Synthesis and Antimicrobial Activity of Novel Benzoxazoles

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A series of 2-(*p*-substituted-benzyl)-5-[[4-(*p*-chloro/fluoro-phenyl)piperazin-1-yl]acetamido]-benzoxazoles were synthesized in need of new compounds for the fight against microbial pathogens. Their structures were elucidated by spectral techniques. These new derivatives, along with previously synthesized 2-(*p*-substituted-benzyl)-5-substituted-benzoxazoles, were evaluated for their antibacterial and antifungal activities against standard strains and drug-resistant isolates in comparison with ampicillin, gentamicin sulfate, ofloxacin, vancomycin, fluconazole, and amphotericin B trihydrate. The minimum inhibitory concentration (MIC) of each compound was determined by a two-fold serial dilution technique. The compounds were found to possess a broad spectrum of antimicrobial activities with MIC values of $32-256 \ \mu g/ml$. Although standard drugs were more active against the pathogenes employed in this study, the activities of the new benzoxazoles and reference drugs against drug-resistant isolates of the microorganisms were largely similar.

Key words: Benzoxazoles, Antibacterial Activity, Antifungal Activity

Introduction

The fight against bacterial infections has resulted in the development of a wide variety of antibiotics. After years of misuse of antibiotics, bacteria have become antibiotic-resistant, resulting in a potential global health crisis. Infectious diseases due to Gram-positive bacteria such as methicillin-resistant Staphylococcus aureus (MRSA), vancomycin-resistant Enterococcus faecalis (VREF), and penicillin-resistant Streptococcus pneumoniae (PRSP) are the leading causes of morbidity and mortality today (Moustafa et al., 2004). Besides, during the past 20 years an increase in invasive fungal infections, particularly in immunosuppressed patients, has been observed which are now considered to be causes of morbidity and mortality as well. Therefore, there is still need for new antifungal and antibacterial agents (Andriole, 1999). Benzoxazoles are the structural isosteres of natural nucleotides and interact easily with the biopolymers so that they constitute an important class of heterocyclic compounds with antimicrobial and antibiotic activity (Prudhomme et al., 1986; Sarma et al., 2003; Haansuu et al., 2001;

Temiz-Arpaci et al., 2002, 2005; Yildiz-Oren et al., 2004; Tekiner-Gulbas et al., 2007). The benzoxazole derivative calcimycin (Fig. 1) is a carboxylic polyether antibiotic from a strain of *Streptomyces chartreusis* (NRRL 3882). It was found to be very active against Gram-positive bacteria including some *Bacillus* and *Micrococcus* strains (Prudhomme et al., 1986).

In the last seven years we have described the synthesis of different derivatives of some 2,5-disubstituted benzoxazoles and their in vitro antimicrobial activity against some Gram-positive and Gram-negative bacteria and the fungus Candida albicans (Temiz-Arpaci et al., 2005, 2008; Oksuzoglu et al., 2007, 2008; Arisoy et al., 2008). In the present study, a new series of 2-(p-substituted-benzyl)-5-[[4-(p-chloro/fluoro-phenyl)piperazin-1-yl]acetamido]-benzoxazoles, 3 - 12. has been synthesized as target compounds and evaluated for their antibacterial and antifungal activities, along with previously synthesized benzoxazole derivatives, against standard strains and drug-resistant isolates in comparison with several reference drugs. Furthermore, structure-activity relationships (SAR) are discussed.

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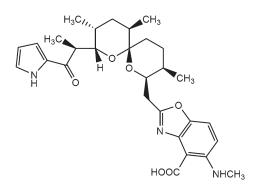


Fig. 1. Chemical structure of calcimycin.

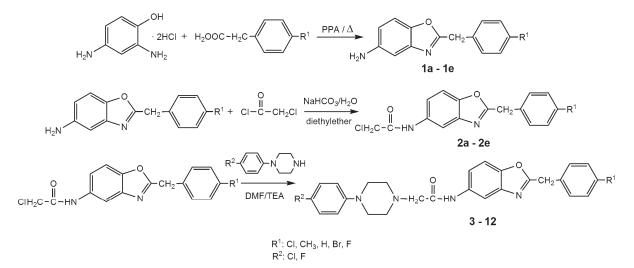
Results and Discussion

We aimed at enhancing the antimicrobial activity of 2,5-disubstituted benzoxazoles by expanding the substituent at the C-5 position. To this end, some new 2-(p-substituted-benzyl)-5-[[4-(pchloro/fluoro-phenyl)piperazin-1-yl]acetamido]benzoxazoles, 3-12, were synthesized. The synthetic route for preparation of the target compounds is shown in Scheme 1. First the 5-amino-2-(*p*-substituted-benzyl)-benzoxazoles 1a - 1ewere obtained by heating appropriate acids with 2,4-diaminophenol in polyphosphoric acid (PPA). The amides 2a-2e were then obtained through the reaction of 5-amino-2-(p-substituted-benzyl)benzoxazoles with chloroacetyl chloride. In the last step the newly synthesized compounds 3-12were prepared from the amides by treating them

with 4-substituted piperazine derivatives. Their structures were elucidated by mass and NMR spectroscopy, respectively, and their purity was analysed through elemental analysis (Table I). The compounds were also evaluated for their antimicrobial activity along with previously synthesized benzoxazole derivatives (Table II).

Compounds 3–12 had same but low antibacterial activity against the bacteria S. aureus and E. faeca*lis* with minimum inhibitory concentration (MIC) values between $128-256 \,\mu \text{g/ml}$ which are higher than those of the standards ampicillin, gentamicin, ofloxacin, and vancomycin. Nevertheless, these compounds had enhanced activity against the drugresistant isolates of these bacteria, in the range of a MIC value of 64 μ g/ml of ampicillin against MRSA (Staphylococcus aureus isolate resistant to methicillin), except for compound 8 which was as potent as gentamicin and vancomycin against the isolate of *E. faecalis* with a MIC value of $32 \mu g/ml$. The new benzoxazole derivatives possessed low antibacterial activity against B. subtillis and its drugresistant isolate with MIC values of $128 \,\mu g/ml$.

Compounds 3-12 provided moderate activity against *P. aeruginosa* and its drug-resistant isolate with MIC values of $64 \mu g/ml$. They had the same activity like ofloxacin against the drug-resistant isolate of *P. aeruginosa*. All derivatives 3-32 possessed low activity against the standard strains of *E. coli* and *K. pneumoniae* in comparison with the standard drugs and, in general, the activities of the previously synthesized derivatives 13-32



Scheme 1. Synthetic pathway of the target compounds 3-12.

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Table I.

	Formula Calculated Found	C ₂₆ H ₂₄ Cl ₂ N ₄ O ₂ C 63.04, H 4.88, N 11.31 C 63.23, H 5.07, N 11.71	C ₅₆ H ₂₄ CIFN ₄ O ₂ C 65.20, H 5.05, N 11.70 C 65.18, H 5.23, N 11.81	C ₂₇ H ₂₇ CIN4O ₂ C 68.27, H 5.73, N 11.80 C 68.00, H 5.68, N 11.99	C 70.72, H 5.94, N 12.22 C 70.72, H 5.94, N 12.22 C 71.12, H 5.67, N 12.26	C ₂₆ H ₂₅ CIN ₄ O ₂ C 67.75, H 5.47, N 12.15 C 67.50, H 5.15, N 11.97	C 70.25, H 5.67, N 12.60 C 70.25, H 5.67, N 12.60 C 70.34, H 5.64, N 12.38	C ₂₆ H ₂₄ CIFN ₄ O ₂ C 65.20, H 5.05, N 11.70 C 65.29, H 4.71, N 11.53
	m/e (%X) (M+H)	$\begin{array}{c} 495(100) \\ 497(70) \\ 499(15) \end{array}$	479(100) 481(37)	475(100) 477(37)	459(100)	461(100) 463(37)	444(100)	479(100) 481(37)
CH2 CH2 K1	13 C NMR (δ in ppm)	34.07, 48.55, 53.21, 62.32, 110.91, 110.97, 117.51, 118.00, 122.96, 129.28, 131.68, 132.49, 134.84, 136.14, 141.61, 147.26, 150.48, 166.37, 168.81			21.30, 34.49, 49.53, 53.39, 62.36, 110.91, 115.81, 116.03, 117.76, 117.83, 117.88, 129.54, 129.89, 132.75, 136.09, 136.85, 141.71, 147.29, 148.60, 155.49, 157.84, 166.87, 168.82		34.88, 49.53, 53.39, 62.37, 110.94, 115.81, 116.03, 117.76, 117.83, 117.93, 127.73, 129.34, 129.68, 135.84, 136.13, 141.71, 147.31, 148.58, 155.50, 157.84, 166.69, 168.83	
R ² – N – H ₂ C – C – HN	$(\delta \text{ in ppm}, J \text{ in Hz})$	2.66–2.68 (4H, t), 3.21 (6H, s), 4.35 (2H, s), 6.94–6.97 (2H, d, $J_{\circ} = 9.2$), 7.22–7.24 (2H, d, $J_{\circ} = 8.8$), 7.42 (4H, s), 7.53–7.56 (H, dd, $J_{\circ} =$ 8.8, $J_{m} = 2.0$), 7.59–7.61 (H, d, $J_{\circ} = 8.4$), 8.08–8.08 (H, d, $J_{m} = 1.6$), 9.92 (H, s)	$\begin{array}{l} 2.67-2.69 \ (\mathrm{4H, t}), \ 3.15-3.17 \ (\mathrm{4H, t}), \\ 3.22 \ (\mathrm{2H, s}), \ 4.35 \ (\mathrm{2H, s}), \ 6.94-7.07 \ (\mathrm{4H, m}), \\ 7.42 \ (\mathrm{4H, s}), \ 7.53-7.56 \ (\mathrm{H, dd}, \ J_o = 8.8, \ J_m = \\ 2.0), \ 7.59-7.62 \ (\mathrm{H, d}, \ J_o = 8.8), \ 8.08-8.09 \\ (\mathrm{H, d}, \ J_m = 2.0), \ 9.92 \ (\mathrm{H, s}) \end{array}$	2.25 (3H, s), 2.63–2.65 (4H, t), 3.19 (6H, s), 4.24 (2H, s), 6.92–6.94 (2H, d, $J_o = 8.8$), 7.12–7.14 (2H, d, $J_o = 8.7$, 7.19–7.23 (4H, m), 7.49–7.52 (H, dd, $J_o = 8.4$, $J_m = 2.0$), 7.55–7.57 (H, d, $J_o = 8.4$), 8.4), 8.04–8.04 (H, d, $J_m = 1.6$), 9.89 (H, s)	2.28 (3H, s), 2.66–2.69 (4H, t), 3.14–3.17 (4H, t), 3.21 (2H, s), 4.26 (2H, s), 6.94–6.97 (2H, m), 7.03–7.07 (2H, m), 7.15–7.16 (2H, d, $J_{\circ} = 7.6$), 7.23–7.25 (2H, d, $J_{\circ} = 8$), 7.51–7.54 (H, dd, $J_{\circ} = 8.8$, $J_{m} = 2.0$), 7.57–7.59 (H, d, $J_{\circ} = 8.8$), 8.07–8.07 (H, d, $J_{m} = 2.0$), 9.91 (H, s)	2.66–2.68 (4H, t), 3.21 (6H, s), 4.32 (2H, s), 6.94–6.96 (2H, d, $J_o = 8.8$), 7.22–7.38 (7H, m), 7.52–7.55 (H, dd, $J_o = 9.2$, $J_m = 2.0$), 7.58–7.60 (H, d, $J_o = 8.8$), 8.08–8.08 (H, d, $J_m = 2.0$), 9.90 (H, s)	$\begin{array}{l} 2.67-2.69 \ (\mathrm{4H}, \ \mathrm{t}), \ 3.15-3.17 \ (\mathrm{4H}, \ \mathrm{t}), \ 3.22 \\ (\mathrm{2H}, \ \mathrm{s}), \ 4.32 \ (\mathrm{2H}, \ \mathrm{s}), \ 6.94-7.07 \ (\mathrm{4H}, \ \mathrm{m}), \\ 7.28-7.38 \ (\mathrm{5H}, \ \mathrm{m}), \ 7.53-7.56 \ (\mathrm{H}, \ \mathrm{d}, \ J_o = 8.8), \\ J_{\mathrm{m}} = 1.6), \ 7.58-7.61 \ (\mathrm{H}, \ \mathrm{d}, \ J_o = 8.8), \ 8.09-8.10 \\ (\mathrm{H}, \ \mathrm{d}, \ J_{\mathrm{m}} = 1.2), \ 9.90 \ (\mathrm{H}, \ \mathrm{s}) \end{array}$	$\begin{array}{l} 2.66-2.69 \ (4H, t), 3.22 \ (6H, s), 4.33 \ (2H, s), \\ 6.94-6.97 \ (2H, d, J_o=8.8), 7.17-7.25 \ (4H, m), \\ 7.41-7.45 \ (2H, m), 7.54-7.57 \ (H, d, J_o=9.2), \\ 7.60-7.62 \ (H, d, J_o=8.4), 8.10 \ (H, s), 9.93 \ (H, s) \end{array}$
	Yield (%)	83	76	75	61	85	73	74
	M.p. [°C]	Cl 150–153	131–134	Cl 148–152	CH ₃ F 148–153	CI 118–122	95-100	CI 130–134
	\mathbb{R}^2		Г	G	т	G	Г	G
	R	CI	CI	CH ₃	CH	Н	Н	ц
	Com- pound	ŝ	4	n	6	F	8	6

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Formula Calculated Found	C ₅₆ H ₂₄ F ₂ N ₄ O ₂ C 67.52, H 5.23, N 12.11 C 67.24, H 5.01, N 11.83	539(77) C ₂₆ H ₃₄ BrCIN ₄ O ₂ 541(100) C 57.85, H 4.48, N 10.38 543(27) C 57.90, H 4.32, N 10.33	523(100) C ₂₆ H ₂₄ BrFN ₄ O ₂ 525(100) C 59.66, H 4.62, N 10.70 C 59.38, H 4.43, N 10.47
m/e (%X) (M+H)	463(100)	539(77) 541(100) 543(27)	523(100) 525(100)
¹³ C NMR (<i>ð</i> in ppm)		33.38, 47.80, 52.46, 61.55, 110.19, 116.73, 117.25, 120.23, 122.21, 128.51, 131.26, 131.44, 134.48, 135.38, 140.86, 146.51, 149.71, 165.51, 168.04	
¹ H NMR $(\delta \text{ in ppm}, J \text{ in Hz})$	$\begin{array}{l} 2.67-2.69 \ (4H, t), \ 3.14-3.17 \ (4H, t), \ 3.21 \ (2H, s), \ 4.33 \ (2H, s), \ 6.94-6.98 \ (2H, m), \ 7.03-7.07 \ (2H, m), \ 7.17-7.21 \ (2H, m), \ 7.41-7.44 \ (2H, m), \ 7.53-7.56 \ (H, \ dd, \ J_{\circ}=9.2, \ J_{\rm m}=2.0), \ 7.59-7.61 \ (H, \ d, \ J_{\circ}=8.4), \ 8.08-8.08 \ (H, \ d, \ J_{\rm m}=2.0), \ 9.91 \ (H, \ s) \end{array}$	$\begin{array}{l} 2.66-2.68 \ (\mathrm{HH}, \mathrm{t}), \ 3.22 \ (\mathrm{6H}, \mathrm{s}), \ 4.33 \ (\mathrm{2H}, \mathrm{s}), \\ 6.94-6.96 \ (\mathrm{2H}, \mathrm{d}, J_o = 8.8), \ 7.22-7.24 \\ (\mathrm{2H}, \mathrm{d}, J_o = 8.8), \ 7.34-7.36 \ (\mathrm{2H}, \mathrm{d}, J_o = 8.4), \\ 7.54-7.62 \ (\mathrm{4H}, \mathrm{m}), \ 8.09-8.10 \ (\mathrm{H}, \mathrm{d}, J_\mathrm{m} = 2.0), \\ 9.93 \ (\mathrm{H}, \mathrm{s}) \end{array}$	2.67–2.69 (4H, t), 3.15–3.17 (4H, t), 3.22 (2H, s), 4.33 (2H, s), 6.94–6.98 (2H, m), 7.03–7.07 (2H, m), 7.34–7.36 (2H, d, $J_{\rm o}$ = 8), 7.54–7.61 (4H, m), 8.08–8.09 (H, d, $J_{\rm m}$ = 1.6), 9.92 (H, s)
Yield (%)		78	
$\begin{array}{cccc} Com- \ R^1 & R^2 & M.p. & Yield \\ pound & & \begin{bmatrix} ^{\circ}C \end{bmatrix} & (\%) \end{array}$	10 F F 96–99 72	11 Br Cl 158–160 78	12 Br F 136–139 80
\mathbb{R}^2	Ц	G	Щ
R	۲ <u>ـ</u>	Br	Br
Com- pound	10	11	12

Table I continued.

were higher than those of the new benzoxazoles. The activities of all compounds against the isolates of these bacteria were as high as those of ampicillin and better than those of gentamicin.

The comparison of the activities of the new benzoxazoles with those of the antifungal drugs fluconazole and amphothericin B showed that the newly synthesized compounds were less inhibitory against *C. albicans* and *C. krusei* with MIC values between $64 \mu \text{g/ml}$ and $256 \mu \text{g/ml}$.

It can be concluded that different substituents at the C-5 position of the 2-(*p*-substitutedbenzyl)-benzoxazole nucleus provided similar antimicrobial activity. The activities of standard drugs against standard strains were higher than those of the benzoxazole derivatives, whereas in most cases the MIC values of standard drugs against the isolates of the bacteria were quite similar to those of the benzoxazoles.

Experimental

Materials and methods

Chemicals and solvents were purchased from Sigma-Aldrich (Taufkirchen, Germany) and Fisher Scientific (Pittsburgh, PA, USA) and were used without further purification. Silica gel HF₂₅₄ chromatoplates (0.3 mm) were used for thin layer chromatography (TLC), and the mobile phase was chloroform/methanol (10:0.5, v/v) for compounds 3-12. Melting points (M.p.) were recorded on a Stuart Scientific SMP 1 (Bibby Scientific Limited, Staffordshire, UK) instrument and are uncorrected. NMR spectra were recorded on a Varian (Palo Alto, CA, USA) Mercury 400 MHz NMR spectrometer in CDCl₃ or dimethylsulfoxide (DMSO d_6); tetramethylsilane (TMS) was used as an internal standard. The mass spectra were recorded on a Waters (Milford, MA, USA) ZQ Micromass LC-MS spectrometer using the ESI(+) method. Elemental analyses were performed on an LECO 932 CHNS (St. Joseph, MI, USA) instrument and were within $\pm 0.4\%$ of theoretical values.

Materials used in the microbiology study were; Mueller Hinton agar (MHA) (Merck, Darmstadt, Germany), Mueller Hinton broth (MHB) (Merck), Sabouraud dextrose agar (SDA) (Merck), RPMI-1640 medium with L-glutamine (Sigma-Aldrich), 3-(N-morpholino)-propane-sulfonic acid (MOPS) (Sigma-Aldrich), 96-well microplates (BD, Franklin Lakes, NJ, USA), transfer pipette (Eppendorf, Hamburg, Germany), am-

Compound	\mathbb{R}^1	\mathbb{R}^2	MIC [µg/ml]														
			0	Gram-negative bacteria					Gram-positive bacteria						Fungi		
			E.c.	E.c.*	K.p.	K.p.*	P.a.	P.a.*	S.a.	S.a.*	E.f.	E.f.*	B.s.	B.s.*	C.a.	C.k.	
3	(p-Cl)Ph-N	Cl	128	128	128	128	64	64	128	64	128	64	128	128	64	64	
4	(p-F)Ph-N	Cl	128	128	128	128	64	64	128	64	128	64	128	128	64	64	
5	(p-Cl)Ph-N	CH_3	128	128	128	128	64	64	128	64	128	64	128	128	128	64	
6	(p-F)Ph-N	CH_3	128	128	128	128	64	64	256	64	128	64	128	128	64	64	
7	(p-Cl)Ph-N	Η	128	128	128	128	64	64	256	64	128	64	128	128	128	64	
8	(p-F)Ph-N	Η	128	128	128	128	64	64	128	64	128	32	128	128	128	64	
9	(p-Cl)Ph-N	F	128	128	128	128	64	64	256	64	128	64	128	128	256	64	
10	(p-F)Ph-N	F	128	128	128	128	64	64	128	64	128	64	128	128	64	64	
11	(p-Cl)Ph-N	Br	128	128	128	128	64	64	128	64	128	64	128	128	128	256	
12	(p-F)Ph-N	Br	128		128	128	64	64	128	64	128	64	128	128	128	64	
13	Ο	Cl	128		128	128	64	64	128	64	128	64	128	128	64	64	
14	CH_2	Cl	64	128	64	128	64	64	32	64	128	64	128	128	128	64	
15	CH ₃ -N	Cl	64		128	128	64	64	128	64	128	64	128	128	64	64	
16	Ph-N	Cl	64	128	64	128	64	64	128	64	128	64	128	256	64	64	
17	Ο	CH_3			128	128	64	64	128	64	128	64	128	128	32	64	
18	CH_2	CH_3	64	128	64	128	64	64	64	64	64	64	128	128	64	64	
19	CH ₃ -N	CH_3	64	128	64	128	64	64	64	64	128	64	128	256	64	64	
20	Ph-N	CH_3	64	128	64	128	64	64	128	64	128	64	128	256	64	64	
21	Ο	Η	128		128	128	64	64	128	64	128	64	128	128	128	64	
22	CH_2	Η	64	128	64	128	64	128	128	64	128	64	128	128	128	64	
23	CH ₃ -N	Η	128		128	128	64	128	256	64	128	64	128	128	128	64	
24	Ph-N	Η	32	128	64	128	64	128	128	64	128	64	128	128	64	64	
25	0	Br	64		128	128	64	128	128	64	128	64	128	128	128	64	
26	CH_2	Br	64		128	128	64	128	128	64	128	64	128	128	128	64	
27	CH ₃ -N	Br	64	128	64	128	64	128	128	64	128	64	128	128	128	64	
28	Ph-N	Br	64		128	128	64	128	256	64	128	64	128	128	128	64	
29	0	F	32	128	64	128	64	64	128	64	128	64	128	128	128	64	
30	CH ₂	F	128		128	128	64	64	128	64	128	64	128	128	128	64	
31	CH ₃ -N	F	32		128	128	64	64	128	64	128	64	128	128	128	64	
32	Ph-N	F	64	128	64	128	64	128	128	64	128	64	128	128	64	64	
Ampicillin			2	128	2	128	n.d.	n.d.	2	64	2	2	0.5	0.5		n.d.	
Genta- micin			0.5	>512	0.5	256	0.5	>512	0.125	32	4	32	0.25	0.125	n.d.	n.d.	
Ofloxacin			< 0.0625	64	0.125	0.5	8	64	0.25	0.25	1	4	0.125	0.25	n.d.	n.d.	
Vanco- mycin			n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	1	1	1	32	n.d.	n.d.	n.d.	n.d.	
Fluco- nazole			n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	1	32	
Ampho- thericin B			n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	0.25	2	

Table II. In vitro antimicrobial activities of newly and previously synthesized benzoxazole derivatives in comparison with reference drugs.

n.d., not determined (microbiological assays were not performed due to following reasons: *P. aeruginosa* is naturally resistant to ampicillin; Gram-negative bacteria employed in the study are naturally resistant to vancomycin; vancomycin is not used in the therapy against *B. subtilis*; antibacterial drugs were not assayed against fungi; antifungal drugs were not assayed against bacteria). E.c., *E. coli* ATCC 25922; E.c.*, *E. coli* isolate (ESBL); K.p., *K. pneumoniae* RSKK 574; K.p.*, *K. pneumoniae*

E.c., E. coli ATCC 25922; E.c.*, E. coli isolate (ESBL); K.p., K. pneumoniae RSKK 574; K.p.*, K. pneumoniae isolate (ESBL); P.a., P. aeruginosa ATCC 25853; P.a.*, P. aeruginosa isolate (resistant to gentamicin); S.a., S. aureus ATCC 29213; S.a.*, S. aureus isolate (MRSA); E.f., E. faecalis ATCC 29212; E.f.*, E. faecalis isolate (VRE); B.s., B. subtillis ATCC 6633; B.s.*, B. subtillis isolate (resistant to ceftriaxon); C.a., C. albicans ATCC 10231; C.k., C. krusei ATCC 6258.

picillin (Mustafa Nevzat Pharmaceuticals, Istanbul, Turkey), gentamicin sulfate (Paninkret, Pinneberg, Germany), ofloxacin (Zhejiang Huangyan East Asia Chemical, Huangyan, Zhejiang, China), vancomycin (Mayne Pharma, Salisbury South, SA, Australia), fluconazole (Sigma-Aldrich), amphotericin B trihydrate (Riedel de Haen, Seelze, Germany), DMSO (Riedel de Haen).

Microorganisms used in the assay were; Klebsiel*la pneumoniae* isolate [extended β -lactamase spectrum (ESBL)], Escherichia coli isolate (ESBL), Enterococcus faecalis isolate [resistant to vancomycin (VRE)], Bacillus subtillis isolate (resistant to ceftriaxon), Pseudomonas aeruginosa isolate (resistant to gentamicin), and Staphylococcus au*reus* isolate [resistant to methicillin (MRSA)], K. pneumoniae RSKK 574, E. coli ATCC 25922, E. faecalis ATCC 29212, P. aeruginosa ATCC 25853, B. subtillis ATCC 6633, S. aureus ATCC 29213, Candida albicans ATCC 10231, and Candida krusei ATCC 6258. Reference strains and clinical isolates were obtained from Gazi University, Faculty of Pharmacy, Department of Pharmaceutical Microbiology, Culture Collection (Ankara, Turkey) and Gazi University Hospital, Microbiology Laboratory (Ankara, Turkey), respectively.

General procedure for the preparation of 5-amino-2-(p-substituted-benzyl)-benzoxazoles (1a-1e)

The 5-amino-2-(p-substituted-benzyl)-benzoxazoles were synthesized by heating 0.02 mol 2,4-diaminophenol hydrochloride with 0.02 mol p-substituted phenyl acetic acid in 25 g polyphosphoric acid (PPA) and stirring for 1-2 h. At the end of the reaction period, the residue was poured into an ice/water mixture, and the solution was neutralized with 10% NaOH. The resulting precipitate was filtered, washed with distilled water, dissolved in boiling ethanol with 0.2 g charcoal, and filtered off. Then distilled water was added slowly to the filtrate in order to stimulate crystallization. The crude compounds 1a-1e were obtained by filtering and drying the crystalline material (Yildiz-Oren et al., 2004; Oksuzoglu et al., 2007, 2008; Temiz-Arpaci et al., 2008; Arisov et al., 2008).

General procedure for the preparation of 5-(2-chloroacetamido)-2-(p-substituted-benzyl)benzoxazoles (**2a**-**2e**)

Chloroacetyl chloride (0.02 mol) was added over a period of 1 h to a stirred, ice-cooled mixture

of 5-amino-2-(*p*-substituted-benzyl)-benzoxazole (0.02 mol), sodium bicarbonate (0.02 mol), diethyl ether (40 ml), and water (20 ml). The mixture was continuously stirred overnight at room temperature and filtered. The precipitate was washed with water, 2 M HCl, and water, respectively, and the crude product was obtained by drying needles *in vacuo* (Arisoy *et al.*, 2008).

General procedure for the preparation of 2-(p-substituted-benzyl)-5-[[4-(p-chloro/fluoro-phenyl)piperazin-1-yl]acetamido]-benzoxazoles (3–12)

0.002 mol 5-(2-chloroacetamido)-2-(p-substituted-benzyl)-benzoxazole were added to 0.002 mol N-(p-chloro/fluoro-phenyl)piperazine and 0.006 mol triethylamine (TEA) solution in 3.5 ml *N*,*N*-dimethylformamide (DMF). The mixture was stirred at room temperature for 24 h. At the end of the reaction time, 5 ml toluene were added, and the reaction medium was evaporated under reduced pressure. The residue was dissolved in chloroform and washed three times with 5% NaOH and then once with distilled water. The solution was dried over anhydrous sodium sulfate, filtered, and evaporated under reduced pressure. The residue was dissolved in ethyl acetate and precipitated by adding *n*-hexane. If necessary, recrystallization was performed. Crystalline material was dried in vacuo. All the compounds 3-12 were prepared as original products. Their structures were supported by spectral data. The mass and ¹H NMR spectra, respectively, and the results of the elemental analyses agree with those of the proposed structures. ¹³C NMR spectra were obtained only for compounds 3, 6, 8, 11. Physical and spectral data of the compounds are reported in Table I.

Microbiological assays

For microbiological assays, ampicillin, gentamicin sulfate, ofloxacin, vancomycin, fluconazole, and amphotericin B trihydrate were dissolved in appropriate solvents recommended by the Clinical and Laboratory Standards Institute (CLSI) guidelines (CLSI, 2006, 2008). Stock solutions of the test compounds were prepared in DMSO.

Bacterial susceptibility testing was performed according to the guidelines of CLSI M100-S18 (CLSI, 2008). MHB was added to each well of the microplates. The bacterial suspensions used for inoculation were prepared at 10⁵ CFU/ml by diluting fresh cultures at McFarland 0.5 density (10⁷ CFU/ml). Two-fold diluted solutions of the compounds were inoculated with bacterial suspensions of 10⁵ CFU/ml (10 μ l inoculum per well to give 10⁴ CFU/ml bacteria in the wells), and the microplates were incubated overnight at 37 °C.

Fungal susceptibility testing was performed according to the guidelines of CLSI M27-A3 (CLSI, 2006). RPMI-1640 medium with L-glutamine buffered to pH 7 with MOPS was added to each well of the microplates. The colonies were suspended in sterile saline, and the resulting suspension was adjusted to McFarland 0.5 density (10^6 CFU/ml). A working suspension was prepared by an 1:100 dilution followed by an 1:20 dilution of the stock suspension. $10 \,\mu$ l of this suspension at 10^3 CFU/ ml were added to the two-fold diluted solution of

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the compounds, and microplates were incubated for 24-48 h at 35 °C.

After incubation, the lowest concentration of the compounds that completely inhibited visible growth was determined and reported as minimum inhibitory concentration (MIC). Control wells contained all components, except the tested compounds, and all experiments were done in triplicate.

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