10.1111/j.1365-2036.2007.03562.x> PMID: 17973645
15. Leonel AJ, Alvarez-Leite JI. Butyrate: implications for intestinal function. *Curr Opin Clin Nutr Metab Care.* 2012; 15(5):474–9. Epub 2012/07/17. <https://doi.org/10.1097/MCO.0b013e32835665fa> PMID: 22797568.
16. Lin HV, Frassetto A, Kowalik EJ Jr., Nawrocki AR, Lu MM, Kosinski JR, et al. Butyrate and propionate protect against diet-induced obesity and regulate gut hormones via free fatty acid receptor 3-independent mechanisms. *PLoS ONE.* 2012; 7(4):e35240. Epub 2012/04/17. <https://doi.org/10.1371/journal.pone.0035240> PMID: 22506074; PMCPMC3323649.
17. Theil PK, Jorgensen H, Serena A, Hendrickson J, Bach Knudsen KE. Products deriving from microbial fermentation are linked to insulinaemic response in pigs fed breads prepared from whole-wheat grain and wheat and rye ingredients. *Br J Nutr.* 2011; 105(3):373–83. <https://doi.org/10.1017/S0007114510003715> PMID: 20923581
18. Ingerslev AK, Theil PK, Hedemann MS, Laerke HN, Bach Knudsen KE. Resistant starch and arabinoxylan augment SCFA absorption, but affect postprandial glucose and insulin responses differently. *Br J Nutr.* 2014; 111(9):1564–76. Epub 2014/02/11. <https://doi.org/10.1017/S0007114513004066> PMID: 24507768.
19. Nielsen TS, Laerke HN, Theil PK, Sorensen JF, Saarinen M, Forssten S, et al. Diets high in resistant starch and arabinoxylan modulate digestion processes and SCFA pool size in the large intestine and faecal microbial composition in pigs. *Br J Nutr.* 2014; 112(11):1837–49. Epub 2014/10/21. <https://doi.org/10.1017/S000711451400302X> PMID: 25327182.
20. Hooda S, Matte JJ, Vasanthan T, Zijlstra RT. Dietary Oat beta-Glucan Reduces Peak Net Glucose Flux and Insulin Production and Modulates Plasma Incretin in Portal-Vein Catheterized Grower Pigs. *J Nutr.* 2010; 140(9):1564–9. <https://doi.org/10.3945/jn.110.122721> PMID: 20660287
21. Regmi PR, van Kempen T, Matte JJ, Zijlstra RT. Starch with High Amylose and Low in Vitro Digestibility Increases Short-Chain Fatty Acid Absorption, Reduces Peak Insulin Secretion, and Modulates Incretin Secretion in Pigs. *J Nutr.* 2011; 141(3):398–405. <https://doi.org/10.3945/jn.110.132449> PMID: 21248198
22. Rerat AA, Vaissade P, Vaugelade P. Absorption kinetics of some carbohydrates in conscious pigs .2. Quantitative aspects. *Br J Nutr.* 1984; 51(3):517–29. <https://doi.org/10.1079/bjn19840057> PMID: 6722092
23. Beckh K, Monnikes H, Loos S, Arnold R, Koop H. Low hepatic clearance of peptide YY (PYY) in the perfused rat liver. *Regul Pept.* 1992; 37(3):205–12. Epub 1992/02/18. [https://doi.org/10.1016/0167-0115\(92\)90615-2](https://doi.org/10.1016/0167-0115(92)90615-2) PMID: 1557512.
24. Zhou J, Hegsted M, McCutcheon KL, Keenan MJ, Xi X, Raggio AM, et al. Peptide YY and proglucagon mRNA expression patterns and regulation in the gut. *Obesity (Silver Spring, Md).* 2006; 14(4):683–9. Epub 2006/06/03. <https://doi.org/10.1038/oby.2006.77> PMID: 16741270.
25. Zhou J, Martin RJ, Tulley RT, Raggio AM, McCutcheon KL, Shen L, et al. Dietary resistant starch upregulates total GLP-1 and PYY in a sustained day-long manner through fermentation in rodents. *Am J Physiol Endocrinol Metab.* 2008; 295(5):E1160–E6. <https://doi.org/10.1152/ajpendo.90637.2008> PMID: 18796545
26. Keenan MJ, Martin RJ, Raggio AM, McCutcheon KL, Brown IL, Birkett A, et al. High-amylose resistant starch increases hormones and improves structure and function of the gastrointestinal tract: a

- microarray study. *Journal of nutrigenetics and nutrigenomics*. 2012; 5(1):26–44. Epub 2012/04/21. <https://doi.org/10.1159/000335319> PMID: 22516953; PMCPCmc4030412.
27. Nilsson AC, Johansson-Boll EV, Björck IME. Increased gut hormones and insulin sensitivity index following a 3-d intervention with a barley kernel-based product: a randomised cross-over study in healthy middle-aged subjects. *Br J Nutr*. 2015; FirstView:1–9. <https://doi.org/10.1017/S0007114515002524> PMID: 26259632
 28. Sandberg JC, Björck IM, Nilsson AC. Rye-Based Evening Meals Favorably Affected Glucose Regulation and Appetite Variables at the Following Breakfast; A Randomized Controlled Study in Healthy Subjects. *PLoS ONE*. 2016; 11(3):e0151985. Epub 2016/03/19. <https://doi.org/10.1371/journal.pone.0151985> PMID: 26990559; PMCPCMC4798690.
 29. Brown AJ, Goldsworthy SM, Barnes AA, Eilert MM, Tcheang L, Daniels D, et al. The orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. *J Biol Chem*. 2003; 278(13):11312–9. <https://doi.org/10.1074/jbc.M211609200> PMID: 12496283
 30. Nielsen TS, Theil P, Purup S, Nørskov N, Bach Knudsen KE. Effects of Resistant Starch and Arabinoxylan on Parameters Related to Large Intestinal and Metabolic Health in Pigs Fed Fat Rich Diets. *J Agric Food Chem*. 2015. <https://doi.org/10.1021/acs.jafc.5b03372> PMID: 26566722
 31. Adam CL, Williams PA, Dalby MJ, Garden K, Thomson LM, Richardson AJ, et al. Different types of soluble fermentable dietary fibre decrease food intake, body weight gain and adiposity in young adult male rats. *Nutr Metab (Lond)*. 2014; 11:36. Epub 2014/08/26. <https://doi.org/10.1186/1743-7075-11-36> PMID: 25152765; PMCPCmc4141268.
 32. Cani PD, Hoste S, Guiot Y, Delzenne NM. Dietary non-digestible carbohydrates promote L-cell differentiation in the proximal colon of rats. *Br J Nutr*. 2007; 98(1):32–7. Epub 2007/03/21. <https://doi.org/10.1017/S0007114507691648> PMID: 17367575.
 33. Kaji I, Karaki S, Tanaka R, Kuwahara A. Density distribution of free fatty acid receptor 2 (FFA2)-expressing and GLP-1-producing enteroendocrine L cells in human and rat lower intestine, and increased cell numbers after ingestion of fructo-oligosaccharide. *J Mol Histol*. 2011; 42(1):27–38. Epub 2010/11/30. <https://doi.org/10.1007/s10735-010-9304-4> PMID: 21113792.
 34. Mortensen K, Christensen LL, Holst JJ, Orskov C. GLP-1 and GIP are colocalized in a subset of endocrine cells in the small intestine. *Regul Pept*. 2003; 114(2–3):189–96. Epub 2003/07/02. [https://doi.org/10.1016/S0167-0115\(03\)00125-3](https://doi.org/10.1016/S0167-0115(03)00125-3) PMID: 12832109.
 35. Habib AM, Richards P, Cairns LS, Rogers GJ, Bannon CA, Parker HE, et al. Overlap of endocrine hormone expression in the mouse intestine revealed by transcriptional profiling and flow cytometry. *Endocrinology*. 2012; 153(7):3054–65. Epub 2012/06/12. <https://doi.org/10.1210/en.2011-2170> PMID: 22685263; PMCPCmc3440453.
 36. Svendsen B, Pedersen J, Albrechtsen NJ, Hartmann B, Torang S, Rehfeld JF, et al. An analysis of cosecretion and coexpression of gut hormones from male rat proximal and distal small intestine. *Endocrinology*. 2015; 156(3):847–57. Epub 2014/12/24. <https://doi.org/10.1210/en.2014-1710> PMID: 25535831.
 37. Cho HJ, Kosari S, Hunne B, Callaghan B, Rivera LR, Bravo DM, et al. Differences in hormone localisation patterns of K and L type enteroendocrine cells in the mouse and pig small intestine and colon. *Cell Tissue Res*. 2015; 359(2):693–8. Epub 2014/11/08. <https://doi.org/10.1007/s00441-014-2033-3> PMID: 25378285.
 38. van der Wielen N, van Avesaat M, de Wit NJ, Vogels JT, Troost F, Masclee A, et al. Cross-species comparison of genes related to nutrient sensing mechanisms expressed along the intestine. *PLoS ONE*. 2014; 9(9):e107531. Epub 2014/09/13. <https://doi.org/10.1371/journal.pone.0107531> PMID: 25216051; PMCPCmc4162619.
 39. Wewer Albrechtsen NJ, Kuhre RE, Torang S, Holst JJ. The intestinal distribution pattern of appetite- and glucose regulatory peptides in mice, rats and pigs. *BMC Res Notes*. 2016; 9:60. Epub 2016/02/03. <https://doi.org/10.1186/s13104-016-1872-2> PMID: 26830025; PMCPCMC4736122.
 40. Keenan MJ, Zhou J, McCutcheon KL, Raggio AM, Bateman HG, Todd E, et al. Effects of resistant starch, a non-digestible fermentable fiber, on reducing body fat. *Obesity (Silver Spring, Md)*. 2006; 14(9):1523–34. Epub 2006/10/13. <https://doi.org/10.1038/oby.2006.176> PMID: 17030963.
 41. Reimer RA, Maurer AD, Eller LK, Hallam MC, Shaykhtudinov R, Vogel HJ, et al. Satiety hormone and metabolomic response to an intermittent high energy diet differs in rats consuming long-term diets high in protein or prebiotic fiber. *J Proteome Res*. 2012; 11(8):4065–74. Epub 2012/07/14. <https://doi.org/10.1021/pr300487s> PMID: 22788871; PMCPCmc3411197.
 42. Psichas A, Sleeth ML, Murphy KG, Brooks L, Bewick GA, Hanyaloglu AC, et al. The short chain fatty acid propionate stimulates GLP-1 and PYY secretion via free fatty acid receptor 2 in rodents. *Int J Obes*. 2015; 39(3):424–9. Epub 2014/08/12. <https://doi.org/10.1038/ijo.2014.153> PMID: 25109781; PMCPCMC4356745.

43. Voortman T, Hendriks HF, Witkamp RF, Wortelboer HM. Effects of long- and short-chain fatty acids on the release of gastrointestinal hormones using an ex vivo porcine intestinal tissue model. *J Agric Food Chem*. 2012; 60(36):9035–42. Epub 2012/07/05. <https://doi.org/10.1021/jf2045697> PMID: 22757966.
44. da Silva CS, Haenen D, Koopmans SJ, Hooiveld G, Bosch G, Bolhuis JE, et al. Effects of resistant starch on behaviour, satiety-related hormones and metabolites in growing pigs. *Animal*. 2014; 8(9):1402–11. <https://doi.org/10.1017/S1751731114001116> PMID: 24845880
45. Adrian TE, Ferri GL, Bacarese-Hamilton AJ, Fuessl HS, Polak JM, Bloom SR. Human distribution and release of a putative new gut hormone, peptide YY. *Gastroenterology*. 1985; 89(5):1070–7. Epub 1985/11/01. PMID: 3840109.
46. Zhang T, Uchida T, Gomez G, Lluís F, Thompson JC, Greeley GH Jr. Neural regulation of peptide YY secretion. *Regul Pept*. 1993; 48(3):321–8. Epub 1993/11/03. PMID: 8278624.
47. Sheikh SP, Holst JJ, Orskov C, Ekman R, Schwartz TW. Release of PYY from pig intestinal mucosa; luminal and neural regulation. *Regul Pept*. 1989; 26(3):253–66. Epub 1989/12/01. Release of PYY from pig intestinal mucosa; luminal and neural regulation. PMID: 2623190.
48. Le Gall M, Serena A, Jorgensen H, Theil PK, Bach Knudsen KE. The role of whole-wheat grain and wheat and rye ingredients on the digestion and fermentation processes in the gut—a model experiment with pigs. *Br J Nutr*. 2009; 102(11):1590–600. <https://doi.org/10.1017/S0007114509990924> PMID: 19635175