We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



185,000

200M



Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

# Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



# Clusterin (APOJ) in Alzheimer's Disease: An Old Molecule with a New Role

Sarah K. Woody and Liqin Zhao

Additional information is available at the end of the chapter

http://dx.doi.org/10.5772/64233

#### Abstract

Clusterin (CLU), initially identified in 1983 as a "clustering factor" in ram rete testis fluid, is a multifaceted protein that was re-discovered and subsequently renamed eight times from 1983 to 1992. CLU exists as multiple protein isoforms including the 80 kDa glycosylated mature/secreted form of CLU (mCLU) and the smaller non-modified nuclear and intracellular forms of CLU (nCLU and icCLU, respectively). These isoforms, which are expressed at the highest levels in the brain, are suggested to play distinct roles in various disease processes such as those involving inflammation and apoptosis. Currently, CLU, also known as apolipoprotein J (APOJ) which belongs to the same protein family as apolipoprotein E (APOE), is the third most significant genetic risk factor for the development of late-onset Alzheimer's disease (LOAD); however, an extensive gap exists in the literature in understanding the physiological roles of CLU in normal brain and the pathogenic mechanisms conferred by CLU polymorphisms in the onset of LOAD. In this chapter, we discuss the status of the current knowledge regarding the generation and regulation of CLU protein isoforms, the clinical evidence and possible mechanisms involved in LOAD, and provide our perspectives for future studies.

**Keywords:** late-onset Alzheimer's disease (LOAD), genetic risk factors, clusterin (CLU), apolipoprotein J (APOJ), apolipoprotein E (APOE)

# 1. Introduction

#### 1.1. Alzheimer's disease: current status and challenges

Alzheimer's disease (AD) currently affects 35 million people worldwide, including 5.4 million Americans; a number that is estimated to triple by the year 2050 [1]. As the prevalence of AD



© 2016 The Author(s). Licensee InTech. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. increases, the AD-associated economic burden will also increase. In 2015, the direct costs associated with the care of AD patients in the United States reached \$226 billion. This number is predicted to reach \$1.1 trillion by the year 2050 making AD one of the most costly chronic illnesses in the world [1]. At present, AD is the sixth leading cause of death in the United States and is the only leading cause of death that cannot be prevented or cured. There are currently five FDA-approved drugs available to treat AD; however, these drugs do not address the underlying cause of AD and provide only temporary therapeutic relief in a fraction of the patients to whom they are administered. An extensive amount of clinical trials aimed at treating AD have been performed in the last 15 years; all of which have failed [2, 3]. These unanticipated challenges combined with the estimated rapid increases in AD prevalence stress the importance of identifying the underlying AD risk mechanisms that would allow prevention, risk reduction, and early intervention in the preclinical stage of AD.

### 1.2. Late-onset AD: complex etiology and risk factors

There are two types of AD: early-onset familial AD (FAD) and late-onset sporadic AD (LOAD). FAD is rare and mostly caused by inherited genetic mutations that result in abnormal overproduction of neurotoxic  $\beta$ -amyloid (A $\beta$ ) peptides. LOAD, the most common form of AD representing 95% of human cases, develops after age 60 and involves a heterogeneous and multifactorial etiology. It is now widely accepted that a person's risk for developing LOAD is primarily influenced by a combination of complex interactions between genetic and environmental risk factors. At present, age remains the most predominant risk factor for LOAD. It is estimated that one in nine (11.1%) senior citizens aged 65 or older have been diagnosed with LOAD; a ratio that increases to one in three (33.3%) by age 85 [1]. The National Institute on Aging (NIA) indicates that the risk of developing LOAD doubles every 5 years past the age of 65 [4]. Additionally, epidemiologic studies from the NIA estimate that the total percentage of senior citizens in the United States will increase by 7% by 2030 making senior citizens the fastest-growing age group in the United States and consequently the most at-risk population [4].

Sex also plays a significant role in the development of LOAD. Of the 5.4 million Americans currently living with AD, approximately 65% are women [1]. It was originally postulated that the higher percentage of women living with AD was due to the increased life span of the female population; however, as the average worldwide life expectancy of men and women differs by only 4 years, this presumption is invalid. A meta-analysis of seven sex-specific clinical studies revealed that women are 1.5 times more likely to develop AD than age-matched men, indicating that the female sex confers AD risk independent of age [5]. In addition to a higher incidence of AD, it is now well established that sex influences both the development and progression of LOAD. For example, female AD patients have been shown to exhibit more severe cognitive decline than men during the progression of AD pathology [6–8]. While the exact mechanisms underlying this sex bias are currently unknown, mounting evidence suggests that female vulnerability to AD is largely associated with the irreversible decline of female sex hormones during the onset of menopause [9–11]. However, despite these findings, the precise molecular mechanisms underlying female vulnerability remain uncharacterized.

Genetic predisposition is another prominent risk factor associated with the development of AD. A long-standing observation in the field of LOAD research is the significantly increased AD risk associated with possession of the human apolipoprotein E  $\varepsilon$ 4 allele (APOE  $\varepsilon$ 4) [12], the most predominant genetic risk factor for LOAD. Possession of the  $\varepsilon$ 4 allele is clinically associated with an increased rate and severity of cognitive decline, a younger age of onset, and altered response to AD treatments [13–16]. Moreover, the  $\varepsilon$ 4 allele has been shown to reduce brain glucose utilization [17], increase neuronal inflammation [18], and is associated with increased A $\beta$  dyshomeostasis [19, 20]. In addition to these data, studies have demonstrated that the APOE  $\varepsilon$ 4-associated AD risk is significantly more pronounced in the female population. For example, a recent clinical study conducted in a cohort of 8084 elderly individuals (healthy controls: n = 5496; MCI cases: n = 2588) demonstrated that the risk of clinical conversion from healthy aging to MCI or from MCI to AD conferred by the  $\varepsilon$ 4 allele was significantly greater in women than in men, a finding that corresponds with several earlier reports [21–25].

In addition to APOE  $\varepsilon$ 4, two of the largest genome-wide association (GWA) studies ever conducted to date have recently identified several other genetic risk factors that confer a significantly increased risk of developing LOAD [26, 27]. Of the genetic risk factors identified, clusterin (CLU), also known as apolipoprotein J (APOJ), was established as the third most predominant genetic risk factor for LOAD. CLU, which belongs to the same protein family as APOE, has been shown to regulate inflammation, oxidative stress, and amyloid homeostasis in the brain. Moreover, a recent study conducted by our laboratory indicated that CLU mRNA and protein expression levels are significantly reduced specifically in female brain during a time period that likely corresponds to the onset of reproductive senescence. These data suggest that, similar to APOE  $\varepsilon$ 4, CLU is also influenced by sex in the brain aging process and the pathogenesis of LOAD [28]. In the following sections, we summarize the current understanding of CLU protein isoforms and their biological functions with specific emphasis on the neuroprotective potential of CLU protein isoforms in the brain.

# 2. Clusterin: from form to function

# 2.1. CLU: discovery and nomenclature

In 1983, Blaschuk et al. identified a high-molecular weight protein in ram rete testis fluid [29]. Further analyses indicated that this unknown protein was capable of eliciting the "clustering" of Sertoli cells, mouse testis TM-4 cells, and erythrocytes resulting in the name clusterin. In 1984, Griswold and colleagues purified a dimeric acidic glycoprotein (DAG) from the Sertoli cells of rat testes [30]. This abundantly expressed but uncharacterized protein was detected at several molecular weights via reducing chromatography (41 and 29 kDa), western blot (27 and 21 kDa), and immunoprecipitation (70 kDa) [30]. In 1988, another study identified a "novel" protein in human serum. This heterodimeric protein had a molecular mass of 80 kDa, was composed of two 40-kDa chains, and was sequentially unique to all other proteins. Furthermore, it was concluded that this protein, which was deposited in the renal glomeruli of patients with glomerulonephritis, was integrally involved in kidney health [31]. As a result of these

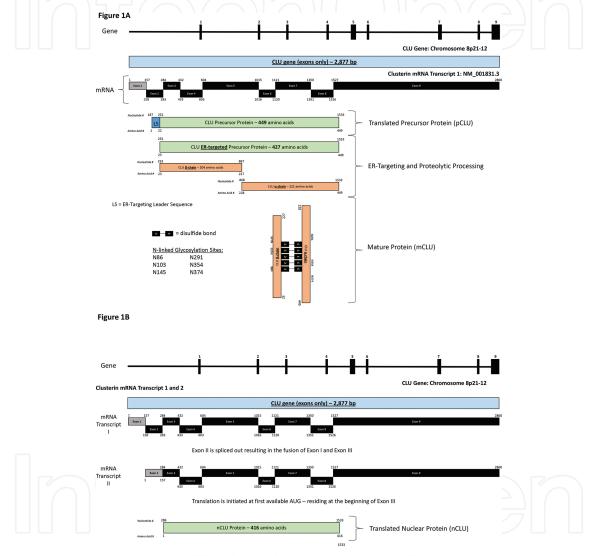
observations, Murphy and colleagues named this protein serum protein 40 kDa,40 kDa (SP-40,40) [31]. In 1990 and the years following, Harmony and colleagues identified and extensively characterized a component of high-density lipoproteins in human plasma which was referred to as apolipoprotein J (APOJ) [32]. However, upon the advent of DNA sequencing technology, it was determined that clusterin, DAG, SP-40,40, and APOJ were in fact the same protein. In the following decade, clusterin was "re-discovered" and subsequently re-labeled with other alternative names including testosterone-repressed prostrate message-2 (TRPM-2) [33], KU70-binding protein 1 (KUB1) [34], complement lysis inhibitor (CLI) [35], and sulphated glycoprotein-2 (SGP2) [36]. In 1992, a forum conducted at Cambridge University officially deemed this diverse protein clusterin (CLU).

### 2.2. CLU: from gene to protein

CLU is a single-copy gene located on the short arm of chromosome 8 (8p21-12) [37, 38] where it spans approximately 18,115 base pairs (bp). Upon the splicing of eight introns, this nineexon product spans approximately 2877 bp, and is transcribed into at least two distinct mRNA transcripts. CLU mRNA transcript 1 (NM\_001831.3), the most extensively characterized transcript, is translated into the mature/secreted isoform of CLU (mCLU) that has been predominantly identified and studied in the field of CLU research. CLU mRNA transcript 1 is initially translated into a 449-amino-acid precursor protein (pCLU, 60 kDa) beginning at a canonical translational start site located at base pair 187 in exon 2. This pCLU protein contains an N-terminal 22-amino-acid endoplasmic reticulum (ER)-targeted signaling peptide (amino acids 1–22 or bp 187–252) and two nuclear localization sequences in exon 3 (amino acids 78– 81 or bp 418–429) and exons 8–9 (amino acids 443–447 or bp 1513–1528). The translated pCLU protein is then targeted to the ER where the 22-amino-acid leader sequence (LS) is cleaved. Following LS cleavage, the peptide bond between R227 and S228 is cleaved resulting in the formation of two individual CLU subunits: the alpha subunit (CLU $\alpha$ , 34–37 kDa) and the beta subunit (CLUβ, 36–39 kDa). These two subunits are subsequently linked by five disulphide bonds to form an anti-parallel heterodimer [39]. N-glycosylation at six glycosylation sites is the final step in the generation of mCLU which, under nonreducing conditions, has a molecular weight of 75-80 kDa (Figure 1A) [40]. Alternatively, complete removal of exon 2 via alternative splicing results in the fusion of exons 1 and 3, thereby creating CLU mRNA transcript 2. In this secondary transcript, translation is initiated at a canonical translational start site located in exon 3. This results in the production of a CLU protein isoform that lacks the ER LS but retains the nuclear localization sequences. This alternative CLU isoform, which is non-ER targeted and unglycosylated, is shuttled between the cytoplasm and the nucleus and is referred to as "nuclear" CLU (nCLU, 49 kDa, Figure 1B). In addition to mCLU and nCLU, emerging evidence indicates that several different splicing variants of CLU also exist. These splicing variants, which are relatively uncharacterized, are suggested to lack portions of exon 2 and/or exon 5 and are generally referred to as "intracellular" isoforms (icCLU, 45, 50, and 53-55 kDa) [41-43].

Initial characterization by Harmony et al. indicated the expression of CLU mRNA in liver, lung, spleen, heart, reproductive tissues, and brain with predominant expression in brain and reproductive tissues [44]. Since this initial characterization, several other research groups,

including our own, have detected CLU mRNA and protein expression in many cell lines and tissue types tested. Moreover, CLU appears to be ubiquitously expressed on the subcellular level with multiple studies demonstrating CLU expression in the cytosol [45], nucleus [41], ER, and Golgi apparatus. Within the brain, CLU expression has been detected within neurons [46], astrocytes [46–48], microglia [49], and within the extracellular space [50]. While initial reports indicated that CLU was solely synthesized and secreted from the astrocytes in a manner similar to APOE [51], our recent findings demonstrate that pure cultures of primary neurons express



No ER-targeting leader sequence in protein – therefore protein is not processed in the ER

**Figure 1.** CLU transcription and translation. In humans, CLU is a single-copy gene located on the short arm of chromosome 8 that is comprised of nine exons spanning approximately 2.8 kb. (**A**) The mCLU isoform is generated from mRNA transcript 1 from a canonical translational start site in exon 2. The resulting precursor protein (pCLU), which contains an N-terminal ER-targeting leader sequence (LS), is transported to the ER where the 22-amino-acid LS is removed. CLU is then cleaved into the alpha and beta subunits and rapidly disulfide bonded and glycosylated to form an anti-parallel, heterodimeric glycoprotein: mCLU. (**B**) Alternatively, the nCLU isoform is generated from mRNA transcript 2. In this transcript, a splicing event removes exon 2 resulting in the fusion of exons 1 and 3. Translation is initiated at the beginning of exon 3 resulting in a truncated CLU isoform that lacks the ER-targeting LS. Therefore, the nCLU isoform, which retains the nuclear localization sequence, bypasses the ER/Golgi apparatus and is shuttled between the cytosol and the nuclear compartment. mCLU, nCLU, and to a lesser extent icCLU isoforms indicating that neurons are also capable of generating de novo CLU. Though the exact physiological functions of CLU remain a mystery, the nearly ubiquitous nature of CLU indicates the significance of this protein in cellular homeostasis.

## 2.3. CLU: transcriptional regulation

Though the gene promoter of CLU is highly conserved across species, the transcriptional regulation of CLU is complex as the predominant CLU transcriptional regulators appear to differ between tissue and cell type. However, despite the controversy in the literature, it is generally agreed that CLU is primarily upregulated by cellular injury, cytotoxic insult, and various stress stimuli [52-54]. For instance, Loisen and colleagues demonstrated that the CLU gene promoter contains an MG132 responsive region and a heat-shock element (HSE) indicating that proteasomal stress directly influences CLU transcription [52]. Another study demonstrated that the CLU gene promoter contains both HSEs and an activator protein-1 (AP-1) response element indicating direct transcriptional regulation by stimuli derived from cellular proliferation and differentiation [54]. In addition to these data, alternative stressrelated transcription factor response elements have been identified in the CLU gene promoter including a cAMP response element (CRE), an AP-2 response element, a specificity protein-1 (SP1) response element, and a glucocorticoid response element (GRE) [33, 53]. It has also been demonstrated that apoptotic stimuli modulates CLU transcription, specifically in cancer. An early study from Cervellera et al. identified a MYB binding site in the 5' flanking region of CLU and that B-MYB, a MYB family member that regulates cellular proliferation and apoptosis, directly bound to and transactivated the CLU gene [55]. CLU transcription is also regulated by several different growth factors including nerve growth factor (NGF) and transforming growth factor beta (TGFβ) [56–58]. For instance, it has been demonstrated that TGFβ induces the upregulation of CLU gene expression by stimulating the interaction between the CLU gene promoter and AP-1 [57]. An extension of these studies demonstrated that TGFβ deficiency resulted in the repression of CLU gene expression via interaction between c-Fos and the CLU gene promoter; an interaction that was abrogated upon cellular stimulation with TGFβ [58].

# 2.4. CLU: posttranslational modification

CLU is regulated by several types of posttranslational modification (PTM), the most predominant type being N-linked glycosylation. As previously indicated, mCLU is N-glycosylated at six different asparagine residues (N86, N103, N145, N291, N354, and N374) during ER-Golgi processing: a modification that comprises approximately 20–25% of the total mass of mCLU [59]. While glycosylation status was originally thought to have little to no impact on CLU function [40, 60], a recent study demonstrated that the chaperone activity of mCLU is dependent upon mCLU glycosylation [61]. This study also demonstrated that the glycosylation of nCLU did not result in chaperone activity indicating that glycosylation-mediated effects are specific to the mCLU isoform. It has also been established that complete deglycosylation of mCLU results in a 70–90% decrease in mCLU chaperone activity and a significant decrease in the number of  $\alpha$ -helices in the secondary structure of CLU. These data suggest that the lack of chaperone activity in deglycosylated mCLU could be, in part, due to the significant changes in secondary structure. Additionally, this study indicates that partially glycosylated mCLU retains chaperone activity suggesting that "core" glycosylation sites are crucial for mCLU function, while peripheral glycosylation may be dispensable [61]. Parallel to these findings, a study by Kang et al. indicated that ER stress, which inhibits protein glycosylation, resulted in rapid retro-translocation of mCLU from the ER yielding several hypo-glycosylated CLU isoforms. These hypo-glycosylated under normal conditions and cleared through proteasomal degradation. However, if the proteasome is chemically inhibited following ER stress, hypo-glycosylated that, contrary to what was originally postulated, glycosylation is crucial for mCLU chaperone activity.

In addition to N-linked glycosylation, CLU is also a primary target for ubiquitination and phosphorylation. It has been demonstrated that nCLU is a target for K63 ubiquitination through the ubiquitin E3 ligase, a product of von Hippel-Lindau (pVHL). However, contrary to the canonical function of protein ubiquitination, K63-linked ubiquitination of nCLU does not target nCLU for destruction, rather it promotes nCLU nuclear translocation for reasons that are currently unknown [63]. Pertaining to CLU phosphorylation, a recent proteomics study which focused on the identification of the serum phospho-proteome has identified three different phosphorylation sites at residues Thr393, Ser394, and Ser39 within the CLU protein. Additionally, a more recent study indicated that treatment of hepatocytes with 10-mM glucose and fructose significantly increased the levels of mCLU serine phosphorylation. This same study demonstrated increased mCLU serine phosphorylation in both the skeletal muscle and the liver of rats that were orally administered high doses of glucose and/or fructose indicating that phosphorylated CLU may interact with or respond to the activation of glucose-sensitive cellular bioenergetic pathways. In addition to ubiquitination and phosphorylation, an early report indicated that CLU is iodinated at 1 of the 12 tyrosine residues within the CLU protein. This iodination occurs within the apical plasma membrane of thyrocytes and is suggested to serve as a mechanism by which the thyroid gland can conserve iodine, which is relatively rare in the body [64]. It is also suggested that CLU activity is regulated by both sialylation [65] and acetylation [66]; however, definitive acetylation or sialylation sites have not been identified.

# 3. CLU in Alzheimer's disease: clinical findings

#### 3.1. CLU polymorphisms in LOAD

Since the initial determination of CLU SNP-associated AD risk by Harold et al. and Lambert et al. [26, 27], there have been approximately 40 independent follow-up meta-analyses and case-control studies that have examined the association between CLU SNPs and AD risk (**Table 1**). These reports were located through a PubMed search focused on topics pertaining to CLU SNPs in AD. Resulting articles were reviewed and those studies which provided a

listing of the CLU SNP(s) studied, population demographics, and a thorough description of cognitive assessment and statistical analysis were included in Table 1. Though conflicting evidence exists, the majority of the studies indicate that genetic variation in CLU increases the risk of developing AD and that this association is independent of APOE £4 status. There are approximately 355 identified SNPs in the CLU gene [67]; however, it appears that the primary risk-conferring CLU SNP is rs11136000. Of the 33 studies summarized in Table 1, 25 studies either include or exclusively focus on the impact of the rs11136000 SNP on AD risk; however, the results are inconsistent. Thirteen studies conclude that possession of rs11136000 does confer increased AD risk [26, 27, 68–77], while ten studies conclude no significant association between rs11136000 and AD [78-85]. Moreover, two studies conclude that possession of the rs11136000 SNP reduces risk of AD development [86, 87]. A possible explanation for these discrepancies may be found by examining the population ethnicities. Of the 13 studies that conclude rs11136000 confers AD risk, 11 studies are performed in a predominantly or exclusively western European or American Caucasian population. Alternatively, nine of the 10 studies that conclude no significant association (NSA) between rs11136000 and AD were performed in Asian, eastern European and Russian, Middle Eastern, or Hispanic populations indicating that the risk associated with the rs11136000 SNP may vary based on population ethnicity. Contrary to these data, two separate studies performed in exclusively German and American Caucasian populations found NSA between rs11136000 and AD risk. Moreover, the notion that rs11136000 does not confer AD risk in Asian populations is contradicted by two independent studies that indicate rs11136000-mediated AD risk in exclusively Chinese populations. As all the presented studies performed in Asian populations are adjusted for age, gender, and APOE status, and are comprised of numerically similar sample sizes, it is difficult to identify the exact reason underlying these discrepancies. One observation is that some studies have divided study populations into much smaller groups based upon the specific nucleotide substitution located at the rs11136000 SNP site (i.e. C, T, A substitution), while others have examined only rs11136000 carriers vs. non-carriers. The failure to stratify study populations based on the rs11136000 allele/genotype would have a significant impact on study outcome as the C allele of rs11136000 is considered the risk-conferring allele, while the A allele and T allele are considered normal and neuroprotective, respectively (i.e. C = risk allele, A = normal, and T = protective). Specifically, studies have indicated that the C allele confers a 1.16fold increased chance of developing LOAD and that 36% of Caucasians carry two copies of this AD-risk variant [26, 27]. Moreover, the C allele is associated with faster cognitive decline in preclinical AD [66] and lower memory scores in healthy elderly controls and elderly AD patients [67]. Young healthy carriers of the C allele exhibit neural hyperactivation in memoryassociated brain regions during working memory tasks [73], neural inefficiency in memoryrelated prefrontal and limbic areas during working memory [88], and reduced coupling between hippocampus and prefrontal cortex during memory processing [89]. Structurally, possession of the C allele is associated with diminished white matter integrity in several brain regions [90] and increased longitudinal ventricular expansion in elderly patients independent of APOE ɛ4 and dementia status [91]. Taken together, these data indicate that the rs11136000 SNP is significantly associated with the development of AD in predominantly Caucasian populations and that the rs11136000 AD-associated risk may be initiated several decades prior to the onset of AD.

In addition to rs11136000, another CLU SNP, rs9331888, which was also identified by Lambert and colleagues in the original GWA studies, has also been repeatedly investigated as an AD risk SNP. Of the 33 studies presented in **Table 1**, seven clinical studies and two meta-analyses examined the association of rs9331888 with AD risk [27, 69, 81, 83, 84, 92–95]. However, similar to that of rs11136000, the results vary and appear to be dependent upon population ethnicity. For instance, two separate meta-analyses conclude that rs9331888 confers AD risk in Caucasian but not Asian populations [92, 95]. However, two separate case-control studies performed in exclusively Chinese populations both indicate that rs9331888 is significantly associated with AD risk [84, 94]. In addition to differing and/or small sample sizes, one possible confounding factor could be sex of the study population. As sex modulates an individual's risk for LOAD, it is likely that stratification of study populations by sex will have a significant impact on the study results.

| CLU gene variant                     | Study and                | Study design and  | Diagnoses criteria   | Major findings   |
|--------------------------------------|--------------------------|---|--|--|
|                                      | year of<br>publication   | subjects  |  |  |
| rs11136000                           | Harold et al.<br>(2009)  | GWA study in European<br>and US population:<br><i>Stage 1 population:</i> AD cases<br>n = 3941, control cases:<br>n = 7848<br><i>Stage 2 population:</i> AD cases<br>n = 2023, control cases:<br>n=2340                                       | criteria for definite AD   | - rs11136000 SNP was<br>significantly associated<br>with the development of<br>LOAD but not the age of<br>onset. |
| rs2279590<br>rs9331888<br>rs11136000 | Lambert et al.<br>(2009) | GWA study in French,<br>Finnish, Italian, Spanish,<br>and Belgian population:<br><i>Stage 1 population:</i> AD cases:<br>n = 2032, control cases:<br>n = 5328<br><i>Stage 2 population:</i> AD cases:<br>n = 3978, control cases:<br>n = 3297 | AD diagnoses: DSM-III-R<br>and NINCDS-ADRDA<br>criteria for probable AD<br>s:Control criteria: Subjects<br>without DMS-III-R<br>dementia criteria and with<br>s:integrity of their cognitive<br>functions (MMSE >25) | - All CLU<br>polymorphisms<br>examined showed a<br>significant association<br>with AD development.               |
| rs2279590<br>rs9331888<br>rs11136000 | Yu et al.<br>(2010)      | Case-control study in Han<br>Chinese population:<br><i>AD cases:</i><br>n = 324, AOO > 65 years, 18<br>females: age = 76.87 ± 5.58<br><i>Control cases:</i><br>n = 388, 211 females:<br>age = 75.93 ± 4.69                                    | ADRDA criteria<br>for probable AD; no family   | - rs2279590 showed   |

| CLU gene variant                  | Study and<br>year of<br>publication | Study design and subjects  | Diagnoses criteria   | Major findings  |
|-----------------------------------|-------------------------------------|--|--|---|
|                                   |                                     |  | and examination and<br>MMSE score > 28<br>Subjects with CHF,<br>MI, T2DM, and AS were<br>excluded from study   | Han Chinese<br>population.  |
| rs11136000                        | Seshadri et al.<br>(2010)           | Three-stage GWA study in<br>a white population:<br>Stage 1 population: Dementia<br>-free subjects at start:<br>n = 8935, AD cases:<br>n = 2033, dementia-free<br>control cases:<br>n = 14,642<br>Stage 2 population: AD cases<br>n = 2032, control cases:<br>n = 5328<br>Stage 3 population: AD cases<br>n = 3333, control cases:<br>n = 6995<br>Independent case-control<br>replication population:<br>Ethnicity — Spanish, AD<br>cases:<br>$n = 1140$ , age = $78.8 \pm 7.9$ ,<br>69.9% female; control cases<br>$n = 1209$ , age = $49.9 \pm 9.2$ ,<br>58.2% female | ADRDA criteria for definite<br>probable, or possible AD;<br>AD pathology confirmed a<br>autopsy<br>:   | significantly associated<br>with increased risk for<br>LOAD in all study<br>populations analyzed.                     |
| rs7982<br>rs7012010<br>rs11136000 | Jun et al.<br>(2010)                | Meta-analysis in nine<br>European white cohorts<br>and five non-European<br>cohorts (African American,<br>Israeli-Arab, and Caribbear<br>Hispanic):<br>AD cases:<br>n = 7070<br><i>Control cases:</i><br>n = 8169  |  | - All CLU<br>polymorphisms<br>examined demonstrated<br>a significant association<br>with AD in only white<br>cohorts. |
| rs11136000                        | Corneveaux et<br>al. (2010)         | GWA study of a European<br>population<br><i>AD cases:</i><br><i>n</i> = 1019, 652 females,<br>367 males<br><i>Control cases:</i>   | AD diagnoses: Clinically<br>diagnosable dementia<br>at time of death and<br>neuropathological<br>confirmation of AD (Braak<br>stage V or VI) upon autops | - rs11136000 SNP was<br>significantly associated<br>with LOAD.  |

| CLU gene variant | Study and<br>year of<br>publication | Study design and subjects  | Diagnoses criteria   | Major findings   |
|------------------|-------------------------------------|--|--|--|
|                  | te                                  | <i>n</i> = 591, 285 females, 306 males   | <i>Control criteria:</i> Without<br>clinically diagnosable<br>dementia at time of death;<br>autopsy confirmation of an<br>absence of<br>neuropathological<br>hallmarks (Braak stage <<br>III)      | ÐN   |
| rs11136000       | Jessen et al.<br>(2010)             | Longitudinal cohort study<br>in German population:<br><i>No memory impairment:</i> 685<br>females, 342 males,<br>age = $79.4 \pm 3.4$<br><i>Memory impairment without</i><br><i>worry:</i> 591 females, 415<br>males, age = $79.8 \pm 3.6$<br><i>Memory impairment with</i><br><i>worry:</i> 273 females, 109<br>males, age = $79.8 \pm 3.5$ | ADRDA criteria<br>for probable AD  | - The rs1113600 AD-risk<br>variant is associated with<br>low plasma CLU levels in<br>cognitively intact<br>healthy controls and<br>numerically (but non-<br>significantly) associated<br>with lowered plasma<br>CLU in AD cases. |
| rs11136000       | Lancaster et al.<br>(2011)          | fMRI study in young<br>Caucasian cohort:<br><i>Subjects:</i><br>n = 43, 22 males,<br>21 females,<br>age 18–51 Subjects were<br>genotyped for<br>rs11136000 SNP and<br>pooled according<br>to genotype:<br>CC = risk group (<br>n = 13) and CT/TT = non-risk<br>group (<br>n = 24/6)  | Inclusion criteria:<br>No personal or family<br>history of neuropsychiatric,<br>neurological, or<br>neurodegenerative<br>disease; no chronic somatic<br>illnesses or history<br>of substance abuse | during working memory<br>tasks in the frontal and  |
| rs11136000       | Schurmann<br>et al. (2011)          | n = 24/6)<br>GWA study on a subset of<br>participants from the<br>German Study on Aging<br>Cognition and Dementia:<br><i>AD cases:</i><br>n = 67, 47 females, 20 males<br>age = 85.3±3.7<br><i>Control cases:</i><br>n = 191, 134 females, 57<br>males, age = 83.7±3.2   |  | - rs11136000 AD-risk<br>variant was associated<br>with low plasma CLU<br>levels.   |

| CLU gene variant  | Study and<br>year of<br>publication | Study design and subjects  | Diagnoses criteria  | Major findings  |
|---|-------------------------------------|--|---|---|
| Rs7982<br>Rs572844<br>rs1532277<br>rs2279590<br>Rs9331888<br>rs10503814 | Komatsu et al.<br>(2011)            | Case-control<br>study in<br>Japanese population:<br>AD cases:<br>n = 180, 101 females,<br>79 males, age = 67.4±6.7<br>Control cases:<br>n = 130, 67 females, 63<br>males, age = 64.4±6.7   | AD diagnoses: NINCDS-<br>ADRDA criteria;<br>subjects had no<br>family history of AD<br><i>Control criteria</i> :<br>No history of dementia or<br>other neuropsychiatric<br>disorders    | - No association was<br>detected between CLU<br>SNPs and AD in a<br>Japanese population.  |
| rs11136000  | Golenkina<br>et al. (2010)          | Cohort study in a Russian<br>population:<br><i>AD cases:</i> Early-onset—<br>n = 214, AOO = 56.9 ± 5.38<br>Late-onset—<br>n = 320, AOO 72.2 ± 5.04<br><i>Control cases:</i><br>Moscow region:<br>n = 343, age range = 35–85,<br>age = 60.96 ± 7.94 Ural<br>region:<br>n = 160, age range = 69–89,<br>age = 73.87 ± 3.87 Siberian<br>region:<br>n = 199, age range = 41–96,<br>age = 61 ± 15.34 | AD diagnoses: NINCDS-<br>ADRDA criteria, ICD-10<br>criteria, and DSM-IV criteria<br><i>Control criteria</i> : Cognitively<br>intact individuals   |   |
| rs881146<br>rs11136000<br>rs17057441<br>rs70120100                      | Lee et al.<br>(2011)                | Nested case-control<br>GWAS in a cohort of<br>Caribbean Hispanic<br>subjects:<br><i>AD cases</i> :<br>n = 549, age of<br>onset = 79.98 ± 8.0<br><i>Control cases</i> :<br>n = 544  | Dementia diagnoses:<br>Diagnoses established on<br>the basis<br>of all available information<br>gathered from initial and<br>follow-up studies<br>AD diagnoses:<br>NINDS-ADRDA criteria | - rs881146 SNP was<br>significantly associated<br>with LOAD -<br>rs11136000 and other<br>SNPs were not<br>significantly associated<br>with LOAD in a<br>Caribbean Hispanic<br>population. |
| rs11136000  | Ma et al.<br>(2011)                 | Case-control study in<br>Chinese Han population:<br>AD cases:<br>n = 127, 73 females, 54<br>males,<br>age = $73.12 \pm 8.58$<br>Control cases:<br>n = 143, 79 females, 64<br>males, age = $73.80 \pm 6.30$   | AD diagnoses: 2007 revised<br>AD diagnoses criteria<br><i>Control criteria</i> :<br>No history of neurological<br>disease and MMSE score ><br>29  | significantly associated with LOAD in Chinese   |

| CLU gene variant                                  | Study and<br>year of<br>publication | Study design and subjects  | Diagnoses criteria   | Major findings  |
|---|-------------------------------------|--|--|---|
| rs11136000  | Braskie et al<br>(2011)             | Brain imaging study of<br>Australian Caucasian<br>population:<br><i>Subjects:</i><br>n = 398, age<br>range = 20–29, mean age =<br>23.6 ± 2.2   | Subject criteria: Healthy,<br>young, right-handed<br>Australian Caucasian twin<br>containing the rs11136000<br>genotype and ventricle size<br>consistent with a healthy<br>adult | matter integrity in the   |
| rs11136000  | Ferrari et al.<br>(2012)            | Case-control study in a<br>Caucasian-American<br>population:<br>AD cases:<br>n = 342, age = 76.78 ± 8.6<br>Control cases:<br>n = 277, age = 70.21 ± 8.6  | <i>AD diagnoses:</i> NINCDS-<br>ADRDA criteria<br><i>Control criteria:</i> Subject<br>within cognitively normal<br>limits on a standard<br>psychometric test                     | - rs11136000 was<br>significantly associated<br>with LOAD.  |
| rs2279590rs11136000                               | Kamboh et al.<br>(2012)             | Case-control study in<br>Caucasian-American<br>population:<br>AD cases:<br>n = 1348,<br>$AOO = 72.6 \pm$<br>6.4, $65.6%$ female<br>Control cases:<br>$n = 1359$ , age = $74.7 \pm 6.5$ ,<br>60.8% female | AD diagnoses: NINCDS-<br>ADRDA criteria for<br>probable or definite AD<br><i>Control criteria:</i><br>Non-demented Caucasian-<br>American over 60 years of<br>age                | - No significant<br>association was observed<br>between CLU SNPs and<br>AD in<br>case-control study.  |
| rs7982  | Karch et al.<br>(2012)              | GWA study in<br>Euro-American population<br><i>AD cases:</i><br>$n = 73$ , age = $87 \pm 7$ , 42% mal<br><i>Control cases:</i><br>$n = 39$ , age = $86 \pm 9$ , 44%<br>male                              | AD diagnoses: Autopsy<br>n:confirmed AD<br><i>Control criteria</i> : Age-<br>ematched cognitively norma<br>controls  | <ul> <li>rs7982 was associated<br/>with disease status.</li> <li>Elevated CLU levels are<br/>classociated with AD<br/>brains.</li> <li>CLU is altered at the<br/>mRNA level in AD<br/>brain.</li> </ul> |
| rs3087554<br>rs9331942<br>rs9331949<br>rs11136000 | Lin et al.<br>(2012)                | Case-control study in<br>Taiwanese population:<br><i>AD cases:</i><br><i>n</i> = 268,<br><i>Control cases:</i><br><i>n</i> = 389   | <i>AD diagnoses:</i> DSM-IV<br>criteria and<br>NINCDS-<br>ADRDA criteria<br><i>Control cases:</i> Assessed via<br>Short Portable Mental<br>Status Questionnaire                  | - rs11136000 was<br>associated with<br>significantly reduced<br>risk for AD.  |
| rs2279590<br>rs9331888                            | Chen et al.<br>(2012)               | Case-control study in<br>southern Chinese<br>population:   | <i>AD diagnoses:</i> NINCDS-<br>ADRDA criteria and no<br>family history of AD  | - rs2279590 and<br>rs11136000 SNPs confer<br>susceptibility to AD in  |

| CLU gene variant | Study and<br>year of<br>publication     | Study design and subjects  | Diagnoses criteria   | Major findings  |
|------------------|---|--|--|---|
| rs11136000       |   | AD cases:<br>n = 462<br>Control cases:<br>n = 350  | <i>Control criteria</i> : Cognitively<br>normal individuals as<br>indicated by CDR scale   | southern Chinese<br>population.   |
| rs9331888        | Xing et al.<br>(2012)                   | Case-control study:<br>AD cases:<br>$n = 104$ , AOO = $\geq 65$ , age<br>$= 80.20 \pm 5.57$ , 63 females, 4<br>males<br>Control cases:<br>$n = 104$ , age = $79.32 \pm 5.37$ , 55<br>females, 46 males                       | AD diagnoses: NINCDS-<br>ADRDA criteria for<br>probable AD<br>1 <i>Control criteria</i> : Confirmed<br>healthy by medical history,<br>medical examination, and<br>8MMSE score > 28 | CLU protein and mRNA  |
| rs11136000       | Klimkowicz-<br>Mrowiec et al.<br>(2012) | Case-control study in a<br>Polish population:<br><i>AD cases:</i><br>$n = 253$ , age = $73.9 \pm 5.8$ , 173<br>females<br><i>Control cases:</i><br>$n = 240$ , age = $73.8 \pm 6.9$ , 133<br>females                         | of AD<br>Control criteria: MMSE >  | - No significant<br>association between<br>rs11136000 and the AD in<br>a Polish population.   |
| 18 CLU SNPS      | Yu et al.<br>(2013)                     | Case-control study in Han<br>Chinese population:<br><i>AD cases:</i><br>$n = 796$ , AOO = $\geq 65$ , age<br>= 74.3 $\pm$ 7.0, 396 females<br><i>Control cases:</i><br>$n = 796$ , age = 73.9 $\pm$ 6.5, 388<br>females      | ADRDA criteria for<br>probable AD. No family<br>history of<br>neurodegenerative<br>disorders or dementia   | - Of the 18 SNPs tested,<br>only the C allele (major<br>allele) of rs9331949 was<br>significantly associated<br>with AD in the Han<br>Chinese population. |
| rs11136000       | Thambisetty et<br>al. (2013)            | Two-part longitudinal<br>study from Baltimore<br>Longitudinal Aging Study<br><i>Study 1 population:</i><br>n = 88, age = 69, age range<br>= 56–86<br><i>Study 2 population:</i><br>n = 599, age = 67.5, age range<br>= 60–93 | inflammation; Subject<br>without cognitive<br>impairment as indicated by<br>NINCDS-ADRDA   | subjects carrying the<br>CLU risk allele exhibit<br>increased rCBF in brain<br>regions intrinsic to   |

| CLU gene variant                                  | Study and<br>year of<br>publication | Study design and subjects  | Diagnoses criteria   | Major findings  |
|---|-------------------------------------|--|--|---|
|   |                                     |  |  | memory decline over<br>non-carriers.  |
| rs11136000  | Pedroza et al.<br>(2014)            | Association study in a whit<br>and black population:<br><i>AD cases:</i><br>n = 44 black,<br>n = 432 white, age = 78.9, ag<br>range = 52.2–91.2<br><i>Control cases:</i><br>n = 224 black,<br>n = 2219 white, age = 78.7,<br>age range = 60.5–96.4 | ADRDA criteria for<br>probable AD<br><i>Control criteria:</i> CDR score  | - The minor allele of<br>rs11136000 may confer<br>enhanced memory in<br>whites.   |
| rs1532278<br>rs2279590<br>rs9331888<br>Rs11136000 | Lu et al.<br>(2014)                 | Case-control study in<br>southern Han Chinese<br>population:<br>AD cases:<br>$n = 499$ , age = $69.990 \pm 9.96$<br>Control cases:<br>$n = 592$ , age = $68.930 \pm$<br>9.390  | Not provided   | - No significant<br>association was detected<br>between CLU SNPs and<br>LOAD in southern Han<br>Chinese population.   |
| rs1532278   | Patel et al.<br>(2014)              | Prospective cohort study in<br>a British Caucasian cohort<br>with Down syndrome:<br><i>Subjects:</i><br>n = 304 Down syndrome<br>patients, age > 16  | nDementia diagnoses: ICD-10<br>research criteria   | - No significant<br>association between<br>rs1532278 and the<br>development of<br>dementia in a cohort of<br>Caucasian Down<br>syndrome patients.   |
| rs11136000  | Lancaster et al.<br>(2015)          | fMRI study in young<br>Caucasian population:<br><i>Subjects:</i><br><i>n</i> = 85, age range = 19–47   | Inclusion criteria: Healthy,<br>right-handed, young<br>Caucasians with no history<br>of mental illness or drug<br>abuse                                  | - Carriers of the<br>rs11136000 risk variant<br>exhibit higher activation<br>levels in memory-related<br>pre-frontal and limbic<br>areas during working<br>memory tasks.  |
| rs11136000  | Sen et al.<br>(2015)                | Case-control study in a<br>Turkish population: <i>AD</i><br><i>cases:</i><br>n = 112, age range = 65–98,<br>age = 73.59 $\pm$ 7.59 <i>Control</i><br><i>cases:</i><br>$n = 106$ , age = 74.04 $\pm$ 5.29   | <i>AD diagnoses:</i> NINCDS-<br>ADRDA criteria for<br>probable AD—no family<br>history of dementia <i>Control</i><br><i>criteria:</i> Cognitively intact | - No significant<br>association was observed<br>between rs11136000 and<br>AD in the entire Turkish<br>population Turkish<br>females carrying the<br>rs11136000 TT genotype<br>exhibited increased<br>BEHAVE-AD scores |

| CLU gene variant | Study and<br>year of<br>publication | Study design and subjects  | Diagnoses criteria   | Major findings   |
|------------------|-------------------------------------|--|--|--|
| rs11136000       | Sohrabifar                          | Case-control study in an   | Not provided   | suggesting a possible<br>association between the<br>TT genotype and female<br>Turkish subjects.<br>- No significant  |
|                  | et al. (2015)                       | Iranian population:<br>$AD \ cases:$<br>n = 160<br>$Control \ cases:$<br>n = 163   |  | association between<br>rs11136000 and AD in an<br>Iranian population.  |
| rs9331888        | Toral-Rios<br>et al. (2015)         | Case-control study in a<br>Mexican population:<br><i>AD cases:</i><br>n = 94, age > 60<br><i>Control cases:</i><br>n = 100, age > 60   | AD diagnoses: NINCDS-<br>ADRDA criteria<br>Control criteria: MMSE<br>≥ 24, no memory<br>complaints,<br>no acute or severe chronic<br>illness   | - No significant<br>association between<br>rs9331888 and AD in a<br>Mexican population.  |
| rs9331888        | Shuai et al.<br>(2015)              | Meta-analysis of 11<br>case-control studies:<br><i>Ethnicities:</i> Caucasian and<br>Asian populations<br><i>AD cases:</i><br>n = 8766<br><i>Control cases:</i><br>n = 11,366      | Study inclusion criteria:<br>(1) Study evaluated<br>rs9331888 SNP and AD rish<br>(2) Case-control design<br>(3) Sufficient study<br>population was provided  | -Significant association<br>between rs9331888 and<br>AD in Caucasian<br>population among<br>allelic, additive, and<br>recessive models.<br>- No association in<br>combined population or<br>only Asian population. |
| rs2279590        | Zhang et al.<br>(2015)              | Meta-analysis of 11<br>case-control studies:<br><i>Ethnicities</i> : Caucasian and<br>Asian populations<br><i>AD cases</i> :<br>n = 8605<br><i>Control cases</i> :<br>n = 12,050   | Study inclusion criteria:<br>(1) Study evaluated<br>rs2279590 SNP and AD rish<br>(2) Case-control design<br>(3) Study provided the<br>number of rs2279590<br>genotypes<br>(4) Study provided OR with<br>a 95% CI | Asian population among<br>additive and recessive<br>models.  |
| rs9331888        | Zhang et al.<br>(2015)              | Meta-analysis of 12<br>case-control studies:<br><i>Ethnicities</i> : Caucasian and<br>Asian populations<br><i>AD cases</i> :<br>n = 16,876<br><i>Control cases</i> :<br>n = 19,295 | Study inclusion criteria:<br>(1) Study evaluated<br>rs9331888 SNP and AD rish<br>(2) Case-control design<br>(3) Study provided the<br>number of SNP genotypes<br>(4) Study provided OR<br>with a 95% CI          | <ul> <li>Significant association<br/>detected in pooled</li> <li>population.</li> <li>Subgroup analysis<br/>demonstrates a<br/>significant association<br/>between rs9331888 and<br/>AD in Caucasian</li> </ul>    |

| CLU gene variant | Study and<br>year of<br>publication | Study design and subjects | Diagnoses criteria | Major findings                       |
|------------------|-------------------------------------|---------------------------|--------------------|--------------------------------------|
|                  |                                     |                           |                    | population but not Asian population. |

Abbreviations: Age of onset (AOO), behavioural pathology in Alzheimer's disease (BEHAVE-AD), the Consortium to Establish a Registry for Alzheimer's Disease (CERAD), Clinical Dementia Rating (CDR), congestive heart failure (CHF), Diagnostic and Statistical Manual of Mental Disorders, Third Edition, Revised (DSM-III-R), Diagnostic and Statistical Manual of Mental Disorders, Fourth Edition (DSM-IV), functional magnetic resonance imaging (fMRI), genome-wide association (GWA), mini-mental state examination (MMSE), National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer's Disease and Related Disorders Association (NINCDS-ADRDA), odds ratio (OR), type 2 diabetes mellitus (T2DM).

Table 1. CLU polymorphisms in AD (2009–2016).

#### 3.2. CLU as an AD biomarker

In 1992, it was suggested that peripheral CLU (then referred to as SGP-2) expression may serve as a potential biomarker for predicting the onset and/or severity of neurodegenerative disorders such as LOAD [96]. Though this concept was proposed over 20 years ago, the possibility of CLU as an AD biomarker is only recently being examined. Since 2010, 10 different studies have been performed with the aim of determining the validity of CLU as an AD biomarker (Table 2). However, the conclusions of these studies are contradictory at best. Of the 10 studies presented in Table 2, six studies conclude that increased plasma CLU levels are associated with increased rate of cognitive decline [97], increased white matter atrophy [98], increased risk for AD [99], and were indicative of greater fibrillar Aβ burden [100]. However, contrary to these findings, four studies conclude that CLU levels are not significantly different between control subjects and subjects with MCI, AD, or dementia, suggesting that peripheral CLU is unreliable as an AD biomarker [101–105]. One primary difference between these studies is the fluid that was analyzed for CLU concentration. The six studies concluding that CLU would be a reliable biomarker utilize plasma samples for analysis, whereas the three of the four studies indicating no difference between control and AD subjects measure serum or platelets. Another key difference between these conflicting reports is the sample size. In three of the four studies concluding that CLU would not be a reliable AD biomarker, the sample size per group is less than 70 subjects, whereas most of the studies indicating the possibility of CLU as a peripheral biomarker contain several hundred subjects per group. Therefore, it is also possible that these differences are the result of inadequate sample size. Despite these discrepancies, these studies collectively suggest that at least plasma CLU could provide a predictive biomarker for determining the risk for AD.

| Study and year of publication | Fluid<br>analyzed | Study design and subjects | Diagnoses criteria    | Major findings           |
|-------------------------------|-------------------|---------------------------|-----------------------|--------------------------|
| Thambisetty et al.            | Plasma            | Prospective cohort study: | AD diagnoses: NINCDS- | -Increased plasma        |
| (2010)                        |                   | Subjects:                 | ADRDA criteria for    | concentration of CLU was |
|                               |                   | n = 844                   | probable AD           |                          |

| Study and year of                       |           | Study design and   | Diagnoses criteria  | Major findings  |
|---|-----------|--|---|---|
| publication                             | analyzed  | subjects   |   |   |
|   | ft(E      | <i>Ethnicity:</i> White European<br>(UK, France, Italy,<br>Finland, Poland, Greece)<br>derived from the<br>KLC-ART and<br>AddNeuroMed<br>cohort studies and the<br>Baltimore Longitudinal<br>Study of Aging  | MCI diagnoses: Subjective<br>memory complaints, CDR<br>scores of less than 1, and<br>evidence of objective<br>memory impairment using<br>the CERAD criteria<br><i>Control criteria</i> : Subjects<br>with no MCI and MMSE ≥<br>28 | fibrillar Aβ burden.  |
| Schrijvers et al.<br>(2011)             | Plasma    | Case-cohort study from<br>the Rotterdam Study in<br>the Netherlands:<br><i>Subjects:</i> 60 individuals<br>with prevalent AD at<br>baseline, a sub-cohort of<br>926 subjects, and an<br>additional 156 subjects<br>diagnosed with AD<br>throughout follow-up<br>time | Study outcome:<br>Prevalent AD<br>AD diagnoses: Severity of<br>AD measured by the<br>MMSE score, and the risk<br>of developing AD during<br>follow-up examinations  |   |
| Ijsselstijn et al. (2011                | )Serum    | Case-control study<br>derived from the<br>Rotterdam Scan Study:<br>AD cases:<br>$n = 43$ , age = $78 \pm 6.5$ , 32<br>females<br>Control cases:<br>$n = 43$ , age = $78 \pm 6.8$ ,<br>32 females   | AD diagnoses: DSM-III R<br>criteria<br>Control criteria: MMSE<br>≥ 28   | - No significant difference<br>in serum CLU levels<br>between pre-symptomatic<br>AD and controls ( <i>p</i> -value =<br>0.54).  |
| Thambisetty et al.<br>(2012)            | Plasma    | Longitudinal cohort<br>study:<br>139 cognitively intact<br>subjects,<br>age = 70.5   | Baseline criteria: Free of<br>clinical diagnosis of<br>dementia at evaluation<br>MCI diagnoses: Petersen<br>criteria<br>Dementia diagnoses:<br>DSM III criteria   | <ul> <li>Higher baseline</li> <li>concentration of plasma</li> <li>was associated with slower</li> <li>rates of brain atrophy.</li> <li>Peripheral concentration</li> <li>of CLU appear to reflect</li> <li>concentrations in AD-</li> <li>vulnerable brain regions.</li> </ul> |
| Mukaetova-<br>Ladinska et al.<br>(2012) | Platelets | Case-control study:<br>AD cases:<br>n = 25, age = 78.08 ± 1.0, 10<br>females<br>Control cases:<br>n = 26, age = 70.81<br>± 1.98, 18 females  | <i>AD diagnoses:</i><br>NINCDS-ADRDA criteria<br>for probable AD<br><i>Control criteria:</i> Subjects<br>with no cognitive and/or<br>neurological problems  | - No significant difference<br>in platelet CLU levels<br>between control and AD<br>patients.  |
| Silajdzic et al. (2012                  | !)Plasma  | Quantitative ELISA assessment of plasma  | <i>AD diagnoses:</i> DSM-IIIR criteria and NINCDS-  | - No significant difference<br>in plasma CLU levels   |

| Study and year of           |                | Study design and  | Diagnoses criteria   | Major findings   |
|-----------------------------|----------------|---|--|--|
| publication                 | analyzed       | subjectsCLU levels: $AD$ cases: $n = 127$ $Dementia$ cases: $n = 82$ $Depression$ cases: $n = 30$ $Control$ cases: $n = 171$                | ADRDA criteria for<br>probable AD<br>VaD diagnoses: DSM-IIIR<br>criteria and<br>NINDS-AIREN criteria for<br>probable dementia<br>DLB diagnoses: Consensus<br>criteria by McKeith and<br>McKhann<br>Control criteria: No<br>memory complaints | between control cases and<br>AD, dementia or<br>depression cases.  |
| Song et al. (2012)          | Plasma         | Longitudinal cohort<br>study—Sydney Memory<br>and Aging Study:<br>$MCI \ cases:$<br>n = 257<br>$Control \ cases:$<br>n = 407                | <i>MCI diagnoses:</i><br>International consensus<br>criteria<br>and CDR > 0.5  | <ul> <li>CLU plasma levels were<br/>negatively correlated with<br/>gray matter volume and<br/>positively correlated with<br/>CSF volume.</li> <li>Higher plasma CLU levels<br/>predict white matter<br/>atrophy over 2 years in<br/>elderly subjects.</li> </ul> |
| Sattlecker et al.<br>(2014) | Whole blood    | Prospective cohort<br>study — AddNeuroMed<br>Biomarker Study:<br>AD cases:<br>n = 331<br>MCI cases:<br>n = 149<br>Control cases:<br>n = 211 | Not provided   | - Increased plasma CLU is<br>significantly associated<br>with increased rate of<br>cognitive decline.  |
| Jongbloed et al.<br>(2015)  | CSF and plasma | Quantitative diagnostic<br>study:<br>AD cases:<br>n = 107<br>MCI cases:<br>n = 50<br>Control cases:<br>n = 67                               | AD diagnoses: NINCDS-<br>ADRDA criteria for<br>probable AD<br><i>MCI diagnoses:</i> Petersen's<br>criteria<br><i>Control criteria:</i><br>Cognitively healthy<br>spouses or<br>relatives of AD group   | <ul> <li>Elevated plasma CLU was<br/>associated with increased<br/>risk for AD and related to<br/>cognitive decline in MCI<br/>patients.</li> <li>Plasma CLU is inversely<br/>related to cognitive decline<br/>in AD patients.</li> </ul>                        |
| Dukic et al. (2016)         | Serum          | Quantitative comparison<br>of serum CLU levels:<br>AD cases:<br>n = 70<br>Dementia cases:<br>n = 67<br>MCI cases:<br>n = 48                 | AD diagnoses: NINCDS-<br>ADRDA criteria<br>VaD diagnoses: NINCS-<br>AIREN criteria<br>MCI diagnoses: Peterson's<br>criteria  | - Serum concentrations of<br>CLU did not differ between<br>groups.   |

| Study and year of | Fluid    | Study design and | Diagnoses criteria            | Major findings |
|-------------------|----------|------------------|-------------------------------|----------------|
| publication       | analyzed | subjects         |                               |                |
|                   | Contro   | Control cases:   | Control criteria: Cognitively |                |
|                   |          | n = 50           | healthy spouses of AD         | and            |
|                   |          |                  | VaD patients                  |                |

Table 2. CLU as an AD biomarker (2010–2016).

# 4. CLU in the brain: mechanisms of action

Of the known CLU isoforms, mCLU is by far the most studied and has been described as a chaperone-like protein that clears misfolded proteins, cellular debris, and protein aggregates from the cytosol and extracellular space [106–113]. However, the nCLU and icCLU isoforms remain relatively uncharacterized. Several reports have suggested that nCLU and icCLU exhibit solely proapoptotic characteristics; however, results vary across laboratories and are inconsistent [41, 42, 114–116]. This section reviews the available literature pertaining to CLU isoforms in the brain with particular emphasis on the molecular mechanisms by which CLU protein isoforms regulate amyloid homeostasis, inflammation, and apoptosis.

### 4.1. CLU and Aβ homeostasis

In the early 1990s, CLU mRNA and protein levels were found to be significantly elevated in AD brain, specifically in the frontal cortex and hippocampus of post-mortem AD brain tissue [117, 118]. Shortly after these discoveries, McGeer et al. demonstrated robust CLU immunoreactivity within senile plaques [119]. It was further demonstrated that mCLUbound soluble Aβ proteins in the cerebral spinal fluid (CSF) [120] and that CLU expression increased the solubility of A $\beta$  and prevented A $\beta$  aggregation [121]. These data strongly suggested that CLU may play an important role in the pathogenesis of AD via regulation of brain amyloid burden. However, contrary to these findings, it has also been demonstrated that increased CLU expression exacerbated A<sub>β</sub>-induced neurotoxicity [122]. Moreover, DeMattos et al. demonstrated that A $\beta$  plaque formation was facilitated by CLU in an animal model of AD suggesting that CLU exerts a negative impact on the brain in the development of AD pathology [123]. These literary contradictions continued to persist until 2007 when a study by Yerbery and colleagues provided a possible explanation for the simultaneously pro- and anti-amyloidogenic effects associated with mCLU [124]. This study indicated that the pro-amyloidogenic effects of mCLU were restricted to conditions in which A $\beta$  was present in a very large molar excess. Under these conditions, mCLU, which functions as a chaperone-like protein to temporarily stabilize misfolded proteins [125], bound to and stabilized A $\beta$  thereby facilitating A $\beta$  aggregation. Alternatively, when mCLU was present at much higher but still substoichiometric levels (i.e. a molar ratio of clusterin:  $A\beta = 1:10$ ), mCLU provided substantial anti-amyloidogenic effects by inhibiting plaque formation [124]. These data suggest that CLU may exhibit neuroprotective characteristics in preclinical or early stages of AD when brain amyloid burden is significantly lower. Alternatively, CLU may exert a negative impact during later stages of AD when brain amyloid burden is extensive, though this hypothesis is yet to be tested. Parallel to this notion, a more recent study performed in rat brains indicated that mCLU prevented Aβ42-induced learning and memory impairments, reduced Aβ42-induced glia inflammation, and reduced Aβ42-mediated neuronal degeneration when Aβ42 oligomers were incubated with mCLU prior to brain injection. However, these effects were not observed in rats injected with pre-formed Aβ42 oligomers and mCLU without pre-incubation indicating that mCLU does prevent Aβ42-induced neurotoxicity prior to extensive Aβ42 oligomerization [126]. In addition to these studies, mCLU has been shown to impact the rate of Aβ42 clearance. A study by Bell and colleagues demonstrated that the rate of Aβ clearance was increased by as much as 83% when bound to CLU. This same study further demonstrated that CLU-bound A $\beta$  is transported across the blood-brain barrier specifically through LRP2-mediated transport, while APOE-bound Aβ was transported through LRP-1 [127]. While the regulation of A $\beta$  by mCLU is relatively well characterized, one question that remains unanswered is whether alternative CLU isoforms exert some impact on amyloid homeostasis. It has been demonstrated that AB toxicity induces the expression of intracellular CLU (icCLU) in neurons; however, the physiological impact of increased icCLU expression was not determined in this study [128]. At present, no literature specifically implicates a role for nCLU/icCLU isoforms in the regulation of Aβ; however, as nCLU/icCLU isoforms are reportedly induced by cellular stress in multiple peripheral cell lines and nCLU is induced upon treatment with exogenous A<sub>β</sub>, it is likely that nCLU/ icCLU isoforms mediate some effect on amyloid homeostasis in the brain; however, more research is needed before a conclusion can be made.

# 4.2. CLU and inflammation

It is well established that persistent inflammation likely caused by the deposition of neurotoxic protein aggregates in the brain is a key component of LOAD [129]. Early studies suggest that CLU inhibits the activation of the complement system in the brain [31, 130–132]. For instance, several early publications indicated that CLU (then referred to as SP-40,40) prevented the formation of the membrane attack complex (MAC), suggesting that increased CLU would suppress initiation of acute inflammation. However, these data were contradicted by a more recent study that demonstrated CLU-mediated activation of the major histocompatibility complex class II (MHC II) antigen in primary cultures of rat microglia. This same study showed that administration of exogenous CLU resulted in the direct activation of microglia in the brain and the subsequent secretion of pro-inflammatory cytokines such as tumour necrosis factor alpha (TNF- $\alpha$ ) indicating that increased CLU expression induces the acute inflammatory response [49]. These findings were corroborated by another study that demonstrated increased CLU staining within reactive microglia in the cortices of rats following cerebral ischemia [133]. Collectively, these studies suggest that increased CLU expression results in the activation of glial cells and the subsequent secretion of pro-inflammatory mediators. Therefore, it is possible that the increased secretion of cytokines such as TNF- $\alpha$  could contribute to chronic inflammation in AD brain; however, this hypothesis requires further testing.

#### 4.3. CLU and apoptosis

Several studies performed in human cancer cell lines have demonstrated that mCLU and nCLU exhibit opposing effects on cell death pathways. mCLU has been shown to protect cells from oxidative stress and inhibit intrinsic apoptosis by interacting with and stabilizing the KU-70-Bax protein complex [134–138]. In contrast, nCLU is suggested to initiate intrinsic apoptotic pathways resulting in rapid cell death [115, 136]. The contrasting functions of mCLU and nCLU appear to also exist in the brain; however, unlike cancer-focused studies, relatively few brainbased investigations have included an examination of the apoptotic characteristics of nCLU. An early study by Schreiber et al. demonstrated that CLU (then referred to as SGP-2) mRNA expression was rapidly and transiently increased in astrocytes, but not CA3 and CA1 neurons, following administration of kainic acid (KA), a neurotoxic seizure-inducing compound [139]. Another study performed in WT, human CLU overexpressing (hCLU-OE) mice and Cluknockout (Clu-/-) mice subjected to middle cerebral artery occlusion (MCAO) indicated that CLU overexpression resulted in reduced brain injury. Specifically, this study demonstrated a 30-50% increase in CLU mRNA expression 7 days post-ischemia in the ischemic brain hemisphere specifically in the penumbral area (the area that separates necrotic from normal brain tissue). Morphometric analysis of the ischemic hemisphere revealed that the penumbra was significantly thinner in hCLU-OE mice and significantly thicker in Clu-/- mice when compared with WT mice indicating an inverse relationship between CLU mRNA expression and brain injury [140]. Collectively, these two studies strongly support a neuroprotective role for CLU in the brain following significant brain injury. In contrast, ethanol-mediated toxicity has been shown to significantly increase CLU expression in the cortex and amygdala. This upregulated CLU, which was shown to interact with Bcl-XL, was translocated to the nucleus upon exposure to ethanol, and was associated with increased cell death suggesting that these effects were mediated by nCLU [135]. Another study performed in neonatal mice subjected to hypoxic-ischemic brain injury indicated that CLU accumulated in dying neurons following brain injury. Moreover, this study indicated that CLU-deficient mice exhibited 50% less brain injury when compared to wild-type controls indicating that CLU expression exacerbates neuronal cell death following brain injury [141]. Collectively, these studies indicate that nCLU protein expression may be associated with increased cell death following traumatic brain injury or in response to cytotoxic stimuli.

# 5. Future perspectives

CLU is currently the third most significant genetic risk factor for the development of LOAD; however, an extensive gap exists in the literature in understanding the neurophysiological and neuropathological functions of CLU. Moreover, the bulk of brain-based CLU research refers to CLU as a single protein with few studies including a characterization of its isoforms. As CLU isoforms appear to mediate different physiological processes, the tendency to focus on the effects of CLU as a singular protein could lead to conflicting reports in the literature that are currently unresolved. Therefore, before researchers can fully ascertain the therapeutic

potential of CLU from a clinical perspective, it is vital that these key deficiencies are addressed at the molecular level.

First, it is crucial that current and future studies strive to examine CLU isoforms individually, with particular emphasis on separating the nCLU and mCLU isoforms. Of the studies published pertaining to CLU in the brain, approximately five studies include an examination of nCLU. While it is possible that nCLU does function to regulate apoptosis, recent findings from our laboratory indicate roughly equivalent expression levels of both mCLU and nCLU in healthy primary cortical neurons suggesting that nCLU may be integrally involved in cellular homeostasis. Moreover, our recent data indicate that a nCLU or icCLU isoform is localized to the mitochondria suggesting that these alternative CLU isoforms may play an important role in the regulation of brain mitochondria function. While these studies are still underway, future work should focus on identifying the exact CLU isoforms expressed in other types of brain cells including astrocytes and microglia. Moreover, these studies should examine the cellular distribution, key protein modulators, and the neurophysiological function of each nCLU/icCLU isoform.

An emerging topic in the study of AD is the impact of sex on the development and progression of LOAD. As previously discussed, the female population is more susceptible to developing LOAD and the risk conferred by genetic factors, such as APOE, is greater in females. Moreover, our recent analyses have demonstrated that CLU expression is significantly reduced in the early aging of female but not male brain during a time that corresponds with the onset of reproductive senescence [28]. These data strongly suggest that CLU expression is modulated, in part, by sex hormone signaling pathways in the brain. Parallel to these findings, our recent studies have revealed that brain CLU isoform expression is regulated via estrogen receptor (ER) signaling. Additionally, we find that testosterone (TT) differentially regulates mCLU and nCLU expression; TT increases mCLU expression and decreases nCLU expression. An extension of these studies revealed that TT-mediated upregulation of mCLU expression results from the aromatization of TT to 17β-estradiol (E2). These data are particularly interesting when considered in the context of sex hormone changes between men and women throughout the aging process. It is well established that menopausal onset results in a significant and irreversible decline in ovarian sex hormones, such as E2. However, TT levels in males gradually decline with age at a rate of approximately 2% per year [142]. Therefore, it is possible that TTmediated upregulation of the neuroprotective mCLU isoform may, in part, contribute to the reduced incidence of AD in men. Likewise, the significant reduction in E2 levels in menopausal and/or postmenopausal women may result in significantly reduced mCLU levels thereby contributing to female vulnerability. While more research is needed to fully elucidate the interactions between sex hormones and neuronal CLU isoforms, these data underscore the importance of including sex as a variable in the study of risk factors that mediate the development of LOAD.

It is particularly interesting that two of the top five genetic risk factors associated with the development of LOAD are members of the apolipoprotein family: APOE and CLU. Therefore, another avenue of research to be considered in the AD field is the possibility of intersecting or overlapping risk pathways mediated by these two genetic factors. Studies have shown that

APOE and CLU share a number of important physiological properties. For instance, they are among the few proteins associated with brain lipoproteins [143, 144]. They interact with a shared set of cell-surface receptors [108] and both APOE and CLU promote neurite outgrowth [145, 146]. Moreover, elimination of either protein in an AD mouse model results in increased accumulation of A $\beta$  [147]. Furthermore, presence of the C allele of the CLU AD-risk SNP has been shown to exacerbate the APOE ɛ4-mediated decrease in brain activity during executive attention tasks in young healthy dementia-free adults [148]. In addition, the genetic variance that results in increased AD risk from both genes is also associated with compromised or reduced protein expression and/or binding capabilities. Our data indicate that APOE protein expression levels are significantly increased in 6-month-old female Clu-/- mice. However, mCLU expression levels are significantly reduced in 6-month-old female human APOE £4 gene targeted-replacement mice when compared to APOE  $\varepsilon$ 3 mice indicating that reduced CLU expression may contribute to APOE ɛ4-mediated AD risk. Collectively, these studies indicate that APOE and CLU could share common risk pathways that contribute to the development of LOAD. Delineation of such pathways will potentially provide valuable insights for an increased understanding of the etiology of LOAD and ultimately help to devise therapeutic strategies to prevent or reduce the risk of developing the disease.

# Author details

Sarah K. Woody<sup>1</sup> and Liqin Zhao<sup>1,2\*</sup>

\*Address all correspondence to: lzhao@ku.edu

1 Department of Pharmacology and Toxicology, School of Pharmacy, University of Kansas, Lawrence, KS, USA

2 Neuroscience Graduate Program, University of Kansas, Lawrence, KS, USA

# References

- [1] Alzheimer's A. 2015 Alzheimer's disease facts and figures. Alzheimer's & Dementia: The Journal of the Alzheimer's Association. 2015;11(3):332–84.
- [2] Why Do All the Large Alzheimer's Drug Trials Fail? [press release]. http:// www.dana.org/News/Why\_Do\_All\_the\_Large\_Alzheimer\_s\_Drug\_Trials\_Fail\_/: The Dana Foundation, July 8, 2013.
- [3] Pharma counts just 3 Alzheimer's drug wins in 13 years (101 losses!) [press release]. http://www.fiercebiotech.com/story/pharma-counts-just-3-alzheimers-drug-wins-13years-101-losses/2012-09-14: FierceBiotech, September 14, 2012.

- [4] National Institute on Aging. Alzheimer's Disease. 2015; 1–8 https://www.nia.nih.gov/ alzheimers/publication/alzheimers-disease-fact-sheet.
- [5] Gao S, Hendrie HC, Hall KS, Hui S. The relationships between age, sex, and the incidence of dementia and Alzheimer disease: a meta-analysis. Archives of General Psychiatry. 1998;55(9):809–15.
- [6] Irvine K, Laws KR, Gale TM, Kondel TK. Greater cognitive deterioration in women than men with Alzheimer's disease: a meta analysis. Journal of Clinical and Experimental Neuropsychology. 2012;34(9):989–98.
- [7] Chapman RM, Mapstone M, Gardner MN, Sandoval TC, McCrary JW, Guillily MD, et al. Women have farther to fall: gender differences between normal elderly and Alzheimer's disease in verbal memory engender better detection of Alzheimer's disease in women. Journal of International Neuropsychology Society. 2011;17(4):654–62.
- [8] Schmidt R, Kienbacher E, Benke T, Dal-Bianco P, Delazer M, Ladurner G, et al. Sex differences in Alzheimer's disease. Neuropsychiatry. 2008;22(1):1–15.
- [9] Zhao L, Mao Z, Brinton RD. A select combination of clinically relevant phytoestrogens enhances estrogen receptor beta-binding selectivity and neuroprotective activities in vitro and in vivo. Endocrinology. 2009;150(2):770–83.
- [10] Zhao L, Brinton RD. Structure-based virtual screening for plant-based ERbeta-selective ligands as potential preventative therapy against age-related neurodegenerative diseases. Journal of Medicinal Chemistry. 2005;48(10):3463–6.
- [11] Zhao L, Wu TW, Brinton RD. Estrogen receptor subtypes alpha and beta contribute to neuroprotection and increased Bcl-2 expression in primary hippocampal neurons. Brain Research. 2004;1010(1–2):22–34.
- Saunders AM, Schmader K, Breitner JC, Benson MD, Brown WT, Goldfarb L, et al. Apolipoprotein E epsilon 4 allele distributions in late-onset Alzheimer's disease and in other amyloid-forming diseases. Lancet. 1993;342(8873):710–1.
- [13] Blacker D, Haines JL, Rodes L, Terwedow H, Go RC, Harrell LE, et al. ApoE-4 and age at onset of Alzheimer's disease: the NIMH genetics initiative. Neurology. 1997;48(1): 139–47.
- [14] Corder EH, Saunders AM, Risch NJ, Strittmatter WJ, Schmechel DE, Gaskell PC, Jr., et al. Protective effect of apolipoprotein E type 2 allele for late onset Alzheimer disease. Nature Genetics. 1994;7(2):180–4.
- [15] Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, et al. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. Science. 1993;261(5123):921–3.

- [16] Liu CC, Kanekiyo T, Xu H, Bu G. Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. Nature Reviews Neurology. 2013;9(2): 106–18.
- [17] Langbaum JB, Chen K, Lee W, Reschke C, Bandy D, Fleisher AS, et al. Categorical and correlational analyses of baseline fluorodeoxyglucose positron emission tomography images from the Alzheimer's disease neuroimaging initiative (ADNI). NeuroImage. 2009;45(4):1107–16.
- [18] Lynch JR, Tang W, Wang H, Vitek MP, Bennett ER, Sullivan PM, et al. APOE genotype and an ApoE-mimetic peptide modify the systemic and central nervous system inflammatory response. The Journal of Biological Chemistry. 2003;278(49): 48529–33.
- [19] Drzezga A, Grimmer T, Henriksen G, Muhlau M, Perneczky R, Miederer I, et al. Effect of APOE genotype on amyloid plaque load and gray matter volume in Alzheimer disease. Neurology. 2009;72(17):1487–94.
- [20] Drzezga A, Riemenschneider M, Strassner B, Grimmer T, Peller M, Knoll A, et al. Cerebral glucose metabolism in patients with AD and different APOE genotypes. Neurology. 2005;64(1):102–7.
- [21] Altmann A, Tian L, Henderson VW, Greicius MD, Alzheimer's Disease Neuroimaging Initiative I. Sex modifies the APOE-related risk of developing Alzheimer disease. Annals of Neurology. 2014;75(4):563–73.
- [22] Farrer LA, Cupples LA, Haines JL, Hyman B, Kukull WA, Mayeux R, et al. Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer disease meta analysis consortium. JAMA. 1997;278(16):1349–56.
- [23] Fleisher A, Grundman M, Jack CR, Jr., Petersen RC, Taylor C, Kim HT, et al. Sex, apolipoprotein E epsilon 4 status, and hippocampal volume in mild cognitive impairment. Archives of Neurology. 2005;62(6):953–7.
- [24] Payami H, Montee KR, Kaye JA, Bird TD, Yu CE, Wijsman EM, et al. Alzheimer's disease, apolipoprotein E4, and gender. JAMA. 1994;271(17):1316–7.
- [25] Payami H, Zareparsi S, Montee KR, Sexton GJ, Kaye JA, Bird TD, et al. Gender difference in apolipoprotein E-associated risk for familial Alzheimer disease: a possible clue to the higher incidence of Alzheimer disease in women. American Journal of Human Genetics. 1996;58(4):803–11.
- [26] Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, et al. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. Nature Genetics. 2009;41(10):1088–93.

- [27] Lambert JC, Heath S, Even G, Campion D, Sleegers K, Hiltunen M, et al. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. Nature Genetics. 2009;41(10):1094–9.
- [28] Zhao L, Mao Z, Woody SK, Brinton RD. Sex differences in metabolic aging of the brain: insights into female suceptibility to Alzheimer's disease. Neurobiology of Aging. 2016;42:69–79.
- [29] Fritz IB, Burdzy K, Setchell B, Blaschuk O. Ram rete testis fluid contains a protein (clusterin) which influences cell-cell interactions in vitro. Biology of Reproduction. 1983;28(5):1173–88.
- [30] Sylvester SR, Skinner MK, Griswold MD. A sulfated glycoprotein synthesized by Sertoli cells and by epididymal cells is a component of the sperm membrane. Biology of Reproduction. 1984;31(5):1087–101.
- [31] Murphy BF, Kirszbaum L, Walker ID, d'Apice AJ. SP-40,40, a newly identified normal human serum protein found in the SC5b-9 complex of complement and in the immune deposits in glomerulonephritis. The Journal of Clinical Investigation. 1988;81(6):1858– 64.
- [32] de Silva HV, Stuart WD, Park YB, Mao SJ, Gil CM, Wetterau JR, et al. Purification and characterization of apolipoprotein J. The Journal of Biological Chemistry. 1990;265(24): 14292–7.
- [33] Wong P, Pineault J, Lakins J, Taillefer D, Leger J, Wang C, et al. Genomic organization and expression of the rat TRPM-2 (clusterin) gene, a gene implicated in apoptosis. The Journal of Biological Chemistry. 1993;268(7):5021–31.
- [34] Yang CR, Yeh S, Leskov K, Odegaard E, Hsu HL, Chang C, et al. Isolation of Ku70binding proteins (KUBs). Nucleic Acids Research. 1999;27(10):2165–74.
- [35] Jenne DE, Tschopp J. Molecular structure and functional characterization of a human complement cytolysis inhibitor found in blood and seminal plasma: identity to sulfated glycoprotein 2, a constituent of rat testis fluid. Proceedings of the National Academy of Sciences of the United States of America. 1989;86(18):7123–7.
- [36] Purrello M, Bettuzzi S, Di Pietro C, Mirabile E, Di Blasi M, Rimini R, et al. The gene for SP-40,40, human homolog of rat sulfated glycoprotein 2, rat clusterin, and rat testosterone-repressed prostate message 2, maps to chromosome 8. Genomics. 1991;10(1): 151–6.
- [37] Dietzsch E, Murphy BF, Kirszbaum L, Walker ID, Garson OM. Regional localization of the gene for clusterin (SP-40,40; gene symbol CLI) to human chromosome 8p12→p21. Cytogenetics and Cell Genetics. 1992;61(3):178–9.
- [38] Tobe T, Minoshima S, Yamase S, Choi NH, Tomita M, Shimizu N. Assignment of a human serum glycoprotein SP-40,40 gene (CLI) to chromosome 8. Cytogenetics and Cell Genetics. 1991;57(4):193–5.

- [39] Burkey BF, deSilva HV, Harmony JA. Intracellular processing of apolipoprotein J precursor to the mature heterodimer. The Journal of Lipid Research. 1991;32(6):1039–48.
- [40] Stewart EM, Aquilina JA, Easterbrook-Smith SB, Murphy-Durland D, Jacobsen C, Moestrup S, et al. Effects of glycosylation on the structure and function of the extracellular chaperone clusterin. Biochemistry. 2007;46(5):1412–22.
- [41] Leskov KS, Klokov DY, Li J, Kinsella TJ, Boothman DA. Synthesis and functional analyses of nuclear clusterin, a cell death protein. The Journal of Biological Chemistry. 2003;278(13):11590–600.
- [42] Prochnow H, Gollan R, Rohne P, Hassemer M, Koch-Brandt C, Baiersdorfer M. Nonsecreted clusterin isoforms are translated in rare amounts from distinct human mRNA variants and do not affect Bax-mediated apoptosis or the NF-kappaB signaling pathway. PloS One. 2013;8(9):e75303.
- [43] Reddy KB, Jin G, Karode MC, Harmony JA, Howe PH. Transforming growth factor beta (TGF beta)-induced nuclear localization of apolipoprotein J/clusterin in epithelial cells. Biochemistry. 1996;35(19):6157–63.
- [44] de Silva HV, Harmony JA, Stuart WD, Gil CM, Robbins J. Apolipoprotein J: structure and tissue distribution. Biochemistry. 1990;29(22):5380–9.
- [45] Nizard P, Tetley S, Le Drean Y, Watrin T, Le Goff P, Wilson MR, et al. Stress-induced retrotranslocation of clusterin/APOJ into the cytosol. Traffic. 2007;8(5):554–65.
- [46] Pasinetti GM, Johnson SA, Oda T, Rozovsky I, Finch CE. Clusterin (SGP-2): a multifunctional glycoprotein with regional expression in astrocytes and neurons of the adult rat brain. The Journal of Comparative Neurology. 1994;339(3):387–400.
- [47] Cordero-Llana O, Scott SA, Maslen SL, Anderson JM, Boyle J, Chowhdury RR, et al. Clusterin secreted by astrocytes enhances neuronal differentiation from human neural precursor cells. Cell Death and Differentiation. 2011;18(5):907–13.
- [48] Zwain IH, Grima J, Cheng CY. Regulation of clusterin secretion and mRNA expression in astrocytes by cytokines. Molecular and Cellular Neurosciences. 1994;5(3):229–37.
- [49] Xie Z, Harris-White ME, Wals PA, Frautschy SA, Finch CE, Morgan TE. Apolipoprotein J (clusterin) activates rodent microglia in vivo and in vitro. Journal of Neurochemistry. 2005;93(4):1038–46.
- [50] Kumita JR, Poon S, Caddy GL, Hagan CL, Dumoulin M, Yerbury JJ, et al. The extracellular chaperone clusterin potently inhibits human lysozyme amyloid formation by interacting with prefibrillar species. Journal of Molecular Biology. 2007;369(1):157–67.
- [51] DeMattos RB, Brendza RP, Heuser JE, Kierson M, Cirrito JR, Fryer J, et al. Purification and characterization of astrocyte-secreted apolipoprotein E and J-containing lipopro-

teins from wild-type and human apoE transgenic mice. Neurochemistry International. 2001;39(5–6):415–25.

- [52] Loison F, Debure L, Nizard P, le Goff P, Michel D, le Drean Y. Up-regulation of the clusterin gene after proteotoxic stress: implication of HSF1-HSF2 heterocomplexes. The Biochemical Journal. 2006;395(1):223–31.
- [53] Michel D, Chatelain G, North S, Brun G. Stress-induced transcription of the clusterin/ APOJ gene. The Biochemical Journal. 1997;328 (Pt 1):45–50.
- [54] Trougakos IP, Gonos ES. Regulation of clusterin/apolipoprotein J, a functional homologue to the small heat shock proteins, by oxidative stress in ageing and age-related diseases. Free Radical Research. 2006;40(12):1324–34.
- [55] Cervellera M, Raschella G, Santilli G, Tanno B, Ventura A, Mancini C, et al. Direct transactivation of the anti-apoptotic gene apolipoprotein J (clusterin) by B-MYB. The Journal of Biological Chemistry. 2000;275(28):21055–60.
- [56] Gutacker C, Klock G, Diel P, Koch-Brandt C. Nerve growth factor and epidermal growth factor stimulate clusterin gene expression in PC12 cells. The Biochemical Journal. 1999;339 (Pt 3):759–66.
- [57] Jin G, Howe PH. Regulation of clusterin gene expression by transforming growth factor beta. The Journal of Biological Chemistry. 1997;272(42): 26620–6.
- [58] Jin G, Howe PH. Transforming growth factor beta regulates clusterin gene expression via modulation of transcription factor c-Fos. European Journal of biochemistry/FEBS. 1999;263(2):534–42.
- [59] Kapron JT, Hilliard GM, Lakins JN, Tenniswood MP, West KA, Carr SA, et al. Identification and characterization of glycosylation sites in human serum clusterin. Protein Science: A Publication of the Protein Society. 1997;6(10):2120–33.
- [60] Dunker AK, Lawson JD, Brown CJ, Williams RM, Romero P, Oh JS, et al. Intrinsically disordered protein. Journal of Molecular Graphics & Modelling. 2001;19(1):26–59.
- [61] Rohne P, Prochnow H, Wolf S, Renner B, Koch-Brandt C. The chaperone activity of clusterin is dependent on glycosylation and redox environment. Cellular Physiology and Biochemistry : International Journal of Experimental Cellular Physiology, Biochemistry, and Pharmacology. 2014;34(5):1626–39.
- [62] Kang SW, Yoon SY, Park JY, Kim DH. Unglycosylated clusterin variant accumulates in the endoplasmic reticulum and induces cytotoxicity. The International Journal of Biochemistry & Cell Biology. 2013;45(2):221–31.

- [63] Xue J, Lv DD, Jiao S, Zhao W, Li X, Sun H, et al. pVHL mediates K63-linked ubiquitination of nCLU. PloS One. 2012;7(4):e35848.
- [64] Lemansky P, Brix K, Herzog V. Subcellular distribution, secretion, and posttranslational modifications of clusterin in thyrocytes. Experimental Cell Research. 1999;251(1): 147–55.
- [65] Ghosh P, Hale EA, Lakshman MR. Plasma sialic-acid index of apolipoprotein J (SIJ): a new alcohol intake marker. Alcohol. 2001;25(3):173–9.
- [66] Nuutinen T, Suuronen T, Kyrylenko S, Huuskonen J, Salminen A. Induction of clusterin/APOJ expression by histone deacetylase inhibitors in neural cells. Neurochemistry International. 2005;47(8):528–38.
- [67] Masoodi TA, Al Shammari SA, Al-Muammar MN, Alhamdan AA, Talluri VR. Exploration of deleterious single nucleotide polymorphisms in late-onset Alzheimer disease susceptibility genes. Gene. 2013;512(2):429–37.
- [68] Braskie MN, Jahanshad N, Stein JL, Barysheva M, McMahon KL, de Zubicaray GI, et al. Common Alzheimer's disease risk variant within the CLU gene affects white matter microstructure in young adults. The Journal of Neuroscience: The Official Journal of the Society for Neuroscience. 2011;31(18):6764–70.
- [69] Chen LH, Kao PY, Fan YH, Ho DT, Chan CS, Yik PY, et al. Polymorphisms of CR1, CLU and PICALM confer susceptibility of Alzheimer's disease in a southern Chinese population. Neurobiology of Aging. 2012;33(1):210 e1–7.
- [70] Corneveaux JJ, Myers AJ, Allen AN, Pruzin JJ, Ramirez M, Engel A, et al. Association of CR1, CLU and PICALM with Alzheimer's disease in a cohort of clinically characterized and neuropathologically verified individuals. Human Molecular Genetics. 2010;19(16):3295–301.
- [71] Ferrari R, Moreno JH, Minhajuddin AT, O'Bryant SE, Reisch JS, Barber RC, et al. Implication of common and disease specific variants in CLU, CR1, and PICALM. Neurobiology of Aging. 2012;33(8):1846 e7–18.
- [72] Jun G, Naj AC, Beecham GW, Wang LS, Buros J, Gallins PJ, et al. Meta-analysis confirms CR1, CLU, and PICALM as alzheimer disease risk loci and reveals interactions with APOE genotypes. Archives of Neurology. 2010;67(12):1473–84.
- [73] Lancaster TM, Baird A, Wolf C, Jackson MC, Johnston SJ, Donev R, et al. Neural hyperactivation in carriers of the Alzheimer's risk variant on the clusterin gene. European Neuropsychopharmacology: The Journal of the European College of Neuropsychopharmacology. 2011;21(12):880–4.
- [74] Ma JF, Liu LH, Zhang Y, Wang Y, Deng YL, Huang Y, et al. Association study of clusterin polymorphism rs11136000 with late onset Alzheimer's disease in Chinese Han

population. American Journal of Alzheimer's Disease and Other Dementias. 2011;26(8): 627–30.

- [75] Schurmann B, Wiese B, Bickel H, Weyerer S, Riedel-Heller SG, Pentzek M, et al. Association of the Alzheimer's disease clusterin risk allele with plasma clusterin concentration. Journal of Alzheimer's Disease : JAD. 2011;25(3):421–4.
- [76] Seshadri S, Fitzpatrick AL, Ikram MA, DeStefano AL, Gudnason V, Boada M, et al. Genome-wide analysis of genetic loci associated with Alzheimer disease. JAMA. 2010;303(18):1832–40.
- [77] Thambisetty M, Beason-Held LL, An Y, Kraut M, Nalls M, Hernandez DG, et al. Alzheimer risk variant CLU and brain function during aging. Biological Psychiatry. 2013;73(5):399–405.
- [78] Golenkina SA, Gol'tsov A, Kuznetsova IL, Grigorenko AP, Andreeva TV, Reshetov DA, et al. Analysis of clusterin gene (CLU/APOJ) polymorphism in Alzheimer's disease patients and in normal cohorts from Russian populations. Molekuliarnaia Biologiia. 2010;44(4):620–6.
- [79] Kamboh MI, Minster RL, Demirci FY, Ganguli M, Dekosky ST, Lopez OL, et al. Association of CLU and PICALM variants with Alzheimer's disease. Neurobiology of Aging. 2012;33(3):518–21.
- [80] Klimkowicz-Mrowiec A, Sado M, Dziubek A, Dziedzic T, Pera J, Szczudlik A, et al. Lack of association of CR1, PICALM and CLU gene polymorphisms with Alzheimer disease in a Polish population. Neurologia i Neurochirurgia Polska. 2013;47(2):157–60.
- [81] Komatsu M, Shibata N, Kuerban B, Ohnuma T, Baba H, Arai H. Genetic association between clusterin polymorphisms and Alzheimer's disease in a Japanese population. Psychogeriatrics : The Official Journal of the Japanese Psychogeriatric Society. 2011;11(1):14–8.
- [82] Lee JH, Cheng R, Barral S, Reitz C, Medrano M, Lantigua R, et al. Identification of novel loci for Alzheimer disease and replication of CLU, PICALM, and BIN1 in Caribbean Hispanic individuals. Archives of Neurology. 2011;68(3):320–8.
- [83] Lu SJ, Li HL, Sun YM, Liu ZJ, Yang P, Wu ZY. Clusterin variants are not associated with southern Chinese patients with Alzheimer's disease. Neurobiology of Aging. 2014;35(11):2656 e9–11.
- [84] Yu JT, Li L, Zhu QX, Zhang Q, Zhang W, Wu ZC, et al. Implication of CLU gene polymorphisms in Chinese patients with Alzheimer's disease. Clinica Chimica Acta: International Journal of Clinical Chemistry. 2010;411(19–20):1516– 9.

- [85] Yu JT, Ma XY, Wang YL, Sun L, Tan L, Hu N, et al. Genetic variation in clusterin gene and Alzheimer's disease risk in Han Chinese. Neurobiology of Aging. 2013;34(7):1921 e17–23.
- [86] Lin YL, Chen SY, Lai LC, Chen JH, Yang SY, Huang YL, et al. Genetic polymorphisms of clusterin gene are associated with a decreased risk of Alzheimer's disease. European Journal of Epidemiology. 2012;27(1):73–75.
- [87] Pedraza O, Allen M, Jennette K, Carrasquillo M, Crook J, Serie D, et al. Evaluation of memory endophenotypes for association with CLU, CR1, and PICALM variants in black and white subjects. Alzheimer's & Dementia: The Journal of the Alzheimer's Association. 2014;10(2):205–13.
- [88] Lancaster TM, Brindley LM, Tansey KE, Sims RC, Mantripragada K, Owen MJ, et al. Alzheimer's disease risk variant in CLU is associated with neural inefficiency in healthy individuals. Alzheimer's & Dementia: The Journal of the Alzheimer's Association. 2015;11(10):1144–52.
- [89] Erk S, Meyer-Lindenberg A, Opitz von Boberfeld C, Esslinger C, Schnell K, Kirsch P, et al. Hippocampal function in healthy carriers of the CLU Alzheimer's disease risk variant. The Journal of Neuroscience: The Official Journal of the Society for Neuroscience. 2011;31(49):18180–4.
- [90] Thambisetty M, An Y, Kinsey A, Koka D, Saleem M, Guntert A, et al. Plasma clusterin concentration is associated with longitudinal brain atrophy in mild cognitive impairment. NeuroImage. 2012;59(1):212–7.
- [91] Roussotte FF, Gutman BA, Madsen SK, Colby JB, Thompson PM, Alzheimer's Disease Neuroimaging I. Combined effects of Alzheimer risk variants in the CLU and ApoE genes on ventricular expansion patterns in the elderly. The Journal of Neuroscience: The Official Journal of the Society for Neuroscience. 2014;34(19):6537–45.
- [92] Shuai P, Liu Y, Lu W, Liu Q, Li T, Gong B. Genetic associations of CLU rs9331888 polymorphism with Alzheimer's disease: a meta-analysis. Neuroscience Letters. 2015;591:160–5.
- [93] Toral-Rios D, Franco-Bocanegra D, Rosas-Carrasco O, Mena-Barranco F, Carvajal-Garcia R, Meraz-Rios MA, et al. Evaluation of inflammation-related genes polymorphisms in Mexican with Alzheimer's disease: a pilot study. Frontiers in Cellular Neuroscience. 2015;9:148.
- [94] Xing YY, Yu JT, Cui WZ, Zhong XL, Wu ZC, Zhang Q, et al. Blood clusterin levels, rs9331888 polymorphism, and the risk of Alzheimer's disease. Journal of Alzheimer's Disease: JAD. 2012;29(3):515–9.

- [95] Zhang S, Li X, Ma G, Jiang Y, Liao M, Feng R, et al. CLU rs9331888 Polymorphism contributes to Alzheimer's disease susceptibility in Caucasian but not East Asian populations. Molecular Neurobiology. 2015.
- [96] Michel D, Chabot JG, Moyse E, Danik M, Quirion R. Possible functions of a new genetic marker in central nervous system: the sulfated glycoprotein-2 (SGP-2). Synapse. 1992;11(2):105–11.
- [97] Sattlecker M, Kiddle SJ, Newhouse S, Proitsi P, Nelson S, Williams S, et al. Alzheimer's disease biomarker discovery using SOMAscan multiplexed protein technology. Alzheimer's & Dementia: The Journal of the Alzheimer's Association. 2014;10(6):724– 34.
- [98] Song F, Poljak A, Crawford J, Kochan NA, Wen W, Cameron B, et al. Plasma apolipoprotein levels are associated with cognitive status and decline in a community cohort of older individuals. PloS One. 2012;7(6):e34078.
- [99] Jongbloed W, van Dijk KD, Mulder SD, van de Berg WD, Blankenstein MA, van der Flier W, et al. Clusterin levels in plasma predict cognitive decline and progression to Alzheimer's disease. Journal of Alzheimer's Disease: JAD. 2015;46(4): 1103–10.
- [100] Schrijvers EM, Koudstaal PJ, Hofman A, Breteler MM. Plasma clusterin and the risk of Alzheimer disease. JAMA. 2011;305(13):1322–6.
- [101] Dukic L, Simundic AM, Martinic-Popovic I, Kackov S, Diamandis A, Begcevic I, et al. The role of human kallikrein 6, clusterin and adiponectin as potential blood biomarkers of dementia. Clinical Biochemistry. 2016;49(3):213–8.
- [102] IJ L, Dekker LJ, Koudstaal PJ, Hofman A, Sillevis Smitt PA, Breteler MM, et al. Serum clusterin levels are not increased in presymptomatic Alzheimer's disease. Journal of Proteome Research. 2011;10(4):2006–10.
- [103] Mukaetova-Ladinska EB, Abdel-All Z, Andrade J, Alves da Silva J, O'Brien JT, Kalaria RN. Plasma and platelet clusterin ratio is altered in Alzheimer's disease patients with distinct neuropsychiatric symptoms: findings from a pilot study. International Journal of Geriatric Psychiatry. 2015;30(4):368–75.
- [104] Mukaetova-Ladinska EB, Abdel-All Z, Dodds S, Andrade J, Alves da Silva J, Kalaria RN, et al. Platelet immunoglobulin and amyloid precursor protein as potential peripheral biomarkers for Alzheimer's disease: findings from a pilot study. Age and Ageing. 2012;41(3):408–12.
- [105] Silajdzic E, Minthon L, Bjorkqvist M, Hansson O. No diagnostic value of plasma clusterin in Alzheimer's disease. PloS One. 2012;7(11):e50237.

- [106] Bailey RW, Dunker AK, Brown CJ, Garner EC, Griswold MD. Clusterin, a binding protein with a molten globule-like region. Biochemistry. 2001;40(39): 11828–40.
- [107] Bartl MM, Luckenbach T, Bergner O, Ullrich O, Koch-Brandt C. Multiple receptors mediate APOJ-dependent clearance of cellular debris into nonprofessional phagocytes.
   Experimental Cell Research. 2001;271(1):130–41.
- [108] Kounnas MZ, Loukinova EB, Stefansson S, Harmony JA, Brewer BH, Strickland DK, et al. Identification of glycoprotein 330 as an endocytic receptor for apolipoprotein J/clusterin. The Journal of Biological Chemistry. 1995;270(22):13070– 5.
- [109] Lakins JN, Poon S, Easterbrook-Smith SB, Carver JA, Tenniswood MP, Wilson MR. Evidence that clusterin has discrete chaperone and ligand binding sites. Biochemistry. 2002;41(1):282–91.
- [110] Law GL, Griswold MD. Activity and form of sulfated glycoprotein 2 (clusterin) from cultured Sertoli cells, testis, and epididymis of the rat. Biology of Reproduction. 1994;50(3):669–79.
- [111] Leeb C, Eresheim C, Nimpf J. Clusterin is a ligand for apolipoprotein E receptor 2 (ApoER2) and very low density lipoprotein receptor (VLDLR) and signals via the Reelin-signaling pathway. The Journal of Biological Chemistry. 2014;289(7): 4161–72.
- [112] Poon S, Rybchyn MS, Easterbrook-Smith SB, Carver JA, Pankhurst GJ, Wilson MR. Mildly acidic pH activates the extracellular molecular chaperone clusterin. The Journal of Biological Chemistry. 2002;277(42):39532–40.
- [113] Wyatt A, Yerbury J, Poon S, Dabbs R, Wilson M. Chapter 6: The chaperone action of Clusterin and its putative role in quality control of extracellular protein folding. Advances in Cancer Reserch. 2009;104:89–114.
- [114] Debure L, Vayssiere JL, Rincheval V, Loison F, Le Drean Y, Michel D. Intracellular clusterin causes juxtanuclear aggregate formation and mitochondrial alteration. Journal of Cell Science. 2003;116(Pt 15):3109–21.
- [115] Dia VP, Mejia EG. Lunasin promotes apoptosis in human colon cancer cells by mitochondrial pathway activation and induction of nuclear clusterin expression. Cancer Letters. 2010;295(1):44–53.
- [116] Kimura K, Asami K, Yamamoto M. Effect of heat shock treatment on the production of variant testosterone-repressed prostate message-2 (TRPM-2) mRNA in culture cells. Cell Biochemistry and Function. 1997;15(4):251–7.
- [117] Lidstrom AM, Bogdanovic N, Hesse C, Volkman I, Davidsson P, Blennow K. Clusterin (apolipoprotein J) protein levels are increased in hippocampus and in frontal cortex in Alzheimer's disease. Experimental Neurology. 1998;154(2):511–21.

- [118] May PC, Lampert-Etchells M, Johnson SA, Poirier J, Masters JN, Finch CE. Dynamics of gene expression for a hippocampal glycoprotein elevated in Alzheimer's disease and in response to experimental lesions in rat. Neuron. 1990;5(6):831–9.
- [119] McGeer PL, Klegeris A, Walker DG, Yasuhara O, McGeer EG. Pathological proteins in senile plaques. The Tohoku Journal of Experimental Medicine. 1994;174(3):269–77.
- [120] Ghiso J, Matsubara E, Koudinov A, Choi-Miura NH, Tomita M, Wisniewski T, et al. The cerebrospinal-fluid soluble form of Alzheimer's amyloid beta is complexed to SP-40,40 (apolipoprotein J), an inhibitor of the complement membrane-attack complex. The Biochemical Journal. 1993;293(Pt 1):27–30.
- [121] Matsubara E, Soto C, Governale S, Frangione B, Ghiso J. Apolipoprotein J and Alzheimer's amyloid beta solubility. The Biochemical Journal. 1996;316(Pt 2):671–9.
- [122] Oda T, Wals P, Osterburg HH, Johnson SA, Pasinetti GM, Morgan TE, et al. Clusterin (APOJ) alters the aggregation of amyloid beta-peptide (A beta 1-42) and forms slowly sedimenting A beta complexes that cause oxidative stress. Experimental Neurology. 1995;136(1):22–31.
- [123] DeMattos RB, O'Dell M A, Parsadanian M, Taylor JW, Harmony JA, Bales KR, et al. Clusterin promotes amyloid plaque formation and is critical for neuritic toxicity in a mouse model of Alzheimer's disease. Proceedings of the National Academy of Sciences of the United States of America. 2002;99(16):10843–8.
- [124] Yerbury JJ, Poon S, Meehan S, Thompson B, Kumita JR, Dobson CM, et al. The extracellular chaperone clusterin influences amyloid formation and toxicity by interacting with prefibrillar structures. FASEB J. 2007;21(10):2312–22.
- [125] Poon S, Easterbrook-Smith SB, Rybchyn MS, Carver JA, Wilson MR. Clusterin is an ATP-independent chaperone with very broad substrate specificity that stabilizes stressed proteins in a folding-competent state. Biochemistry. 2000;39(51):15953–60.
- [126] Cascella R, Conti S, Tatini F, Evangelisti E, Scartabelli T, Casamenti F, et al. Extracellular chaperones prevent Abeta42-induced toxicity in rat brains. Biochimica et Biophysica Acta. 2013;1832(8):1217–26.
- [127] Bell RD, Sagare AP, Friedman AE, Bedi GS, Holtzman DM, Deane R, et al. Transport pathways for clearance of human Alzheimer's amyloid beta-peptide and apolipoproteins E and J in the mouse central nervous system. Journal of Cerebral Blood Flow and Metabolism: Official Journal of the International Society of Cerebral Blood Flow and Metabolism. 2007;27(5):909–18.
- [128] Killick R, Ribe EM, Al-Shawi R, Malik B, Hooper C, Fernandes C, et al. Clusterin regulates beta-amyloid toxicity via Dickkopf-1-driven induction of the wnt-PCP-JNK pathway. Molecular Psychiatry. 2014;19(1):88–98.
- [129] Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, et al. Inflammation and Alzheimer's disease. Neurobiology of Aging. 2000;21(3):383–421.

- [130] Choi NH, Mazda T, Tomita M. A serum protein SP40,40 modulates the formation of membrane attack complex of complement on erythrocytes. Molecular Immunology. 1989;26(9):835–40.
- [131] Choi NH, Nakano Y, Tobe T, Mazda T, Tomita M. Incorporation of SP-40,40 into the soluble membrane attack complex (SMAC, SC5b-9) of complement. International Immunology. 1990;2(5):413–7.
- [132] Kirszbaum L, Bozas SE, Walker ID. SP-40,40, a protein involved in the control of the complement pathway, possesses a unique array of disulphide bridges. FEBS Letters. 1992;297(1–2):70–6.
- [133] Van Beek J, Chan P, Bernaudin M, Petit E, MacKenzie ET, Fontaine M. Glial responses, clusterin, and complement in permanent focal cerebral ischemia in the mouse. Glia. 2000;31(1):39–50.
- [134] Jun HO, Kim DH, Lee SW, Lee HS, Seo JH, Kim JH, et al. Clusterin protects H9c2 cardiomyocytes from oxidative stress-induced apoptosis via Akt/ GSK-3beta signaling pathway. Experimental and Molecular Medicine. 2011;43(1): 53–61.
- [135] Kim N, Han JY, Roh GS, Kim HJ, Kang SS, Cho GJ, et al. Nuclear clusterin is associated with neuronal apoptosis in the developing rat brain upon ethanol exposure. Alcoholism, Clinical and Experimental Research. 2012;36(1):72–82.
- [136] Miyake H, Chi KN, Gleave ME. Antisense TRPM-2 oligodeoxynucleotides chemosensitize human androgen-independent PC-3 prostate cancer cells both in vitro and in vivo. Clinical Cancer Research: An Official Journal of the American Association for Cancer Research. 2000;6(5):1655–63.
- [137] Trougakos IP, Lourda M, Antonelou MH, Kletsas D, Gorgoulis VG, Papassideri IS, et al. Intracellular clusterin inhibits mitochondrial apoptosis by suppressing p53-activating stress signals and stabilizing the cytosolic Ku70-Bax protein complex. Clinical Cancer Research: An Official Journal of the American Association for Cancer Research. 2009;15(1):48–59.
- [138] Zhang H, Kim JK, Edwards CA, Xu Z, Taichman R, Wang CY. Clusterin inhibits apoptosis by interacting with activated Bax. Nature Cell Biology. 2005;7(9):909–15.
- [139] Schreiber SS, Tocco G, Najm I, Baudry M. Seizure activity causes a rapid increase in sulfated glycoprotein-2 messenger RNA in the adult but not the neonatal rat brain. Neuroscience Letters. 1993;153(1):17–20.
- [140] Wehrli P, Charnay Y, Vallet P, Zhu G, Harmony J, Aronow B, et al. Inhibition of post-ischemic brain injury by clusterin overexpression. Nature Medicine. 2001;7(9):977–9.

- [141] Han BH, DeMattos RB, Dugan LL, Kim-Han JS, Brendza RP, Fryer JD, et al. Clusterin contributes to caspase-3-independent brain injury following neonatal hypoxiaischemia. Nature Medicine. 2001;7(3):338–43.
- [142] Brawer MK. Testosterone replacement in men with andropause: an overview. Reviews in urology. 2004;6(Suppl 6):S9–15.
- [143] Elliott DA, Weickert CS, Garner B. Apolipoproteins in the brain: implications for neurological and psychiatric disorders. Clinical Lipidology. 2010;51(4):555–73.
- [144] Koch S, Donarski N, Goetze K, Kreckel M, Stuerenburg HJ, Buhmann C, et al. Characterization of four lipoprotein classes in human cerebrospinal fluid. Journal Lipid Research. 2001;42(7):1143–51.
- [145] Kang SW, Shin YJ, Shim YJ, Jeong SY, Park IS, Min BH. Clusterin interacts with SCLIP (SCG10-like protein) and promotes neurite outgrowth of PC12 cells. Experimental Cell Research. 2005;309(2):305–15.
- [146] Nathan BP, Bellosta S, Sanan DA, Weisgraber KH, Mahley RW, Pitas RE. Differential effects of apolipoproteins E3 and E4 on neuronal growth in vitro. Science. 1994;264(5160):850–2.
- [147] DeMattos RB, Cirrito JR, Parsadanian M, May PC, O'Dell MA, Taylor JW, et al. ApoE and clusterin cooperatively suppress Abeta levels and deposition: evidence that ApoE regulates extracellular Abeta metabolism in vivo. Neuron. 2004;41(2):193–202.
- [148] Green AE, Gray JR, Deyoung CG, Mhyre TR, Padilla R, Dibattista AM, et al. A combined effect of two Alzheimer's risk genes on medial temporal activity during executive attention in young adults. Neuropsychologia. 2014;56:1–8.





IntechOpen