

On acute gene expression changes after ventral root replantation

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Replantation of avulsed spinal ventral roots has been show to enable significant and useful regrowth of motor axons in both experimental animals and in human clinical cases, making up an interesting exception to the rule of unsuccessful neuronal regeneration in central nervous system. Compared to avulsion without repair, ventral root replantation seems to rescue lesioned motoneurons from death. In this study we have analyzed the acute response to ventral root avulsion and replantation in adult rats with gene arrays combined with cluster analysis of gene ontology search terms. The data show significant differences between rats subjected to ventral replantation compared to avulsion only. Even though number of genes related to cell death is similar in the two models after 24 h, we observed a significantly larger number of genes related to neurite growth and development in the rats treated with ventral root replantation, possibly reflecting the neuroregenerative capacity in the replantation model. In addition, an acute inflammatory response was observed after avulsion, while effects on genes related to synaptic transmission were much more pronounced after replantation than after avulsion alone. These data indicate that the axonal regenerative response from replantation is initiated at an earlier stage than the possible differences in terms of neuron survival. We conclude that this type of analysis may facilitate the comparison of the acute response in two types of injury.

Keywords: ventral root, regeneration, inflammation, spinal cord, gene expression

INTRODUCTION

The pioneering studies of Ramon y Cajal demonstrated that injured neurons in the central nervous system (CNS) have a strong capacity to extend new axons into the peripheral nervous system (PNS; Cajal, 1928). This observation was reinvestigated by Aguayo and colleagues (Richardson et al., 1980; Aguayo et al., 1987). Cajal (1928) described that motor axons that had been severed due to spinal cord injury had the capacity to regrow to neighboring ventral roots. This observation was later confirmed with electron microscopy and intracellular labeling with horseradish peroxidase (Risling et al., 1983; Lindå et al., 1992). The regrowing axons penetrate through a highly unusual CNS environment which lacks a blood-brain barrier (BBB) function (Risling et al., 1989) but possesses a high content of cells bearing neurotrophin receptors (Frisen et al., 1992, 1998; Risling et al., 1992) and matrix molecules (Risling et al., 1993; Deckner et al., 2000). A clinical counterpart to this injury is a ventral root avulsion at the border between the CNS and PNS, typically caused by a high energy trauma such as a motorbike accident causing excessive trauma to the shoulder and head resulting in stretching and rupturing of ventral roots. Ventral root avulsion is not followed by spontaneous regrowth, since the avulsed roots are widely separated from the spinal cord inside the subarachnoid space or even pulled to a position outside the vertebral channel. Replantation of avulsed spinal ventral roots into the spinal cord has been shown to be enable significant and useful regrowth

of motor axons in both experimental animals and in human clinical cases (Carlstedt et al., 1986, 1995, 2009; Cullheim et al., 1989, 1999).

The results from such treatment in humans are less successful in older patients and good reinnervation in distal muscles like in the hand is seldom possible. In a recent case report Carlstedt et al. (2009) described a preadolescent boy with complete brachial plexus avulsion injury that was treated by replantation of five ventral roots. Shoulder muscle recovery started 8-10 months after the spinal cord operation. At 12-15 months, elbow function began to recover, followed 2 years postoperatively by forearm, wrist, and intrinsic hand muscle activity. This led to recovery of hand motor function without restoration of sensation. Bilateral motor cortex activity and activation of the sensory cortex on use of the affected hand was demonstrated by fMRI. The findings of that case study suggested that the restored hand function might rely on cortical sensory programs established before the injury (Carlstedt et al., 2009). Although the time needed for recovery seems very long, it cannot be excluded that the final result is dependent on several biological programs that are elicited in the early acute stage.

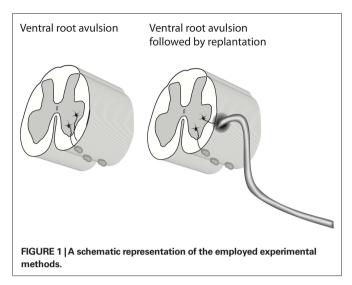
Previous studies indicate that ventral root replantation can be neuroprotective for motoneurons (Hoang and Havton, 2006; Eggers et al., 2010). If so, how rapidly after the avulsion should surgical intervention take place? In this study we have performed a gene array in the acute phase with subsequent cluster analysis to evaluate whether this type of analysis may be used to identify time points for critical events after ventral root avulsion and replantation.

MATERIALS AND METHODS

All animal experiments were approved by the local ethic's committee in Stockholm. Six adult Sprague-Dawley rats were anesthetized by Isoflurane inhalation and the lumbosacral spinal cord was exposed. The left L5 ventral root was identified and avulsed by gentle traction of the root. In three of the animals the root was replanted into the lateral funiculus (Figure 1). The wound was closed in layers. After 24 h the animals were euthanized with 0.5 ml pentobarbital (40 mg/ml) and the inferior vena cava was cut open. The lumbosacral spinal cord was rapidly dissected out, meninges and rootlets removed. Thereafter the spinal cord segment L5 was immersed in RNAlater® (Qiagen, Crawley, West Sussex, UK). Lumbar spinal cord specimens from three unoperated animals were collected and treated accordingly and used as controls. For microarray analysis, samples comprising the left ventral quadrant (lesion side) were used. RNA samples were analyzed at the Karolinska Institutet core facility for Bioinformatics and Expression Analysis¹, where target preparation and hybridization to the microarray were completed. RNA was labeled with biotin to produce the final target according to Affymetrix standard procedures2. Labeled cRNA was then hybridized to the Affymetrix Rat Gene ST 1.0 array. Each of the 27,342 genes are represented on the array by approximately 26 probes spread across the full length of the gene. This 700,000 unique 25-mer oligonucleotide design is supposed to provide an accurate picture of gene expression. After probing and scanning, the quality of the images was checked. All arrays passed the Affymetrix quality control check. Following normalization, the change in gene expression between the three controls, the three avulsed, and the three avulsed and replanted rats was compared using an unpaired t-test and fold change values. Lists of genes that passed the selected significance level were uploaded to the Database for Annotation, Visualization, and Integrated Discovery (DAVID)³ for functional annotation and detection of enriched functional-related gene groups and enriched biological themes, particularly GO terms (Dennis et al., 2003; Huang da et al., 2009).

RESULTS

Thus, about 30,000 transcripts were analyzed in the three groups (replanted, avulsed only, and controls). The material was evaluated with regard to signal intensity and fold change (signal ratio in cross comparison tests). *p*-Values for the three intensity values for each gene in each group were calculated. Around 2000 genes in each comparison (replantation vs avulsion; replantation vs control; avulsion vs control) had a *p*-value of stronger than <0.05. The aim of the study was to perform a broad comparison of biological themes, based on a comparison of the about 2000 genes in each group that had a strong *p*-value. It would therefore be difficult to validate the findings by examination of individual genes. Instead, the lists of significantly regulated genes from each group were evaluated with



the aid of the DAVID. It was possible to group responding genes into functional-related gene groups (GO - Gene Ontology search terms). All significantly regulated genes, both up- and down-regulated, were used in this part of the analysis in order to identify the most actively regulated GO families. Distinctive profiles for each of the examined conditions were apparent when the families of genes related to development, differentiation, inflammatory response, apoptosis, neurogenesis, and synaptic transmission were compared. For example, a number of genes related to neurogenesis were found to be up-regulated in animals subjected to avulsion combined with replantation (Figure 2). A distinct change in expression of genes related to synaptic transmission and conduction of nerve impulses was also observed in the replanted animals, but not after avulsion only. A response related to inflammation was more obvious after avulsion than replantation. No obvious difference with regard to cell death systems was found.

The differences were also assessed by cluster analysis of enriched biological themes, including pathways. The Enrichment score is based on the mean value of the –log of the *p*-values (EASEscore) for the members in that cluster. A cut-off value of 2.0 was used, which corresponds to a *p*-value <0.01. Thus, a higher Enrichment score value indicates a low *p*-value and therefore means that the cluster is significant. The most enriched clusters (most significantly regulated) observed after replantation were related to development and differentiation as well as synaptic transmission (**Table 1**). Avulsion injury only, induced large shifts in genes related to inflammation, signal transduction, and proliferation (**Table 2**). Enriched themes in a comparison between replantation and avulsion were development/differentiation and synaptic transmission (**Table 3**).

DISCUSSION

The biological changes initiated after a proximal motor axon lesion are very complex. Spinal motoneurons seem to have a stronger potential than average for axonal regrowth. However, the interaction between the motoneurons and the environment appears to be very complex. The outcome of untreated ventral root avulsion may be paralysis, loss of sensory function, and neurogenic pain. Clinical studies show that replantation with subsequent motor reinnervation can reduce neuropathic pain as well (Htut et al., 2006).

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³http://david.abcc.ncifcrf.gov

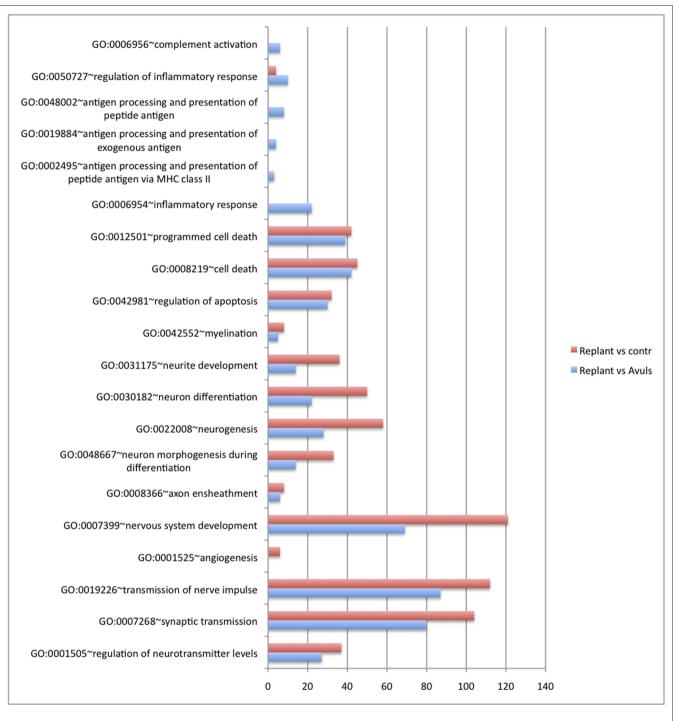


FIGURE 2 | A diagram illustrating the number of significantly regulated genes in selected categories after avulsion and avulsion plus replantation. The *x*-axis indicates the number of genes that have undergone significant changes in expression. Note that this number includes both up- and down-

regulated genes. The number of cell death genes is similar in the two groups, while genes related to neurite development and neurogenesis show a much more prominent response after replantation than after avulsion only. The inflammatory response is more obvious after avulsion than after replantation.

The replantation will result in a limited but unavoidable spinal cord injury as well as disruption of the BBB (Sjögren et al., 1991). Many of the events (Risling et al., 1992, Frisen et al., 1998, Lindholm et al., 2001, 2002, 2004) that have been observed after lesions in the ventral funiculus can be assumed to occur also after this type

of lesion. The replantation is performed to the lateral aspect of the spinal cord, a position more accessible for surgery than the ventral funiculus. It has been shown that regrowing axons are capable of finding a new route inside the spinal cord and navigate to the replanted root (Cullheim et al., 1989), while other axons use the pia Table 1 | Replantation vs control. Cluster analysis of regulated genes,employing the Database for Annotation, Visualization, and IntegratedDiscovery (DAVID). It was possible to group responding genes intofunctional-related gene groups (GO – Gene Ontology search terms) andcluster analysis of enriched biological themes. The Enrichment score is basedon the mean value of the –log of the *p*-values (EASEscore) for the membersin that cluster. A cut-off value of 2.0 was used, which corresponds to a*p*-value <0.01. The most enriched cluster contains themes for neurite</td>development and neuron differentiation.

Term	Count	<i>p</i> -Value
FUNCTIONAL GROUP 1 ENRICHMENT SCORE:	8.67	
GO:0007399 ~ nervous system development	121	1,20E-24
GO:0030182 ~ neuron differentiation	50	6,24E-11
GO:0048699 ~ generation of neurons	53	2,54E-10
GO:0031175 ~ neurite development	36	3,36E-09
GO:0048667 ~ neuron morphogenesis	33	4,35E-09
during differentiation		
GO:0032990 ~ cell part morphogenesis	36	1,42E-06
GO:0048468 ~ cell development	104	1,73E-05
GO:0000902 ~ cell morphogenesis	47	3,63E-04
GO:0032989 ~ cellular	47	3,63E-04
structure morphogenesis		
FUNCTIONAL GROUP 2 ENRICHMENT SCORE:	7.64	
GO:0007269 ~ neurotransmitter secretion	27	2,61E-12
GO:0045055 ~ regulated secretory pathway	28	1,35E-11
GO:0048489 ~ synaptic vesicle transport	18	4,79E-10
GO:0006887 ~ exocytosis	26	1,72E-09
GO:0003001 ~ generation of a signal	29	2,31E-09
involved in cell–cell signaling		
GO:0045045 ~ secretory pathway	40	4,46E-08
GO:0032940 ~ secretion by cell	42	4,81E-07
GO:0016079 ~ synaptic vesicle exocytosis	11	6,48E-07
GO:0016192 ~ vesicle-mediated transport	47	1,21E-04
GO:0051649 ~ establishment of	74	3,80E-04
cellular localization		
FUNCTIONAL GROUP 3 ENRICHMENT SCORE:	5.79	
GO:0044255 ~ cellular lipid	62	1,75E-07
metabolic process		
GO:0008610 ~ lipid biosynthetic process	37	3,74E-07
GO:0006694 ~ steroid biosynthetic process	18	1,35E-06
GO:0016125 ~ sterol metabolic process	18	1,98E-06
GO:0008202 ~ steroid metabolic process	24	6,15E-05
FUNCTIONAL GROUP 4 ENRICHMENT SCORE:	3.68	
GO:0048878 ~ chemical homeostasis	37	3,37E-05
GO:0006873 ~ cellular ion homeostasis	30	1,50E-04
GO:0019725 ~ cellular homeostasis	33	1,81E-03
FUNCTIONAL GROUP 5 ENRICHMENT SCORE:	3.19	
GO:0046942 ~ carboxylic acid transport	18	2,35E-04
GO:0015849 ~ organic acid transport	18	2,63E-04
GO:0006865 ~ amino acid transport	14	6,20E-04
GO:0015837 ~ amine transport	14	4,56E-03
FUNCTIONAL GROUP 6 ENRICHMENT SCORE:	2.54	
GO:0006631 ~ fatty acid metabolic process	24	1,43E-04
GO:0006633 ~ fatty acid biosynthetic process	10	7,74E-03
GO:0016053 ~ organic acid biosynthetic process	10	2,21E-02

 Table 2 | Avulsion vs control. A cluster analysis of the response to avulsion.

 Themes related to inflammatory response are enriched in this analysis.

Term	Count	<i>p</i> -Value
FUNCTIONAL GROUP 1 ENRICHMENT SCORE:	7.63	
GO:0007166 ~ cell surface receptor	172	8,11E-11
linked signal transduction		
GO:0050907 ~ detection of chemical	94	3,71E-08
stimulus during sensory perception		
GO:0007600 ~ sensory perception	102	4,19E-06
FUNCTIONAL GROUP 2 ENRICHMENT SCORE:	4.39	
GO:0009617 ~ response to bacterium	15	6,72E-07
GO:0032496 ~ response to lipopolysaccharide	6	2,70E-04
GO:0002237 ~ response to molecule	6	3,80E-04
of bacterial origin		
FUNCTIONAL GROUP 3 ENRICHMENT SCORE:	4.15	
GO:0042127 ~ regulation of cell proliferation	44	7,94E-06
GO:0048522 ~ positive regulation of	64	1,89E-04
cellular process		,
GO:0008284 ~ positive regulation of	24	2,31E-04
cell proliferation		
FUNCTIONAL GROUP 4 ENRICHMENT SCORE:	3 99	
GO:0002526 ~ acute inflammatory response	19	2,41E-07
GO:0002443 ~ leukocyte mediated immunity	13	4,06E-06
GO:0002460 ~ adaptive immune response based	17	4,00L 00
on somatic recombination of immune receptors	13	9,91E-06
built from immunoglobulin superfamily domains	10	0,01L-00
GO:0002449 ~ lymphocyte mediated immunity	12	2,87E-05
GO:0050776 ~ regulation of immune response	12	2,87E-05
GO:0002682 ~ regulation of immune	15	1,64E-04
system process	15	1,042-04
GO:0051240 ~ positive regulation of multicellular	16	3,55E-04
organismal process	10	3,55L-04
GO:0006956 ~ complement activation	10	3,71E-04
GO:0050778 ~ positive regulation of	10	9,04E-04
immune response	12	3,04L-04
GO:0002684 ~ positive regulation of	12	1,21E-03
immune system process	12	1,212-03
GO:0002455 ~ humoral immune response	6	1.49E-03
mediated by circulating immunoglobulin	0	1,402-00
GO:0006958 ~ complement activation,	6	1,49E-03
classical pathway	0	1,432-03
1 ,	2.20	
FUNCTIONAL GROUP 5 ENRICHMENT SCORE:		7,94E-06
GO:0042127 ~ regulation of cell proliferation	44	,
GO:0008285 ~ negative regulation of	23	2,39E-04
cell proliferation	40	
GO:0048523 ~ negative regulation of	49	8,88E-02
cellular process		
FUNCTIONAL GROUP 6 ENRICHMENT SCORE:		4 405 65
GO:0048534 ~ hemopoietic or lymphoid	19	1,43E-03
organ development		o · ·
GO:0002520 ~ immune system development	19	2,12E-03
GO:0030097 ~ hemopoiesis	17	4,19E-03
GO:0030099 ~ myeloid cell differentiation	11	5,12E-03
		(Continued)

Table 2 | Continued

Term	Count	<i>p</i> -Value	
FUNCTIONAL GROUP 7 ENRICHMENT SCORE: 2.08			
GO:0007067 ~ mitosis	15	2,73E-03	
GO:0000087 ~ M phase of mitotic cell cycle	15	3,52E-03	
GO:0022403 ~ cell cycle phase	21	8,20E-03	
GO:0000279 ~ M phase	17	9,70E-03	
GO:0000074 ~ regulation of progression	20	1,94E-02	
through cell cycle			
GO:0051726 ~ regulation of cell cycle	20	2,15E-02	

mater as a conduit to enter the replanted root (Risling et al., 1991). It seems possible that the replanted ventral root, with its reactive Schwann cells, may have a trophic and neuroprotective effect on injured motoneurons. Previous studies indicate that ventral root replantation can be neuroprotective for motoneurons (Hoang and Havton, 2006; Eggers et al., 2010). Thus, replantation appears to elicit a number of important events in the spinal cord that may be partly beneficial with respect to regrowth to the replanted root. The acute changes in gene expression that were detected in the present study should therefore be assumed to be related to the initiation of a large number of biological changes that occur after this type of injury and surgery. Important themes that were affected were related to neurite growth, cell death, inflammation, and synaptic transmission.

Micro arrays have been employed to identify changes in gene expression after ventral root avulsion (Hu et al., 2002; Yang et al., 2006). The study by Hu et al. (2002) revealed increases in the expression of genes coding for proteins involved in the apoptosis cascades, as well as decreases in expression of genes related to energy metabolism, transporter proteins, ion channels, and receptors. It was also shown that cathepsins, metalloproteinases, and proteasome-related protein products were highly up-regulated in motor neurons following axotomy. The study by Yang et al. (2006) showed a decreased expression of genes that are known to facilitate neuronal survival and axonal regeneration.

Careful mapping of gene regions regulating neurodegeneration has revealed that the gene response linked to neurodegeneration after avulsion is closely related to T cell infiltration and major histocompatibility complex class II expression on microglia (Lidman et al., 2003; Olsson et al., 2005). The kinetics of the avulsion lesion has been summarized by Olsson et al. (2005): During the first few days following the injury, a first phase of microglial activation can be detected. After approximately 1 week the activated cells begin to express MHC class II. This is paralleled by a sparse lymphocyte infiltration and commencing death of axotomized cells. During the second and third post-operative weeks approximately 50–75% of the axotomized cells disappear.

These studies have generated important data relevant for understanding of how degeneration of motoneurons is initiated following avulsion injury. The present study, however, is to our knowledge the first direct comparison of the response to avulsion and replantation based on micro arrays with aid of cluster analysis. The results suggest that this type of gene ontology comparison can be a powerful and effective way to deal with the **Table 3 | Avulsion vs replantation.** A cluster analysis of the response to avulsion compared to the response to replantation after avulsion. Themes relating to synaptic transmission are enriched.

	Count	<i>p</i> -Value
ENRICHMENT SCORE: 6.62		
GO:0031644 ~ regulation of neurological	38	1,88E-10
system process		
GO:0051969 ~ regulation of transmission of	35	1,72E-09
nerve impulse		
GO:0044057 ~ regulation of system process	43	2,09E-06
GO:0010646 ~ regulation of	79	4,82E-03
cell communication		
ENRICHMENT SCORE: 6.17		
GO:0006873 ~ cellular ion homeostasis	52	3,77E-08
GO:0019725 ~ cellular homeostasis	56	7,93E-07
GO:0048878 ~ chemical homeostasis	56	1,01E-05
ENRICHMENT SCORE: 5.55		
GO:0006836 ~ neurotransmitter transport	32	1,65E-14
GO:0046903 ~ secretion	41	1,23E-07
GO:0007269 ~ neurotransmitter secretion	16	3,01E-07
GO:0003001 ~ generation of a signal involved	21	6,62E-06
in cell-cell signaling		
GO:0032940 ~ secretion by cell	31	8,37E-06
GO :0006887 ~ exocytosis	21	2,31E-05
GO:0048489 ~ synaptic vesicle transport	11	1,40E-04
GO:0016079 ~ synaptic vesicle exocytosis	8	5,76E-04
GO:0016192 ~ vesicle-mediated transport	33	1,71E-01
ENRICHMENT SCORE: 3.96		
GO:0050905 ~ neuromuscular process	15	2,12E-05
GO:0008344 ~ adult locomotory behavior	14	7,91E-05
GO:0007628 ~ adult walking behavior	8	7,47E-04
ENRICHMENT SCORE: 3.48		
GO:0051046 ~ regulation of secretion	34	1,62E-05
GO:0017157 ~ regulation of exocytosis	12	2,14E-04
GO:0051049 ~ regulation of transport	48	3,77E-04
GO:0060341 ~ regulation of cellular localization	33	4,03E-04
GO:0060627 ~ regulation of	18	4,53E-04
vesicle-mediated transport		
GO:0051047 ~ positive regulation of secretion	18	9,21E-04
GO:0051050 ~ positive regulation of transport	27	2,11E-03
ENRICHMENT SCORE: 3.43		
GO:0034728 ~ nucleosome organization	16	1,434E-05
GO:0065004 ~ protein-DNA complex assembly	16	2,063E-05
GO:0006334 ~ nucleosome assembly	15	2,560E-05
GO:0034622 ~ cellular macromolecular	25	1,072E-02
complex assembly		
GO:0006325 ~ chromatin organization	22	9,082E-02
ENRICHMENT SCORE: 2.90		
GO:0006865 ~ amino acid transport	16	2,082E-04
	10	4,010E-04
GO:0015837 ~ amine transport		.,
GO:0015837 ~ amine transport GO:0046942 ~ carboxylic acid transport	20	1,491F-03
GO:0015837 ~ amine transport GO:0046942 ~ carboxylic acid transport GO:0015849 ~ organic acid transport	20 20	1,491E-03 1,617E-03

(Continued)

Table 3 | Continued

Term	Count	<i>p</i> -Value
GO:0015807 ~ L-amino acid transport	7	7,361E-03
ENRICHMENT SCORE: 2.40		
GO:0007612 ~ learning	14	3,83E-04
GO:0007613 ~ memory	11	2,21E-03
GO:0008542 ~ visual learning	8	2,29E-03
GO:0009416 ~ response to light stimulus	17	2,66E-03
GO:0045471 ~ response to ethanol	9	1,87E-01

complex information that is generated by micro arrays. The data appears compatible with the notion that replantation initiates a significant growth response. The data of the present study show that the number of regulated genes related to neurite formation is much higher in the replanted animals, whereas the number of regulated cell death genes is similar in the two situations. Our data suggest that the inflammatory response is more prominent in animals subjected to avulsion without replantation. These data indicate that the axonal regenerative response from replantation is initiated at an earlier stage than possible differences in neuroprotective effects. Inflammatory changes that may precede degeneration seem be more pronounced already at 24 h in the animals subjected to avulsion only, although infiltration with T cells, other lymphocytes, and macrophages has not reached a maximum at this stage (cf. Olsson et al., 2005). Thus, this part of the inflammatory response may be dependent on resident cells, such as microglia. It should also be pointed out that a regulation

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of genes for inflammatory molecules may relate to events that not are regarded as a component in the classical inflammatory response. For example the complement cascade molecule C1q has been shown to function as a molecular tag for synaptic removal. The molecular interactions that localize C1q specifically to weak synapses are unknown. However, microglia, the resident immune cells of the brain, produce large amounts of the receptors for C1q and C3 and thus are likely to be responsible for the removal of unwanted synapses (Eroglu and Barres, 2010). It should not be excluded that similar mechanisms could be activated during the early stage following ventral root avulsion and could be involved in the removal of excitatory glutamate synapses (Lindå et al., 2000).

In a recent study we have observed differences between rotational and penetrating brain injuries with the same type of gene arrays and gene ontology analysis (Risling et al., 2010). Interestingly, the regenerative response that was observed in the present study after replantation could not be detected 24 h after a penetrating brain injury (Risling et al., 2010), further emphasizing the unique response following the replantation. It is concluded that this type of gene expression analysis can be of value for the examination of time points for critical events after neurotrauma and that the events after a superficial lesion in the spinal cord has some differences compared to a superficial lesion in the brain. We suggest that gene ontology can be a valuable tool that can be used to establish the time window for a delayed ventral root replantation, since the method seems capable to detect important differences between a regenerative and a nonregenerative response.

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial

or financial relationships that could be construed as a potential conflict of interest.

Received: 08 September 2010; accepted: 21 December 2010; published online: 04 January 2011.

Citation: Risling M, Ochsmann T, Carlstedt T, Lindå H, Plantman S, Rostami E, Angeria M and Sköld MK (2011) On acute gene expression changes after ventral root replantation. Front. Neur. 1:159. doi: 10.3389/fneur.2010.00159

This article was submitted to Frontiers in Neurotrauma, a specialty of Frontiers in Neurology.

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