

**The association between blood copper concentration and biomarkers related to cardiovascular disease  
risk – analysis of 206 individuals in the Northern Finland Birth Cohort 1966**

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**Short title:** Copper and biomarkers of cardiovascular disease risk

44    **Abstract**

45    **Background:** Copper is an abundant trace element in humans where alterations in the circulating concentration  
46    could inform on chronic disease aetiology. To date, data are lacking to study how copper may associate with  
47    cardiovascular disease (CVD) risk factors in young and healthy population. Molecular evidence suggests an  
48    important role of copper in liver metabolism, an essential organ in maintaining cardiovascular health and  
49    inflammation, therefore supporting copper as an associated biomarker of the risk.

50    **Objective:** We performed a cross-sectional analysis to examine the possible associations between blood  
51    copper levels and risk factors for CVD and pre-inflammatory process.

52    **Design:** The data has been collected from a sub-sample set of the Northern Finland Birth Cohort 1966  
53    (NFBC1966) at 31 years.

54    **Participants:** The study included 206 individuals, 116 men and 90 women. To reduce environmental  
55    individual variations affecting both copper and the metabolic profile in the study sample, the participants were  
56    selected as: i) being born in Finnish Lapland and ii) living in their birth place for the last five years preceding  
57    blood sampling.

58    **Main outcome measures:** Fasting blood copper concentration was measured by inductively coupled plasma  
59    mass spectrometer. The CVD risk factors included 6 metabolic clusters (30 cardiovascular and pro-  
60    inflammatory factors) assessed by nuclear magnetic resonance. Multivariate linear regression analysis was  
61    performed to test the linear association between blood copper and 6 metabolic clusters for CVD risk.  
62    Associations were assessed under correction for multiple testing.

63    **Results:** Copper (Cu) levels were comparable in men and women, with no difference between sexes (*p-value*  
64    <0.60). In multiple regression models, sex adjusted, copper was associated with 9 metabolites from 4 metabolic  
65    clusters. After adjustment with BMI, copper was associated with 4 metabolites from 3 metabolic clusters:  
66    glutamine, beta-hydroxybutyrate, alpha-1-acid glycoprotein (AGP) and high-sensitive C-reactive protein (hs-  
67    CRP). After correction for multiple testing, Cu was found positively associated with only 2 biomarkers of  
68    inflammation including AGP [*p*=0.04] and hs-CRP [*p*=0.0001].

69    **Conclusions:** Considering the strength and limitation of the study design, the present study does not support  
70    evidence for an independent role of copper on biomarkers for CVD risk. Nevertheless, we are reporting a  
71    robust association of copper with the inflammatory load that is important to consider in light with the

inflammatory component of chronic health. In addition, the association of copper with metabolites may be attributable to BMI or environmental factors associated to it, and warrants further research in large population samples.

**Keywords:** Copper, metabolic profile, young adult, cardiovascular disease risk, metabolomics, inflammatory biomarkers

## Introduction

Cardiovascular disorders and diseases are a major cause of death where scientific efforts are needed to identify early risk factors and potential preventive biomarkers in non-affected populations. After Iron and Zinc, Copper (Cu) is the third most important trace element circulating in the body [1,2] originating principally from dietary sources *i.e.* food and water [2].  $\text{Cu}^{++}$  can be extremely toxic due to its oxidoreductase function [3,4] and is transported in the circulation by the carrier-protein ceruloplasmin [5]. There is contrasting evidence for the influence of Cu on pro- or anti-oxidative pathways and subsequent risk of atherosclerosis in humans [6]. To date Cu plays a role in nineteen enzymatic reactions involved in oxidative stress and metabolic homeostasis [4] including the Cu/Zn dependent superoxide dismutase (SOD), the cytochrome *c* oxidase and the ceruloplasmin [4]. In addition, there is no clinical threshold defining a healthy range of fasting blood copper. While modulation of dietary intake of copper could be implemented as a strategy to influence cardiometabolic health [7,8], fundamental questions remains to be solved before clear recommendation can be announced. A wider knowledge of the metabolic pathways linking copper to altered cardiovascular health can be needed. Following the advent of wider metabolomic profiling [9] we are gaining a dimension in the molecular pathways involved aside from the classical markers of cardiovascular disease especially LDL-C, triglycerides and HDL-C [10]. This includes variations in lipoprotein structure and composition, some amino-acids, ketone bodies and pro-inflammatory factors [11]. To date 30 metabolites are suggested to influence cardiovascular health and might be influenced by copper concentration of copper metabolism related pathways [10].

In line with the biological functions of copper and the current bodies of evidence, the present study aimed at testing the hypothesis, in a non-affected sample of young and healthy adults at 31 years, that fasting blood copper is associated with CVD risk biomarkers as defined from previous large scale metabolomic study [10]. Identifying potential new biomarkers such as copper might help detecting very early disease state with the aim

100 to also inform on the molecular pathway involved. The objectives of the present study were therefore to  
101 characterise blood concentration in a young and healthy non-affected sample with cardiovascular disease and  
102 test the linear association of copper with known risk factors established from the study of the metabolome and  
103 inflammatory load related to the etiology of chronic cardio-metabolic diseases.

104

105 **Materials and Methods**

106 *Study design and material*

107 The study was based on the 31-year follow-up of the Northern Finland Birth Cohort (NFBC1966) [12]. Briefly  
108 the NFBC1966 is a prospective birth cohort of 12,058 (96% of total population) live-born individuals in the  
109 province of Oulu (65 degree North) and Lapland in year 1966 [12,13]. The inclusion criteria for the present  
110 study were i) being born in Finnish Lapland and ii) living in their birthplace for the last five years preceding  
111 fasting blood sampling and iii) complete case analysis. Due to economic constraint, copper was measured only  
112 in 126 males and 122 females. The selection criteria aimed at reducing environmental variations known to  
113 affect both copper and the metabolic profile. We excluded pregnant women (N=6), oral contraceptive users  
114 (N=16), and hs-CRP>10 mg/L (N=6) (acute inflammation) as these factors are associated with variation in the  
115 metabolic profile [14]. The complete case analysis data consisted of N=206 subjects (N=20 subjects missing  
116 with one or more of the covariates and metabolomics data examined). The anthropometric, socio-demographic  
117 and lifestyle characteristics of the population are described in Table 1 (see [15] for detailed protocols). All  
118 participants gave written informed consents and the Ethics Committee of Northern Ostrobothnia hospital  
119 district and the University of Oulu (Finland) approved the study.

120 This study was performed on a healthy adult population at 31 years with overnight fasted blood samples (~7-  
121 8 hours). The overnight fasted state was chosen to reduce inter-individual variability on the potential outcomes  
122 also limiting the influence on the oxidative phosphorylation/fatty-acid oxidation with the synthesis of ketone  
123 body or any circulating metabolite examined in the study. The alcohol consumption of the participants  
124 described in the **Table 1** was obtained from the postal questionnaire data on the frequency of consumption of  
125 various alcoholic beverages during the previous six months at 31 years [15]. Participant's invited for the  
126 clinical examination was requested to abstain from alcohol consumption before blood sampling. There was no  
127 difference in copper concentrations with the categories of alcohol consumption examined in our study (*p*-

128 value=0.46). In addition, participants included in the study have not reported history of liver disease or any  
129 other metabolic diseases.

130

### 131 *Biochemical analyses*

132 The present measure of Cu concentration was performed in total blood as plasma or serum Cu concentrations  
133 are considered less precise indicators for the total body copper status [16], where previous report failed to  
134 observe changes in plasma Cu or serum Cu concentrations with Cu supplementation or Cu intake [16].

135 12-hour overnight fasted blood samples were collected in metal-free tubes following an immediate freezing in  
136 -80°C for further analyses. An aliquot of 0.3 mL of blood sample was diluted ten times with an alkaline solution  
137 containing Triton X-100 and ethylenediaminetetraacetic acid disodium salt dehydrate (EDTA) [17]. For  
138 calibration the standard addition procedure was performed. Measurement of copper in prepared solutions were  
139 made by an Octapole Reaction System (ORS) Inductively Coupled Plasma Mass Spectrometer (ICP-MS,  
140 7500ce, Agilent) equipped with an ASX-510 Autosampler (Cetac). Instrumental conditions: Babington  
141 nebulizer, Scott-type spray chamber, spray chamber temperature 5 °C, plasma gas flow rate 15 L/min, carrier  
142 gas flow rate 0.8 L/min, make-up gas flow rate 0.1 L/min, RF power 1500 W, reaction cell gas helium 4,3  
143 mL/min, isotope monitored <sup>65</sup>Cu. Analytical precision was 5%. The accuracy of results was checked by  
144 analysing Seronorm Trace Elements Whole Blood L-1 (SERO AS, Norway) as a reference material. The  
145 measured values were in good agreement with reference values.

### 146 *Inflammatory biomarkers*

147 Serum high-sensitivity C-reactive protein (hs-CRP, mg/L) concentration was measured by  
148 immunoenzymometric assay (Medix Biochemica, Espoo, Finland). The intra- and inter-assay coefficients of  
149 variation (CV) were 4.2% and 5.2%.

150 Active plasminogen activator inhibitor-1 (active PAI-1, ng/ml) concentration was measured by multiplex array  
151 method using non-magnetic human CVD/cytokine panel (SPR349, Merck Millipore, USA) according to  
152 manufacturer's instructions. The intra- and inter-assay CV were 6.8% and 18.6%.

153 Alpha-1-acid glycoprotein (AGP) and other metabolite measurements: Alpha-1-acid glycoprotein (mmol/L)  
154 was extracted along with a group of 123 metabolites in NFBC1966. The quantification of the metabolomics  
155 biomarkers including AGP was performed by Proton Nuclear Magnetic Resonance (NMR) as described in

156 Würtz *et al.*, (2015) [10]. The metabolomics platform is based on three molecular windows (lipoprotein, low-  
157 molecular-weight metabolite data and lipid) for each sample [10].

158 *Grouping of the 6 clusters of biomarkers:* The circulating metabolites were reported as strong predictors of  
159 CVD risk as the conventional lipid risk factors, and were markers of CVD onset during more than a decade  
160 follow-up [10]. In addition, the biomarker associations were replicated in independent cohorts with varying  
161 baseline characteristics including age and ethnicity, and were consistent across different metabolite profiling  
162 platforms [10]. Therefore, to reduce the number of hypotheses tested and to account for the small sample size  
163 of the current study we chose to select only the sentinel metabolites from the six metabolic clusters available  
164 by the NMR metabolic methods (see supplementary figure from the article Würtz *et al.*, (2015, figure 2) [10]).  
165 In total 30 metabolites showing significant association with cardiovascular events were selected [10]. The  
166 metabolites and inflammatory biomarkers were categorised under 6 clusters as reported in Würtz *et al.*, (2015,  
167 supplementary figure 2 from the article [10]). Cluster 1, amino acids including branched-chain amino acids,  
168 aromatic amino acids (alanine, glutamine, isoleucine, phenyl alanine, tyrosine), cluster 2, glycolysis related  
169 metabolites including ketone bodies (glucose, lactate, pyruvate, glycerol, beta-hydroxy butyrate), cluster 3,  
170 fatty acids (total fatty acids, monounsaturated fatty acids (MUFA), saturated fatty acids(SFA)), cluster 4,  
171 apolipoproteins (apolipoprotein B, apolipoprotein A1), cluster 5, lipids including total lipid concentration in  
172 lipoprotein subclasses (triglycerides, extremely large very large density lipoprotein (VLDL), very large VLDL,  
173 large VLDL, medium VLDL, small VLDL, very small VLDL, intermediate-density lipoprotein (IDL), large  
174 low density lipoprotein (LDL), medium LDL, small LDL, large HDL, medium high density lipoprotein (HDL),  
175 small HDL) and cluster 6, inflammatory biomarkers (hs-CRP, alpha-1-acid glycoprotein (AGP), activePAI-  
176 1).

177

### 178 ***Statistical analysis***

179 Statistical analyses were conducted using SAS software version 9.4 (SAS Institute Inc., Cary, USA). Due to  
180 incomplete information on one or more of the metabolites with copper concentration, 20 subjects were  
181 excluded from the study. The difference between complete and incomplete data set was examined by Kruskal-  
182 Wallis test and Pearson's Chi-square test. Normality for all variables was assessed and natural log-  
183 transformation was done accordingly. The comparisons of Cu concentration between groups were analysed

184 using Student's *t*-test for continuous variables and Chi-square for categorical. A multiple linear regression  
185 analyses using standardised score (z-scores; mean=0; SD=1) was then performed to study the association  
186 between copper concentration and the metabolites (model 1, unadjusted). The linear model was adjusted for  
187 sex (model 2), and further adjusted for BMI (model 3). *P*-values were adjusted for multiple testing using the  
188 Benjamin and Hochberg (1995) false discovery rate (FDR) approach separately for each cluster (n=6) [18].  
189 The statistical tests were two-sided and significance was considered at  $p<0.05$ .

190

## 191 **Results**

192 **Characteristics of participants.** General characteristics of the study participants are summarised in **Table 1**.  
193 The data analysis was performed on N=206 individuals (116 men and 90 women). We found no statistically  
194 significant differences between men and women for anthropometric, lifestyle and socio-demographic  
195 characteristics ( $p$ -value>0.05). In addition, we found no difference between complete and incomplete data  
196 analysis (data not shown).

197 **Copper and CVD risk markers.** **Figure 1** shows results from the multivariable regression analyses of the  
198 association between copper and six metabolic clusters and inflammatory biomarkers examined in the study. In  
199 model 1, unadjusted, copper was positively associated with metabolites phenylalanine, very small VLDL, IDL,  
200 alpha-1-acid glycoprotein and hs-CRP ( $p$ -value<0.05). In addition, in model 1, copper was negatively  
201 associated with metabolite glutamine ( $p$ -value<0.05). In model 2, adjusted for sex, the strength of association  
202 was almost identical between Cu and the above mentioned metabolites. In addition, the strength of the  
203 association between Cu and few other additional metabolites attained significant effect after adjustment for  
204 sex including, apolipoprotein A1, large LDL, and large HDL. In model 3, after adjustment for BMI, the  
205 association between Cu and all the other metabolites were attenuated, except for glutamine, beta-hydroxy  
206 butyrate, alpha-1-acid glycoprotein and hs-CRP. After correction for multiple testing using FDR-approach, Cu  
207 was found to be only positively associated with inflammatory biomarkers, alpha-1-acid glycoprotein ( $\beta=0.14$ ,  
208  $P$ -value correction=0.04) and hs-CRP ( $\beta=0.28$ ,  $P$ -value correction=0.0001). Copper was not associated with  
209 PAI-1 in any of the three models examined in our study ( $P$ -value>0.05).

210

## 211 **Discussion**

212 Copper is both a pro-oxidant and antioxidant [6]. Copper plays a role in nineteen enzymatic reactions involved  
213 in oxidative stress and metabolic homeostasis [4] including the Cu/Zn dependent superoxide dismutase (SOD),  
214 the cytochrome *c* oxidase and the ceruloplasmin [4]. On the other hand, copper ions participate in radical  
215 reactions such as conversion of superoxide to hydrogen peroxide and hydroxyl radicals, and catalyse the  
216 oxidative modification of LDL in vitro [19,20]. High serum Cu has been reported as an independent risk factor  
217 for CVD disease in multiple epidemiological studies [6,21,22] which show the duplicitous nature of copper.  
218 However, these results should be interpreted with caution as the studies were performed in wider age groups,  
219 no adjustment for BMI/inflammatory biomarkers, small population size or in diseased populations [6,21,22].

#### 220 *Copper and circulating metabolites – BMI as a main effect modulator?*

221 From the sex-adjusted model, we observed copper to be associated with 9 metabolites (phenylalanine, very  
222 small VLDL, IDL, apolipoprotein A1, large LDL, large HDL, glutamine, alpha-1-acid glycoprotein and hs-  
223 CRP) in the multiple regression analyses. However, adjustment for BMI in model 3 strongly attenuated these  
224 associations suggesting an important modification either by BMI or the environmental factors associated to it.  
225 The environmental factors include an unhealthy diet, sedentary behaviour, and physical inactivity. Adjustment  
226 of these factors in the analyses would have provided information on whether the attenuation of the association  
227 between copper and metabolites were entirely based on BMI or confounded by the environmental factors  
228 associated to it. Only one previous investigation on 231 participants has reported significant association  
229 between serum copper and classical markers (total cholesterol, LDL-cholesterol) of CVD risk adjusting for  
230 BMI [23]. Few studies has reported an increase in serum copper concentrations in overweight and obese  
231 participants compared with normal weight participants [24,25]. Another study (N=189) has reported an  
232 increase in copper concentrations with categories of BMI [26]. However, no difference were observed between  
233 total blood copper concentration and BMI categories in our study ( $p$ -value=0.07, **Table 1**). Only limited  
234 information is available in the literature on the influence of BMI on copper concentration in healthy young  
235 population. These results may show that increase in Cu concentrations and link to CVD risk may be influenced  
236 by lifestyle factors and future studies are warranted to test this association.

#### 237 *Copper and inflammation* (Figure 2)

238 Copper plays an essential role in regulating reactive oxygen and nitrogen species (ROS/RNS) concentration  
239 and, variations in Cu metabolism could modulate various inflammatory pathways [27]. Copper was found to



240 be positively associated with alpha-1-acid glycoprotein and hs-CRP in our study. In fact, the present data  
241 supports a role for Cu in modulating systemic inflammation with a robust association between Cu and  
242 inflammatory biomarkers. Our results confirm the previously reported association between copper and  
243 inflammation [23,26], thus adding to the body of evidence linking copper with the alteration of inflammatory  
244 biomarkers.

245 Alpha-1-acid glycoprotein and CRP are acute-phase inflammatory biomarkers, and may play a critical role in  
246 the aetiology of low-grade inflammation status as they are frequently reported in relation to obesity and CVD  
247 events [28]. In our cross-sectional study we sought to identify the elements that promote systemic inflammation  
248 in relation to copper which may offer insights into the development of CVD, thus suggesting targets for CVD  
249 prevention. However, the association between copper and inflammatory biomarkers, should be interpreted with  
250 caution in the light of interplay between the factors copper, oxidative stress and inflammation (**Figure 2**).  
251 Under inflammatory conditions, serum copper levels are increased and trigger oxidative stress responses that  
252 activate inflammatory responses [27]. Interestingly, copper dyshomeostasis, oxidative stress and inflammation  
253 are commonly present in several chronic diseases including obesity, neurodegenerative diseases and CVD  
254 (**Figure 2**). There are still controversies on the causal and consequential interconnections of copper, oxidative  
255 stress and inflammation which warrants for further epidemiological studies to carefully interpret the above  
256 mentioned associations (**Figure 2**).

257

#### 258 *Copper and plasminogen activator inhibitor-1*

259 Among the list of other risk markers for cardiovascular events, it has been proposed that impairment of the  
260 fibrinolytic system detected in plasma, due to increased plasminogen activator inhibitor-1 (PAI-1)  
261 concentration, could predict the complications of CVD [29]. PAI-1 is responsible for the specific and rapid  
262 inhibition of fibrinolytic proteases, thus maintaining proper homeostasis and stringent control with the  
263 fibrinolysis system, thrombosis and vascular remodelling [30]. However, active PAI-1 is inherently unstable  
264 and readily undergoes a significant structural rearrangement and converts to an inactive, latent form that can  
265 no longer inhibit fibrinolytic proteases [31]. Metal ion ligands such as copper are reported to modulate the  
266 stability of PAI-1 [31]. Copper binds to PAI-1 with high affinity, resulting in significant acceleration in the

267 rate of latency conversion [31]. Few studies has reported an association of copper ions in interfering with the  
268 stability of PAI-1 concentration in the circulation [31,32].

269 Previous investigation on healthy women (N=16) has reported a significant decrease in PAI-1 concentration  
270 with Cu supplementation [16]. This is in contrast from our report where Cu concentration was not associated  
271 with PAI-1 in our study. This may be because of inclusion of both sexes and higher sample size in our study  
272 when compared to the previous investigation in women only [16]. In addition, previous investigation has not  
273 accounted for rigorous statistical procedures including multiple testing and Cu in our study was measured from  
274 the blood samples in normal physiological state rather than from supplementation.

275

### 276 *Strengths and limitations*

277 The present analysis supported evidence for an association of Cu and an atherogenic metabolic profile that  
278 might be influenced by sex and body mass index in humans. Unfortunately, the accurate assessment of Cu  
279 concentration requires expensive ICP-MS analysis on blood samples collected on metal-free tubing, which  
280 will limit the opportunity for a large-scale data analysis. The strength of the present approach lies in the fact  
281 that the exploration is performed in a representative sample of young adult subjects with equal number of  
282 males and females. Moreover, the study sample was selected as regards geographical area for living in order  
283 to limit dietary and environmental exposure. In addition, the total blood copper concentration measured in the  
284 present study was consistent with previously published values suggestive of a healthy physiological range in  
285 the NFBC1966 participants [33]. We were unable to quantify other indicators related to Cu status such as  
286 ceruloplasmin and the Cu-dependent enzymes (erythrocyte superoxide dismutase (SOD), cytochrome C  
287 oxidase activity) which could have provided additional information on the copper metabolism.

288 We have measured the circulating metabolites from serum which are considered to be more sensitive and  
289 reproducible in biomarker detection when compared to plasma samples [34]. Metabolomics gives an  
290 instantaneous snapshot of the metabolism and cannot be translated into a dynamic map of metabolite traffic  
291 on biochemical routes. However, we should be aware that we measured serum concentrations which may not  
292 necessarily reflect intracellular processes. As extensive metabolomics data are increasingly available, we  
293 expect in future comprehensive metabolic profiles from large epidemiologic studies which will elucidate  
294 molecular mechanism and pathways in relation to copper and CVD risk.

295 We have examined a wide range of cardiovascular risk factors in relation to copper concentration. However,  
296 the cross sectional study design does not ensure causality. The possibility of uncontrolled or unknown  
297 confounders cannot be ruled out. Interestingly, one genome wide association study (GWAS) was performed  
298 for blood copper concentration [35]. The data on 2603 Australian adults reported two genome wide significant  
299 variants, rs1175550 and rs2769264 associated with blood copper. To ascertain potential causal association  
300 between blood copper and the metabolic profile observed in our present analysis, we search for association  
301 between the Cu SNPs from [35] on the summary statistics provided from the last GWAS on nuclear magnetic  
302 resonance metabolomics profile [36]. Unfortunately, both Cu-SNPs did not show association for any of the  
303 metabolites from a large GWAS catalogue, therefore suggesting a non-causal association. In addition, it is  
304 important to mention that the present study was performed using a limited study sample and further large  
305 sample population studies are needed to confirm an association between copper and biomarkers of  
306 cardiovascular disease risk. Finally, the study was performed on an ethnically homogeneous population of the  
307 Northern Finland which reduces the ability to generalise our results and warrants for multicentre studies to  
308 confirm if these results could be translated to the general population.

309

### 310 ***Conclusion***

311 Our data together with others support an association between Cu and non-specific biomarkers of inflammation  
312 which may be related to ongoing proinflammatory process *in vivo* and warrants further research to understand  
313 the role of Cu in the aetiology of chronic diseases. The study warrants replication of the results in future large  
314 epidemiological investigations with different population characteristics which could help in early intervention  
315 and prevention of cardiovascular disease risk.

316

317 **Competing Interests** - None.

318 **Acknowledgements** - We thank the entire NFBC1966 study team, including the research staff and all others  
319 involved in the data collection and processing and those involved in the oversight and management of the  
320 study. We acknowledge late Professor Paula Rantakallio for launch of Northern Finland Birth Cohort 1966  
321 and initial data collection, Sarianna Vaara for data collection and Tuula Ylitalo for administration. The authors

322 thank all the participants of NFBC1966 study. The technical assistance of Professor Milena Horvat and Dr.  
323 Janja Tratnik (Jozef Stefan Institute, Ljubljana, Slovenia) is gratefully acknowledged.

324 **Funding:** This work was supported by the Academy of Finland (MR.J, grant number 24300796, 24302031);  
325 Biocenter Oulu Doctoral Program (S.P), Juho Vainio Foundation (S.P), and Orion Research Foundation sr  
326 (S.P); The Finnish Medical Foundation (T.P), and Sigrid Jusélius Foundation (T.P); European Union's  
327 Horizon 2020 research and innovation programme (MR.J, S.S, S.P, grant number 633595) for the  
328 DynaHEALTH action and LifeCycle EU, and European Community's Seventh Framework Programme  
329 FP7/2007-2013-Environment (including Climate Change) FP7-ENV-2008-1- under Grant Agreement No:  
330 226534-ArcRisk (A.R and K.A). No funding bodies had any role in study design, data collection and analysis,  
331 decision to publish, or preparation of the manuscript.

332 **Contributions:** SP, AR and SS designed the analysis. SP conducted the analysis and wrote the manuscript  
333 with guidance from TP, AR, KA and SS. DM was responsible for blood metal concentration measurements in  
334 this analysis. All authors contributed intellectually to the manuscript and approved the final version.

335

## 336 Cited literature

337 [1] S Lutsenko. Human copper homeostasis: a network of interconnected pathways, Curr.Opin.Chem.Biol. 14  
338 (2010) 211-217.

339 [2] M Olivares, R Uauy. Copper as an essential nutrient, Am.J.Clin.Nutr. 63 (1996) 6S.

340 [3] ES LeShane, U Shinde, JM Walker, AN Barry, NJ Blackburn, M Ralle, et al. Interactions between copper-  
341 binding sites determine the redox status and conformation of the regulatory N-terminal domain of ATP7B,  
342 J.Biol.Chem. 285 (2010) 6327-6336.

343 [4] RA Festa, DJ Thiele. Copper: an essential metal in biology, Curr.Biol. 21 (2011) 877.

344 [5] E Ehrenwald, GM Chisolm, PL Fox. Intact human ceruloplasmin oxidatively modifies low density  
345 lipoprotein, J.Clin.Invest. 93 (1994) 1493-1501.

346 [6] JT Salonen, R Salonen, H Korpela, S Suntioinen, J Tuomilehto. Serum copper and the risk of acute  
347 myocardial infarction: a prospective population study in men in eastern Finland, Am.J.Epidemiol. 134 (1991)  
348 268-276.

349 [7] LM Klevay. Coronary heart disease: the zinc/copper hypothesis, Am.J.Clin.Nutr. 28 (1975) 764-774.

350 [8] RA DiSilvestro, EL Joseph, W Zhang, AE Raimo, YM Kim. A randomized trial of copper supplementation  
351 effects on blood copper enzyme activities and parameters related to cardiovascular health, Metabolism. 61  
352 (2012) 1242-1246.

353 [9] M Ala-Korpela. Potential role of body fluid <sup>1</sup>H NMR metabonomics as a prognostic and diagnostic tool,  
354 Expert Rev.Mol.Diagn. 7 (2007) 761-773.

355 [10] P Wurtz, AS Havulinna, P Soininen, T Tynkkynen, D Prieto-Merino, T Tillin, et al. Metabolite profiling  
356 and cardiovascular event risk: a prospective study of 3 population-based cohorts, Circulation. 131 (2015) 774-  
357 785.

358 [11] P Wurtz, AJ Kangas, P Soininen, DA Lawlor, G Davey Smith, M Ala-Korpela. Quantitative Serum  
359 Nuclear Magnetic Resonance Metabolomics in Large-Scale Epidemiology: A Primer on -Omic Technologies,  
360 Am.J.Epidemiol. 186 (2017) 1084-1096.

361 [12] P Rantakallio. Groups at risk in low birth weight infants and perinatal mortality, Acta Paediatr.Scand. 193  
362 (1969) Suppl 193:1+.

363 [13] A Taanila, GK Murray, J Jokelainen, M Isohanni, P Rantakallio. Infant developmental milestones: a 31-  
364 year follow-up, Dev.Med.Child Neurol. 47 (2005) 581-586.

365 [14] Q Wang, P Wurtz, K Auro, L Morin-Papunen, AJ Kangas, P Soininen, et al. Effects of hormonal  
366 contraception on systemic metabolism: cross-sectional and longitudinal evidence, Int.J.Epidemiol. 45 (2016)  
367 1445-1457.

368 [15] S Palaniswamy, E Hypponen, DM Williams, J Jokelainen, E Lowry, S Keinanen-Kiukaanniemi, et al.  
369 Potential determinants of vitamin D in Finnish adults: a cross-sectional study from the Northern Finland birth  
370 cohort 1966, BMJ Open. 7 (2017) 013161.

371 [16] S Bugel, A Harper, E Rock, JM O'Connor, MP Bonham, JJ Strain. Effect of copper supplementation on  
372 indices of copper status and certain CVD risk markers in young healthy women, Br.J.Nutr. 94 (2005) 231-236.

373 [17] BARANY EBBA , BERGDAHL INGVAR A. , SCHÜTZ ANDREJS, SKERFVING STAFFAN and  
374 OSKARSSON AGNETA. Inductively Coupled Plasma Mass Spectrometry for Direct Multi-element Analysis  
375 of Diluted Human Blood and Serum, Journal of Analytical Atomic Spectrometry. (1997) 1005-1009.

376 [18] Benjamini, Y., & Hochberg, Y. Controlling the False Discovery Rate: A Practical and Powerful Approach  
377 to Multiple Testing, Journal of the Royal Statistical Society. Series B (Methodological). 57 (1995) 289–300.

378 [19] I Bremner. Manifestations of copper excess, Am.J.Clin.Nutr. 67 (1998) 1073S.

379 [20] GA Ferns, DJ Lamb, A Taylor. The possible role of copper ions in atherogenesis: the Blue Janus,  
380 Atherosclerosis. 133 (1997) 139-152.

381 [21] A Reunanen, P Knekt, RK Aaran. Serum ceruloplasmin level and the risk of myocardial infarction and  
382 stroke, Am.J.Epidemiol. 136 (1992) 1082-1090.

383 [22] N Leone, D Courbon, P Ducimetiere, M Zureik. Zinc, copper, and magnesium and risks for all-cause,  
384 cancer, and cardiovascular mortality, Epidemiology. 17 (2006) 308-314.

385 [23] S Bo, M Durazzo, R Gambino, C Berutti, N Milanese, A Caropreso, et al. Associations of dietary and  
386 serum copper with inflammation, oxidative stress, and metabolic variables in adults, J.Nutr. 138 (2008) 305-  
387 310.

388 [24] R Tungtrongchitr, P Pongpaew, B Phonrat, A Tungtrongchitr, D Viroonudomphol, N Vudhivai, et al.  
389 Serum copper, zinc, ceruloplasmin and superoxide dismutase in Thai overweight and obese,  
390 J.Med.Assoc.Thai. 86 (2003) 543-551.

391 [25] SC Lima, RF Arrais, CH Sales, MG Almeida, KC de Sena, VT Oliveira, et al. Assessment of copper and  
392 lipid profile in obese children and adolescents, *Biol.Trace Elem.Res.* 114 (2006) 19-29.

393 [26] M Ghayour-Mobarhan, A Taylor, SA New, DJ Lamb, GA Ferns. Determinants of serum copper, zinc and  
394 selenium in healthy subjects, *Ann.Clin.Biochem.* 42 (2005) 364-375.

395 [27] TC Pereira, MM Campos, MR Bogo. Copper toxicology, oxidative stress and inflammation using  
396 zebrafish as experimental model, *J.Appl.Toxicol.* 36 (2016) 876-885.

397 [28] MA Connelly, EG Gruppen, JD Otvos, RPF Dullaart. Inflammatory glycoproteins in cardiometabolic  
398 disorders, autoimmune diseases and cancer, *Clin.Chim.Acta.* 459 (2016) 177-186.

399 [29] C Song, S Burgess, JD Eicher, CJ O'Donnell, AD Johnson. Causal Effect of Plasminogen Activator  
400 Inhibitor Type 1 on Coronary Heart Disease, *J.Am.Heart Assoc.* 6 (2017) 10.1161/JAHA.116.004918.

401 [30] I Diebold, D Kraicun, S Bonello, A Gorlach. The 'PAI-1 paradox' in vascular remodeling,  
402 *Thromb.Haemost.* 100 (2008) 984-991.

403 [31] JC Bucci, CS McClintock, Y Chu, GL Ware, KD McConnell, JP Emerson, et al. Resolving distinct  
404 molecular origins for copper effects on PAI-1, *J.Biol.Inorg.Chem.* 22 (2017) 1123-1135.

405 [32] JC Bucci, MB Trelle, CS McClintock, T Qureshi, TJ Jorgensen, CB Peterson. Copper(II) Ions Increase  
406 Plasminogen Activator Inhibitor Type 1 Dynamics in Key Structural Regions That Govern Stability,  
407 *Biochemistry.* 55 (2016) 4386-4398.

408 [33] P Kiilholma, M Gronroos, P Liukko, P Pakarinen, H Hyora, R Erkkola. Maternal serum copper and zinc  
409 concentrations in normal and small-for-date pregnancies, *Gynecol.Obstet.Invest.* 18 (1984) 212-216.

410 [34] Z Yu, G Kastenmuller, Y He, P Belcredi, G Moller, C Prehn, et al. Differences between human plasma  
411 and serum metabolite profiles, *PLoS One.* 6 (2011) e21230.

412 [35] DM Evans, G Zhu, V Dy, AC Heath, PA Madden, JP Kemp, et al. Genome-wide association study  
413 identifies loci affecting blood copper, selenium and zinc, *Hum.Mol.Genet.* 22 (2013) 3998-4006.

414 [36] J Kettunen, A Demirkan, P Wurtz, HH Draisma, T Haller, R Rawal, et al. Genome-wide study for  
415 circulating metabolites identifies 62 loci and reveals novel systemic effects of LPA, *Nat.Comm.* 7 (2016)  
416 11122.

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429 **Figures and Tables**  
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431 **Figure 1** shows the association of copper concentration with metabolic clusters examined in the  
432 study. The \* represents the metabolites that were significant after the BH-false discovery rate  
433 correction.

434 **Figure 2** represents the relationship between copper and inflammation.

435 **Legend for Figure 2:** Copper and inflammation. Copper induces oxidative stress through imbalance  
436 in antioxidant system that increase reactive oxygen and nitrogen species (ROS, RNS), which might  
437 induce the activation of signaling pathways that upregulate proinflammatory cytokines leading to  
438 inflammation. However, inflammation is a hallmark of several chronic diseases including obesity,  
439 cardiovascular and neurodegenerative diseases. There are still controversies on causal and  
440 consequential interconnections between copper, oxidative stress and inflammation: those are  
441 mentioned throughout the text and represented by dotted lines. hs-CRP - high sensitivity C-reactive  
442 protein.

443 **Table 1:** Descriptive statistics and fasting blood copper concentration by anthropometric, lifestyle  
444 and socio-demographic factors.

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**Table 1:** Descriptive statistics and fasting blood copper concentration by anthropometric, lifestyle and socio-demographic factors.

	<b>Total (N=206 – 195)**</b>	<b>[Cu] ng/mL Mean (SD)</b>	<b>P-value*</b>
<b>Age (years)</b>	31		
<b>Men (n %)</b>	116 (56.31)	835.66 (154.49)	
<b>Women (n %)</b>	90 (43.69)	883.52 (146.49)	0.60
<b>BMI (kg/m<sup>2</sup>) [Mean (SD)]</b>	25.23 (4.34)		
Normal weight (n %)	110 (53.4)	835.63 (137.78)	
Overweight and Obese (n %)	96 (46.6)	880.57 (165.38)	0.07
<b>Socioeconomic status (n %)<sup>1</sup></b>			
I + II (Professional)	24 (12.18)	894.50 (178.19)	
III (Skilled worker)	56 (28.43)	876.91 (123.76)	
IV (Unskilled worker)	66 (33.50)	822.35 (169.22)	0.16
V (Farmer)	9 (4.57)	886.22 (127.96)	
VI (Other)	42 (21.32)	876.86 (150.09)	
<b>Diet score<sup>2</sup> (n %)</b>			
0-1	55 (27.50)	879.42 (145.59)	
2-3	116 (58.00)	862.82 (156.39)	0.10
4-5	29 (14.50)	805.34 (151.61)	
<b>Smoking status<sup>3</sup> (n %)</b>			
No smoker	109 (55.61)	835.59 (144.38)	
Smokers	87 (44.39)	888.56 (159.82)	0.32
<b>Alcohol consumption (g/day)<sup>4</sup> (n %)</b>			
Abstainer	14 (7.18)	900.29 (198.01)	
Low risk drinker	172 (88.21)	854.66 (149.28)	0.46
At-risk drinker	9 (4.62)	891.78 (148.97)	

Values are presented as mean (standard deviation) or number (%).

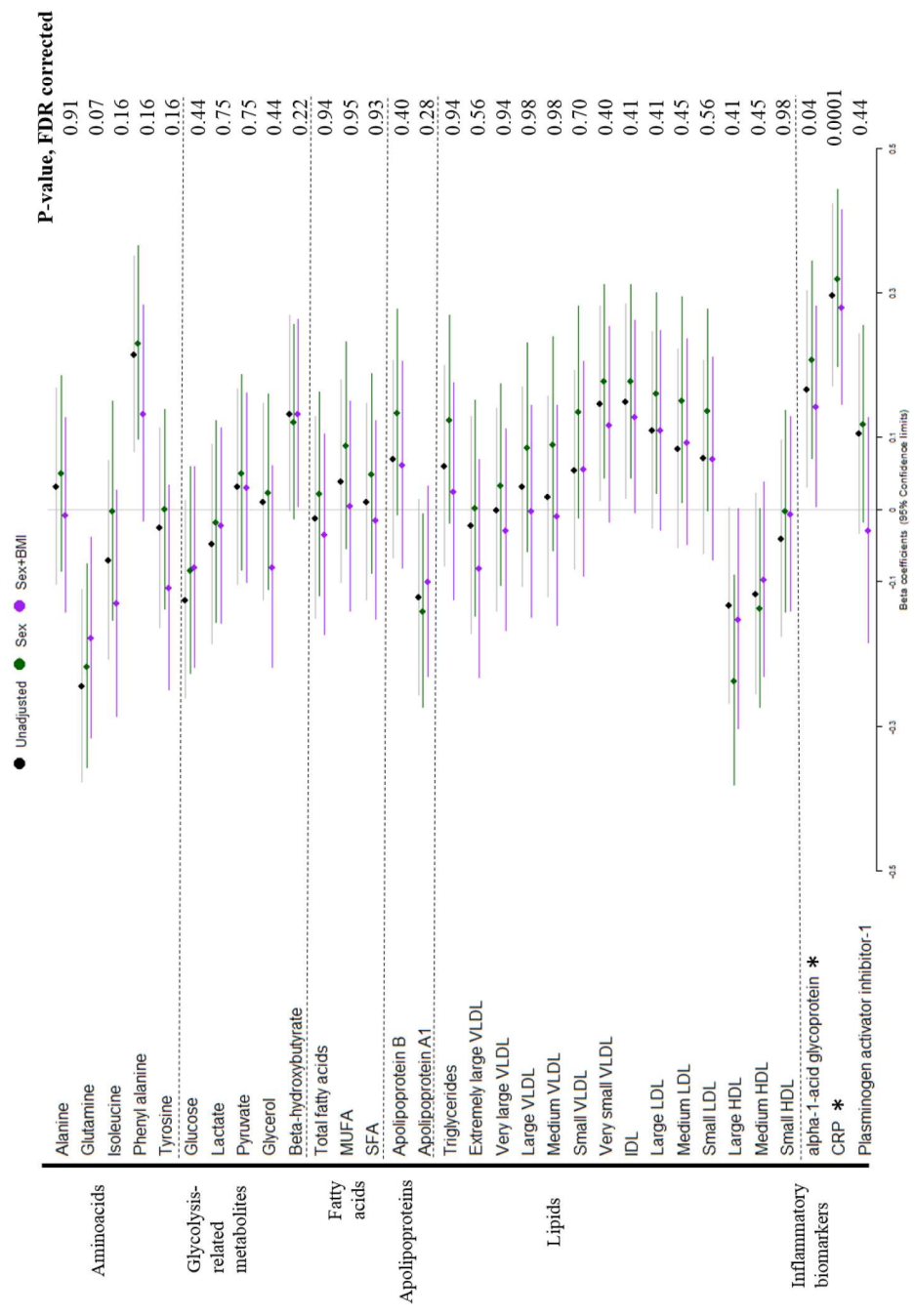
\*P-value was calculated using Student's *t*-test for continuous variables and ANOVA for categorical variables.

\*\* N varies due to missing data for some of the variables

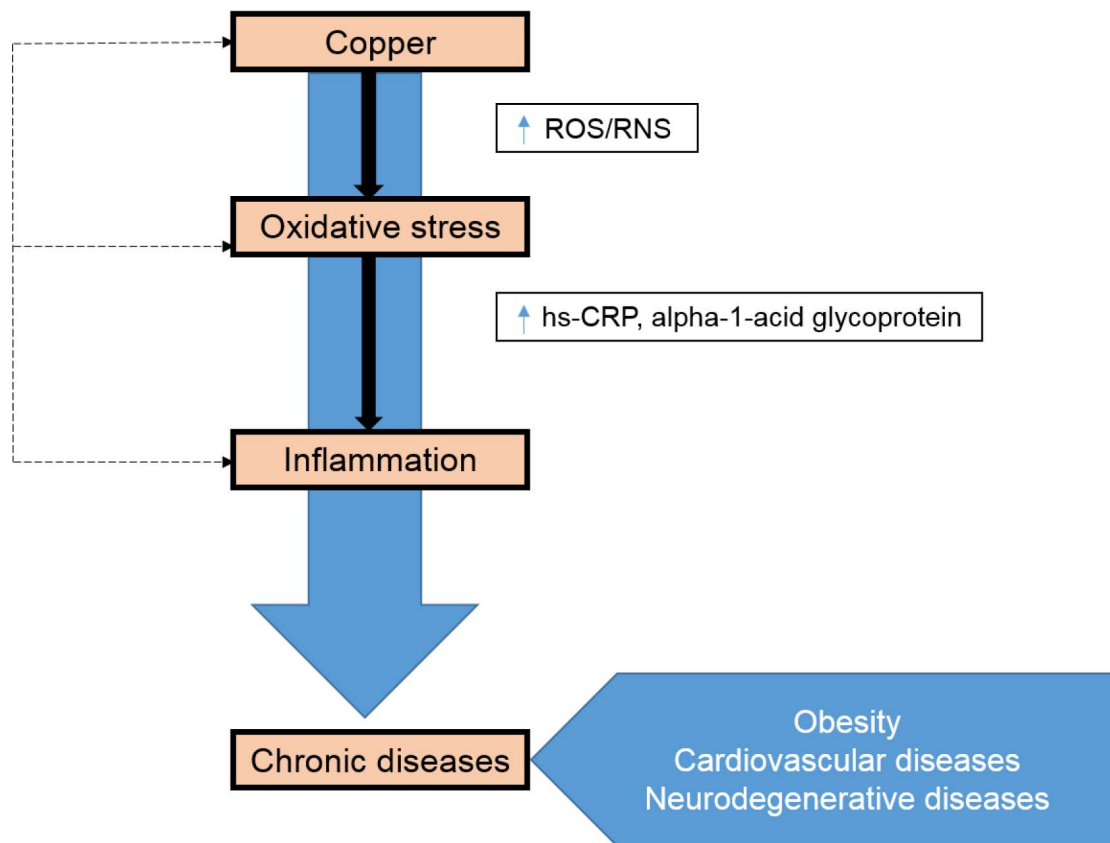
1. Data available on N=197 individuals; N=9 observations missing with socioeconomic position
2. Data available on N=200 individuals; N=6 observations missing with diet score
3. Data available on N=196 individuals; N=10 observations missing with smoking status
4. Data available on N=195 individuals; N=11 observations missing with alcohol consumption; Alcohol classification according to WHO sex-specific classification as abstainer, low risk drinker ( $\leq 20$  g/day and  $\leq 40$  g/day for women and men, respectively) or at-risk drinker ( $> 20$ g/day and  $> 40$ g/day for women and men, respectively).



**Figure 1** shows the association of copper concentration with metabolic clusters examined in the study. The \* represents the metabolites that were significant after the BH-false discovery rate correction.



**Figure 2** represents the relationship between copper and inflammation.



**Legend for Figure 2:** Copper and inflammation. Copper induces oxidative stress through imbalance in antioxidant system that increase reactive oxygen and nitrogen species (ROS, RNS), which might induce the activation of signaling pathways that upregulate proinflammatory cytokines leading to inflammation. However, inflammation is a hallmark of several chronic diseases including obesity, cardiovascular and neurodegenerative diseases. There are still controversies on causal and consequential interconnections between copper, oxidative stress and inflammation: those are mentioned throughout the text and represented by dotted lines. hs-CRP - high sensitivity C-reactive protein.