



Subject-specific biomechanical analysis to estimate locations susceptible to osteoarthritis - finite element modeling and MRI follow-up of ACL reconstructed patients

Journal:	<i>Journal of Orthopaedic Research</i>
Manuscript ID	JOR-21-0088.R1
Wiley - Manuscript type:	Research Article (Member)
Date Submitted by the Author:	n/a
Complete List of Authors:	<p>Bolcos, Paul; University of Eastern Finland Faculty of Science and Forestry, Applied Physics</p> <p>Mononen, Mika; University of Eastern Finland, Department of Applied Physics</p> <p>Roach, Koren; University of California San Francisco, Radiology and Biomedical Imaging</p> <p>Tanaka, Matthew; University of California-San Francisco, Radiology and Biomedical Imaging</p> <p>Suomalainen, Juha-Sampo; Kuopio University Hospital, Department of Clinical Radiology</p> <p>Mikkonen, Santtu; University of Eastern Finland, Department of Applied Physics</p> <p>Nissi, Mikko; University of Eastern Finland, Department of Applied Physics; University of Oulu, Research Unit of Medical Imaging, Physics and Technology</p> <p>Töyräs, Juha; University of Eastern Finland, Department of Applied Physics; Kuopio University Hospital, Diagnostic Imaging Centre; The University of Queensland, School of Information Technology and Electrical Engineering</p> <p>Link, Thomas; University of California San Francisco, Department of Radiology and Biomedical Imaging</p> <p>Souza, Richard; University of California, San Francisco, Physical Therapy and Rehabilitation Science; University of California, San Francisco, Radiology and Biomedical Imaging</p> <p>Majumdar, Sharmila; University of California, San Francisco, Department of Radiology</p> <p>Ma, Benjamin; UCSF</p> <p>Li, Xiaojuan; Cleveland Clinic, Program of Advanced Musculoskeletal Imaging, Department of Biomedical Engineering, Lerner Research Institute</p> <p>Korhonen, Rami; University of Eastern Finland, Department of Physics and Mathematics; Kuopio University Hospital, Diagnostic Imaging Centre</p>
Areas of Expertise:	Biomechanics, Post-traumatic osteoarthritis, Articular cartilage
Keywords:	Osteoarthritis - Post Traumatic, Finite Element Analysis, Cartilage, Biomechanics, Gait, Reconstruction



SCHOLARONE™
Manuscripts

Subject-specific biomechanical analysis to estimate locations susceptible to osteoarthritis - finite element modeling and MRI follow-up of ACL reconstructed patients

Paul O. Bolcos^{1, *}, Mika E. Mononen¹, Koren E. Roach², Matthew S. Tanaka², Juha-Sampo Suomalainen³, Santtu Mikkonen¹, Mikko J. Nissi^{1,5}, Juha Töyräs^{1,6,7}, Thomas M. Link², Richard Souza², Sharmila Majumdar², Benjamin Ma², Xiaojuan Li⁴, Rami K Korhonen^{1,3}

¹Department of Applied Physics, University of Eastern Finland, Kuopio, Finland

²Department of Radiology and Biomedical Imaging, University of California San Francisco, San Francisco, Unites States of America

³Department of Clinical Radiology, Kuopio University Hospital, Kuopio, Finland

⁴Department of Biomedical Engineering, Cleveland Clinic, Cleveland, Unites States of America

⁵Research Unit of Medical Imaging, Physics and Technology, University of Oulu, Oulu, Finland

⁶School of Information Technology and Electrical Engineering, The University of Queensland, Brisbane, Australia

⁷Diagnostic Imaging Centre, Kuopio University Hospital, Kuopio Finland

Word count (Introduction through Discussion) = 4754

Word count (Abstract) = 241

*Corresponding author:

Paul Octavian Bolcos

Department of Applied Physics, University of Eastern Finland

POB 1627, FI-70211 Kuopio, Finland

Tel. +358 45 2290653

E-mail: paul.bolcos@uef.fi

31 Abstract

32 The aims of this case-control study were to: (1) Identify cartilage locations and volumes at risk
33 of osteoarthritis using subject-specific finite element (FE) models; (2) Quantify the
34 relationships between the simulated biomechanical parameters and T_2 and $T_{1\rho}$ relaxation times
35 of MRI.

36 We created subject-specific FE models for 7 patients with anterior cruciate ligament (ACL)
37 reconstruction and 6 controls based on a previous proof-of-concept study. We identified
38 locations and cartilage volumes susceptible to osteoarthritis, based on maximum principal
39 stresses and absolute maximum shear strains in cartilage exceeding thresholds of 7 MPa and
40 32%, respectively. The locations and volumes susceptible to osteoarthritis were compared
41 qualitatively and quantitatively against 2-year longitudinal changes in T_2 and $T_{1\rho}$ relaxation
42 times.

43 The degeneration volumes predicted by the FE models, based on excessive maximum principal
44 stresses, were significantly correlated ($r=0.711$, $p<0.001$) with the degeneration volumes
45 determined from T_2 relaxation times. There was also a significant correlation between the
46 predicted stress values and changes in T_2 relaxation time ($r=0.649$, $p<0.001$). Absolute
47 maximum shear strains and changes in $T_{1\rho}$ relaxation time were not significantly correlated.

48 Five out of seven patients with ACL reconstruction showed excessive maximum principal
49 stresses in either one or both tibial cartilage compartments, in agreement with follow-up
50 information from MRI. Expectedly, for controls, the FE models and follow-up information
51 showed no degenerative signs.

52 Our results suggest that the presented modelling methodology could be applied to prospectively
53 identify ACL reconstructed patients at risk of biomechanically driven osteoarthritis,
54 particularly by the analysis of maximum principal stresses of cartilage.

For Peer Review

1. Introduction

Anterior cruciate ligament (ACL) rupture is a common sports-related knee joint injury. It has been shown that patients with ACL rupture have a high risk of developing osteoarthritis (OA)^{1,2}. Given the contribution of ACL in the knee joint stability, ACL ruptures are often surgically reconstructed using tissue grafts. However, postoperative studies have shown that knee OA may develop even in the short-term after ACL reconstruction (ACLR)^{2,3}. It has been reported that almost half of patients with ACLR have signs of articular cartilage degeneration at 1-year follow-up². Identifying patients that are at low or high risk of developing OA would be useful, as this may improve our mechanistic understanding of OA and promote the development of post-operative strategies for delaying and/or preventing the onset and progression of OA.

There are several methods to clinically assess knee OA using self-assessment questionnaires, such as Knee Osteoarthritis Outcome Score (KOOS), or using imaging methods, such as Kellgren-Lawrence (KL) or whole-organ resonance magnetic imaging (WORMS)⁴. However, these methods provide limited information on cartilage integrity or composition and are susceptible to intra- and interobserver variability⁵. Measurement of T_2 and $T_{1\rho}$ relaxation times offers a quantitative assessment of local articular cartilage composition⁶. Several studies have related the T_2 relaxation time with collagen network integrity and arrangement^{7,8} and the $T_{1\rho}$ relaxation time with proteoglycan (PG) content^{9,10}.

A possible mechanism that may lead to OA in patients with ACLR is altered knee joint biomechanics, leading to abnormal stresses or strains experienced by articular cartilage^{11,12}. These abnormal stresses and strains have been evaluated experimentally^{13,14} and computationally^{13,14} *ex vivo*, and by finite element (FE) modeling of human joints^{15,16}. In a clinical setting, the FE model generation and computational solution should be as fast as

possible. In terms of model complexity, several studies have shown that FE models with simplified geometry and motion¹⁷, cartilage material properties¹⁸ and ligament formulation¹⁹ produce similar, if not the same, results as more computationally demanding approaches. In a recent proof-of-concept study, we showed that these simpler FE models can predict areas susceptible to OA in agreement with follow-up information²⁰.

The aims of this study can be divided into two sub-aims: **(1)** Identify locations and cartilage volumes in the knee joints of patients with ACLR and controls at risk of developing OA, using the biomechanical modeling methodology from a previous proof-of-concept study²⁰. Excessive maximum principal stresses were assumed to lead to collagen network degeneration^{20,21} and excessive absolute maximum shear strains were assumed to cause PG loss^{22,23}. **(2)** Quantify the ability of the simulated maximum principal stresses and maximum shear strains to predict 2-year longitudinal changes in T_2 and $T_{1\rho}$ relaxation times, respectively.

We examined the following hypothesis: **(1)** Collagen network damage simulated via excessive maximum principal stresses is strongly related to longitudinal changes in the collagen-sensitive T_2 relaxation time; **(2)** PG loss simulated via excessive absolute maximum shear strains is strongly related to longitudinal changes in PG-sensitive $T_{1\rho}$ relaxation time. To our knowledge, the only FE modeling study aiming at predicting knee OA progression for a relatively high number of subjects is a recent study by Mononen et al.¹⁵ (n=21). In contrast to that study, we believe this is the first study in which subject-specific FE models of the knee joint, in terms of both geometry and motion, were created for a cohort and were qualitatively and quantitatively verified against follow-up MR imaging assessment methods.

2. Materials and methods

The workflow of the study is shown in Figure 1. This study is a level II prospective cohort study and includes 13 subjects: 7 patients with ACLR and 6 healthy controls. FE models for each subject were generated, with knee joint geometries manually segmented from the 3D-FSE (CUBE) MRI sequences (Figure 1a) and knee joint motions obtained from motion capture data (Figure 1b). The FE models included femoral and tibial cartilages, menisci, and cruciate and collateral ligaments (Figure 1c). The simulation results of the models were then compared against follow-up T_2 and $T_{1\rho}$ information, as well as WOMS and KOOS grades (Figure 1e-f, Table 1).

2.1. Patient demographics and acquired data

MR imaging and motion capture were performed at the University of California, San Francisco (UCSF) (Table 1). All subjects gave informed consent and data acquisition was approved by and carried out in accordance with the rules and regulations of the Institutional Review Board under the Human Research Protection Program at UCSF. More details on the patient demographics and measurement setup are provided in the Supplementary materials.

T_2 and $T_{1\rho}$ relaxation time mapping was done using a two-parametric non-linear exponential fit with Aedes plugin (<http://aedes.uef.fi>) for Matlab and custom scripts at both 1- and 3-year follow-up time points. Each compartment of the tibial cartilage was manually segmented from the T_2 and $T_{1\rho}$ mapped images.

Co-registering the 1-year and 3-year MRI images may be possible. However, it would introduce uncertainties due to the MRI slice thickness of 4 mm and the discrepancy in the slice location between the two follow-up points. Also, at the 3-year follow-up timepoint, there may be regions showing changes in cartilage thickness, complicating co-registering the two MRI image stacks. Instead, T_2 and $T_{1\rho}$ relaxation times above 60ms were assumed to indicate

collagen network damage and PG loss, respectively^{24–27} (see more below). Some sensitivity analysis for this threshold has also been done in a previous study¹⁶. Please also find more clarification on the threshold selection in the Supplementary materials.

2.2. FE models

The methodology used to generate the FE models for each subject was identical to that applied in our previous study²⁰. Details of the FE model generation process, including segmentation, mesh generation, and motion and material implementation, are presented in Supplementary materials. Detailed material properties for each soft tissue are shown in the Supplementary materials, Table s1.

To identify locations prone to collagen network degeneration, the maximum principal stress (tensile stress) distribution was calculated by taking the peak centroid value of the maximum principal stress in each element throughout the entire stance phase of gait. Similarly, to identify location prone to PG loss, the distribution of absolute maximum shear strains was calculated by taking the peak centroid value of the absolute maximum shear strain in each element throughout the entire stance phase of gait. Maximum principal stresses above 7 MPa threshold were assumed to indicate collagen network degeneration^{20,21}. Absolute maximum shear strains above 32% threshold were assumed to indicate PG loss^{22,23}. These thresholds were based on previous experimental and computational studies^{14,22,28–38}. They also match values reported in a recent review article by Jørgensen³⁹. Some sensitivity analysis for these thresholds has also been done in previous studies¹⁶.

2.3. Comparison between FE model results and T_2 / $T_{1\rho}$ relaxation times

Qualitative comparison

The FE model results were qualitatively compared against T_2 and $T_{1\rho}$ relaxation times using axial views (Figure 1d). For the FE model, the axial view indicates the distribution maximum principal stresses or absolute maximum shear strains (centroid values of elements) on the

superficial tibial cartilage. To determine the T_2 and $T_{1\rho}$ values on the tibial cartilage surface we used a custom Matlab script. An example of process for obtaining the axial view is provided in the Supplementary materials, Figure s4.

Quantitative comparison

Volume comparison. To compare volumes susceptible to collagen degeneration and PG loss predicted by the FE models at the 1-year follow-up time point against the cartilage volumes with changes in T_2 and $T_{1\rho}$ relaxation times between the 1- and 3-year follow-up time points, the following steps were taken:

1. For the FE models, in each compartment a volume-of-interest (VOI) was defined as the total volume of elements exceeding the 7 MPa or 32% thresholds and taken as the degenerated percentage of the total volume of the compartment.
2. For T_2 and $T_{1\rho}$ relaxation times at both the 1- and 3-year follow-up time points, in each compartment, the VOI was defined as the total volume exceeding the 60ms threshold^{24–27,40} and similarly taken as the degenerated percentage of the total volume of the compartment. The volume of tissue assumed to be degenerated between the time points was calculated by subtracting the VOI at the 3-year time point from the VOI at the 1-year time point.

Value comparison. To assess the relationship between the **maximum principal** stress and the change in T_2 relaxation times between the 1- and 3-year follow-up time points, the following steps were taken:

1. For the FE models, the location and value of peak **maximum principal** stress was determined for each joint compartment from the entire stance phase (one location and one centroid value). Then, the average value of all element centroids around the peak value was determined. Henceforth, we will refer to the calculated ‘average centroid value around the peak’ as ‘peak value’ (values are almost the same for all patients, see the Supplementary materials).

2. For the T_2 maps, T_2 values at the 1- and 3-year follow-up time points were determined at the areas corresponding (defined as the average of 10 pixels) to those determined in the step 1. The change in T_2 relaxation times between the 1- and 3-year time points was computed.

Similar steps were applied to assess the relationship between the peak absolute maximum shear strain and the change in $T_{1\rho}$ relaxation times between the 1- and 3-year follow-up time points. More information is provided in the Supplementary materials. Please note that for the quantitative analysis, both sets of participants are combined.

Other verification methods

We also compared the peak maximum principal stresses against 3-year WOMBS grades (grades from 0 to 3 at the 3-year follow-up, details are provided in the Supplementary materials). Additionally, we compared the biomechanical parameters (peak maximum principal stresses and absolute shear strains predicted at the 1-year time point) and the measured changes in MRI parameters (2-year longitudinal change in T_2 , $T_{1\rho}$ and WOMBS) against the 2-year longitudinal changes in KOOS grades (details are provided in the Supplementary materials).

Statistical analysis

We assessed the relationship between the predicted and potentially degenerated cartilage volumes. Performing a normality test revealed that the data was not normally distributed, and thus Spearman's correlation was used. Bivariate least square linear fits were calculated using a method that takes into account the uncertainties in the variables⁴¹. This method is known to be robust for outliers and extreme observations. Similarly, the Spearman's correlation coefficients and a bivariate least square linear fits were calculated between the peak values of maximum principal stress and absolute maximum shear strain and the change in T_2 and $T_{1\rho}$

199 relaxation times, respectively. Furthermore, bivariate least square linear functions⁴¹ were fitted
200 between the simulated biomechanical parameters and the relaxation times.

201 To evaluate if there are statistically significant differences in **maximum principal** stress and
202 absolute maximum shear strain, and the change in T_2 and $T_{1\rho}$ relaxation times, between patients
203 with ACLR and healthy controls, we used the Mann-Whitney U-test, since the data was not
204 normally distributed. All statistical tests were carried out in MATLAB, using custom scripts.

For Peer Review

3. Results

3.1. Comparison between the FE model results and T_2 and $T_{1\rho}$ relaxation times

Qualitative comparison

We compared the **maximum principal** stress distributions on the joint surfaces at the 1-year follow-up time point and changes in the T_2 relaxation time distributions at the 1- and 3-year follow-up time points for the 7 patients with ACLR (Figure 2) and the 6 controls (Figure 3). Both for the patients and controls there is a good qualitative correspondence between the simulated **maximum principal** stresses and the longitudinal changes in T_2 relaxation times. For Patients 1-3, 5 and 7 with ACLR, the articular cartilage locations with excessive **maximum principal** stresses matched the areas with changes in T_2 relaxation times (Figure 2). For Patients 4 and 6, only small **maximum principal** stresses (less than 5 MPa) and changes in T_2 relaxation times (less than 5ms) were observed. As was expected, small **maximum principal** stresses and negligible changes in T_2 relaxation times were observed for all healthy control subjects (Figure 3). For the 7 patients with ACLR, we also compared the sagittal sections (see Supplementary materials, Figure s5).

In terms of absolute maximum shear strains, the correspondence between the FE model results and MRI findings was not evident. For Patient 1, PG loss was predicted via excessive absolute maximum shear strains on the lateral tibial cartilage with a similar distribution as the **maximum principal** stresses. $T_{1\rho}$ relaxation times were also increased between the 1- and 3-year follow-up time points in the same regions (not shown). For Patients 2-7, absolute maximum shear strains did not exceed the assumed degeneration threshold (see below), while the $T_{1\rho}$ relaxation time showed a similar distribution and values as the T_2 relaxation time.

Quantitative comparison – Cartilage Volumes

The cartilage volumes with collagen network degeneration predicted by the FE models matched with the volumes of degenerated cartilage estimated based on the change in T_2 relaxation times (Figure 4a and Supplementary materials, Figure s6). There was a significant positive correlation between the predicted and measured degenerated cartilage volumes ($r=0.711$, $p<0.001$). Using bivariate least squares, we obtained a linear fit with an $R^2=0.946$.

The cartilage volumes with assumed PG loss predicted by the FE models did not match with the cartilage volumes with PG loss estimated based on the change in $T_{1\rho}$ relaxation times (see Supplementary materials, Figures s6 and s7). There was a non-significant and weak correlation between the volumetric PG loss predicted by the FE models and that estimated by the change in $T_{1\rho}$ relaxation times ($r=0.279$, $p=0.168$).

Quantitative comparison - FE model and MRI peak values

There was a positive correlation between peak maximum principal stresses at the 1-year time point and local changes in T_2 relaxation times between the 1- and 3-year follow-up time points (Figure 4b), ($r=0.649$, $p<0.001$). Using bivariate least squares, we obtained a linear fit with an $R^2=0.906$. Again, there was no statistically significant correlation between peak absolute maximum shear strains and changes in $T_{1\rho}$ relaxation times ($r=0.280$, $p=0.160$, see Supplementary materials, Figure s8).

Quantitative comparison - FE model and WOMBS

When grouping the subjects by WOMBS grade (details provided in the Supplementary materials), we could differentiate patients roughly in three different risk groups for the progression of OA (low, moderate and high risk) (Figure 5 and Supplementary materials, Figure s9). Patients experiencing less than 7 MPa maximum principal stress showed no or minor tissue alterations based on WOMBS and T_2 relaxation times. On the other hand, patients experiencing stresses between 7 and 10 MPa were at a higher risk and those experiencing over

10 MPa **maximum principal** stresses had the most severe tissue changes during the follow-up, both in terms of WORMS and T_2 relaxation times.

Quantitative comparison – KOOS

We did not find any statistically significant correlations between the KOOS grades and biomechanical or MRI parameters (see Supplementary materials for more details).

3.2. Comparison between patients with ACLR and healthy controls

The median **maximum principal** stress value was significantly higher in patients with ACLR (7.20 MPa) than in healthy controls (5.30 MPa) (Figure 6, $p=0.008$). Similarly, the median change in T_2 relaxation times between 1- and 3-year follow-up time points was significantly higher in ACLR patients (5.3ms) than controls (1.61ms) (Figure 6, $p<0.001$). The **maximum principal** stress and the change in T_2 relaxation time showed large patient-specific variability in the ACLR group (standard deviation of 2.77 MPa and 12.45ms, respectively). Obviously, this was not the case for the control group where subjects mostly remained healthy (standard deviation of 0.54 MPa and 0.98ms, respectively). In terms of absolute maximum shear strains, there was no statistically significant difference between patients with ACLR and healthy controls (Figure 6, $p=0.439$). On the other hand, between the two groups, there was a statistically significant difference in the change of $T_{1\rho}$ relaxation time between the 1- and 3-year follow-up time points (Figure 6, $p<0.001$). The distribution, range and values for the change in $T_{1\rho}$ relaxation times were similar to those in T_2 relaxation times (not shown). Patients with ACLR had significantly lower KOOS grades at both 1- and 3-year follow-up time points than controls. More information is presented in the Supplementary materials..

4. Discussion

In this study, FE models were created for seven patients with ACLR and six healthy controls. The knee geometry was obtained from manually segmented high-resolution MRI and the knee joint motion was obtained from motion capture. The methodology used to generate the FE models was identical to that described in a previous proof-of-concept study⁴⁰. We identified locations in each compartment of the tibial cartilage at risk of biomechanically-driven OA due to excessive **maximum principal** stresses and absolute maximum shear strains. Then, we compared these FE model predictions qualitatively and quantitatively with follow-up MRI findings. The location and volume of cartilage at risk for collagen degeneration predicted by the FE model, through excessive **maximum principal** stresses, matched the cartilage volumes of increased T_2 values during the follow-up in **85% of subjects**. Furthermore, there was a positive correlation between the predicted volume and values of maximum principal stresses and T_2 relaxation times. There were also significant differences in **maximum principal** stresses, and T_2 and $T_{1\rho}$ relaxation times, but not in absolute maximum shear strains, between ACLR patients and healthy controls. **Our results suggest that the proposed FE modeling workflow with simplified geometries (i.e. only the tibiofemoral joint), loading conditions (i.e. directly from motion capture without musculoskeletal modelling) and materials (i.e. without time-dependent degeneration mechanisms), can identify patients at risk of developing biomechanically driven collagen degeneration in OA.**

Collagen network. Both qualitatively and quantitatively, the articular cartilage locations with excessive **maximum principal** stresses matched the areas with changes in T_2 relaxation times. For almost all patients the posterior aspect of the cartilage area showed high **maximum principal** stresses. In the same posterior aspect, T_2 relaxation times were increased between the follow-up time points. The posterior site was also noted in other studies^{2,42,43} and is attributed to valgus collapse^{42,43}, particularly for the lateral compartment. Importantly, all subjects in both

groups predicted to be at **high-risk** or at **low-risk** of OA onset and development by the FE model (maximum principal stresses above 10 MPa or below 7 MPa, respectively) matched the follow-up MRI information (change in T_2 relaxation time above 23 ms or below 10ms, respectively; WORMS 3 or WORMS 0/1, respectively). This highlights the importance of patient-specific analysis. In the future, more subjects will be added from other patient groups.

Using the 7 MPa and 60ms thresholds as collagen network degeneration indicators, a good match was found between the degenerated volumes predicted by the FE models and those determined from T_2 maps (Figure 4a and Supplementary materials, Figure s6). Some differences between the predictions and MRI findings may be influenced by the assumed degeneration threshold values, which are likely patient-specific, or other limitations listed below.

There were significant differences between patients with ACLR and healthy controls in maximum principal stresses and changes in T_2 and $T_{1\rho}$ relaxation times. The predicted maximum principal stresses showed that some patients with ACLR are at a higher risk of developing OA than others, in agreement with previous ACLR studies^{1,2}. Relaxation times showed similar patient-specificity. The distribution of peak maximum principal stresses and changes in T_2 and $T_{1\rho}$ relaxation times were more clustered in controls with a small standard deviation, and the values were below the assumed degeneration thresholds. This was to be expected, since one of the criteria for the patient selection of healthy controls was that they should not show any degenerative signs at any time point.

Interestingly, using 7 MPa^{30,44} and 10 MPa³⁹ as maximum principal stress thresholds, we could differentiate patients whose knees were evaluated by WORMS roughly in three different risk groups for the progression of OA. Therefore, our proposed and relatively straightforward biomechanical method might be applicable for clinical risk assessment. However, it should be

noted that the rough division to these three risk categories was made by the authors, though based on the results, and apply at the moment only to these studied patient groups (patients with ACLR and controls). Therefore, this grouping cannot be generalized yet. There are several factors in the model that may need to be adjusted to other patient groups and are listed in limitations below.

PG loss. Both qualitatively and quantitatively, the connection between absolute shear strain and increased $T_{1\rho}$ relaxation time was not evident (see Supplementary materials, Figures s6-s8 and Table s2), though increased values in both of these parameters and WORMS can be seen especially in patients 1, 2 and 7. For patients 1 and 2, please refer to our previous proof-of-concept study²⁰, where the relationship between maximum shear strains and $T_{1\rho}$ relaxation times was examined in more detail. These patients had full-depth cartilage defects, which were included and analyzed also in detail in a recent proof-of-concept mechanobiological modelling study²⁸. The results of that study showed that for these two patients the simulated shear strain driven PG loss is highly localized around the cartilage lesion²⁸. In that study, both $T_{1\rho}$ and T_2 relaxation times were increased in the vicinity of the lesion between 1- and 3-year follow-up time points. Combined, the results indicate that 1) at least on a compartment level, absolute maximum shear strain may not indicate PG loss or changes in tissue integrity, and 2) that highly localized strain levels may be more important and could indicate highly localized cell death and PG loss, even though global strain levels would not change.

Similar to T_2 relaxation times, there were significant differences between patients with ACLR and controls in terms of $T_{1\rho}$. Generally, the values of $T_{1\rho}$ were ~10ms higher than those of T_2 at both 1- and 3-year follow-up time points. The range and distribution of $T_{1\rho}$ were similar to those of T_2 . This would suggest that both T_2 and $T_{1\rho}$ may be more indicative of the overall state of articular cartilage integrity than collagen or PG separately^{7,45}.

Relation with patient reported outcomes. We did not find any statistically significant correlations between the KOOS grades and the FE model predictions, or between the KOOS grades and the MRI measures (see Supplementary materials, Table s3). This suggests that there are other factors than excessive stresses or altered integrity of cartilage that cause symptoms and reduced quality of life, which is not surprising. On the other hand, in the group level there was a significant difference in the KOOS grades and the patients with ACLR showed a [greater](#) variability in KOOS [grades than controls](#). This emphasizes patient-specificity consistent with the biomechanical and MRI results.

Limitations. This study has a few limitations that warrant discussion. They are briefly listed below and expanded upon in the Supplementary materials.

The study was limited to 13 subjects, which is more than in most biomechanical modelling studies but still relatively low. Some discrepancies between the biomechanical parameters and MRI follow-up findings may be attributed to the still relatively low number of patients and may affect the correlation analysis. Generation of subject-specific FE models requires a lot of manual work and time in segmentation, meshing and making the models converge, typically taking at minimum one week per subject from an experienced researcher. However, the FE models were able to distinguish between different patients' risk levels and were in agreement with the follow-up information in 85% of the subjects. To increase the number of patients, the model generation and simulation should become faster. Therefore, the used methodology should be coupled with semi-automatic or fully automatic segmentation techniques³⁸ or even fully automated model generation methods¹⁵. [For instance, in a recently developed atlas-based method¹⁵, the model generation for one patient takes only a few minutes. In the future, AI-based methods could even eliminate the need for FE models, however a large amount of data is needed to properly train such models.](#)

The FE models did not include all muscle forces via musculoskeletal modelling and the patellofemoral joint. Other post-ACLR rehabilitation exercises, such as cutting or single-leg squat, were not included in this study. Constant stress and strain thresholds to estimate cartilage degeneration were used in the models. These thresholds, and material properties of cartilage, could be adjusted in the future at least according to age, gender or physical activity⁴⁶, particularly as the incidence to ACL rupture is typically higher in younger patients^{2,47}. Other mechanisms for OA onset and development, such as underloading and/or inflammation⁴⁸ in early ACLR follow-up, were not considered in the FE models. Implementing all these aforementioned properties can be done in the future, but would increase the model complexity and increase time to obtain the results. This would take the methodology further away from clinical application.

With improved imaging techniques, maybe tissue level properties can in the future be obtained in a personalized manner. However, this is currently time consuming and even with this approach several model parameters need to be assumed from literature^{49,50}. One relatively simple approach to obtain patient-specific tissue level information could be, e.g., to fine tune the degeneration thresholds in the model to capture the experimentally detected changes in cartilage structure during the follow-up of patients. This is one of our long-term goals requiring fast modeling workflow and a lot of subject-specific data from various subject groups.

Only the tibial cartilage was analyzed. This choice was mainly related to the quantitative MRI. In particular, the T_2 relaxation time can show susceptibility to the orientation of the magnetic field. In a recent study⁷, it was shown that the T_2 relaxation time shows the highest dependence on the tissue orientation. Therefore, analysis of the convex and round shape of the femoral condyles and patellar groove may introduce uncertainties^{8,26}. From the clinical point of view, almost half of patients with ACLR show both early^{2,3} and long term¹ signs of OA in the tibial cartilage compartment.

The resolution of the T_2 and $T_{1\rho}$ relaxation time maps was low with a slice thickness of 4 mm and a pixel size of 0.55x0.55mm. This may lead to partial volume effects that could lead to inaccuracies in T_2 and $T_{1\rho}$ values, especially interfaces with high relaxation time differences (i.e. cartilage interface with synovial fluid or bone)^{6,8}. Higher resolution may not be feasible as this would drastically increase the image acquisition time.

Clinical application. Our results suggest that the presented relatively straightforward FE modelling method can be used to identify patients at different risk levels of developing biomechanically driven OA in agreement with MRI follow-up information. This method would be particularly useful in assessing the effects of surgical interventions, such as ACLR, on OA onset and progression. In the future, the methodology could be used to identify and evaluate optimal non-surgical management and post-ACLR rehabilitation strategies for avoiding or delaying the disease progression. FE models with simplified geometry (i.e. only tibiofemoral joint), motion (i.e. directly from motion capture without musculoskeletal modelling) and materials (i.e. without time-dependent degeneration mechanisms) could provide a pathway towards clinical application.

Competing interests

The authors declare no competing interests.

Author contribution

POB contributed to the study design, analysis and interpretation of the data, and preparation and revision of the manuscript. **MEM** contributed to the study design, analysis and interpretation of the data, and preparation and revision of the manuscript. **MJN** contributed to the study design, analysis and interpretation of the data, MRI analysis and interpretation, as well as the preparation and revision of the manuscript. **KER, MST, JSS, TML, RS** and **ShMa** contributed to the clinical MRI data collection, analysis and interpretation of the data, and

revision of the manuscript. **SaMi** and **JT** contributed to the data analysis, statistical interpretation, and revision of the manuscript. **BM** and **XL** contributed to the study design, analysis and interpretation of the data, and revision of the manuscript. **RKK** contributed to the study design, analysis and interpretation of the data, and preparation and revision of the manuscript. All authors have read and approved the final submitted manuscript.

Acknowledgements

Financial support from University of Eastern Finland’s Doctoral Programme in Science, Technology and Computing (SCITECO), Academy of Finland (grants no. 269315, 285909, 305138, 307932, 324529, 324994, 328920), Sigrid Juselius foundation, Business Finland (grant no. 3455/31/2019) and National Institutes of Health (NIH/NIAMS P50 AR060752) are acknowledged. CSC-IT Center for Science, Finland, is acknowledged for computing resources.

References

1. Risberg MA, Oiestad BE, Gunderson R, et al. 2016. Changes in Knee Osteoarthritis, Symptoms, and Function After Anterior Cruciate Ligament Reconstruction. *Am. J. Sports Med.* 44(5):1215–1224 Available from: <http://journals.sagepub.com/doi/10.1177/0363546515626539>.
2. Culvenor AG, Collins NJ, Guermazi A, et al. 2015. Early Knee Osteoarthritis Is Evident One Year Following Anterior Cruciate Ligament Reconstruction: A Magnetic Resonance Imaging Evaluation. *Arthritis Rheumatol.* 67(4):946–955 Available from: <http://doi.wiley.com/10.1002/art.39005>.
3. Williams A, Winalski CS, Chu CR. 2017. Early articular cartilage MRI T2 changes after anterior cruciate ligament reconstruction correlate with later changes in T2 and cartilage thickness. *J. Orthop. Res.* 35(3):699–706 [cited 2018 Mar 20] Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5823014/pdf/nihms941492.pdf>.
4. Li X, Majumdar S. 2013. Quantitative MRI of articular cartilage and its clinical applications. *J. Magn. Reson. Imaging* 38(5):991–1008 [cited 2018 Mar 20] Available from: <http://www.ncbi.nlm.nih.gov/pubmed/24115571>.
5. Wang Y, Wluka AE, Jones G, et al. 2012. Use magnetic resonance imaging to assess articular cartilage. *Ther. Adv. Musculoskelet. Dis.* 4(2):77–97 [cited 2018 Oct 11] Available from: <http://www.ncbi.nlm.nih.gov/pubmed/22870497>.
6. Nissi MJ, Salo E-N, Tiitu V, et al. 2016. Multi-parametric MRI characterization of enzymatically degraded articular cartilage. *J. Orthop. Res.* 34(7):1111–1120 [cited 2018 Dec 17] Available from: <http://www.ncbi.nlm.nih.gov/pubmed/26662555>.
7. Hänninen N, Rautiainen J, Rieppo L, et al. 2017. Orientation anisotropy of quantitative

- 455 MRI relaxation parameters in ordered tissue. *Sci. Rep.* 7(1):9606 [cited 2018 Dec 17]
456 Available from: <http://www.ncbi.nlm.nih.gov/pubmed/28852032>.
- 457 8. Nissi MJ, Töyräs J, Laasanen MS, et al. 2004. Proteoglycan and collagen sensitive
458 MRI evaluation of normal and degenerated articular cartilage. *J. Orthop. Res.*
459 22(3):557–564 [cited 2018 May 21] Available from:
460 <http://doi.wiley.com/10.1016/j.orthres.2003.09.008>.
- 461 9. Duvvuri U, Kudchodkar S, Reddy R, Leigh JS. 2002. T1ρ relaxation can assess
462 longitudinal proteoglycan loss from articular cartilage in vitro. *Osteoarthr. Cartil.*
463 10(11):838–844 [cited 2018 Apr 13] Available from:
464 <http://www.ncbi.nlm.nih.gov/pubmed/12435327>.
- 465 10. Wheaton AJ, Dodge GR, Borthakur A, et al. 2005. Detection of changes in articular
466 cartilage proteoglycan by T1ρ magnetic resonance imaging. *J. Orthop. Res.* 23(1):102–
467 108 [cited 2018 Apr 13] Available from:
468 <http://www.ncbi.nlm.nih.gov/pubmed/15607881>.
- 469 11. Gardinier ES, Manal K, Buchanan TS, Snyder-Mackler L. 2013. Altered loading in the
470 injured knee after ACL rupture. *J. Orthop. Res.* 31(3):458–464 [cited 2017 Sep 26]
471 Available from: <http://doi.wiley.com/10.1002/jor.22249>.
- 472 12. Konrath JM, Saxby DJ, Killen BA, et al. 2017. Muscle contributions to medial
473 tibiofemoral compartment contact loading following ACL reconstruction using
474 semitendinosus and gracilis tendon grafts. *PLoS One* 12(4):e0176016 [cited 2017 Sep
475 7] Available from: <http://dx.plos.org/10.1371/journal.pone.0176016>.
- 476 13. Wilson W, van Burken C, van Donkelaar C, et al. 2006. Causes of mechanically
477 induced collagen damage in articular cartilage. *J. Orthop. Res.* 24(2):220–228 [cited
478 2018 May 18] Available from: <http://doi.wiley.com/10.1002/jor.20027>.

- 479 14. Henao-Murillo L, Ito K, van Donkelaar CC. 2018. Collagen Damage Location in
480 Articular Cartilage Differs if Damage is Caused by Excessive Loading Magnitude or
481 Rate. *Ann. Biomed. Eng.* 46(4):605–615 [cited 2018 Jul 10] Available from:
482 <http://link.springer.com/10.1007/s10439-018-1986-x>.
- 483 15. Mononen ME, Liukkonen MK, Korhonen RK. 2019. Utilizing Atlas-Based Modeling
484 to Predict Knee Joint Cartilage Degeneration: Data from the Osteoarthritis Initiative.
485 *Ann. Biomed. Eng.* 47(3):813–825 [cited 2019 May 6] Available from:
486 <http://link.springer.com/10.1007/s10439-018-02184-y>.
- 487 16. Orozco GA, Bolcos P, Mohammadi A, et al. 2020. Prediction of local fixed charge
488 density loss in cartilage following ACL injury and reconstruction: A computational
489 proof-of-concept study with MRI follow-up. *J. Orthop. Res.* :jor.24797 [cited 2020 Jul
490 31] Available from: <https://onlinelibrary.wiley.com/doi/abs/10.1002/jor.24797>.
- 491 17. Bolcos PO, Mononen ME, Mohammadi A, et al. 2018. Comparison between kinetic
492 and kinetic-kinematic driven knee joint finite element models. *Sci. Rep.* 8(1):17351
493 [cited 2018 Nov 28] Available from: [http://www.nature.com/articles/s41598-018-](http://www.nature.com/articles/s41598-018-35628-5)
494 [35628-5](http://www.nature.com/articles/s41598-018-35628-5).
- 495 18. Klets O, Mononen ME, Tanska P, et al. 2016. Comparison of different material models
496 of articular cartilage in 3D computational modeling of the knee: Data from the
497 Osteoarthritis Initiative (OAI). *J. Biomech.* 49(16):3891–3900 Available from:
498 <https://linkinghub.elsevier.com/retrieve/pii/S0021929016311241>.
- 499 19. Orozco GA, Tanska P, Mononen ME, et al. 2018. The effect of constitutive
500 representations and structural constituents of ligaments on knee joint mechanics. *Sci.*
501 *Rep.* 8(1):2323 [cited 2018 Mar 20] Available from:
502 <http://www.nature.com/articles/s41598-018-20739-w>.

- 503 20. Bolcos PO, Mononen ME, Tanaka MS, et al. 2019. Identification of locations
504 susceptible to osteoarthritis in patients with anterior cruciate ligament reconstruction:
505 Combining knee joint computational modelling with follow-up T1 ρ and T2 imaging.
506 Clin. Biomech. [cited 2019 Sep 12] Available from:
507 [https://www.sciencedirect.com/science/article/pii/S0268003318310295?via%3Dihub#s](https://www.sciencedirect.com/science/article/pii/S0268003318310295?via%3Dihub#s0100)
508 0100.
- 509 21. Mononen ME, Tanska P, Isaksson H, Korhonen RK. 2018. New algorithm for
510 simulation of proteoglycan loss and collagen degeneration in the knee joint: Data from
511 the osteoarthritis initiative. J. Orthop. Res. 36(6):1673–1683 [cited 2018 Mar 20]
512 Available from: <http://doi.wiley.com/10.1002/jor.23811>.
- 513 22. Bonnevie ED, Delco ML, Jasty N, et al. 2016. Chondrocyte death and mitochondrial
514 dysfunction are mediated by cartilage friction and shear strain. Osteoarthr. Cartil.
515 24:S46 [cited 2018 May 14] Available from:
516 <http://linkinghub.elsevier.com/retrieve/pii/S1063458416001266>.
- 517 23. Hashimoto S, Nishiyama T, Hayashi S, et al. 2009. Role of p53 in human chondrocyte
518 apoptosis in response to shear strain. Arthritis Rheum. 60(8):2340–2349 [cited 2018
519 Oct 17] Available from: <http://www.ncbi.nlm.nih.gov/pubmed/19644890>.
- 520 24. Bolbos RI, Ma CB, Link TM, et al. 2008. In Vivo T1 ρ Quantitative Assessment of
521 Knee Cartilage After Anterior Cruciate Ligament Injury Using 3 Tesla Magnetic
522 Resonance Imaging. Invest. Radiol. 43(11):782–788 [cited 2018 Apr 13] Available
523 from: <https://insights.ovid.com/crossref?an=00004424-200811000-00004>.
- 524 25. Li X, Kuo D, Theologis A, et al. 2011. Cartilage in anterior cruciate ligament-
525 reconstructed knees: MR imaging T1 ρ and T2--initial experience with 1-year
526 follow-up. Radiology 258(2):505–14 [cited 2018 Mar 20] Available from:

<http://pubs.rsna.org/doi/10.1148/radiol.10101006>.

26. Surowiec RK, Lucas EP, Fitzcharles EK, et al. 2014. T2 values of articular cartilage in clinically relevant subregions of the asymptomatic knee. *Knee Surgery, Sport. Traumatol. Arthrosc.* 22(6):1404–1414 [cited 2018 Aug 23] Available from: <http://link.springer.com/10.1007/s00167-013-2779-2>.
27. Van Rossom S, Smith CR, Zevenbergen L, et al. 2017. Knee Cartilage Thickness, T1ρ and T2 Relaxation Time Are Related to Articular Cartilage Loading in Healthy Adults. *PLoS One* 12(1):e0170002 [cited 2018 May 22] Available from: <http://dx.plos.org/10.1371/journal.pone.0170002>.
28. Orozco GA, Tanska P, Florea C, et al. 2018. A novel mechanobiological model can predict how physiologically relevant dynamic loading causes proteoglycan loss in mechanically injured articular cartilage. *Sci. Rep.* 8(1):1–16.
29. Danso EK, Honkanen JTJ, Saarakkala S, Korhonen RK. 2014. Comparison of nonlinear mechanical properties of bovine articular cartilage and meniscus. *J. Biomech.* 47(1):200–206 Available from: <http://linkinghub.elsevier.com/retrieve/pii/S0021929013004387>.
30. Mononen ME, Tanska P, Isaksson H, Korhonen RK. 2016. A Novel Method to Simulate the Progression of Collagen Degeneration of Cartilage in the Knee: Data from the Osteoarthritis Initiative. *Sci. Rep.* 6(1):21415 Available from: <http://www.nature.com/articles/srep21415>.
31. Mootanah R, Imhauser CW, Reisse F, et al. 2014. Development and validation of a computational model of the knee joint for the evaluation of surgical treatments for osteoarthritis. *Comput. Methods Biomech. Biomed. Engin.* 17(13):1502–1517 [cited 2018 Oct 5] Available from:

- 551 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4047624/>.
- 552 32. Hosseini SM, Veldink MB, Ito K, van Donkelaar CC. 2013. Is collagen fiber damage
553 the cause of early softening in articular cartilage? *Osteoarthr. Cartil.* 21(1):136–143
554 [cited 2018 Feb 6] Available from:
555 <https://www.sciencedirect.com/science/article/pii/S1063458412009648>.
- 556 33. Kempson GE. 1982. Relationship between the tensile properties of articular cartilage
557 from the human knee and age. *Ann. Rheum. Dis.* 41(5):508–511 [cited 2018 Oct 23]
558 Available from: <http://www.ncbi.nlm.nih.gov/pubmed/7125720>.
- 559 34. Ewers BJ, Dvoracek-Driksna D, Orth MW, Haut RC. 2001. The extent of matrix
560 damage and chondrocyte death in mechanically traumatized articular cartilage explants
561 depends on rate of loading. *J. Orthop. Res.* 19(5):779–784 [cited 2018 Sep 25]
562 Available from:
563 <https://www.sciencedirect.com/science/article/pii/S0736026601000067>.
- 564 35. Kelly PA, O'Connor JJ. 1996. Transmission of rapidly applied loads through articular
565 cartilage. Part 2: Cracked cartilage. *Proc. Inst. Mech. Eng. H.* 210(1):39–49 [cited
566 2018 May 18] Available from:
567 http://journals.sagepub.com/doi/10.1243/PIME_PROC_1996_210_389_02.
- 568 36. Wilson W, Huyghe JM, van Donkelaar CC. 2006. A composition-based cartilage
569 model for the assessment of compositional changes during cartilage damage and
570 adaptation. *Osteoarthr. Cartil.* 14(6):554–560 [cited 2019 May 6] Available from:
571 <https://www.sciencedirect.com/science/article/pii/S1063458405003675>.
- 572 37. Gardiner BS, Woodhouse FG, Besier TF, et al. 2016. Predicting Knee Osteoarthritis.
573 *Ann. Biomed. Eng.* 44(1):222–233 [cited 2017 Sep 21] Available from:
574 <http://link.springer.com/10.1007/s10439-015-1393-5>.

- 575 38. LaValley MP, Lo GH, Price LL, et al. 2017. Development of a clinical prediction
576 algorithm for knee osteoarthritis structural progression in a cohort study: value of
577 adding measurement of subchondral bone density. *Arthritis Res. Ther.* 19(1):95 [cited
578 2018 Oct 5] Available from: [http://arthritis-](http://arthritis-research.biomedcentral.com/articles/10.1186/s13075-017-1291-3)
579 [research.biomedcentral.com/articles/10.1186/s13075-017-1291-3](http://arthritis-research.biomedcentral.com/articles/10.1186/s13075-017-1291-3).
- 580 39. Jørgensen AEM, Kjær M, Heinemeier KM. 2017. The effect of aging and mechanical
581 loading on the metabolism of articular cartilage. *J. Rheumatol.* 44(4):410–417 [cited
582 2018 Mar 27] Available from:
583 <http://www.jrheum.org/content/44/4/410><http://www.jrheum.org/alerts><http://www.jrheum.org/faq>http://www.jrheum.org/reprints_permissions<http://www.jrheum.org>.
- 585 40. Stahl R, Luke A, Li X, et al. 2009. T1rho, T2 and focal knee cartilage abnormalities in
586 physically active and sedentary healthy subjects versus early OA patients—a 3.0-Tesla
587 MRI study. *Eur. Radiol.* 19(1):132–143 [cited 2018 Aug 23] Available from:
588 <http://link.springer.com/10.1007/s00330-008-1107-6>.
- 589 41. Francq BG, Govaerts BB. 2014. Measurement methods comparison with errors-in-
590 variables regressions. From horizontal to vertical OLS regression, review and new
591 perspectives. *Chemom. Intell. Lab. Syst.* 134:123–139.
- 592 42. Lee TQ, Morris G, Csintalan RP. 2003. The Influence of Tibial and Femoral Rotation
593 on Patellofemoral Contact Area and Pressure. *J. Orthop. Sports Phys. Ther.*
594 33(11):686–693.
- 595 43. Kozanek M, Van De Velde SK, Gill TJ, Li G. 2008. The contralateral knee joint in
596 cruciate ligament deficiency. *Am. J. Sports Med.* 36(11):2151–2157 [cited 2019 Nov
597 15] Available from: <http://journals.sagepub.com/doi/10.1177/0363546508319051>.
- 598 44. Bolcos PO, Mononen ME, Tanaka MS, et al. 2018. Prediction of failure locations in

- 599 the knees of patients with ACL reconstruction-comparison against follow-up T1 ρ and
600 T2 maps. 220–228 p. Available from: <http://aedes.uef.fi>.
- 601 45. Blumenkrantz G, Majumdar S. 2007. Quantitative magnetic resonance imaging of
602 articular cartilage in osteoarthritis. *Eur. Cell. Mater.* 13:76–86 [cited 2018 Dec 20]
603 Available from: <http://www.ncbi.nlm.nih.gov/pubmed/17506024>.
- 604 46. Peters AE, Akhtar R, Comerford EJ, Bates KT. 2018. The effect of ageing and
605 osteoarthritis on the mechanical properties of cartilage and bone in the human knee
606 joint. *Sci. Rep.* 8(1):5931 [cited 2018 Dec 19] Available from:
607 <http://www.nature.com/articles/s41598-018-24258-6>.
- 608 47. Patterson B, Culvenor AG, Barton CJ, et al. 2020. Poor functional performance 1 year
609 after ACL reconstruction increases the risk of early osteoarthritis progression. *Br. J.*
610 *Sports Med.* 54(9):246–253 [cited 2020 Sep 4] Available from: <http://bjsm.bmj.com/>.
- 611 48. Eskelinen ASA, Tanska P, Florea C, et al. 2020. Mechanobiological model for
612 simulation of injured cartilage degradation via proinflammatory cytokines and
613 mechanical. *PLoS Comput. Biol.* 16(6):e1007998 [cited 2020 Sep 14] Available from:
614 <https://doi.org/10.1371/journal.pcbi.1007998>.
- 615 49. Räsänen LP, Mononen ME, Lammentausta E, et al. 2016. Three dimensional patient-
616 specific collagen architecture modulates cartilage responses in the knee joint during
617 gait. *Comput. Methods Biomech. Biomed. Engin.* 19(11):1225–1240 [cited 2019 May
618 9] Available from:
619 <http://www.tandfonline.com/doi/full/10.1080/10255842.2015.1124269>.
- 620 50. Räsänen LP, Tanska P, Mononen ME, et al. 2016. Spatial variation of fixed charge
621 density in knee joint cartilage from sodium MRI – Implication on knee joint mechanics
622 under static loading. *J. Biomech.* 49(14):3387–3396 [cited 2018 Sep 27] Available

623 from:
624 <https://www.sciencedirect.com/science/article/pii/S0021929016309861?via%3Dihub>.

625

For Peer Review

Figure Captions

Figure 1. Workflow of the study. a) Knee joint MR image segmentation; b) Knee joint rotations and ground reaction forces from motion capture; c) FE model overview, with geometry from a) and motion from b); d) Axial view comparison between the FE model and T_2 and $T_{1\rho}$ maps; e) Sagittal view comparison between the FE model and T_2 and $T_{1\rho}$ relaxation times maps; f) Quantitative evaluation of degenerated volumes estimated from the FE model and MRI. Also, correlations between the predicted biomechanical parameters and changes in T_2 and $T_{1\rho}$ relaxation times between 1-year and 3-year follow-up time points were computed.

Figure 2. Axial view of the **maximum principal** (tensile) stress distribution at the 1-year follow-up time point and the T_2 relaxation time distributions at the 1- and 3-year follow-up time points for patients with ACLR. Peak values for tensile stresses during the stance phase are given for each compartment. The values for the T_2 relaxation time at the 1- and 3-year time points are given for the same location as peak tensile stresses. Note: Both left and right knees are shown in the same coordinate system, with medial compartment on the left.

Figure 3. Axial view of the **maximum principal** (tensile) stress distribution at the 1-year follow-up time point and the T_2 relaxation time distribution at the 3-year follow-up time point in healthy controls. Peak values for tensile stresses are given for each compartment. The values for the T_2 relaxation time are given for the same location as peak tensile stresses. Note: The T_2 relaxation time distribution at the 1-year follow-up time point is not presented, since it is almost identical with the 3-year follow-up time point. Note: Both left and right knees are shown in the same coordinate system, with medial compartment on the left.

Figure 4. a) Correlation between the predicted degenerated volumes from the FE models at the 1-year time point and those estimated from T_2 between the 1-year and 3-year follow-up time points. b) Correlation between the peak values of **maximum principal** (tensile) stresses at the

1-year time point and changes in the T_2 relaxation time between the 1- and 3-year follow-up time points. Note: The values for the tensile stress and changes in the T_2 relaxation time are taken in the same location.

Figure 5. Correlation between the peak values of **maximum principal** (tensile) stresses at the 1-year time point and changes in the T_2 relaxation time between the 1- and 3-year follow-up time points. Data points were grouped according to WORMS grades at the 3-year follow-up time point. The green, yellow and red areas indicate compartments at **low**, **moderate** and **high** risk of developing OA.

Figure 6. Box plots of the peak values of **maximum principal** (tensile) stresses and absolute maximum shear strains during the stance phase of gait at the 1-year time point and changes in the T_2 and $T_{1\rho}$ relaxation times between the 1- and 3-year follow-up time points for patients with ACLR and healthy controls. Mann-Whitney U-test p-values are also provided.

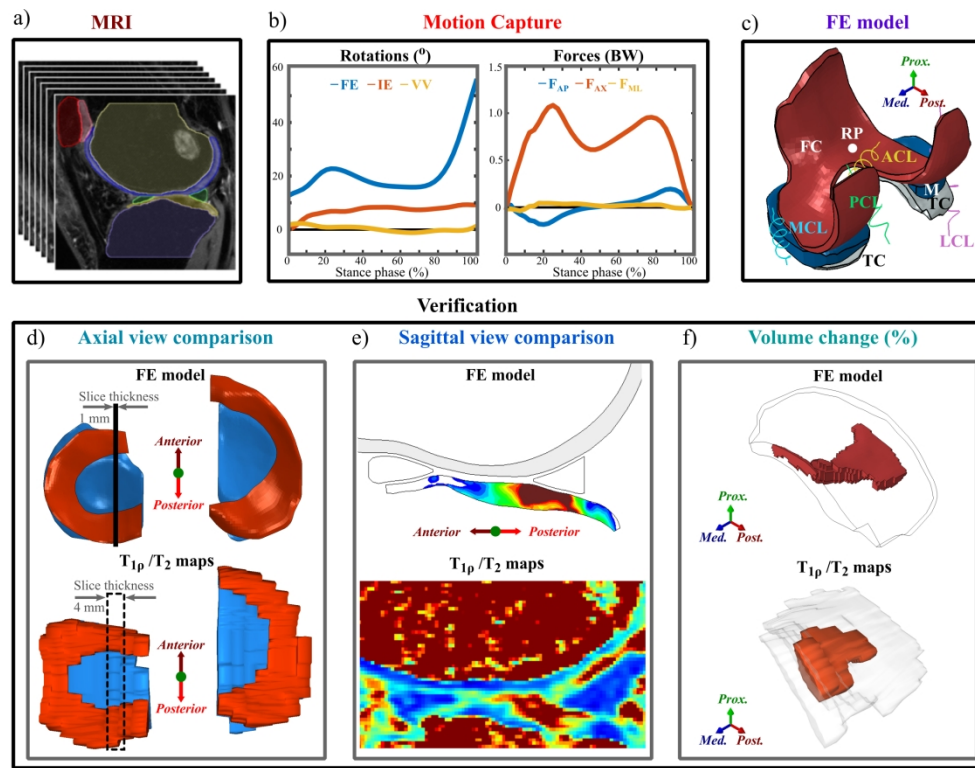
Table Captions

Table 1. Demographics and measured data. A detailed description of measurement protocols can be found in Supplementary materials.

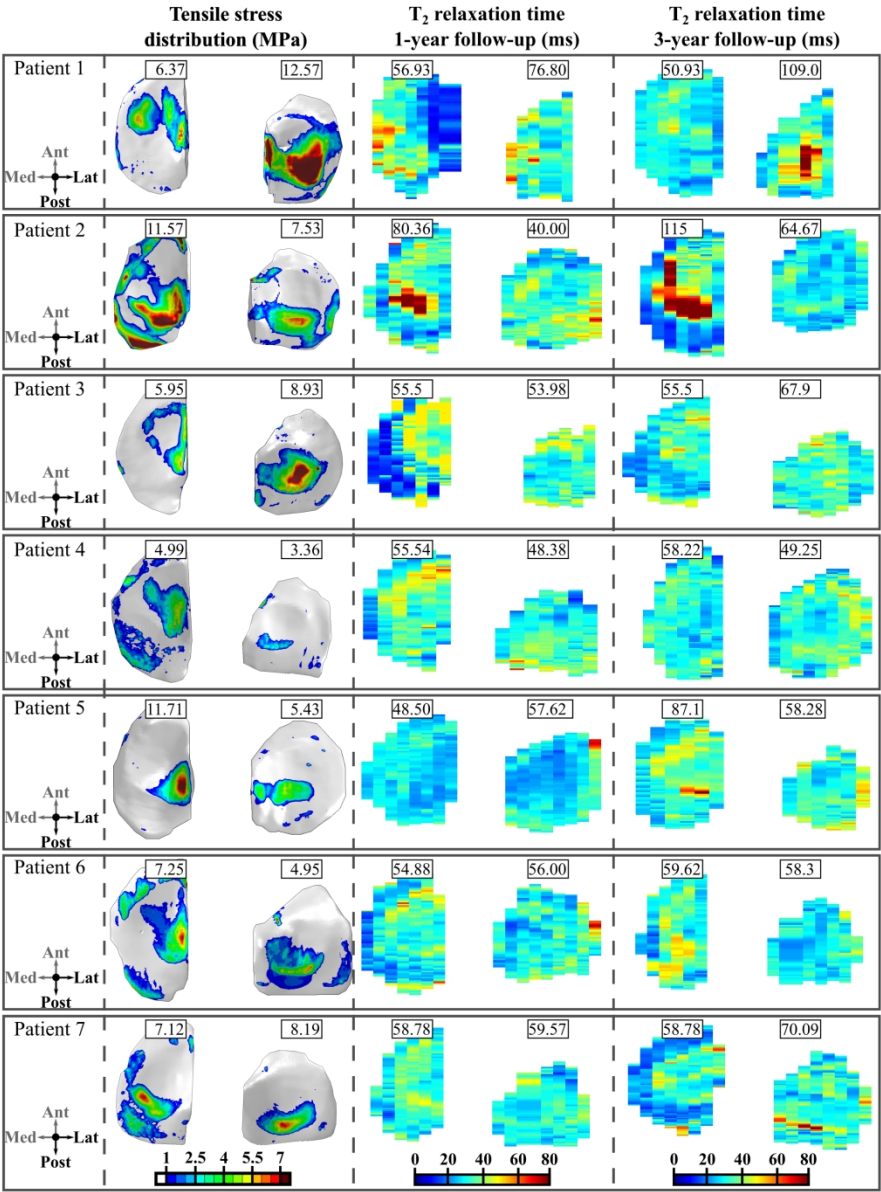
Tables and Captions

Table 1. Demographics and measured data. A detailed description of the measurement protocols can be found in Supplementary material.

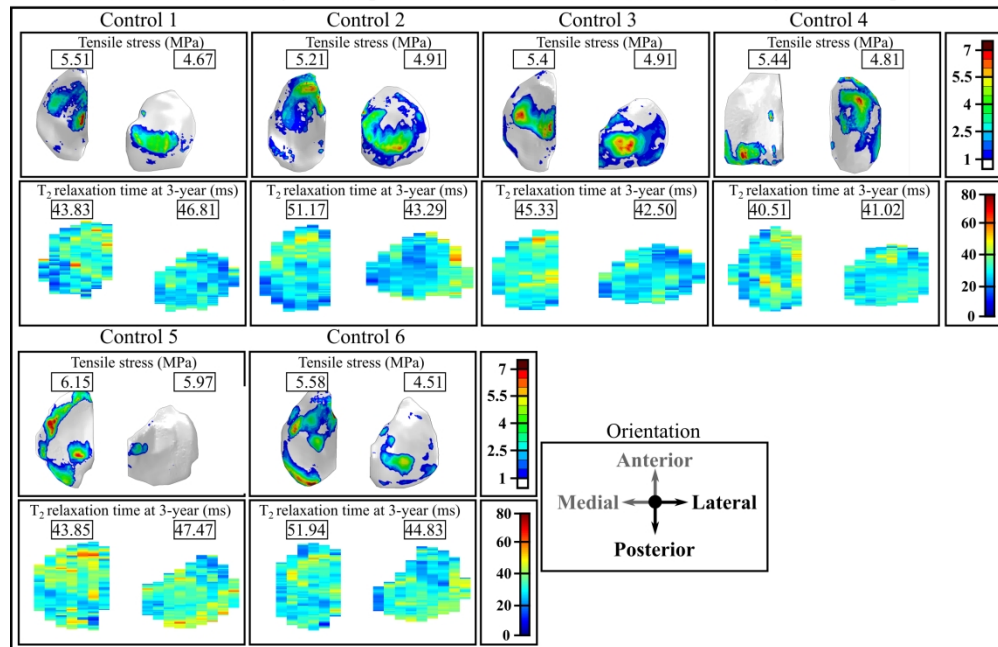
Demographics	ACLR (n=7)	Controls (n=6)
Age (years)	37.8±6.5	31.3±0.37
Gender (male: female)	5:2	3:3
Weight (kg)	70.60±13.43	68.3±10.3
Height (m)	1.74±0.11	1.69±0.11
Measured data	Time points and usage	
	1-year follow-up	3-year follow-up
3D-FSE CUBE MRI (3D fat-saturated, intermediate-weighted, fluid-sensitive fast-spin-echo)	FE geometry + WORMS	WORMS
Sagittal 3D MAPSS MRI sequence ^{52,62}	T ₂ /T _{1ρ} relaxation time	T ₂ /T _{1ρ} relaxation time
Knee injury Osteoarthritis Outcome Score (KOOS)	Verification	Verification
Motion Capture	FE motion	-



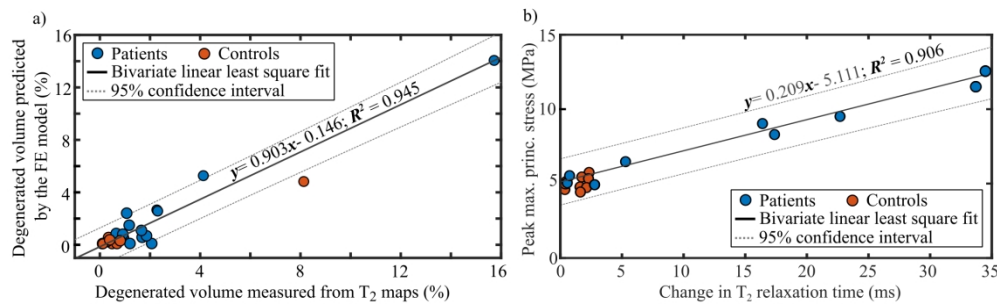
Workflow of the study. a) Knee joint MR image segmentation; b) Knee joint rotations and ground reaction forces from motion capture; c) FE model overview, with geometry from a) and motion from b); d) Axial view comparison between FE model and T_2 and $T_{1\rho}$ maps; e) Sagittal view comparison between FE model and T_2 and $T_{1\rho}$ relaxation times maps; f) Quantitative evaluation of simulated and actual degenerated volumes. Also, the correlation between predicted biomechanical parameters and the change in T_2 and $T_{1\rho}$ relaxation values between 1-year and 3-year follow-up time points was computed.



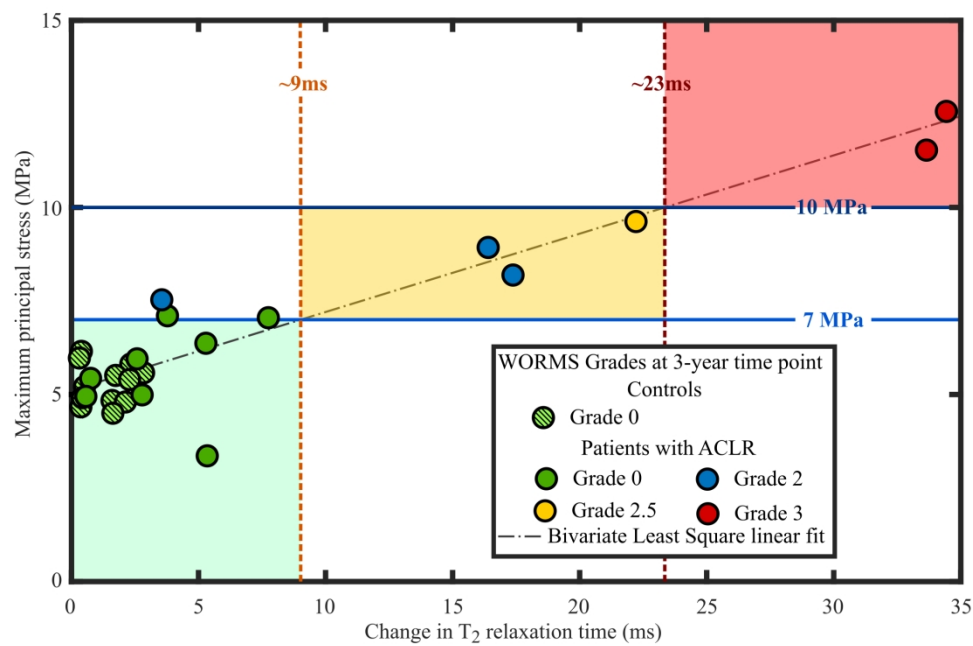
Axial view of the maximum principal (tensile) stress distribution at the 1-year follow-up time point and the T₂ relaxation time distributions at the 1- and 3-year follow-up time points for patients with ACLR. Peak values for tensile stresses during the stance phase are given for each compartment. The values for the T₂ relaxation time at the 1- and 3-year time points are given for the same location as peak tensile stresses. Note: Both left and right knees are shown in the same coordinate system, with medial compartment on the left.

Tensile stress distribution at 1-year time point and T_2 relaxation time distribution at 3-year follow-up for controls

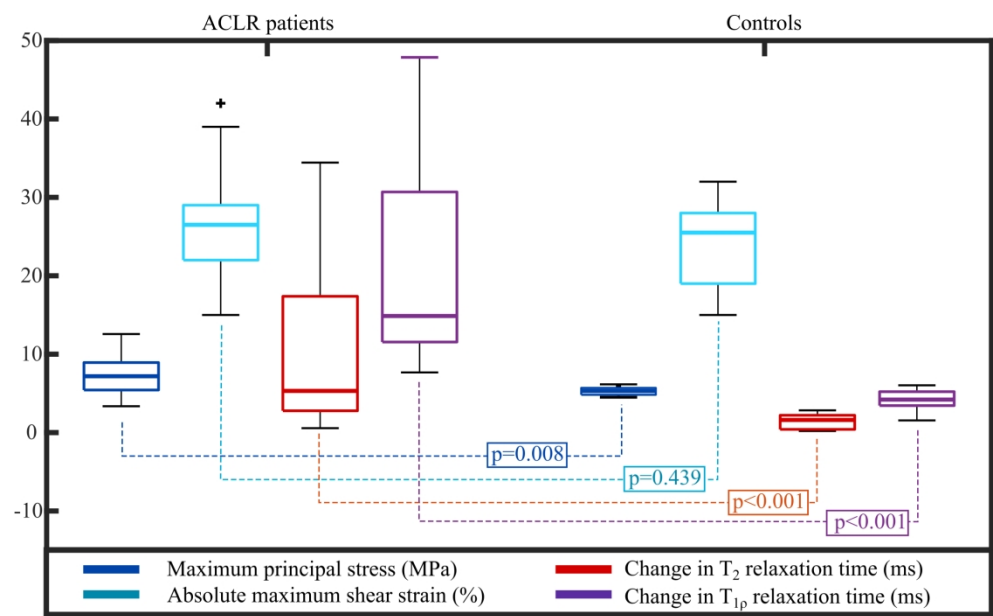
Axial view of the maximum principal (tensile) stress distribution at the 1-year follow-up time point and the T_2 relaxation time distribution at the 3-year follow-up time point in healthy controls. Peak values for tensile stresses are given for each compartment. The values for the T_2 relaxation time are given for the same location as peak tensile stresses. Note 1: The T_2 relaxation time distribution at the 1-year follow-up time point is not presented, since it is almost identical with the 3-year follow-up time point. Note 2: Both left and right knees are shown in the same coordinate system, with medial compartment on the left.



a) Correlation between the predicted degenerated volumes from the FE models at the 1-year time point and those estimated from T₂ between the 1-year and 3-year follow-up time points. b) Correlation between the peak values of maximum principal (tensile) stresses at the 1-year time point and changes in the T₂ relaxation time between the 1- and 3-year follow-up time points. Note: The values for the tensile stress and changes in the T₂ relaxation time are taken in the same location.



Correlation between the peak values of maximum principal (tensile) stresses at the 1-year time point and changes in the T2 relaxation time between the 1- and 3-year follow-up time points. Data points were grouped according to WORMS grades at the 3-year follow-up time point. The green, yellow and red areas indicate compartments at low, moderate and high risk of developing OA.



Box plots of the peak values of maximum principal (tensile) stresses and absolute maximum shear strains during the stance phase of gait at the 1-year time point and changes in the T₂ and T_{1ρ} relaxation times between the 1- and 3-year follow-up time points for patients with ACLR and healthy controls. Mann-Whitney U-test p-values are also provided.