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# Determinants of Exhaled Breath Condensate pH in a Large Population With Asthma

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*Background:* Exhaled breath condensate (EBC) pH is 2 log orders below normal during acute asthma exacerbations and returns to normal with antiinflammatory therapy. However, the determinants of EBC pH, particularly in stable asthma, are poorly understood. We hypothesized that patients with severe asthma would have low EBC pH and that there would be an asthma subpopulation of patients with characteristically low values.

*Methods:* We studied the association of EBC pH with clinical characteristics in 572 stable subjects enrolled in the Severe Asthma Research Program. These included 250 subjects with severe asthma, 291 with nonsevere asthma, and 31 healthy control subjects.

*Results:* Overall, EBC in this population of stable, treated study subjects was not lower in severe asthma (8.02; interquartile range [IQR], 7.61-8.41) or nonsevere asthma (7.90; IQR, 7.52-8.20) than in control subjects (7.9; IQR, 7.40-8.20). However, in subjects with asthma the data clustered below and above pH 6.5. Subjects in the subpopulation with pH < 6.5 had lower fraction of exhaled NO (FENO) values (FENO =  $22.6 \pm 18.1$  parts per billion) than those with pH  $\geq 6.5$  (39.9  $\pm 40.2$  parts per billion; P < .0001). By multiple linear regression, low EBC pH was associated with high BMI, high BAL neutrophil counts, low prebronchodilator FEV<sub>1</sub> ratio, high allergy symptoms, race other than white, and gastroesophageal reflux symptoms.

*Conclusion:* Asthma is a complex syndrome. Subjects who are not experiencing an exacerbation but have low EBC pH appear to be a unique subpopulation. *CHEST 2011; 139(2):328–336* 

**Abbreviations:** AQLQ = Asthma Quality of Life Questionnaire; EBC = exhaled breath condensate; FENO = fraction of exhaled nitric oxide; GER = gastroesophageal reflux; GSNO = S-nitrosoglutathione;  $PC_{20}$  = provocative concentration of methacholine causing a 20% fall in FEV<sub>1</sub>; SARP = Severe Asthma Research Program

The airway can be acidified in a variety of inflammatory lung diseases.<sup>1-10</sup> The airway epithelium has a pH buffering function analogous to that of the renal tubular epithelium: Acid originating in the airway, or inhaled, is buffered by pH regulatory enzymes.<sup>2,6,11,12</sup> In turn, these enzymes are regulated. For example, airway buffering by glutaminase may be inhibited by T helper cells 1 cytokines, possibly for innate host defense by airway acid.<sup>11-13</sup> In cystic fibrosis, the airway is chronically somewhat acidic.<sup>8,10</sup> In the case of asthma, the proximal airway is acidified minimally, if at all, at baseline,<sup>1,14,15</sup> and it becomes more acidic<sup>12,13</sup> in the context of acute exacerbations caused by intercurrent viral infections.<sup>1,16</sup> Here, we tested the hypothesis that there is a subpopulation of stable, severe asthma patients with low exhaled breath condensate (EBC) pH. This primary outcome variable was studied in a large cohort of subjects in the Severe Asthma Research Program (SARP).

Determinants of EBC pH are multifactorial.<sup>1,11,17-19</sup> Changes in deaerated EBC pH reflect changes in subglottic airway acid production and buffering.<sup>2,9,17,20</sup> For example, EBC pH values measured through an endotracheal tube after intubation are the same as those measured through a mouthpiece before intubation,<sup>20</sup> and subjects with asthma intubated for respiratory failure have low EBC pH that normalizes with corticosteroid treatment.<sup>9</sup> Of note, EBC pH values do not vary with bronchodilator administration, hyperventilation, or hypoventilation.<sup>14,20</sup>

Asthma is likely not exclusively one homogeneous disease but is a collection of different biochemical and immune responses to environmental challenges that have, as a common pathway, airway inflammation and reversible bronchoconstriction.<sup>21,22</sup> We report that low pH is associated with phenotypic features such as high BMI, low baseline predrug FEV, high airway neutrophil percent identified by BAL, non-white race, and gastroesophageal reflux (GER) symptoms. Further, we confirm<sup>1,5,23</sup> that there is a cluster of subjects with EBC pH values < 6.5 and that these have a low fraction of exhaled nitric oxide (FENO) and, particularly in severe asthma, increased allergy symptoms. We conclude that low EBC pH may ultimately be useful as a biomarker to identify (1) specific subpopulations of asthma patients, and (2) exacerbations of asthma.<sup>1,16</sup> EBC pH does not discriminate between severe and nonsevere subgroups of stable asthma, but rather is one biomarker that may be useful for identifying an asthma subpopulation.

### MATERIALS AND METHODS

The Severe Asthma Research Program is a multicenter collaborative study sponsored by the National Heart, Lung, and Blood Institute.<sup>24</sup> The goal of the study is to characterize as fully as

DOI: 10.1378/chest.10-0163

possible the distinguishing features of severe asthma, as defined by the American Thoracic Society Consensus Meeting and adopted by the SARP Steering Committee.24,25 All subjects with severe asthma are maintained long-term on high-dose inhaled and/or systemic corticosteroid treatment and remain symptomatic when compared with a large cohort of subjects with nonsevere asthma and with a smaller cohort of control subjects enrolled for comparison.<sup>24,25</sup> The methods of evaluation have been published.<sup>24</sup> All subjects undergo lung function testing, including maximum bronchodilator challenge; they also undergo an extensive, questionnaire-based history, including allergy history, and those subjects who do not have a specific contraindication undergo methacholine challenge and flexible bronchoscopy. Subjects also undergo allergen skin testing (Greer; Lenoir, North Carolina) for tree mix (white ash, American beech, red birch, American elm, shagbark hickory, red oak, cottonwood), timothy grass, ragweed, Alternaria, Cladosporium, Aspergillus, Dermatophagoides farinae, Dermatophagoides pteronyssinus, weed mix (cocklebur, lambs quarter, pigweed, English plantain, Russian thistle), dog, cat, and cockroach. Most subjects in the first phase of SARP, including all patients at the Wake Forest, North Carolina; Emory, Georgia; Cleveland, Ohio; Virginia; and Wisconsin sites, where there was a specific interest in EBC analysis, underwent EBC analysis. The protocol was approved by the Institution Review Board of each participating institution.

#### EBC Collection

EBC was collected over 5 to 10 min at baseline on the initial visit of the SARP protocol as previously described. Samples were collected using the R-tube (Respiratory Research, Inc; Charlottesville, Virginia) at a collection temperature of  $-20^{\circ}$ C. For this study, EBC was shipped on dry ice at variable time intervals to the University of Virginia for deaeration (at ambient temperature<sup>26</sup>) and pH measurement by investigators blinded to the identity of characteristics of the subject. Note that deaeration and storage at  $-20^{\circ}$ C yields reproducible pH results, even in intercontinental shipping,<sup>20,27,28</sup> across pH range 5-8 (n = 15); repeat assays give an  $r^2 = 0.975.^{27}$  Samples were deaerated using argon as previously described for 7 min, and stable pH recorded as previously described.<sup>1</sup>

#### Data Analysis

All SARP data are collected, stored, and analyzed in the SARP Data Coordinating Center at National Jewish Hospital, Denver, Colorado. Data were further analyzed in the Department of Public Health Sciences at University of Virginia, Charlottesville, Virginia.

By univariate analysis, we studied the relationship between a continuous pH value and risk factors. We computed the correlation between pH value and continuous risk factors (eg, FEV1 and BMI, numbers of positive skin tests, presence of BAL neutrophils). We carried out *t* test and nonparametric rank-sum test for two-sample comparison (eg, GER) and analysis of variance for categorical variables with more than two levels (eg, allergy symptoms in spring and winter). We then used multiple linear regression to study the independent relationship of the continuous pH value with these risk factors, adjusting for other confounding variables. We studied variables associated with dichotomized pH determined by (1) the strongest positive and negative predictive value for identifying GER symptoms (identified by receiver operator curve analysis), and (2) distributions observed to be derived at pH 6.5. The statistical analyses were carried out in SAS 9.1 (SAS Institute Inc; Cary, North Carolina). The plots were drawn in S-Plus software (Insightful Corp; Seattle, Washington). Results are presented as mean  $\pm$  SD or median and range.

Manuscript received January 19, 2010; revision accepted August 9, 2010.

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<sup>\*</sup>A complete list of study group participants is available in e-Appendix 1.

Funding/Support: This work was supported by the National Institutes of Health [Grants NIH/NHLBI: 2RO1, HL 69170 NHLBI Severe Asthma Research Program].

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FIGURE 1. Exhaled breath condensate pH population distribution in normal control subjects (green line), subjects with stable mild to moderate asthma (blue dashed line), and subjects with stable severe asthma (red dotted line). Note the distribution of some subjects with subjects substantially below pH 6.5. Individual subjects are shown as hash marks on the x-axis.

## RESULTS

EBC was collected from 573 subjects aged 6 through 70 years between July 2003 and April 2007. This cohort included 249 subjects with severe asthma (aged  $37.8 \pm 15.6$  years; 39% men), 293 subjects with nonsevere asthma (aged  $32.2 \pm 13.4$  years; 32% men), and 31 healthy control subjects (aged  $28.7 \pm 9.6$  years; 32% men) (e-Tables 1, 2).

In this cohort, EBC pH has a median of 7.94, with interquartile range (IQR) 7.55 to 8.30. Because the pH value is not normally distributed, we used the Wilcoxon rank-sum test to perform the two-sample comparison. The deaerated pH of subjects with asthma (median, 7.94; IQR, 7.56-8.30) did not differ from that of control subjects (7.90; IQR, 7.40-8.20; P = .80); pH in subjects with nonsevere asthma (7.90; IQR, 7.52-8.20) was lower than in subjects with severe asthma (8.02; IQR, 7.61-8.41). Patients with asthma had a bimodal distribution, with a subpopulation having low pH (Figs 1-4).

We initially studied the relationship between a continuous pH value and other risk factors by univariate analysis. We found that low pH value was significantly correlated with GER symptoms (P = .007), race (nonwhite; P = .002), history of pneumonia (P = .04), allergic symptoms without cold or flu (P = .04), acute or recurrent sinusitis (P = .03), nasal polyps (P = .01), not taking any form of cortico steroid (P = .02), receiving anti-IgE therapy (P = .04), and more severe allergy symptoms, particularly in the spring (P = .0001). Low pH value was also associated with high BMI (P = .0009), low baseline prebronchodilator FEV<sub>1</sub> (P = .04), low Asthma Quality

of Life Questionnaire (AQLQ) Emotion Score, and low AQLQ Environmental Score (P = .002).

We then adopted multiple linear regression to study the relationship between the continuous pH level and risk factors (Table 1). We included all the variables that were significant in the univariate analysis. In this multiple linear regression model including all subjects who underwent bronchoscopy (n = 161;overall  $R^2 = 0.32$ ), we found that pH value was negatively associated with BAL neutrophil count, BMI, baseline prebronchodilator FEV<sub>1</sub>, and GER symptoms, and positively associated with prebronchodilator  $FEV_1$ . White patients had a 0.42 U higher pH value than other races (Table 1) (P = .0006). These differences were not affected by a priori National Heart, Lung, and Blood Institute guidelines asthma severity assignment. Of note, the association of low pH with GER symptoms was confounded by a strong association between GER symptoms and BMI (P < .0001). In terms of pathophysiology, it is reasonable to speculate that in some patients, high BMI could

Table 1—Relation of pH and Risk Factors Using Multiple Linear Variable Analysis

Risk Factor	Estimateª	SE	P Value
Baseline prebronchodilator FEV <sub>1</sub>	0.16	0.06	.001
BAL neutrophils, %	-0.037	0.010	.0005
BMI	-0.018	0.008	.03
GER symptoms	-0.41	0.15	.009
Race (white vs non-white)	$0.42^{b}$	0.12	.0006

N = 161; overall  $R^2 = 0.32$ . GER = gastroesophageal reflux.

<sup>a</sup>Slope of relationship (continuous variables) or difference (dichotomous variables).

<sup>b</sup>White pH was 0.42 units higher than non-white (primarily black).



FIGURE 2. The relation between pH and phenotypic features. A, BMI. B, Fraction of BAL neutrophils. C, FEV<sub>1</sub>. D, GER symptoms. Asthma severity is denoted by the following: green dot, control; blue X, not severe; red circle, severe. Note that there is a natural distribution of these data above and below pH 6.5. GER = gastroesophageal reflux.

lead to GER, which, in turn, could contribute to low EBC pH.

In secondary analyses, we observed natural clusters of EBC pH in asthma above and below 6.5 (Figs 1, 2). This distribution had been previously reported.<sup>1,5,23</sup> We therefore also compared patient characteristics dichotomized above and below pH 6.5 (Figs 1-4; Tables 2, 3). We found that FENO was lower in those subjects with EBC pH < 6.5 (P < .0001), consistent with a previous report.<sup>23</sup> Very low EBC pH was also associated with low provocative concentration of methacholine causing a 20% fall in FEV<sub>1</sub> (PC<sub>20</sub>) (P = .03) and, with severe asthma, allergic symptoms in the spring (P = .006) and winter (P = .03) (Fig 4). There was a trend toward association with number of positive allergy skin tests, but this did not achieve significance (Table 2).

## DISCUSSION

Asthma is increasingly appreciated to be a multifactorial symptom complex.<sup>21,22,29</sup> Genetic and metabolomic markers may ultimately be important tools for patient classification leading to more targeted treatment. For example, it is envisioned that response to therapies such as  $\beta_2$ -agonists, corticosteroids, and leukotriene modifiers may be at least in part genetically determined. Optimal treatment could be targeted based on genetics and/or biochemical markers for responsiveness to these treatments.<sup>29</sup>

EBC pH is a metabolomic marker that is low (acidic) during acute asthma exacerbations. Further, decreased airway pH recapitulates many features of asthma.<sup>1,2,7,30-33</sup> Airway pH appears to be normally distributed in the general population, measured at a retail outlet store,34 though outliers with low pH might be explained if some of the subjects had unreported asthma, GER, or intercurrent infections.<sup>11,12,16,19</sup> Here, we report that EBC pH in a large cohort with wellcharacterized severe and nonsevere asthma is, on the whole, normal. In fact, values for subjects with severe asthma were higher than those previously reported for normal black subjects,<sup>23</sup> and could represent a corticosteroid treatment effect. Consistent with a previous report,<sup>33</sup> EBC pH was higher in whites than non-whites.

In secondary analysis, we found that among subjects with asthma there was a cluster of subjects with pH < 6.5. There was also the previously observed threshold at 7.8, but this was not nearly as distinct in our population of closely monitored, heavily treated patients as that at 6.5. This threshold at pH 6.5 has also previously been noted.<sup>1,5,23</sup> It could reflect the point between pKas (the logarithm to the base 10 of the proton concentration in solution at which the acid is 50% protonated) of weak acids and bases. Strikingly, the variable most strongly associated with pH < 6.5 was low FENO, consistent with a previous report that FENO decreases with pH < 6.5.<sup>23</sup> FENO has been used as a nonspecific marker of inflammation, so the



FIGURE 3. The relationship between pH and allergy symptoms in spring and winter. Asthma severity is denoted by symbols: red dot, normal; blue X, not severe; green circle, severe.

robust association of low FENO with low pH may be counterintuitive. However, this association is not completely surprising for two reasons. First, pH < 6.5 approaches the pKa (the logarithm to the base 10 of the proton concentration in solution at which the acid is 50% protonated) of nitrite.<sup>7</sup> In vitro and in vivo, low airway lining fluid pH can lead to nitrite protonation, forming nitrous acid, which rapidly leads to nitric oxide formation.<sup>1,7,10</sup> Although this will acutely increase FENO,<sup>7</sup> the result of chronically low pH will be depletion of nitrite and other airway nitrogen oxides.<sup>1,7,35</sup> Additionally, S-nitrosoglutathione (GSNO) metabolism increases FENO acutely in vivo but depletes airway GSNO levels long-term, decreasing FENO; indeed, GSNO metabolism is increased in many patients with asthma.<sup>32,33,36-39</sup> Single nucleotide polymorphisms in GSNO metabolic enzyme, GSNO reductase, are predictive of asthma,<sup>38,40</sup> and mice deficient in GSNO reductase are protected from experimental asthma.<sup>41</sup> Note in this regard that GSNO reductase activity can be induced by allergen exposure and can result in increased methacholine responsiveness.<sup>36</sup> Both increased allergic symptoms in general (note that samples were obtained in and out of season) and



FIGURE 4. The relationship between pH and response to GERD symptoms by questionnaire (yes/no). Asthma severity is denoted by symbols: red, normal; blue, not severe; green, severe. GERD = gastroesophageal reflux disease.

low methacholine  $PC_{20}$  were characteristics of this low-pH, low-FENO phenotype. Increased GSNO reductase activity in asthma can be associated not only with decreased airway nitric oxide levels but also with increased production of formic acid, an important

Table 2—Relation Between Dichotomous pH (Cutpoint6.5) and Continuous Risk Factors

Risk Factor	$pH \le 6.5,$ Mean (SD) n = 39	pH > 6.5, Mean (SD) n = 533	P Value
BAL pure neutrophils, %	$11.4 (15.3)^{a}$	$2.6 (4.1)^{b}$	.18
BAL eosinophils, %	$0.99 (1.18)^{a}$	$0.97 (2.13)^{b}$	.98
BMI	32.1 (11.4)	28.6(8.1)	.07
Baseline prebronchodilator FEV <sub>1</sub>	2.30 (0.74)	2.45 (0.92)	.24
Baseline prebronchodilator FEV <sub>1</sub> /FVC ratio	0.73 (0.10)	0.72 (0.12)	.34
Methacholine PC <sub>20</sub>	4.1(5.1)	6.8(13.5)	.03
AQLQ emotion score	4.3(1.7)	4.7(1.7)	.21
AQLQ environmental score	4.1(1.5)	4.6(1.6)	.08
IgE	253(436)	344(761)	.29
Log eosinophils: blood	-0.72(0.51)	-0.67(0.50)	.63
No. positive skin tests	2.4(2.9)	3.2 (3.0)	.09
Age at diagnosis of asthma, y	13.1 (11.6)	13.7 (14.0)	.78
Age of onset of allergic symptoms without cold or	9.8 (8.1)	11.3 (10.9)	.32
flu, y			
Average FENO value	22.6 (18.1)	39.8(40.2)	<.0001

AQLQ = asthma quality of life questionnaire; FENO = fraction of exhaled nitric oxide;  $PC_{20}$  = provocative concentration of methacholine causing a 20% fall in FEV<sub>1</sub>.

<sup>a</sup>Only seven subjects had data available.

<sup>b</sup>Only 157 subjects had data available.

determinant of asthmatic airway acidification.<sup>18,42,43</sup> Therefore, the subset of patients with increased GSNO reductase activity might be predicted to have low FENO and low EBC pH. One way or another, this subset appears to be biochemically unique and may be a subset in which the metabolomically targeted therapy (for example, with inhaled base<sup>7,35</sup>) may be particularly beneficial.

It should be noted that airway EBC pH was unrelated to inhaled corticosteroid dose. However, the use of corticosteroids will raise EBC pH<sup>1</sup> acutely during exacerbation. The severe asthmatic group, defined by high-dose inhaled or systemic corticosteroid use, had and overall tendency to have a slightly higher pH.

EBC pH was lower in patients with GER symptoms. A subpopulation of patients with chronic cough who have consistently low EBC pH on serial studies at home appear to be those patients whose cough can be successfully treated with high-dose proton pump inhibitor therapy.<sup>18</sup> However, the finding is based on a subjective questionnaire. Prospective studies will be needed to determine whether a specific EBC pH will be useful in predicting responses to antacid-class therapies in asthma.

In multiple linear regression, low pH was also independently associated with high BAL neutrophil counts, high BMI, low log blood eosinophil count, and low baseline predrug FEV<sub>1</sub>. With regard to the association between EBC pH and BAL neutrophilia, note that this relationship (1) suggests but does not prove that distal airway/alveolar lung lining fluid

	$pH \leq 6.5$ ,	pH > 6.5,	
	No. (%)	No. (%)	0371
KISK Factor	(n = 39)	(n = 533)	P value
Pneumonia history	20 (53)	218(42)	.23
Visited ED for breathing	10(26)	161(30)	.72
problems in last 12 mo			
Use theophylline	2(5)	40(8)	.76
Asthma symptoms caused by physical activity	23 (61)	307 (61)	>.99
Asthma symptoms caused by animal exposure	24 (63)	301 (62)	>.99
Asthma symptoms caused by physical exercise	26 (70)	416 (83)	.07
Asthma symptoms caused by respiratory infections	37 (95)	451 (90)	.41
Allergic symptoms without cold or flu	35(90)	455~(90)	99.<
GER symptoms	10 (28)	109 (21)	.40
History of cats in the home	8 (21)	104 (20)	.84
History of rodents in the home	0 (0)	12(2)	>.99
History of acute or recurrent sinusitis	18 (47)	205 (39)	.31
History of CPAP or bilevel pressure ventilation use	1 (3)	15(3)	>.99
History of immunotherapy for asthma	2(5)	23(5)	.69
History of nasal polyp	5(14)	63 (12)	.79
Contraceptive use	3 (13)	74(24)	.31
Herbal or natural supplement treatment	0 (0)	25 (6)	.24
Allergy symptoms in spring			.006
None	3(8)	43(8)	
Mild	4(10)	148(29)	
Moderate	15(38)	218(42)	
Severe	17(44)	109(21)	
Allergy symptoms in winter			.03
None	5(13)	110(21)	
Mild	8 (21)	160(31)	
Moderate	11(28)	152(29)	
Severe	15(38)	94(18)	
Taking steroid	31 (79)	394(74)	.57
Taking oral steroid	9(24)	102 (20)	.54
Anti-IgE therapy	3(8)	23(4)	.41
Use leukotriene modifiers	14(37)	190 (37)	>.99
Any long-acting β-agonist use	25 (66)	333 (66)	>.99
Race			.23
White	21 (54)	336 (63)	
Black	16(41)	150(28)	
Other	2(5)	48(9)	

 Table 3—Relation Between Dichotomous pH

 (Cutpoint 6.5) and Discrete Risk Factors

No normal control patients have  $pH \leq 6.5$ . CPAP = continuous positive airway pressure. See Table 1 for expansion of other abbreviation.

may influence EBC pH values, and (2) suggests that EBC pH is influenced by neutrophils. Evidence suggests that the distal airway, which is sampled by BAL, may be more acidic than the proximal airway.<sup>35</sup> In the aggregate, subjects with allergies and allergy symptoms could theoretically receive more corticosteroid treatments—and/or have better adherence leading to higher BMI, more GER, lower lung function, and/or relatively more airway neutrophilia. A chronic, undetected infection could also play a role in the neutrophilic inflammation and low pH. $^{9,12,16}$  Low EBC pH was associated with modest evidence for flow limitation, consistent with a previous report, $^{5}$  although a decrease in FEV<sub>1</sub> per se does not cause a change in EBC pH, $^{41}$  and the negative relationship between pH and FEV<sub>1</sub>/FVC ratio suggests that low pH does not predict obstruction. The determinants of airway pH in stable asthma appear to be multifactorial.

It will be important prospectively to study the associations identified in this study, particularly including the subpopulations with (1) low EBC and low FENO, and (2) low EBC pH and increased BAL neutrophils. These subpopulations may be amenable to targeted, steroid-sparing therapy. Indeed, it may be worthwhile to consider interventional trials—for example, with antacid therapy, inhaled alkaline buffer, inhaled nitrogen oxides, and/or exercise and weight loss—in which patients are stratified by EBC pH.

#### Acknowledgments

Author contributions: Dr Liu: contributed to data analysis and manuscript preparation. Dr Teague: contributed to study design and data analysis. Dr Erzurum: contributed to study design and manuscript preparation. Dr Fitzpatrick: contributed to study design, data collection, and manuscript preparation. Ms Mantri: contributed to data analysis and interpretation. Dr Dweik: contributed to study design. Dr Bleecker: contributed to study design. Dr Meyers: contributed to study design and data analysis. Dr Busse: contributed to study design and manuscript preparation. Dr Calhoun: contributed to study design and data analysis. Dr Castro: contributed to study design. Dr Chung: contributed to study design and data analysis. Dr Curran-Everett: contributed to study design, data collection, and data analysis. Dr Israel: contributed to study design and data collection. Dr Jarjour: contributed to study design and data collection. Dr Moore: contributed to study design, data collection, and data analysis. Dr Peters: contributed to study design and data analysis. Dr Wenzel: contributed to study design and data analysis. Dr Hunt: contributed to data collection, data analysis, and manuscript preparation. Dr Gaston: contributed to study design, data analysis, and manuscript preparation. Financial/nonfinancial disclosures: The authors have reported to CHEST the following conflicts of interest: Dr Busse has provided consultancy/advisory board services to Alexon, AstraZeneca, Altair, Amgen, Abbot Laboratories, Asthmatx, Inc, Boehringer-Ingelheim, Bristol-Meyer Squibb Company, Centocor, Dainippon Sumitomo, Funxional Therapeutics, Ltd, GlaxoSmithKline, Pfizer, Genentech, Johnson & Johnson, Merck, Novartis, Schering-Plough, TEVA, and Wyeth; received lecture fees from Merck; received industry-sponsored grants from Novartis, MedImmune, AstraZeneca, Ception, and GlaxoSmithKline; and received National Institutes of Health (NIH) grant support from NIH-NIAID and NIH-NHLBI. Dr Chung has received grant funding from GlaxoSmithKline, MRC (UK), and Wellcome Trust, and has been renumerated for participation at advisory board meetings of GlaxoSmithKline,

Merck, and Gilead. Dr Jarjour has received pharmaceutical

company grant money from GlaxoSmithKline and Merck and has been a consultant to GlaxoSmithKline, Genentech, and Asthmatx. Dr Hunt is a shareholder in Respiratory Research, Inc and in Airbase Pharmaceuticals and is cofounder of Respiratory Research, Inc, which manufactures EBC collection equipment and has financial interest in EBC pH testing. He is an inventor of EBC pH assays, and his university owns IP on EBC pH testing. He speaks regarding EBC pH testing (no honorarium). He advises for Pulse Health, Inc, a company that studies exhaled breath for oxidant stress. He has received grant money from NIH and the UVA Philip Morris Tobacco Research Fund, and Respiratory Research, Inc. He is the founder of AirBase Therapeutics, LLC. Dr Gaston is a shareholder in Respiratory Research, Inc and in Airbase Pharmaceuticals. The remaining authors have reported to CHEST that no potential conflicts of interest exist with any companies/organizations whose products or services may be discussed in this article.

Additional information: The e-Appendix and e-Tables can be found in the Online Supplement at http://chestjournal.chestpubs.org/content/139/2/328/suppl/DC1

## References

- Hunt JF, Fang K, Malik R, et al. Endogenous airway acidification. Implications for asthma pathophysiology. *Am J Respir Crit Care Med.* 2000;161(3 pt 1):694-699.
- 2. Hunt JF, Gaston B, Ricciardolo F. Acid stress in the airway. *J Allergy Clin Immunol*. 2004;84(3):731-765.
- Jack CI, Calverley PM, Donnelly RJ, et al. Simultaneous tracheal and oesophageal pH measurements in asthmatic patients with gastro-oesophageal reflux. *Thorax*. 1995;50(2): 201-204.
- Karnad DR, Mhaisekar DG, Moralwar KV. Respiratory mucus pH in tracheostomized intensive care unit patients: Effects of colonization and pneumonia. *Crit Care Med.* 1990;18(7): 699-701.
- Kostikas K, Papatheodorou G, Ganas K, Psathakis K, Panagou P, Loukides S. pH in expired breath condensate of patients with inflammatory airway diseases. *Am J Respir Crit Care Med.* 2002;165(10):1364-1370.
- Paget-Brown A, Hunt JF, Gaston B. Tracheal fluid pH in human preterm infants. *Eur Respir J.* 2007;30(5):840-842.
- Gaston B, Kelly R, Urban P, et al. Buffering airway acid decreases exhaled nitric oxide in asthma. J Allergy Clin Immunol. 2006;118(4):817-822.
- Tate S, MacGregor G, Davis M, Innes JA, Greening AP. Airways in cystic fibrosis are acidified: detection by exhaled breath condensate. *Thorax*. 2002;57(11):926-929.
- Walsh BK, Mackey DJ, Pajewski T, Yu Y, Gaston BM, Hunt JF. Exhaled-breath condensate pH can be safely and continuously monitored in mechanically ventilated patients. *Respir Care*. 2006;51(10):1125-1131.
- Yoon SS, Coakley R, Lau GW, et al. Anaerobic killing of mucoid Pseudomonas aeruginosa by acidified nitrite under conditions characteristic of the cystic fibrosis airway. J Clin Invest. 2006;116(2):436-446.
- Hunt JF, Erwin E, Palmer L, et al. Expression and activity of pH-regulatory glutaminase in the human airway epithelium. *Am J Respir Crit Care Med.* 2002;165(1):101-107.
- Carraro S, Doherty J, Zaman K, et al. S-nitrosothiols regulate cell-surface pH buffering by airway epithelial cells during the human immune response to rhinovirus. *Am J Physiol Lung Cell Mol Physiol*. 2006;290(5):L827-L832.
- 13. Gaston B, Adjei A, Amoah AG, et al. Breath condensate pH and ammonia levels are low in patients with active pulmonary tuberculosis. *Conference of the Am Thorac Soc.* 2005:A574.
- Nicolaou NC, Lowe LA, Murray CS, Woodcock A, Simpson A, Custovic A. Exhaled breath condensate pH and childhood

asthma: unselected birth cohort study. *Am J Respir Crit Care Med.* 2006;174(3):254-259.

- Coop C, Hagan LL, Dice JP. Exhaled breath condensate pH in the evaluation of asthma. *Allergy Asthma Proc.* 2008; 29(1):51-54.
- Hunt J, Ngamtrakulpanit L, Vaughan J, et al. Airway acidbase disturbance in rhinovirus cold. *Eur Respir J.* 2003; 45(Suppl):566S.
- Wells K, Vaughan J, Pajewski TN, et al. Exhaled breath condensate pH assays are not influenced by oral ammonia. *Thorax*. 2005;60(1):27-31.
- Greenwald R, Fitzpatrick AR, Erzurum S, et al. The severe asthma phenotype is characterized by elevated levels of formate in exhaled breath condensate [abstract]. Am J Respir Crit Care Med. 2009.
- Hunt J, Yu Y, Burns J, et al. Identification of acid reflux cough using serial assays of exhaled breath condensate pH. *Cough*. 2006;2(1):3-6.
- Vaughan J, Ngamtrakulpanit L, Pajewski TN, et al. Exhaled breath condensate pH is a robust and reproducible assay of airway acidity. *Eur Respir J.* 2003;22(6):889-894.
- Wenzel SE. Asthma: defining of the persistent adult phenotypes. *Lancet*. 2006;368(9537):804-813.
- Holgate ST, Polosa R. The mechanisms, diagnosis, and management of severe asthma in adults. *Lancet*. 2006;368(9537): 780-793.
- Hauswirth DW, Sundy JS, Mervin-Blake S, et al. Normative values for exhaled breath condensate pH and its relationship to exhaled nitric oxide in healthy African Americans. J Allergy Clin Immunol. 2008;122(1):101-106.
- 24. Moore WC, Bleecker ER, Curran-Everett DC, et al. Characterization of the severe asthma phenotype by the National Heart, Lung, and Blood Institute's Severe Asthma Research Program. J Allergy Clin Immunol. 2007;119(2): 405-413.
- Wenzel SE, Busse WW; National Heart, Lung, and Blood Institute's Severe Asthma Research Program. Severe asthma: lessons from the Severe Asthma Research Program. J Allergy Clin Immunol. 2007;119(1):14-21.
- Koczulla AR, Noeske S, Herr C, et al. Ambient temperature impacts on pH of exhaled breath condensate. *Respirology*. 2010;15(1):155-159.
- Ngamtrakulpanit L, Yuanlin Y, Adjei A, Amoah G, Gaston B, Hunt J. Identification of intrinsic airway acidification in pulmonary tuberculosis. *Clob J Health Sci.* 2010;2(1):106-110.
- Kullmann T, Barta I, Horváth I. Effect of freezing on exhaled breath condensate pH. *Respir Med.* 2007;101(12):2566.
- Weiss ST, Litonjua AA, Lange C, et al. Overview of the pharmacogenetics of asthma treatment. *Pharmacogenomics J*. 2006;6(5):311-326.
- Ahmed T, Ali JM, al-Sharif AF. Effect of alkali nebulization on bronchoconstriction in acute bronchial asthma. *Respir Med.* 1993;87(3):235-236.
- Holma B. Influence of buffer capacity and pH-dependent rheological properties of respiratory mucus on health effects due to acidic pollution. *Sci Total Environ*. 1985;41(2):101-123.
- Honma S, Tanaka H, Teramoto S, Igarashi T, Abe S. Effects of naturally-occurring acid fog on inflammatory mediators in airway and pulmonary functions in asthmatic patients. *Respir Med.* 2000;94(10):935-942.
- Martínez D, Vermeulen M, von Euw E, et al. Extracellular acidosis triggers the maturation of human dendritic cells and the production of IL-12. *J Immunol*. 2007;179(3): 1950-1959.
- Paget-Brown AO, Ngamtrakulpanit L, Smith A, et al. Normative data for pH of exhaled breath condensate. *Chest.* 2006; 129(2):426-430.

- Shin H-W, Shelley DA, Fitzpatrick A, Gaston B, George SC. Airway nitric oxide is reduced after PBS inhalation in asthma. *J Appl Physiol*. 2007;102(3):1028-1033.
- Snyder AH, McPherson ME, Hunt JF, Johnson M, Stamler JS, Gaston B. Acute effects of aerosolized S-nitrosoglutathione in cystic fibrosis. *Am J Respir Crit Care Med.* 2002;165(7): 922-926.
- Gaston B, Sears S, Woods J, et al. Bronchodilator S-nitrosothiol deficiency in asthmatic respiratory failure. *Lancet.* 1998;351(9112):1317-1319.
- Wu H, Romieu I, Sienra-Monge JJ, et al. Genetic variation in S-nitrosoglutathione reductase (GSNOR) and childhood asthma. J Allergy Clin Immunol. 2007;120(2):322-328.
- Staab CA, Hellgren M, Höög J-O. Medium- and short-chain dehydrogenase/reductase gene and protein families: Dual

functions of alcohol dehydrogenase 3: implications with focus on formaldehyde dehydrogenase and S-nitrosoglutathione reductase activities. *Cell Mol Life Sci.* 2008;65(24):3950-3960.

- 40. Moore PE, Ryckman KK, Williams SM, Patel N, Summar ML, Sheller JR. Genetic variants of GSNOR and ADRB2 influence response to albuterol in African-American children with severe asthma. *Pediatr Pulmonol.* 2009;44(7):649-654.
- 41. Que LG, Liu L, Yan Y, et al. Protection from experimental asthma by an endogenous bronchodilator. *Science*. 2005; 308(5728):1618-1621.
- 42. Henderson EM, Gaston B. SNOR and wheeze: the asthma enzyme? *Trends Mol Med.* 2005;11(11):481-484.
- 43. Que LG, Yang Z, Stamler JS, Lugogo NL, Kraft M. S-nitrosoglutathione reductase: an important regulator in human asthma. Am J Respir Crit Care Med. 2009;180(3):226-231.