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### Recent advances in the understanding of severe cutaneous adverse reactions

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### Abstract

Severe cutaneous adverse reactions (SCARs) encompass a heterogeneous group of delayed hypersensitivity reactions, which are most frequently caused by drugs. Our understanding of several aspects of SCAR syndromes has evolved considerably over the previous decade. This review explores evolving knowledge on the immunopathogenic mechanisms, pharmacogenomic associations, *in-vivo* and *ex-vivo* diagnostics for causality assessment and medication cross-reactivity data related to SCAR syndromes. Given the rarity and severity of these diseases, multidisciplinary collaboration through large international, national and/or multicentre networks to collect prospective data on patients with SCAR syndromes should be prioritized. This will further enhance a systematised framework for translating epidemiological, clinical, and immunopathogenetic advances into preventive efforts and improved outcomes for patients.

### What's already known about this topic?

- Severe cutaneous adverse reactions (SCARs) encompass a heterogeneous group of delayed hypersensitivity reactions, which are most frequently caused by drugs.
- The designation SCAR most commonly includes Stevens-Johnson syndrome (SJS), toxic epidermal necrolysis (TEN), SJS-TEN overlap, drug reaction with eosinophilia and systemic symptoms (DRESS)/drug-induced hypersensitivity syndrome (DIHS or HSS) and acute generalised exanthematous pustulosis (AGEP).
- The pathogenesis underlying T-cell mediated delayed hypersensitivity reactions involves interactions between small molecule drugs, HLA Class I molecules and T-cell receptors.

### What does this review add?

- The rapid evolution of pharmacogenomic discoveries associating severe T-cell mediated drug hypersensitivity syndromes have created the promise of prevention. This has led either to universal HLA screening prior to drug prescription (*e.g.* HLA-B\*57:01 and abacavir) or specific recommendations regarding HLA genotyping before prescription of drugs in susceptible populations (*e.g.* HLA-B\*15:02 and carbamazepine).
- Knowledge of the immunopathogenesis of SCAR and key novel and non-mutually exclusive mechanisms by which drugs activate T-cells has evolved.
- In-vivo and ex-vivo diagnostics are being increasingly employed to aid causality assessment.
- Knowledge of cross-reactivity between structurally-related medications is still rudimentary;
   however, this knowledge may avoid precipitating subsequent severe episodes and minimise unwarranted restriction of therapeutic options.

### Introduction

Severe cutaneous adverse reactions (SCARs) encompass a heterogeneous group of delayed hypersensitivity reactions, most frequently caused by drugs, which are associated with significant morbidity and mortality. SCARs include Stevens-Johnson syndrome (SJS), toxic epidermal necrolysis (TEN), drug reaction with eosinophilia and systemic symptoms (DRESS)/drug-induced hypersensitivity syndrome (DIHS or HSS) and acute generalised exanthematous pustulosis (AGEP). The clinical, biochemical and histological characteristics of these syndromes are summarised in Table 1.

Our understanding of several aspects of SCAR syndromes has evolved considerably over the previous decade. The recent 2016 UK guidelines on the management of SJS/TEN in adults highlighted many areas of evolving research.<sup>4</sup> The aim of this review article is to provide a complementary review of emerging immunopathogenic mechanisms, established pharmacogenomic associations, *in-vivo* and *ex-vivo* causality assessment tools and medication cross-reactivity data related to SCAR syndromes.

### **Immunopathogenesis of SCAR**

Medications are the causative agents in greater than 85% of SCARs in adults,<sup>5</sup> with frequently implicated drugs being antimicrobials, aromatic antiepileptic drugs and antimetabolite agents, particularly, allopurinol and its derivatives.<sup>4,5</sup> Regardless of the causal medications, T-cell mediated delayed hypersensitivity reactions, triggered by interactions between small molecule drugs, human leucocyte antigen (HLA) Class I molecules and T-cell receptors (TCR), underlie the pathogenesis of most SCARs. Increasing knowledge suggests that carriage of specific HLA risk allele(s) are necessary but not sufficient factors in initiating the immunopathogenesis cascade.<sup>6</sup> Currently three non-mutually exclusive models have been proposed: the hapten/pro-hapten, the pharmacologic interaction (PI) and the altered peptide repertoire models (Fig. 1). The resultant effector immune mechanisms (*e.g.*,

eosinophil-mediated injury in DRESS<sup>7</sup>, CD8+ cytotoxic T-cell mediated injury in SJS/TEN<sup>4</sup> and the cytotoxic peptide 15kdal granulysin that has been identified as a key molecule produced by CD8+ T cells, natural killer (NK) T cells and NK cells that is responsible for the disseminated keratinocyte death in SJS/TEN<sup>8</sup>) in turn contribute to characteristic clinical manifestations of each condition (Table 1). Of note, Bellon *et al.*'s study suggests that the overexpression of endogenous damage-associate molecular patterns (DAMPs) or alarmins in SJS/TEN support the involvement of the innate immune system in the pathogenesis of delayed hypersensitivity reactions, suggesting an extension of the T-cell mediated hypothesis.<sup>9</sup> Indeed, several innate immune components have been investigated in the aetiopathogenesis of SJS/TEN. Morel and colleagues' study revealed that the innate receptor CD94/NKG2C is expressed by NK cells and cytotoxic T lymphocytes and might be involved in triggering degranulation in response to HLA-E in patients with SJS/TEN.<sup>10</sup> A further study by the same authors determined that upregulation of the innate immune molecules, α-defensins 1-3 in T cells, may be involved in the pathogenesis of SJS/TEN.<sup>11</sup> There is accumulating data to suggest that humoral and cellular components of the innate immune response may be involved in the pathogenesis of delayed cutaneous hypersensitivity reactions.<sup>12</sup>

Higher plasma concentrations of the drug and/or its metabolites, caused by the individual's *in-vivo* absorption, distribution, metabolism and elimination enzyme (ADME) activities, or by way of drugdrug interactions, increase the risk for many hypersensitivity reactions. <sup>13,14</sup> This apparent dose-dependency seen in severe T-cell mediated adverse drug reactions (ADRs) supports that small molecules are non-covlaently interacting with an immune receptor. For instance, elevated serum levels of oxypurinol, an active metabolite of allopurinol, which has a long plasma half-life, increases the risk of allopurinol hypersensitivity. <sup>14</sup> Impaired renal function leading to high plasma concentrations of oxypurinol is also directly correlated with disease severity and mortality. <sup>14</sup> Historically, certain types of trimethoprim-sulfamethoxazole hypersensitivity reactions were more likely in those with N-acetyl transferase (NAT) 2 slow-acetylator genotypes. <sup>15</sup> Collectively, the paradigm has been shifting towards an interplay between ADME enzymatic activities and immunologic mechanisms being responsible for the initiation of hypersensitivity responses, <sup>16</sup> further triggered by yet-to-be-determined insults (such as viral infections), leading to polarisation toward distinct cytokine profiles and effector pathways. Further studies are required to explore this evolving concept of hypersensitivity and drug concentration-dependent relationships.

### The role of herpes virus reactivation

Heterologous immunity is a longstanding concept that has recently gained renewed interest to explain both individual susceptibility and tissue specificity of SCAR. In this model, the effector memory T-cells generated during the course of a remote infection and maintained by latency or re-exposure to the infectious agent cross-react with drug modified proteins, thereby highlighting the role of

infectious agents, such as chronic persistent DNA viruses including Human Herpes viruses (HHV), in SCAR pathogenesis.<sup>16</sup>

The concept of heterologous immunity in the immunopathogenesis of SCAR should not be confused with the reactivation of HHV, in particular human herpes virus 6 (HHV-6), which is known to be associated with DRESS. 17-20 Reactivation of Epstein-Barr virus (EBV), cytomegalovirus (CMV), HHV-6 and human herpes virus 7 (HHV-7) has been reported to occur in DRESS syndrome typically 2-3 weeks following the original syndrome and in the absence of re-exposure to the drug. It appears to correlate with the immune dysregulation occurring during DRESS syndrome and in particular, regulatory T-cell dysfunction. The reported proportion of patients with HHV-6 reactivation in DRESS varies according to the specific implicated drug and is between 36% and 62%. 18,21 HHV-6 reactivation, as measured by a rise in HHV-6 IgG titres and plasma HHV-6 DNA levels, typically occurs 2-3 weeks after the onset of the rash. 22 This temporal association suggests a complex interaction between HHV and the immunopathogenesis of DRESS. 24 Furthermore, reactivation of HHV have also been associated with the development of more severe disease. 19,21-24 The development of autoimmune diseases, such as systemic lupus erythematosus, type 1 diabetes mellitus and autoimmune thyroiditis, is a late complications of DRESS that has been associated with herpes virus reactivation. 20,25-27

Reactivation of the other herpes viruses, which include HHV-7, EBV and CMV have also been reported to occur in association with DRESS. <sup>22,28,29</sup> Indeed, sequential reactivation of herpes viruses during the course of DRESS has been described in a similar sequence to that in graft-versus-host disease (GVHD): HHV-6 and/or EBV, followed by HHV-7 and subsequently by CMV. <sup>29</sup> Viral reactivation may also explain the prolonged clinical symptoms, multi-organ involvement and systemic inflammation following discontinuation of the offending drug. <sup>22,29-31</sup>

DRESS has been reported in the setting of immune reconstitution inflammatory syndrome (IRIS). IRIS describes an inflammatory processes that occurs soon after the initiation of highly active antiretroviral therapy (HAART) in patients with Human Immunodeficiency Virus (HIV) and is associated with an increase in CD4+ cell count and/or decrease in HIV viral load.<sup>32</sup> IRIS occurs as a result of immune recovery and it results in the host recognising pre-existing or latent infections.<sup>33</sup> DRESS may be considered a form of immune constitution whereby unregulated immune activation occurs against reactivated herpes viruses.<sup>32</sup>

For SJS/TEN however, there is weaker evidence, only at case report level, for its association with HHV-6 reactivation and this could also be secondary to phenotype misattribution of viral reactivation associated with the profound immunosuppression secondary to the protracted clinical course and significant courses of immunosuppressants, such as ciclosporin used in SJS/TEN.<sup>34,35</sup> The role of CMV has been proposed in the development of AGEP,<sup>36</sup> however evidence from European Study of Severe Cutaneous Adverse Reactions (EuroSCAR) study failed to find such an association.<sup>37</sup> Testing

for herpes virus reactivation in SCAR syndromes may assist in clarifying the diagnosis in cases where the cutaneous and other clinical findings are non-specific, and may also be of prognostic value.<sup>31,21,38</sup>

### Recent advances in pharmacogenomics in SCAR

Individuals with certain HLA genotypes carry higher risks of developing SCAR syndromes. Over the last decade, clinically significant pharmacogenomics associations have been discovered, leading to specific recommendations regarding HLA genotyping before prescription of drugs to reduce the risks in susceptible populations. However, for common causal medications, in particular, antibiotics, very few clinically meaningful HLA associations exist.<sup>39</sup> Medications that are considered to have strong pharmacogenomic associations with severe T-cell mediated ADRs, of which routine genetic screening prior to their prescription have already or in future may soon become the standard of clinical practice are presented herein (Table 2).

### Abacavir

Abacavir (ABC), an antiretroviral drug used in combination therapy to treat HIV, is associated with hypersensitivity syndrome (HSS) in 5% (range 0 – 14%) of patients. <sup>40</sup> The hypersensitivity syndrome associated with ABC is differentiated from DRESS/DIHS in that the median time to presentation with fever and malaise is 8 days with latency periods as short as 1 day and rash, which does not occur in up to 30%, is often a late feature of the presentation. The skin involvement in ABC HSS is typically a mild to moderate exanathem without evidence of blistering or epidermal detachment. De-challenge after withdrawal of drug occurs rapidly with disappearance of the fever, malaise and even skin rash within 72 hours of abacavir withdrawal. HLA-B\*57:01 was found to be a significant risk allele for ABC-HSS by two independent groups. 41,42 The lack of specificity of clinical symptoms and signs associated with ABC HSS in HIV positive individuals led to a high clinical false positive rate and an apparent lack of sensitivity of HLA-B\*57:01 for ABC HSS. This was particularly apparent in ethnicities with a lower prevalence of HLA-B\*57:01 such as African Americans. ABC patch testing was found to be a sensitive and specific means to identify true immunologically-mediated ABC HSS. 43,44 A randomised double-blind controlled trial with a co-primary endpoint of clinically and immunologically (patch-test) confirmed ABC HSS demonstrated the clinical utility of HLA-B\*57:01 screening to completely eliminate immunologically-mediated cases of ABC-HSS in those of European ancestry. 45 A case-control study confirmed the generalizability of this utility to African Americans. 46 Several factors favoured successful translation of HLA-B\*57:01 screening into routine clinical practice including: 100% negative predictive value, low numbers (n=30) needed to test to prevent one case of true-immunologically mediated ABC HSS, generalisability of the test across all ethnic groups and availability of cost-effective quality-assured laboratory methods with rapid turnaround times. 46-48

### Carbamazepine

Carbamazepine is an aromatic amine anticonvulsant and is associated with cutaneous adverse reactions in up to 10% of patients.<sup>49</sup> Although two digit HLA associations had been previously described between allopurinol SJS/TEN and sulfa antimicrobial SJS/TEN, the association between HLA-B\*15:02 and carbamazepine SJS/TEN in a Taiwanese population was the first four digit association for SJS/TEN and the strongest overall for SJS/TEN in the literature to-date.<sup>50</sup> A recent meta-analysis showed that HLA-B\*15:02 is strongly associated with carbamazepine-induced SJS/TEN in Han Chinese and Southeast Asians who carry high allele frequency (pooled Odds Ratio (OR) 113.4, 95% CI 51.2 – 251.0, p<1×10<sup>-5</sup>).<sup>51</sup> However, such association was lacking in Japanese,<sup>52-54</sup> Koreans,<sup>55</sup> and Caucasians,<sup>56,57</sup> in whom the allele carrier frequency was estimated to be <1%<sup>58</sup>. HLA-B\*15:02 testing provides positive predictive value (PPV) of 1.8% and negative predictive value (NPV) of 100% respectively in susceptible populations, with proven cost-effectiveness for screening.<sup>51,59,60</sup>

Although HLA-B\*15:02 is a risk variant strongly associated with carbamazepine SJS/TEN, there is no evidence to suggest that it is associated with hypersensitivity syndrome (HSS) or maculopapular exathems.<sup>51,58</sup>

Unlike HLA-B\*15:02, HLA-A\*31:01 is common with allele carrier frequencies >3% across many ethnic groups<sup>58</sup>. HLA-A\*31:01 was shown to be associated with all SCAR phenotypes across populations including Han Chinese, Japanese, Koreans and Caucasians.<sup>52,55,58,61,62</sup> However, HLA-A\*31:01 showed a stronger association with DRESS (pooled OR 13.2, 95% CI 8.4 – 20.8, p<0.001) over SJS/TEN (pooled OR 3.94, 95% CI 1.4 – 11.5, p=0.01).<sup>58,63</sup> This effect was particularly noted in populations where HLA-B\*15:02 carriage is prevalent where it is likely that the strong association between HLA-B\*15:02 and carbamazepine SJS/TEN overshadows that of HLA-A\*31:01. In contrast, in Europeans, the higher frequency of the HLA-A\*31:01 allele appears to overshadow the effect of the uncommon HLA-B\*15:02 allele. <sup>47,51</sup>

Regulatory agencies such as the US Food and Drug Administration (FDA) and the European Medicines Agency have issued recommendations regarding genotyping before initiation of carbamazepine in certain at-risk populations.<sup>64</sup> Genetic testing for HLA-B\*15:02 is recommended in Han Chinese, Southeast and South Asians or in patients whose ethnic origin is unknown (Level A). HLA-A\*31:01 testing may be considered in patients of all ancestries (level B); however, there is no current recommendation for routine screening for HLA-A\*31:01 before initiation of carbamazepine therapy. In patients who are positive for HLA-B\*15:02, alternatives to carbamazepine should be used, preferably avoiding all aromatic amine anticonvulsants since SJS/TEN has been more weakly associated with HLA-B\*15:02 with these drugs in Southeast Asians. In the case of HLA-A\*31:01 positivity, ideally, alternative first-line medication to carbamazepine should be used in carbamazepine naïve individuals unless there are no identifiable alternatives, in which case patients should be followed with extremely close monitoring for the first signs of evolving SCAR. <sup>58</sup>

### Allopurinol

Allopurinol accounts for up to 5% of all cases with SCAR.<sup>65</sup> An association between allopurinol induced SCAR (SJS/TEN and HSS phenotypes) and HLA-B\*58:01 genotype was first described in Taiwanese Han Chinese population.<sup>66</sup> Thereafter, studies in other ethnic groups including, Han Chinese from mainland China<sup>67,68</sup> and Hong Kong,<sup>69</sup> Thai,<sup>70</sup> Koreans,<sup>71,72</sup> Japanese,<sup>54</sup> and Europeans<sup>73,74</sup> have replicated similar associations, although the strength of association was much weaker with a lower negative predictive value in Japanese and Europeans, likely owing to different allele frequencies across ethnic groups. The NPV of HLA-B\*58:01 screening for allopurinol induced SCAR in Southeast Asian populations is 100%.<sup>75</sup>A modelling study from Singapore showed that routine genetic screening to prevent an episode of SCAR, even in high risk populations, did not appear to be cost-effective.<sup>76</sup> The extreme short and long-term morbidity and mortality that is in particular associated with SJS/TEN, the lack of comparably inexpensive treatment options to allopurinol, the development of newer and less expensive molecular assays for HLA-B\*58:01 and the availability of a prospective screening study suggesting a significantly reduced incidence of allopurinol SCAR with HLA-B\*58:01 screening in Taiwan suggest that further attention and implementation of HLA-B\*58:01 screening may be warranted.

### Causality assessment through clinical, in vivo and ex vivo testing

Assigning drug causality is often difficult in SCAR syndromes, especially when multiple agents are implicated, in particular, antimicrobials.<sup>78</sup> Conversely, in situations of a single implicated drug (*e.g.* carbamazepine, allopurinol), utilisation of appropriate clinical algorithms is often sufficient to assign causality,<sup>5,79</sup> especially in histologically confirmed cases.<sup>80,81</sup> Drug causality may be clinically established through several different validated methods/algorithms, each with own strengths and limitations (Table 3). Nonetheless, *in vivo* and *ex vivo* diagnostics are being increasingly employed to aid causality and management of patients with SCAR.<sup>82</sup> Guidelines exist for the recommended concentrations of drugs to be used in *in vivo* testing for delayed hypersensitivity,<sup>83,84</sup> although universal consensus has not been established.

### Patch testing

Patch testing (PT) involves the application of an implicated and/or potentially cross-reactive drug with a control vehicle (petroleum jelly) to skin for 48 hours<sup>82</sup> and subsequently read after 48-96 hours and if possible 7 days. The safety of PT in SCAR has been increasingly demonstrated.<sup>85-91</sup> Systemic (but non-life threatening) reactions have been reported infrequently with PT, although mostly for antituberculosis drugs in HIV patients.<sup>92-96</sup> The recommendations have been to perform skin testing at least 6 weeks post-resolution of SCAR.<sup>97</sup> The sensitivity of patch testing appears highest for ABC HSS (87%)<sup>43,44</sup> and DRESS (31.6%-58%) and lowest for SJS/TEN (20%-24%) and AGEP

(18%). 85,86,90 The sensitivity also appears to be affected by the investigated drug, highest for abacavir, anticonvulsants and beta-lactam antibiotics, 7 in particular for abacavir (87%), amoxicillin (up to 44.4%), and lowest for vancomycin (9.1%), trimethoprim-sulfamethoxazole (8.6%), macrolides (4.8%), hepatitis C antivirals 9 and cephalosporins (4.4%). The use of oral provocation after a negative PT should be used with caution in patients with SCAR, considering the low sensitivity of PT.

### Intradermal testing

Intradermal testing (IDT) utilising 0.02-0.05 ml of the highest non-irritant concentration of drug, has been reported in DRESS and other SCAR phenotypes in a number of small series. <sup>86,99-101</sup> IDT with delayed readings has been utilised extensively for T-cell mediated hypersensitivity, in particular for non-SCAR phenotypes related to beta-lactams. <sup>102,103</sup> IDT avoids the inconvenience of patch testing and reactions will often occur within 6-24 hours. Barbaud *et al.* demonstrated in a small cohort of predominately beta-lactam SCAR that IDT appeared to have a greater sensitivity than PT when performed following negative PT and was not associated with adverse events. <sup>86</sup> Guidelines also support the use of IDT following negative PT in patients with SCAR, outside of SJS/TEN. <sup>83</sup> IDT is often limited by the availability of a sterile injectable formulation of the investigated drug. Like PT, oral provocation testing after a negative IDT should be undertaken with caution.

### Ex vivo diagnostics

The stimulation of patient peripheral blood mononuclear cells (PBMCs) to measure T-cell responses in the setting of drug-associated SCAR has been increasingly investigated in research and clinical settings. Whilst responses have been detected out to 20 years post-index event, a blood sample from 'acute bleeds' or in the early recovery phase is likely to display greater sensitivity. 104-106 The lymphocyte transformation test (LTT), 99,107 which typically incubates investigated drugs with PBMCs for 5-7 days or longer, measures T-cell responses to a variety of drugs (e.g. antimicrobials, anticonvulsants, analgesics and diuretics) via a stimulation index. 104,107-117 Enzyme-linked immunospot assay (ELISpot) has been primarily employed for antiretroviral and antimicrobial hypersensitivity and SCAR syndromes, 118-121 especially when in vivo testing has been negative. 94,99,113,122,123 Variability in testing methods, incubation periods (1 vs. 2 vs. 5 days), costimulation factors (e.g. IL-7/IL-15) and measured outputs (e.g. granulysin, IFN-γ, TNF-α) make comparison within and between testing modalities difficult. 8,106,124,125 The known drug epitopes are unknown for most T-cell mediated hypersensitivity syndromes <sup>126,127</sup> Currently LTT or ELISpot should not be employed to exclude a suspected drug due to low sensitivity (24-70% 125,128 and 60%-80%, 125 respectively.) Whilst LTT has demonstrated a higher sensitivity in other types of anticonvulsant hypersensitivity (70-90%), <sup>129</sup> lower rates have still been noted in lamotrigine-SJS. <sup>130</sup>

Indeed, Polak and colleagues' study compared the lymphocyte proliferation assay (LPA) against combination INF-γ and IL-4 drug ELISpot assays in patients with delayed-type drug hypersensitivity

reactions in the acute phase. In their study, the assays demonstrated a test specificity of 95%, 83% and 92% for LPA, INF-γ and IL-4, respectively. During acute drug hypersensitivity reactions, the sensitivity of combined measurement of drug-specific INF-γ and IL-4 cytokines was greater than that of LPA (82% *vs.* 50%). Thus, these investigators determined that *in vitro* assays of drug-specific INF-γ and IL-4 production may be more sensitive than LPA for the detection of drug-specific T-cells in the acute setting. <sup>131</sup> Further, a recent study by Haw *et al.* concluded that cytokine assays (INF-γ and IL-4) are superior to LPA in identifying the causative drug in the paediatric population; however, these investigators suggested that when combined, they offer even greater utility in the diagnosis and post-recovery of delayed-type hypersensitivity reactions. <sup>132</sup>

The sensitivity and hence NPV of *ex-vivo* testing in the future is likely to be enhanced by coutilisation of flow cytometry and intracellular cytokine staining methods. 118,133-136

### The importance of drug cross-reactivity between structurally-related drugs

Structurally-related drugs can cause cross-reactions with SCAR. Although the specific epitopes remain elusive with regards to drug-self peptide responses, it is recognized that the immune system may recognise structural similarities. Knowledge regarding the likelihood of cross-reactivity between drugs is important as exposure to structurally similar compounds after an index reaction can precipitate another severe episode. On the contrary, excessive avoidance of medications with low risk of cross-reactivity can lead to unwarranted restriction on therapeutic options that can adversely impact upon clinical care.

### Beta-lactams

All beta-lactams (penicillins, cephalosporins, carbapenems and monobactams) share the core beta-lactam structure but with differing side-chains (Fig. 2). Evolving evidence to date suggests that side chain structures are commonly implicated in beta-lactam cross-reactivity for most immediate and delayed reactions. Table 4 further provides a list of commonly prescribed beta-lactams which share similar side chain structures.

### Cephalosporins

R1 side-chains of cephalosporins (Fig. 2) are highly conserved and have been demonstrated to promote cross-reactions with penicillins containing similar structures. This is particularly true between aminopenicillins (amoxicillin, ampicillin and bacampillin) and aminocephalosporins (cephalexin and cefaclor), with recent studies demonstrating that the cross-reactivity rates between the amino compounds may be as high as 18.7%. On the contrary, patients with delayed aminopenicillin allergy have recently been shown to have complete absence of cross-reactivity and good tolerance to therapeutic challenge to non-amino cephalosporins (cefuroxime and ceftriaxone). 137

Overall, low rates of cross-reactivity exist between penicillins and third and fourth generation cephalosporins of dissimilar side chain structures (1.1% vs. 10.9% for first and second generation cephalosporins which share similar side chains).<sup>138</sup>

Further, an interesting *in vitro* study by El-Ghaiesh *et al.* in eight cystic fibrosis patients with delayed hypersensitivity reactions to piperacillin, compared to five tolerant controls, demonstrated the critical role of drug-specific CD4+ and CD8+ T-cell clones in pathogenesis, which did not cross-react to a multitude of penicillins and cephalosporins including those that share similar side chain to piperacillin (*e.g.* cefoperazone). This study highlights the drug-specific nature of T-cell mediated hypersensitivity reactions as well as the highly complex nature of cross-reactivity to other beta-lactams, with some unknown mechanisms in addition to 'structural similarities,' likely further contributing to its pathogenesis.<sup>113</sup>

### Carbapenems and monobactams

Although a cross-reactivity rate of 5.5% to imipenem has been previously reported in penicillinallergic patients. A more recent study involving 204 patients demonstrated that none of the patients with delayed penicillin hypersensitivity cross-reacted to imipenem, meropenem or ertapenem, and all tolerated therapeutic doses of drug challenge. In view of the reportedly low (<1%) rates of cross-reactivity to carbapenems in patients with immediate penicillin hypersensitivity reactions, the true cross-reactivity rates in delayed reactions are likely very low (<1%) and therefore, carbapenems may be judiciously considered in patients who have limited therapeutic options.

In contrast, virtually zero percent cross-reactivity to aztreonam has been consistently demonstrated in patients with delayed penicillin hypersensitivity reactions. <sup>137,143</sup> The only caveat is that aztreonam should be avoided in patients with ceftazidime allergy due to side-chain similarities.

It should also be noted that although cross-reactivity rates between penicillins and later generation cephalosporins or carbapenems are low, the vast majority of patients included in these studies had benign skin reactions and few patients with definitive SCAR phenotypes were represented. As such, considerable caution should be taken when prescribing beta-lactam antibiotics to patients with SCAR.

### Aromatic anticonvulsants

Commonly prescribed aromatic anticonvulsants include carbamazepine, oxcarbazepine, lamotrigine, phenytoin and phenobarbital. Cross-reactivity between these structurally related aromatic anticonvulsants was originally thought to be mediated by arene oxides, toxic metabolites produced through cytochrome P450 pathway. However, it is now clear that poor metabolisers (*e.g.* CYP2C9\*3) are at higher risk for SCAR associated with some anticonvulsants such as phenytoin. Earlier studies suggested that approximately 70% will experience some degree of cross-reactivity between aromatic anticonvulsants. There is also evidence suggesting that HLA-B\*15:02 and other B75 serotype HLA alleles confer risk of developing SJS/TEN to other aromatic anticonvulsants,

however, to a much lesser degree compared to carbamazepine. <sup>151,152</sup> What is currently unclear is the extent to which HLA cross-reactivity occurs since cases of HLA-B\*15:02 positive individuals who have reacted to one aromatic amine anticonvulsant but tolerated another (despite the association of HLA-B\*15:02 with all aromatic amine anticonvulsant SCAR) have been well-described. Additionally Seitz *et al.* also noted that 21.7% of patients with carbamazepine hypersensitivity also displayed cross-reactivity to tricyclic antidepressants. <sup>150</sup> However, this has not been substantiated as an effect that is seen *in-vivo* and in the case of HSS to carbamazepine, recommendations would not dictate avoidance of tricyclic antidepressants. In patients with SCAR to aromatic anticonvulsants, valproate, gabapentin, pregabalin and levetiracetam are safe alternatives. <sup>153,154</sup>

### **Conclusion and Future Directions**

Recent advances in the knowledge of SCAR syndromes have provided us with a better understanding of immunopathogenic mechanisms, including the potential role of pre-existing cross-reactive T cell responses to viral infections, the discovery of important pharmacogenomic associations, which have become the standard of care, the use of clinical and laboratory methods for causality assessment and the knowledge of drug cross-reactivity mechanisms. Further knowledge on how precisely drugs activate T-cells, the pathomechanism for the generally very low positive predictive value of an HLA risk allele for a specific drug toxicity, more specific pharmacogenomic associations and future mechanistic information including cellular and molecular signatures will be key for pre-clinical prediction and prevention of drug toxicity as well as for enabling personalised approaches to prevention, early intervention and treatment of high morbidity and mortality diseases such as SJS/TEN. As highlighted in this review, numerous aspects of SCAR syndromes merit further interdisciplinary research. Finally, given the overall rarity but high morbidity and mortality of SCAR, collaboration through large international, national and multicentre networks to collect prospective data and biobank samples will further enhance a systematised framework for translating discovery into prevention and improved outcomes for patients.

### References

- 1. Thong BY, Tan TC. Epidemiology and risk factors for drug allergy. *Br J Clin Pharmacol* 2011; **71**:684-700.
- 2. Roujeau JC. Clinical heterogeneity of drug hypersensitivity. *Toxicology* 2005; **209**:123-9.
- 3. Roujeau JC, Stern RS. Severe adverse cutaneous reactions to drugs. *N Engl J Med* 1994; **331**:1272-85.
- 4. Creamer D, Walsh SA, Dziewulski P *et al.* U.K. guidelines for the management of Stevens-Johnson syndrome/toxic epidermal necrolysis in adults 2016. *Br J Dermatol* 2016; **174**:1194-227.

- 5. Sassolas B, Haddad C, Mockenhaupt M *et al.* ALDEN, an algorithm for assessment of drug causality in Stevens-Johnson Syndrome and toxic epidermal necrolysis: comparison with case-control analysis. *Clin Pharmacol Ther* 2010; **88**:60-8.
- 6. Pavlos R, Mallal S, Phillips E. HLA and pharmacogenetics of drug hypersensitivity. *Pharmacogenomics* 2012; **13**:1285-306.
- 7. Posadas SJ, Padial A, Torres MJ *et al.* Delayed reactions to drugs show levels of perforin, granzyme B, and Fas-L to be related to disease severity. *J Allergy Clin Immunol* 2002; **109**:155-61.
- 8. Chung WH, Hung SI, Yang JY *et al*. Granulysin is a key mediator for disseminated keratinocyte death in Stevens-Johnson syndrome and toxic epidermal necrolysis. *Nat Med* 2008; **14**:1343-50.
- 9. Bellon T, Alvarez L, Mayorga C *et al.* Differential gene expression in drug hypersensitivity reactions: induction of alarmins in severe bullous diseases. *Br J Dermatol* 2010; **162**:1014-22.
- 10. Morel E, Escamochero S, Cabanas R *et al.* CD94/NKG2C is a killer effector molecule in patients with Stevens-Johnson syndrome and toxic epidermal necrolysis. *J Allergy Clin Immunol* 2010; **125**:703-10.
- 11. Morel E, Alvarez L, Cabanas R *et al.* Expression of alpha-defensin 1-3 in T cells from severe cutaneous drug-induced hypersensitivity reactions. *Allergy* 2011; **66**:360-7.
- 12. Bellon T, Blanca M. The innate immune system in delayed cutaneous allergic reactions to medications. *Curr Opin Allergy Clin Immunol* 2011; **11**:292-8.
- 13. Grossman I. ADME pharmacogenetics: current practices and future outlook. *Expert Opin Drug Metab Toxicol* 2009; **5**:449-62.
- 14. Chung WH, Chang WC, Stocker SL *et al.* Insights into the poor prognosis of allopurinol-induced severe cutaneous adverse reactions: the impact of renal insufficiency, high plasma levels of oxypurinol and granulysin. *Ann Rheum Dis* 2015; **74**:2157-64.
- 15. Pirmohamed M, Alfirevic A, Vilar J *et al.* Association analysis of drug metabolizing enzyme gene polymorphisms in HIV-positive patients with co-trimoxazole hypersensitivity. *Pharmacogenetics* 2000; **10**:705-13.
- 16. White KD, Chung WH, Hung SI *et al*. Evolving models of the immunopathogenesis of T cell-mediated drug allergy: The role of host, pathogens, and drug response. *J Allergy Clin Immunol* 2015; **136**:219-34.
- 17. Pavlos R, Mallal S, Ostrov D *et al*. Fever, rash, and systemic symptoms: understanding the role of virus and HLA in severe cutaneous drug allergy. *J Allergy Clin Immunol Pract* 2014; **2**:21-33.
- 18. Kardaun SH, Sekula P, Valeyrie-Allanore L *et al.* Drug reaction with eosinophilia and systemic symptoms (DRESS): an original multisystem adverse drug reaction. Results from the prospective RegiSCAR study. *Br J Dermatol* 2013; **169**:1071-80.

- 19. Picard D, Janela B, Descamps V *et al.* Drug reaction with eosinophilia and systemic symptoms (DRESS): a multiorgan antiviral T cell response. *Sci Transl Med* 2010; **2**:46-62.
- 20. Descamps V, Mahe E, Houhou N *et al.* Drug-induced hypersensitivity syndrome associated with Epstein-Barr virus infection. *Br J Dermatol* 2003; **148**:1032-4.
- 21. Tohyama M, Hashimoto K, Yasukawa M *et al.* Association of human herpesvirus 6 reactivation with the flaring and severity of drug-induced hypersensitivity syndrome. *Br J Dermatol* 2007; **157**:934-40.
- 22. Shiohara T, Inaoka M, Kano Y. Drug-induced hypersensitivity syndrome (DIHS): a reaction induced by a complex interplay among herpesviruses and antiviral and antidrug immune responses. *Allergol Int* 2006; **55**:1-8.
- 23. Chen YC, Chiang HH, Cho YT *et al*. Human herpes virus reactivations and dynamic cytokine profiles in patients with cutaneous adverse drug reactions a prospective comparative study. *Allergy* 2015; **70**:568-75.
- 24. Ahluwalia J, Abuabara K, Perman MJ *et al*. Human herpesvirus 6 involvement in paediatric drug hypersensitivity syndrome. *Br J Dermatol* 2015; **172**:1090-5.
- 25. Chiou CC, Chung WH, Hung SI *et al*. Fulminant type 1 diabetes mellitus caused by drug hypersensitivity syndrome with human herpesvirus 6 infection. *J Am Acad Dermatol* 2006; **54**(Suppl. 2):S14-7.
- 26. Aota N, Hirahara K, Kano Y *et al.* Systemic lupus erythematosus presenting with Kikuchi-Fujimoto's disease as a long-term sequela of drug-induced hypersensitivity syndrome. A possible role of Epstein-Barr virus reactivation. *Dermatology* 2009; **218**:275-7.
- 27. Funck-Brentano E, Duong T, Family D *et al.* Auto-immune thyroiditis and drug reaction with eosinophilia and systemic symptoms (DRESS) associated with HHV-6 viral reactivation. *Ann Dermatol Venereol* 2011; **138**:580-5.
- 28. Seishima M, Yamanaka S, Fujisawa T *et al.* Reactivation of human herpesvirus (HHV) family members other than HHV-6 in drug-induced hypersensitivity syndrome. *Br J Dermatol* 2006; **155**:344-9.
- 29. Kano Y, Hiraharas K, Sakuma K *et al.* Several herpesviruses can reactivate in a severe drug-induced multiorgan reaction in the same sequential order as in graft-versus-host disease. *Br J Dermatol* 2006; **155**:301-6.
- 30. Harding DJ, Subramaniam K, MacQuillan G *et al.* Severe drug-induced hypersensitivity syndrome with a shared HLA-B allele. *Med J Aust* 2012; **197**:411-3.
- 31. Shiohara T, Iijima M, Ikezawa Z *et al*. The diagnosis of a DRESS syndrome has been sufficiently established on the basis of typical clinical features and viral reactivations. *Br J Dermatol* 2007; **156**:1083-4.
- 32. Almudimeegh A, Rioux C, Ferrand H *et al.* Drug reaction with eosinophilia and systemic symptoms, or virus reactivation with eosinophilia and systemic symptoms as a manifestation

- of immune reconstitution inflammatory syndrome in a patient with HIV? *Br J Dermatol* 2014; **171**:895-8.
- 33. Kano Y, Ushigome Y, Horie C *et al*. Immune reconstitution inflammatory syndrome observed in the setting of drug-induced hypersensitivity syndrome/drug reaction with eosinophilia and systemic symptoms (DIHS/DRESS). *Clin Transl Allergy* 2014; **4**:148.
- 34. Peppercorn AF, Miller MB, Fitzgerald D *et al*. High-level human herpesvirus-6 viremia associated with onset of Stevens-Johnson syndrome: report of two cases. *J Burn Care Res* 2010; **31**:365-8.
- 35. Teraki Y, Murota H, Izaki S. Toxic epidermal necrolysis due to zonisamide associated with reactivation of human herpesvirus 6. *Arch Dermatol* 2008; **144**:232-5.
- 36. Haro-Gabaldon V, Sanchez-Sanchez-Vizcaino J, Ruiz-Avila P *et al.* Acute generalized exanthematous pustulosis with cytomegalovirus infection. *Int J Dermatol* 1996; **35**:735-7.
- 37. Sidoroff A, Dunant A, Viboud C *et al.* Risk factors for acute generalized exanthematous pustulosis (AGEP)-results of a multinational case-control study (EuroSCAR). *Br J Dermatol* 2007; **157**:989-96.
- 38. Eshki M, Allanore L, Musette P *et al.* Twelve-year analysis of severe cases of drug reaction with eosinophilia and systemic symptoms: a cause of unpredictable multiorgan failure. *Arch Dermatol* 2009; **145**:67-72.
- 39. Aung AK, Haas DW, Hulgan T *et al*.Pharmacogenomics of antimicrobial agents. *Pharmacogenomics* 2014; **15**:1903-30.
- 40. Symonds W, Cutrell A, Edwards M *et al*. Risk factor analysis of hypersensitivity reactions to abacavir. *Clin Ther* 2002; **24**:565-73.
- 41. Mallal S, Nolan D, Witt C *et al.* Association between presence of *HLA-B\* 5701*, *HLA-DR7*, and *HLA-DQ3* and hypersensitivity to HIV-1 reverse-transcriptase inhibitor abacavir. *Lancet* 2002; **359**:727-32.
- 42. Hetherington S, Hughes AR, Mosteller M *et al.* Genetic variations in *HLA-B* region and hypersensitivity reactions to abacavir. *Lancet* 2002; **359**:1121-2.
- 43. Shear NH, Milpied B, Bruynzeel DP *et al.* A review of drug patch testing and implications for HIV clinicians. *AIDS* 2008; **22**:999-1007.
- 44. Phillips EJ, Wong GA, Kaul R *et al.* Clinical and immunogenetic correlates of abacavir hypersensitivity. *AIDS* 2005; **19**:979-81.
- 45. Mallal S, Phillips E, Carosi G *et al.* HLA-B\* 5701 screening for hypersensitivity to abacavir. *N Engl J Med* 2008; **358**:568-79.
- 46. Saag M, Balu R, Phillips E *et al*. High sensitivity of human leukocyte antigen-b\* 5701 as a marker for immunologically confirmed abacavir hypersensitivity in white and black patients. *Clin Infect Dis* 2008; **46**:1111-8.

- 47. Phillips E, Mallal S. Successful translation of pharmacogenetics into the clinic. *Mol Diagn Ther* 2009; **13**:1-9.
- 48. Ruiz-Iruela C, Padullés-Zamora N, Podzamczer-Palter D *et al.* HLA-B\* 57: 01 genotyping in the prevention of hypersensitivity to abacavir: 5 years of experience. *Pharmacogenet Genomics* 2016; 26:390-6.
- 49. Marson AG, Al-Kharusi AM, Alwaidh M *et al*. The SANAD study of effectiveness of carbamazepine, gabapentin, lamotrigine, oxcarbazepine, or topiramate for treatment of partial epilepsy: an unblinded randomised controlled trial. *Lancet* 2007; **369**:1000-15.
- 50. Chung WH, Hung SI, Hong HS *et al.* Medical genetics: a marker for Stevens-Johnson syndrome. *Nature* 2004; **428**:486.
- 51. Yip V, Marson A, Jorgensen A *et al.* HLA genotype and carbamazepine-induced cutaneous adverse drug reactions: a systematic reveiw. *Clin Pharmacol Ther* 2012; **92**:757-65.
- 52. Ozeki T, Mushiroda T, Yowang A *et al.* Genome-wide association study identifies HLA-A\* 3101 allele as a genetic risk factor for carbamazepine-induced cutaneous adverse drug reactions in Japanese population. *Hum Mol Gen* 2011; **20**:1034-41.
- 53. Ikeda H, Takahashi Y, Yamazaki E *et al.* HLA Class I markers in Japanese patients with carbamazepine-induced cutaneous adverse reactions. *Epilepsia* 2010; **51**:297-300.
- 54. Kaniwa N, Saito Y, Aihara M *et al.* HLA-B locus in Japanese patients with anti-epileptics and allopurinol-related Stevens-Johnson syndrome and toxic epidermal necrolysis. *Pharmacogenomics* 2008; **9**:1617-22.
- 55. Kim SH, Lee KW, Song WJ *et al*. Carbamazepine-induced severe cutaneous adverse reactions and HLA genotypes in Koreans. *Epilepsy Res* 2011; **97**:190-7.
- 56. Lonjou C, Thomas L, Borot N *et al.* A marker for Stevens-Johnson syndrome: ethnicity matters. *Pharmacogenomics J* 2006; **6**:265-8.
- 57. Alfirevic A, Jorgensen AL, Williamson PR *et al.* HLA-B locus in Caucasian patients with carbamazepine hypersensitivity. *Pharmacogenomics* 2006; **7**:813-8.
- 58. Amstutz U, Shear NH, Rieder MJ *et al.* Recommendations for HLA-B\* 15:02 and HLA-A\* 31:01 genetic testing to reduce the risk of carbamazepine-induced hypersensitivity reactions. *Epilepsia* 2014; **55**:496-506.
- 59. Chen P, Lin JJ, Lu CS *et al*. Carbamazepine-induced toxic effects and HLA-B\* 1502 screening in Taiwan. *N Engl J Med* 2011; **364**:1126-33.
- 60. Locharernkul C, Shotelersuk V, Hirankarn N. HLA-B\* 1502 screening: time to clinical practice. *Epilepsia* 2010; **51**:936-8.
- 61. McCormack M, Alfirevic A, Bourgeois S *et al.* HLA-A\* 3101 and carbamazepine-induced hypersensitivity reactions in Europeans. *N Engl J Med* 2011; **364**:1134-43.
- 62. Hung SI, Chung WH, Jee SH *et al*. Genetic susceptibility to carbamazepine-induced cutaneous adverse drug reactions. *Pharmacogenet Genomics* 2006; **16**:297-306.

- 63. Genin E, Chen D, Hung S *et al*. HLA-A\* 31:01 and different types of carbamazepine-induced severe cutaneous adverse reactions: an international study and meta-analysis. *Pharmacogenomics J* 2014; **14**:281-8.
- 64. Ferrell PB, McLeod HL. Carbamazepine, HLA-B\* 1502 and risk of Stevens-Johnson syndrome and toxic epidermal necrolysis: US FDA recommendations. *Pharmacogenomics* 2008: **9**:1543-6.
- 65. Roujeau JC, Kelly JP, Naldi L *et al*. Medication use and the risk of Stevens–Johnson syndrome or toxic epidermal necrolysis. *N Engl J Med* 1995; **333**:1600-8.
- 66. Hung SI, Chung WH, Liou LB *et al.* HLA-B\* 5801 allele as a genetic marker for severe cutaneous adverse reactions caused by allopurinol. Proc Natl Acad Sci USA 2005; **102**:4134-9.
- 67. Cheng L, Xiong Y, Qin C *et al.* HLA-B\* 58:01 is strongly associated with allopurinol-induced severe cutaneous adverse reactions in Han Chinese patients: a multicentre retrospective case—control clinical study. *Br J Dermatol* 2015; **173**:555-8.
- 68. Cao ZH, Wei ZY, Zhu QY *et al.* HLA-B\* 58:01 allele is associated with augmented risk for both mild and severe cutaneous adverse reactions induced by allopurinol in Han Chinese. *Pharmacogenomics* 2012; **13**:1193-201.
- 69. Chiu M, Hu M, Ng M *et al.* Association between HLA-B\* 58:01 allele and severe cutaneous adverse reactions with allopurinol in Han Chinese in Hong Kong. *Br J Dermatol* 2012; **167**:44-9.
- 70. Tassaneeyakul W, Jantararoungtong T, Chen P *et al.* Strong association between HLA-B\* 5801 and allopurinol-induced Stevens–Johnson syndrome and toxic epidermal necrolysis in a Thai population. *Pharmacogenetic Genomics* 2009; **19**:704-9.
- 71. Jung JW, Song WJ, Kim YS *et al.* HLA-B58 can help the clinical decision on starting allopurinol in patients with chronic renal insufficiency. *Nephrol Dial Transplant* 2011; **26**:3567-72.
- 72. Kang HR, Jee YK, Kim YS *et al.* Positive and negative associations of HLA class I alleles with allopurinol-induced SCARs in Koreans. *Pharmacogenet Genomics* 2011; **21**:303-7.
- 73. Lonjou C, Borot N, Sekula P *et al*. A European study of HLA-B in Stevens–Johnson syndrome and toxic epidermal necrolysis related to five high-risk drugs. *Pharmacogenet Genomics* 2008; **18**:99-107.
- 74. Gonçalo M, Coutinho I, Teixeira V *et al.* HLA-B\* 58:01 is a risk factor for allopurinol-induced DRESS and Stevens–Johnson syndrome/toxic epidermal necrolysis in a Portuguese population. *Br J Dermatol* 2013; **169**:660-5.
- 75. Ko TM, Tsai CY, Chen SY *et al*. Use of HLA-B\* 58:01 genotyping to prevent allopurinol induced severe cutaneous adverse reactions in Taiwan: national prospective cohort study. *Br*Med J 2015: **351**: 4848.

- 76. Dong D, Tan-Koi WC, Teng GG *et al*. Cost–effectiveness analysis of genotyping for HLA-B\* 5801 and an enhanced safety program in gout patients starting allopurinol in Singapore. *Pharmacogenomics* 2015; **16**:1781-93.
- 77. Cheng L, Zhang L, Gao L *et al.* Genotyping HLA-B\* 5801 for allopurinol-induced severe cutaneous adverse reactions: an accurate and prompt method. *Clin Transl Sci* 2015; **8**:834-6.
- 78. Trubiano JA, Aung AK, Nguyen M *et al*. A comparative analysis between antibiotic- and nonantibiotic-associated delayed cutaneous adverse drug reactions. *J Allergy Clin Immunol Pract* 2016; **4**: 1187-93.
- 79. Kardaun SH, Sekula P, Valeyrie-Allanore L *et al.* Drug reaction with eosinophilia and systemic symptoms (DRESS): an original multisystem adverse drug reaction. Results from the prospective RegiSCAR study. *Br J Dermatol* 2013; **169**:1071-80.
- 80. Cho YT, Liau JY, Chang CY *et al.* Co-existence of histopathological features is characteristic in drug reaction with eosinophilia and systemic symptoms and correlates with high grades of cutaneous abnormalities. *J Eur Acad Dermatol Venereol* 2016; **30**: 2077-84.
- 81. Yawalkar N, Pichler WJ. Immunohistology of drug-induced exanthema: clues to pathogenesis. *Curr Opin Allergy Clin Immunol* 2001; **1**:299-303.
- 82. Rive CM, Bourke J, Phillips EJ. Testing for drug hypersensitivity syndromes. *Clin Biochem Rev* 2013; **34**:15-38.
- 83. Brockow K, Garvey LH, Aberer W *et al.* Skin test concentrations for systemically administered drugs an ENDA/EAACI Drug Allergy Interest Group position paper. *Allergy* 2013: **68**:702-12.
- 84. Brockow K, Romano A, Blanca M *et al.* General considerations for skin test procedures in the diagnosis of drug hypersensitivity. *Allergy* 2002; **57**:45-51.
- 85. Pinho A, Coutinho I, Gameiro A *et al*. Patch testing a valuable tool for investigating non-immediate cutaneous adverse drug reactions to antibiotics. *J Eur Acad Dermatol Venereol* 2016; **31**: 280-7.
- 86. Barbaud A, Collet E, Milpied B *et al*. A multicentre study to determine the value and safety of drug patch tests for the three main classes of severe cutaneous adverse drug reactions. *Br J Dermatol* 2013; **168**:555-62.
- 87. Hassoun-Kheir N, Bergman R, Weltfriend S. The use of patch tests in the diagnosis of delayed hypersensitivity drug eruptions. *Int J Dermatol* 2016; **55**:1219-24.
- 88. Charfi O, Lakhoua G, Sahnoun R *et al.* DRESS syndrome following levofloxacin exposure with positive patch-test. *Therapie* 2015; **70**:547-9.
- 89. Fathallah N, Slim R, Rached S *et al*. Carbamazepine-induced DRESS with severe eosinophilia confirmed by positive patch test. *Dermatitis* 2014; **25**:282-4.
- 90. Santiago F, Goncalo M, Vieira R *et al.* Epicutaneous patch testing in drug hypersensitivity syndrome (DRESS). *Contact Dermatitis* 2010; **62**:47-53.

- 91. Tchen T, Reguiai Z, Vitry F *et al.* Usefulness of skin testing in cutaneous drug eruptions in routine practice. *Contact Dermatitis* 2009; **61**:138-44.
- 92. Lehloenya RJ, Todd G, Wallace J *et al.* Diagnostic patch testing following tuberculosis-associated cutaneous adverse drug reactions induces systemic reactions in HIV-infected persons. *Br J Dermatol* 2016; **175**:150-6.
- 93. Shebe K, Ngwanya MR, Gantsho N *et al.* Severe recurrence of drug rash with eosinophilia and systemic symptoms syndrome secondary to rifampicin patch testing in a human immunodeficiency virus-infected man. *Contact Dermatitis* 2014; **70**:125-7.
- 94. Bensaid B, Rozieres A, Nosbaum A *et al*. Amikacin-induced drug reaction with eosinophilia and systemic symptoms syndrome: delayed skin test and ELISPOT assay results allow the identification of the culprit drug. *J Allergy Clin Immunol* 2012; **130**:1413-4.
- 95. Giorgini S, Martinelli C, Tognetti L *et al*. Use of patch testing for the diagnosis of abacavir-related hypersensitivity reaction in HIV patients. *Dermatol Ther* 2011; **24**:591-4.
- 96. Lin YT, Chang YC, Hui RC *et al.* A patch testing and cross-sensitivity study of carbamazepine-induced severe cutaneous adverse drug reactions. *J Eur Acad Dermatol Venereol* 2013; **27**:356-64.
- 97. Demoly P, Adkinson NF, Brockow K *et al.* International Consensus on drug allergy. *Allergy* 2014; **69**:420-37.
- 98. Federico A, Aitella E, Sgambato D *et al.* Telaprevir may induce adverse cutaneous reactions by a T cell immune-mediated mechanism. *Ann Hepatol* 2015; **14**:420-4.
- 99. Cabanas R, Calderon O, Ramirez E *et al.* Piperacillin-induced DRESS: distinguishing features observed in a clinical and allergy study of 8 patients. *J Investig Allergol Clin Immunol* 2014; **24**:425-30.
- 100. Arruti N, Villarreal O, Bernedo N *et al.* Positive allergy study (intradermal, patch, and lymphocyte transformation tests) in a case of isoniazid-induced DRESS. *J Investig Allergol Clin Immunol* 2016; **26**:119-20.
- 101. Perrin-Lamarre A, Petitpain N, Trechot P *et al*. Glycopeptide-induced cutaneous adverse reaction: results of an immunoallergic investigation in eight patients. *Ann Dermatol Venereol* 2010; **137**:101-5.
- 102. Torres MJ, Sanchez-Sabate E, Alvarez J *et al*. Skin test evaluation in nonimmediate allergic reactions to penicillins. *Allergy* 2004; **59**:219-24.
- 103. Joint Task Force on Practice Parameters, American Academy of Allergy, Asthma and Immunology, American College of Allergy Asthma and Immunology et al. Drug allergy: an updated practice parameter. Ann Allergy Asthma Immunol 2010; 105:259-73.
- 104. Kano Y, Hirahara K, Mitsuyama Y *et al.* Utility of the lymphocyte transformation test in the diagnosis of drug sensitivity: dependence on its timing and the type of drug eruption. *Allergy* 2007; **62**:1439-44.

- 105. Nagao-Dias AT, Teixeira FM, Coelho HL. Diagnosing immune-mediated reactions to drugs. *Allergol Immunopathol (Madr)* 2009; **37**:98-104.
- 106. Fu M, Gao Y, Pan Y *et al.* Recovered patients with Stevens-Johson syndrome and toxic epidermal necrolysis maintain long-lived IFN-gamma and sFasL memory response. *PLoS One* 2012; **7**:e45516.
- 107. Jurado-Palomo J, Cabanas R, Prior N *et al*. Use of the lymphocyte transformation test in the diagnosis of DRESS syndrome induced by ceftriaxone and piperacillin-tazobactam: two case reports. *J Investig Allergol Clin Immunol* 2010; **20**:433-6.
- 108. Gomez E, Torres MJ, Mayorga C *et al.* Immunologic evaluation of drug allergy. *Allergy Asthma Immunol Res* 2012; **4**:251-63.
- 109. Torres MJ, Mayorga C, Blanca M. Nonimmediate allergic reactions induced by drugs: pathogenesis and diagnostic tests. *J Investig Allergol Clin Immunol* 2009; **19**:80-90.
- 110. Beeler A, Zaccaria L, Kawabata T *et al.* CD69 upregulation on T cells as an in vitro marker for delayed-type drug hypersensitivity. *Allergy* 2008; **63**:181-8.
- 111. Britschgi M, Pichler WJ. Acute generalized exanthematous pustulosis, a clue to neutrophil-mediated inflammatory processes orchestrated by T cells. *Curr Opin Allergy Clin Immunol* 2002; **2**:325-31.
- 112. Dias de Castro E, Leblanc A, Sarmento A *et al.* An unusual case of delayed-type hypersensitivity to ceftriaxone and meropenem. *Eur Ann Allergy Clin Immunol* 2015; **47**:225-7.
- 113. El-Ghaiesh S, Monshi MM, Whitaker P *et al*. Characterization of the antigen specificity of T-cell clones from piperacillin-hypersensitive patients with cystic fibrosis. *J Pharmacol Exp Ther* 2012; **341**:597-610.
- 114. Kardaun SH, de Monchy JG. Acute generalized exanthematous pustulosis caused by morphine, confirmed by positive patch test and lymphocyte transformation test. *J Am Acad Dermatol* 2006; **55**(Suppl. 2):S21-3.
- 115. Ogasawara K, Tomitsuka N, Kobayashi M *et al*. Stevens-Johnson syndrome associated with intravenous acetazolamide administration for evaluation of cerebrovascular reactivity. Case report. *Neurol Med Chir* (*Tokyo*) 2006; **46**:161-3.
- 116. Kanny G, Pichler W, Morisset M *et al.* T cell-mediated reactions to iodinated contrast media: evaluation by skin and lymphocyte activation tests. *J Allergy Clin Immunol* 2005; **115**:179-85.
- 117. Romano A, Torres MJ, Di Fonso M, *et al.* Delayed hypersensitivity to cefazolin: report on a case involving lymphocyte transformation studies with different cephalosporins. *Ann Allergy Asthma Immunol* 2001; **87**:238-42.

- 118. Keane NM, Pavlos RK, McKinnon E *et al.* HLA Class I restricted CD8+ and Class II restricted CD4+ T cells are implicated in the pathogenesis of nevirapine hypersensitivity. *AIDS* 2014; **28**:1891-901.
- 119. Keane NM, Roberts SG, Almeida CA *et al*. High-avidity, high-IFNgamma-producing CD8 T-cell responses following immune selection during HIV-1 infection. *Immunol Cell Biol* 2012; **90**:224-34.
- 120. Esser S, Jablonka R, Heinemann FM *et al.* Detection of abacavir hypersensitivity by ELISpot method. *Inflamm Allergy Drug Targets* 2012; **11**:227-34.
- 121. Lucas A, Lucas M, Strhyn A *et al.* Abacavir-reactive memory T cells are present in drug naive individuals. *PLoS One* 2015; **10**:e0117160.
- 122. Rozieres A, Hennino A, Rodet K *et al.* Detection and quantification of drug-specific T cells in penicillin allergy. *Allergy* 2009; **64**:534-42.
- 123. Khalil G, El-Sabban M, Al-Ghadban S *et al.* Cytokine expression profile of sensitized human T lymphocytes following in vitro stimulation with amoxicillin. *Eur Cytokine Netw* 2008; **19**:131-41.
- 124. Scheibenbogen C, Romero P, Rivoltini L *et al*. Quantitation of antigen-reactive T cells in peripheral blood by IFNgamma-ELISPOT assay and chromium-release assay: a four-centre comparative trial. *J Immunol Methods* 2000; **244**:81-9.
- 125. Porebski G, Pecaric-Petkovic T, Groux-Keller M *et al.* In vitro drug causality assessment in Stevens-Johnson syndrome alternatives for lymphocyte transformation test. *Clin Exp Allergy* 2013; **43**:1027-37.
- 126. Yun J, Marcaida MJ, Eriksson KK *et al.* Oxypurinol directly and immediately activates the drug-specific T cells via the preferential use of HLA-B\*58:01. *J Immunol* 2014; **192**:2984-93.
- 127. Yun J, Mattsson J, Schnyder K *et al.* Allopurinol hypersensitivity is primarily mediated by dose-dependent oxypurinol-specific T cell response. *Clin Exp Allergy* 2013; **43**:1246-55.
- 128. Schrijvers R, Gilissen L, Chiriac AM *et al.* Pathogenesis and diagnosis of delayed-type drug hypersensitivity reactions, from bedside to bench and back. *Clin Transl Allergy* 2015; **5**:31.
- 129. Elzagallaai AA, Knowles SR, Rieder MJ *et al.* In vitro testing for the diagnosis of anticonvulsant hypersensitivity syndrome: a systematic review. *Mol Diagn Ther* 2009; **13**:313-30.
- 130. Tang YH, Mockenhaupt M, Henry A *et al.* Poor relevance of a lymphocyte proliferation assay in lamotrigine-induced Stevens-Johnson syndrome or toxic epidermal necrolysis. *Clin Exp Allergy* 2012; **42**:248-54.
- 131. Polak ME, Belgi G, McGuire C *et al*. In vitro diagnostic assays are effective during the acute phase of delayed-type drug hypersensitivity reactions. *Br J Dermatol* 2013; **168**:539-49.
- 132. Haw WY, Polak ME, McGuire C *et al*. In vitro rapid diagnostic tests for severe drug hypersensitivity reactions in children. *Ann Allergy Asthma Immunol* 2016; **117**:61-6.

- 133. Mizukawa Y, Yamazaki Y, Teraki Y *et al*. Direct evidence for interferon-gamma production by effector-memory-type intraepidermal T cells residing at an effector site of immunopathology in fixed drug eruption. *Am J Pathol* 2002; **161**:1337-47.
- 134. Mayorga C, Sanz ML, Gamboa P *et al*. In vitro methods for diagnosing nonimmediate hypersensitivity reactions to drugs. *J Investig Allergol Clin Immunol* 2013; **23**:213-25.
- 135. Almeida CA, Martin AM, Nolan D *et al*. Cytokine profiling in abacavir hypersensitivity patients. *Antivir Ther* 2008; **13**:281-8.
- 136. Nishio D, Izu K, Kabashima K *et al*. T cell populations propagating in the peripheral blood of patients with drug eruptions. *J Dermatol Sci* 2007; **48**:25-33.
- 137. Romano A, Gaeta F, Valluzzi RL *et al*. Cross-reactivity and tolerability of aztreonam and cephalosporins in subjects with a T cell–mediated hypersensitivity to penicillins. *J Allergy Clin Immunol* 2016; **138**:179-86.
- 138. Buonomo A, Nucera E, Pecora V *et al*. Cross-reactivity and tolerability of cephalosporins in patients with cell-mediated allergy to penicillins. *J Investig Allergol Clin Immunol* 2014; **24**:331-7.
- 139. Schiavino D, Nucera E, Lombardo C *et al.* Cross-reactivity and tolerability of imipenem in patients with delayed-type, cell-mediated hypersensitivity to β-lactams. *Allergy* 2009; **64**:1644-8.
- 140. Romano A, Gaeta F, Valluzzi R *et al.* Absence of cross-reactivity to carbapenems in patients with delayed hypersensitivity to penicillins. *Allergy* 2013; **68**:1618-21.
- 141. Romano A, Viola M, Guéant-Rodriguez RM *et al.* Imipenem in patients with immediate hypersensitivity to penicillins. *N Engl J Med* 2006; **354**:2835-7.
- 142. Atanasković-Marković M, Gaeta F, Medjo B *et al*. Tolerability of meropenem in children with IgE-mediated hypersensitivity to penicillins. *Allergy* 2008; **63**:237-40.
- 143. Buonomo A, Nucera E, De Pasquale T *et al*. Tolerability of aztreonam in patients with cell-mediated allergy to β-lactams. *Int Arch Allergy Immunol* 2010; **155**:155-9.
- 144. Aihara M. Pharmacogenetics of cutaneous adverse drug reactions. *J Dermatol* 2011; **38**:246-54.
- 145. Romano A, Pettinato R, Andriolo M *et al.* Hypersensitivity to aromatic anticonvulsants: in vivo and in vitro cross-reactivity studies. *Curr Pharm Des* 2006; **12**:3373-81.
- 146. Krivoy N, Taer M, Neuman MG. Antiepileptic drug-induced hypersensitivity syndrome reactions. *Curr Drug Saf* 2006; **1**:289-99.
- 147. Chung WH, Chang WC, Lee YS *et al.* Genetic variants associated with phenytoin-related severe cutaneous adverse reactions. *JAMA* 2014; **312**:525-34.
- 148. Sierra NM, Garcia B, Marco J *et al*. Cross hypersensitivity syndrome between phenytoin and carbamazepine. *Pharm World Sci* 2005; **27**:170-4.

- 149. Wang X, Lang S, Shi X *et al.* Cross-reactivity of skin rashes with current antiepileptic drugs in Chinese population. *Seizure* 2010; **19**:562-6.
- 150. Seitz CS, Pfeuffer P, Raith P *et al*. Anticonvulsant hypersensitivity syndrome: cross-reactivity with tricyclic antidepressant agents. *Ann Allergy Asthma Immunol* 2006; **97**:698-702.
- 151. Man CB, Kwan P, Baum L *et al.* Association between HLA-B\*1502 allele and antiepileptic drug-induced cutaneous reactions in Han Chinese. *Epilepsia* 2007; **48**:1015-8.
- 152. He N, Min FL, Shi YW *et al.* Cutaneous reactions induced by oxcarbazepine in Southern Han Chinese: incidence, features, risk factors and relation to HLA-B alleles. *Seizure* 2012; **21**:614-8.
- 153. Yang CY, Dao RL, Lee TJ *et al*. Severe cutaneous adverse reactions to antiepileptic drugs in Asians. *Neurology* 2011; **77**:2025-33.
- 154. Alvestad S, Lydersen S, Brodtkorb E. Cross-reactivity pattern of rash from current aromatic antiepileptic drugs. *Epilepsy Res* 2008; **80**:194-200.
- 155. Viard I, Wehrli P, Bullani R *et al.* Inhibition of toxic epidermal necrolysis by blockade of CD95 with human intravenous immunoglobulin. *Science* 1998; **282**:490-3.
- 156. Mockenhaupt M. Epidemiology of cutaneous adverse drug reactions. *Chem Immunol Allergy* 2012; **97**:1-17.
- 157. Roujeau JC. Stevens-Johnson syndrome and toxic epidermal necrolysis are severity variants of the same disease which differs from erythema multiforme. *J Dermatol* 1997; **24**:726-9.
- 158. Schwartz RA, McDonough PH, Lee BW. Toxic epidermal necrolysis: Part I. Introduction, history, classification, clinical features, systemic manifestations, etiology, and immunopathogenesis. *J Am Acad Dermatol* 2013; **69**:173.e1-13.
- 159. Mockenhaupt M. The current understanding of Stevens-Johnson syndrome and toxic epidermal necrolysis. *Expert Rev Clin Immunol* 2011; **7**:803-13.
- 160. Walsh S, Diaz-Cano S, Higgins E *et al.* Drug reaction with eosinophilia and systemic symptoms: is cutaneous phenotype a prognostic marker for outcome? A review of clinicopathological features of 27 cases. *Br J Dermatol* 2013; **168**:391-401.
- 161. Ortonne N, Valeyrie-Allanore L, Bastuji-Garin S *et al.* Histopathology of drug rash with eosinophilia and systemic symptoms syndrome: a morphological and phenotypical study. *Br J Dermatol* 2015; **173**:50-8.
- 162. Szatkowski J, Schwartz RA. Acute generalized exanthematous pustulosis (AGEP): A review and update. *J Am Acad Dermatol* 2015; **73**:843-8.
- 163. Fernando SL. Acute generalised exanthematous pustulosis. *Australas J Dermatol* 2012; **53**:87-92.
- 164. Thienvibul C, Vachiramon V, Chanprapaph K. Five-year retrospective review of acute generalized exanthematous pustulosis. *Dermatol Res Pract* 2015; **2015**:260928.

- 165. Speeckaert MM, Speeckaert R, Lambert J *et al.* Acute generalized exanthematous pustulosis: an overview of the clinical, immunological and diagnostic concepts. *Eur J Dermatol* 2010; **20**:425-33.
- 166. Halevy S. Acute generalized exanthematous pustulosis. *Curr Opin Allergy Clin Immunol* 2009; **9**:322-8.
- 167. Amstutz U, Ross CJ, Castro-Pastrana LI *et al.* HLA-A\* 31:01 and HLA-B\* 15:02 as Genetic Markers for Carbamazepine Hypersensitivity in Children. *Clin Pharmacol Ther* 2013; **94**:142-9.
- 168. Cheung YK, Cheng SH, Chan EJ *et al.* HLA-B alleles associated with severe cutaneous reactions to antiepileptic drugs in Han Chinese. *Epilepsia* 2013; **54**:1307-14.
- 169. Khanna D, Khanna PP, FitzGerald JD *et al.* 2012 American College of Rheumatology Guidelines for Management of Gout Part II: Therapy and Anti-inflammatory Prophylaxis of Acute Gouty Arthritis. *Arthritis Care Res* 2012; **64**:1447-61.
- 170. Arimone Y, Begaud B, Miremont-Salame G *et al.* Agreement of expert judgment in causality assessment of adverse drug reactions. *Eur J Clin Pharmacol* 2005; **61**:169-73.
- 171. Hayashi PH. Drug-Induced Liver Injury Network Causality Assessment: Criteria and Experience in the United States. *Int J Mol Sci* 2016; **17**:201.
- 172. Agbabiaka TB, Savovic J, Ernst E. Methods for causality assessment of adverse drug reactions: a systematic review. *Drug Saf* 2008; **31**:21-37.
- 173. Naranjo CA, Busto U, Sellers EM *et al*. A method for estimating the probability of adverse drug reactions. *Clin Pharmacol Ther* 1981; **30**:239-45.
- 174. Trubiano J, Phillips E. Antimicrobial stewardship's new weapon? A review of antibiotic allergy and pathways to 'de-labeling'. Curr Opin Infect Dis 2013; **26**:526-37.
- 175. Pavlos R, Mallal S, Ostrov D *et al*. T cell-mediated hypersensitivity reactions to drugs. *Annu Rev Med* 2015; **66**:439-54.

Table 1. Summary of the clinical manifestations and histopathological features of SCAR syndromes.

SCAR	Effector	Clinical	Investigation	Histopathologi	Latency	Common
syndrome	mechanisms	manifestations	findings	cal features	period	causal drugs

SJS/TEN	CD8+	SJS and TEN are a	Abnormal liver,	Subepidermal	1-4	Carbamazepine
555/1121	cytotoxic T	disease continuum; the	renal and	blister; spectrum	weeks. 156	Phenytoin
	lymphocyte	differentiation is based	respiratory	of changes	,159	Lamotrigine
	mediated Fas-		function.			
		upon the percentage of		ranging from		Allopurinol
	FasL and	body surface area of	Haematological,	lichenoid		Nevirapine
	granulysin-	skin detachment. 156-158	metabolic, fluid &	reaction pattern		NSAID*
	mediated	Acute onset of blisters	electrolyte	with apoptotic		Sulfonamides
	apoptosis. <sup>4,8,155</sup>	and erosions affecting	complications.	keratinocytes,		Sulfasalazine <sup>159</sup>
	- , -	the skin, and mucous		partial to full		
		membranes; often		thickness		
		associated severe		epidermal		
		systemic complications		necrosis. <sup>158</sup>		
		with significant				
	0)	morbidity and long-				
		term sequelae. 159				
DRESS	T-cell	Clinical presentation is	Haematological	Multiple	2-6	Carbamazepine
	mediated	heterogeneous:	abnormalities,	histological	weeks. <sup>79</sup>	Phenytoin
	perforin-	widespread	most commonly	patterns		Lamotrigine
	granzyme B as	exanthematous	eosinophilia and	including:		Allopurinol
	well as Fas/Fas	eruption, facial	atypical	interface		Sulfonamides
	L-dependent	oedema, fever and	lymphocytes.	reaction,		Vancomycin
	cell death. <sup>7,160</sup>	lymphadenopathy. 18,160,	Abnormal liver,	apoptotic		Minocycline
		161	renal, respiratory	keratinocytes,		Amoxicillin <sup>79</sup>
		High variability in	and other organ	parakeratosis,		
	-	disease severity; some	function. 18,160,161	spongiosis. 161		
		patients have modest				
		systemic symptoms,				
		while others develop				
		significant morbidity				
		due to internal				
		involvement. <sup>160</sup>				
AGEP	Activation and	Acute onset of	Neutrophilia +/-	Spongiform	1–5	Amoxicillin
AGEF		widespread non-	_	subcorneal	days. 163	Quinolones
	proliferation of	•	eosinophilia, abnormal		days.	
	specific CD4 and CD8 T-	follicular sterile	renal/liver	and/or intradermal		Sulfonamides Terbinafine
		pustules overlying				
	cells,	erythematous	function,	pustules with		Hydroxychloroq
	perforin/granzy	oedematous skin,	hypocalcaemia. <sup>162</sup>	marked oedema		uine
	me B and Fas	starting in the	-3.	of the papillary		Diltiazem <sup>166</sup>
	ligand	intertriginous areas,		dermis and		

mechanis	ms to often associated with	polymorphous	
induce	fever. <sup>162-164</sup>	perivascular	
apoptosis	. 162	infiltrate. 165	
+			

AGEP: acute generalised exanthematous pustulosis, CD: cluster of differentiation, DRESS: drug rash with eosinophilia and systemic symptoms, NSAID: Non-steroidal anti-inflammatory drug, SJS: Stevens-Johnson syndrome, TEN: toxic epidermal necrolysis.

Table 2. The rapeutic recommendations where evidence exists for strong HLA associations for various adverse drug reaction phenotypes $^{\dagger}$ .

Medications	HLA	Phenotype	Populations		Selected
_			studied	Therapeutic	references
				recommendation	
Abacavir	HLA-B*57:01	HSS	All	HLA-B*57:01 testing prior to abacavir prescription and avoid abacavir use in HLA- B*57:01 positive individuals	41,42,45-48
Carbamazepine	HLA-B*15:02 <sup>§</sup>	SJS/TEN	Han Chinese (China, Hong Kong, Taiwan), Thai, Malaysian, Indian (South Asians)	Avoid carbamazepine in all HLA-B*15:02 positive individuals <sup>††</sup> Screening currently recommended for at risk populations (Han Chinese, southeast and south Asians) or unknown ethnicity	51,58,64,167,168
Carbamazepine	HLA-A*31:01	DRESS/HSS >SJS/TEN	Han Chinese, Japanese, Korean, Caucasian	If alternative therapeutic agent exists, avoid carbamazepine in all carbamazepine naïve HLA-A*31:01 positive individuals	51,58,63 52,55,61 ,62,64

Allopurinol	HLA-B*58:01	DRESS/HSS	Han Chinese	Avoid allopurinol use in	54,66-74
		and SJS/TEN	(China and	the setting of allopurinol	
			Hong Kong),	naïve HLA-B*58:01	
			Thai, Korean,	posivite individuals	
			Japanese,	Widespread guidelines	
			European	for screening prior to use	
				have not been issued	

SCAR: severe cutaneous adverse reactions, HLA: human leucocyte antigen, HSS: hypersensitivity syndrome, SJS: Stevens-Johnson syndrome, TEN: toxic epidermal necrolysis, DRESS: drug reaction with eosinophilia and systemic symptoms.

Table 3. Three major approaches to drug causality assessment in severe cutaneous adverse drug reactions,

Method	Description	Strengths	Weaknesses	Selected
				references
Global	Inference of causality by	Consensus opinion by	Subjective, influenced	5,170,171
introspection	expert clinical judgement.	a group of experts.	by the experience,	
		Often serves as the	knowledge and biases of	
		gold standard in	the assessor(s).	
		causality assessment.	Poor reproducibility.	

<sup>&</sup>lt;sup>†</sup> For any individual carrying an HLA risk allele, if they have already tolerated the drug for  $\geq 12$  continuous weeks currently or in the past, then it is safe for them to continue the drug or for the drug to be reinstituted in the future.

Sample SJS/TEN is also associated with other B75 serotype HLA alleles such as HLA-B\*15:21, B\*15:08, B\*15:11 and potentially B\*15:30 and B\*15:31, therefore additional caution should be exerted for carbamazepine use if these HLA types are identified.

<sup>&</sup>lt;sup>††</sup>Although a much weaker association exists between HLA-B\*15:02 and other aromatic amine anticonvulsants, such as oxcarbamazepine, eslicarbamzepine, lamotrigine, phenytoin and fosphenytoin, consideration should be given to choosing an alternative non-aromatic anticonvulsant in the case of identified HLA-B\*15:02+.

The American College of Rheumatology Guidelines for Management of Gout (2012) have recommended HLA-B\*58:01 testing prior to allopurinol prescription in specific populations including 1) those of increased risk (Southeast Asian) and 2) Subpopulations with increase risk based on advanced chronic renal failure (stage 3). 169

Bayesian	Uses clinical and	Allows simultaneous	Time consuming and	172
approach	epidemiological data to	assessment of multiple	highly technical.	
	transform a prior into a	causes.		
	posterior probability.	Previous knowledge of		
		the culprit drug profile		
7		is not required.		
Drug	Collection of specific data	Structured and	Clinical utility may be	5,172
causality	points followed by	standardised method.	limited in cases where	
algorithms	problem solving		more than one drug is	
(see A & B)	operations resulting in an	Reproducible and	administered.	
(SCC II & D)	objective assessment of	transparent.		
	probability.		Clinical judgement may	
	productify.		be required at various	
-			stages.	
-			Some algorithms may	
			not be able to identify	
2	_		novel ADRs or first	
	R		cases of ADRs.*	
(A) Naranjo	Consists of 10 questions	Well-validated.	Classifies >90% of	173
Scale	and yields a final	Widely used and	suspected adverse drug	
	assessment of causality	quick/simple tool.	reactions as 'possible.'	
	as: 'definite', 'probable',		Does not take into	
	'possible' or 'doubtful'		account drug-drug	
2	that a drug administered		interactions.	
	in therapeutic doses			
	caused an adverse event.			
(B) ALDEN	Specific algorithm for	Developed by experts	Only validated for	5
	assessing drug causality	in SJS/TEN.	SJS/TEN.	
+	in SJS and TEN.	Validated on cases		
_	The final assessment of	enrolled in the		
_	causality is expressed as	EuroSCAR study in a		
	'very probable'.	case-control analysis.		
	'probable', 'possible',			
	'unlikely' or 'very			
	unlikely.'			
L	drug reaction ALDEN, Algor			<u> </u>

ADR: Adverse drug reaction, ALDEN: Algorithm for drug causality for epidermal necrolysis, SJS:

Stevens-Johnson syndrome, TEN: toxic epidermal necrolysis.

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Table 4. Beta-lactams with similar R1 side-chain structures (adapted from Trubiano et al. 174)

Penicillin G	Amoxicillin	Ampicillin	Ceftriaxone	Cefoxitin	Ceftamandole	Ceftazidime
Cephaloridine	Cefadroxil	Cefaclor	Cefotaxime	Cephaloridine	Cefonicid	Aztreonam
Cephalothin	Cefprozil	Cephalexin	Cefpodoxime	Cephalothin		
Cefoxitin	Cefatrizine	Cephadrine	Cefditoren			
	Cephalexin	Cephaloglycin	Ceftizoxime			
	"	Loracarbef	Cefmenoxime			
	7		Cefepime			

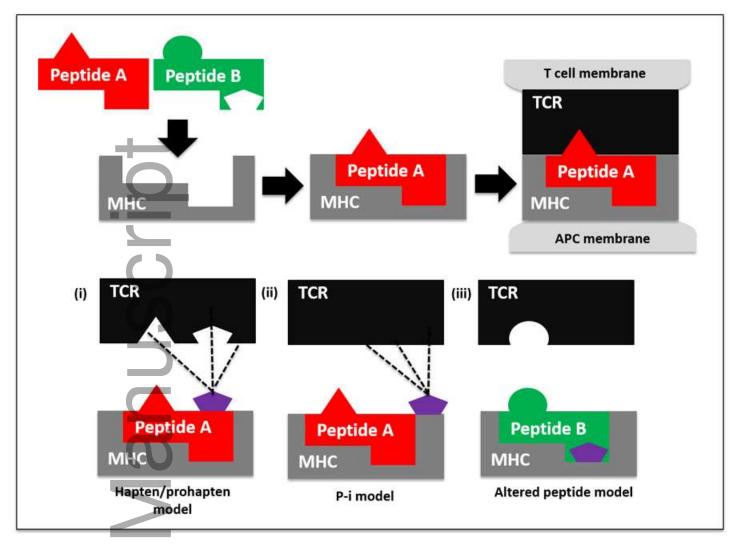
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**Figure 1.** Proposed models of T-cell receptor (TCR), major histocompatability complex (MHC), drug interactions: In the **hapten/prohapten model (i)** a drug (e.g., penicillin) binds covalently to an endogenous peptide (e.g., albumin), forming a new molecule. Antigen presenting cells process and present it as short peptide fragments within the MHC binding cleft, some of which (peptide A) include drug epitopes (purple pentagon). If recognized by a TCR, a drug-specific immune response can ensue. In the **pharmacological-interaction (P-I) model (ii)** the drug binds non-covalently to certain MHC molecules or TCRs, stimulating specific TCR and thus generating drug-reactive T-cells. In the **altered peptide repertoire model (iii)** a drug (e.g., abacavir) binds non-covalently to the binding pocket of a MHC molecule (e.g. HLA-B\*57:01), altering its conformation and allowing a new array of self-peptides (peptide B) to stably occupy it and stimulate T-cells. This can lead to drug-induced activation of autoimmunity (e.g., abacavir hypersensitivity reaction.) Adapted from Pavlos *et al.* <sup>175</sup>

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**Figure 2.** Basic structures of beta-lactams (adapted from Trubiano *et al.*<sup>174</sup>). R denotes side chains. Cephalosporins have two side chains, R1 and R2. However, R2 is lost during hydrolysis.

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