



## Polychlorinated Biphenyls as Hormonally Active Structural Analogues

James D. McKinney<sup>1</sup> and Chris L. Waller<sup>2</sup>

<sup>1</sup>Health Effects Research Laboratory, U.S. Environmental Protection Agency, Research Triangle Park, NC 27711 USA; <sup>2</sup>Center for Molecular Design, School of Medicine, Washington University, St. Louis, MO 63130 USA

Among the environmental chemicals that may be able to disrupt the endocrine systems of animals and humans, the polychlorinated biphenyls (PCBs) are a chemical class of considerable concern. One possible mechanism by which PCBs may interfere with endocrine function is their ability to mimic natural hormones. These actions reflect a close relationship between the physicochemical properties encoded in the PCB molecular structure and the responses they evoke in biological systems. These physicochemical properties determine the molecular reactivities of PCBs and are responsible for their recognition at biological acceptors and receptors, as well as for triggering molecular mechanisms that lead to tissue response. "Coplanarity" of PCB phenyl rings and "laterality" of chlorine atoms are important structural features determining specific binding behavior with proteins and certain toxic responses in biological systems. We compare qualitative structure-activity relationships for PCBs with the limited information on the related non-coplanar chlorinated diphenyl ethers, providing further insights into the nature of the molecular recognition processes and support for the structural relationship of PCBs to thyroid hormones. Steroidlike activity requires conformational restriction and possibly hydroxylation. We offer some simple molecular recognition models to account for the importance of these different structural features in the structure-activity relationships that permit one to express PCB reactivities in terms of dioxin, thyroxine, and estradiol equivalents. The available data support the involvement of PCBs as mimics of thyroid and other steroidal hormones. The potential for reproductive and developmental toxicity associated with human exposure to PCBs is of particular concern. **Key words:** dioxin/thyroxine/estradiol equivalents, PCBs, structure-activity relationships. *Environ Health Perspect* 102:290-297(1994)

There is growing concern (1) that a large number of man-made chemicals that have been released into the environment can disrupt the endocrine systems of animals and humans. Among the chemicals of concern are the persistent, bioaccumulative organohalogen compounds that include some pesticides and industrial chemicals such as polychlorinated aromatics, other synthetic products, and some metals. Possible mechanisms by which these chem-

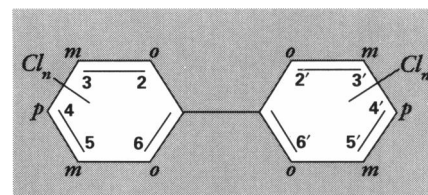
icals may interfere with endocrine function include their ability to mimic natural hormones (both as agonists and antagonists), including recognition of their specific binding sites, ability to react directly or indirectly with hormone structure to alter it, and ability to alter the pattern of hormone synthesis or modulate hormone receptor numbers and affinities. The polychlorinated biphenyls (PCBs) are known or suspected to express biological activities through most, if not all, of these mechanisms.

PCBs are mixtures of chlorinated aromatic chemicals manufactured by chlorination of biphenyl under suitable conditions for varying the percentage by weight of chlorine incorporated. The chemical structure (Fig. 1) of PCBs can be represented in a general way, where  $n, n'$  is the number of chlorine atoms replacing hydrogen atoms in each ring. A total of 209 different congeners are theoretically possible, but less than half that number appear to account for nearly all of the environmental contamination attributable to PCBs. If potential toxicity, environmental prevalence, and relative abundance in animal tissues are considered as criteria, the number of environmentally threatening congeners decreases to fewer than 50. Furthermore, fewer than 25 of these may account for most of the total PCB burden in tissues of invertebrates, fish, birds, and mammals (2).

PCBs were discovered before the turn of the century, and their usefulness in industry as, for example, dielectric and heat-exchange fluids with desirable physical properties, was recognized early. Their chemical resistance and neutral organic chemical nature have led to widespread distribution in the environment, including food webs. Now PCBs are almost entirely restricted to use in closed systems. Human exposure to PCBs has resulted largely from the consumption of food, but occupational exposure also results from inhalation and skin absorption. PCBs accumulate in the fatty tissues of humans and other animals. In turn, PCBs have caused toxic effects in humans and animals. The skin and liver are major sites of pathologic change, and other targets include the gastrointestinal tract, the immune system, and the nervous

system. Studies in animals suggest that some PCB congeners may be carcinogenic and that they can promote the carcinogenicity of other chemicals. Such toxicological properties, both qualitatively and quantitatively, often are congener specific with remarkable dependence on number and position of chlorine substitutions. Structural patterns can thus be used to discriminate among PCB congeners on the basis of toxic potential, if not entirely on toxicity per se (3,4).

The toxic effects of PCBs appear to involve at least three types of basic mechanisms of action: 1) reversible interaction (binding) of the PCB with specific molecular sites of action such as receptors, enzymes, etc., 2) irreversible covalent interaction (binding) between the PCB (probably a reactive metabolite) and target molecules (particularly macromolecules such as DNA and proteins), and 3) accumulation of highly lipid-soluble, metabolically stable PCBs in lipid-rich tissues or tissue compartments. The same or different structural properties may be involved in the expression of these toxic effects. Of particular interest to this discussion is the use of



**Figure 1.** General structure for PCBs. *o, m, p* denote ortho, meta, and para positions, respectively.

Address correspondence to J.D. McKinney, Pharmacokinetics Branch (MD-74), Environmental Toxicology Division, Health Effects Research Laboratory, U.S. EPA, Research Triangle Park, NC 27711 USA. C.L. Waller is currently at the Health Effects Research Laboratory, U.S. EPA, Research Triangle Park, NC.

We thank our many co-workers for their contributions over the years. The use of computer graphics facilities at the Center for Molecular Design, Washington University, St. Louis, and similar facilities at the University of California at San Francisco and San Diego for molecular graphics and modeling studies is also acknowledged. C.L.W. acknowledges the support of NIH training grant T32HL07275. This manuscript has been reviewed in accordance with the policy of the Health Effects Research Laboratory, U.S. EPA, and approved for publication. Approval does not signify that the contents necessarily reflect the views and policies of the agency, nor does mention of tradenames or commercial products constitute endorsement or recommendation for use.

Received 12 November 1993; accepted 3 January 1994.

structure–activity relationship (SAR) approaches to study the reversible interaction of PCB congeners with specific molecular sites in biological systems that normally control or mediate hormonal activities in the body. In the study of SARs, the basic philosophy is to draw conclusions by analogy, assuming that similarity of PCBs with respect to certain chemical properties/reactivities will result in similar biological responses (5). Thus, the problem is one of determining these properties and how they are linked to specific biological activities of interest.

### PCB Chemical Reactivities Underlying Toxicity

PCB chemical reactivities are encoded within the chemical structure of the PCB molecule. Because chemical reactivities underlie biological activities (6), it is important to understand how structure determines the important reactivities and, in turn, how these reactivities are linked to toxicity. One of the more important applications of SAR approaches is to help guide mechanistic research and permit testing of mechanistic hypotheses (7). Simplifying and classifying relevant chemical reactivities can be an important first approach to studying molecular mechanisms of toxicity.

### Stacking Interactions and Coplanarity of Structure

The importance of coplanarity of PCB structure (Fig. 2) in producing dioxinlike toxic effects has been recognized for some time (8). The chemical reactivity (9) associated with this structural feature is the ability to undergo stacking-type molecular interactions with other planar aromatic ring systems such as the heme system in hemoproteins. Such stacking interactions between aromatic ring systems have also been referred to as a charge-transfer or donor-acceptor interaction and can be viewed in a simplistic sense as a “Velcro-type” interaction in which the planar rings facilitate the “sticking together” of the two pieces. It should be pointed out here that the term “coplanar” PCB as widely used in this context is a misnomer in the sense that all PCBs are basically non-coplanar in nature (8). The important distinguishing property of the non-*ortho*-substituted PCBs is that the increase in the conformational enthalpy ( $\Delta H$ ) required for the non-*ortho*-substituted biphenyl to achieve a coplanar state is sufficiently offset (outweighed) by the favorable change in binding free energy ( $\Delta G$ ) associated with formation of a stacking biphenyl-receptor complex. All *ortho*-substituted PCBs are likely to undergo binding interactions in which a coplanar state is never fully achieved.

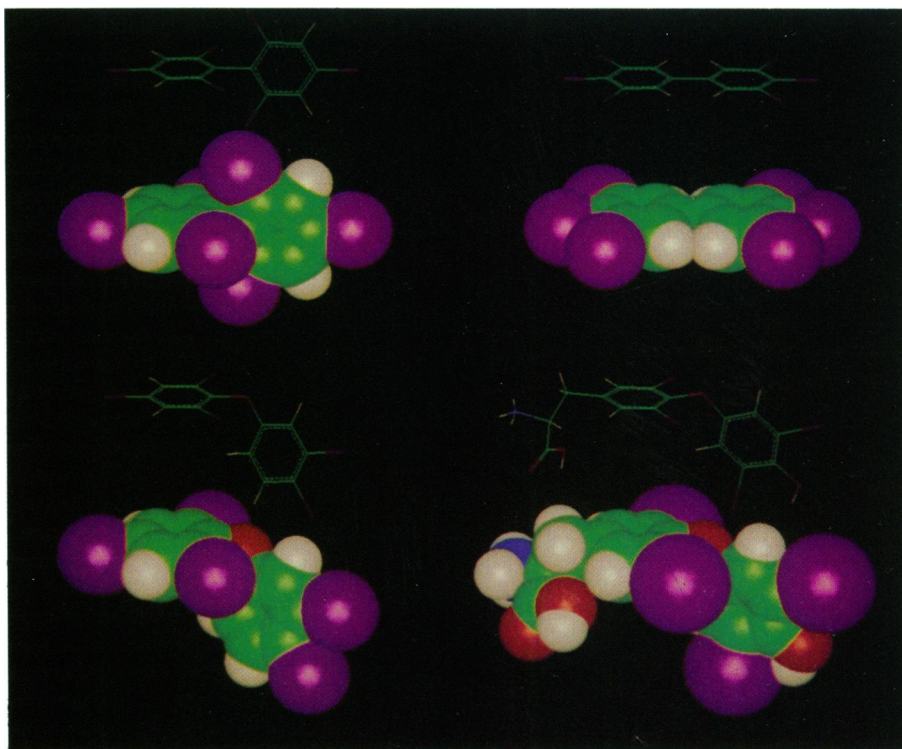
### Cleft-type Interactions and Lateral Chlorine Substitution

Lateral (*meta*, *para* substitutions) chlorination (Fig. 2) has also been recognized (10–12) as an important structural feature in PCB binding interactions and toxicity. Halogen atoms like chlorine contain many electrons and are thus highly polarizable. Many appropriately placed polarizability interactions in a protein-binding site can be quite effective in binding a PCB molecule. The most polarizable chlorines are those contained in the lateral positions of the PCB molecule. This can be envisioned as a molecular cleft-type interaction between the highly polarizable lateral halogen atoms and the hydrophobic interior of the cleft provided by amino acid side chains that converge on the halogen substituents. Similar binding interactions could involve chlorines in nonlateral (*ortho* substitutions) positions of the molecule, but there is not clear evidence that such interactions are important in PCB binding and toxicity (3). An additional factor associated with the polarizable chlorines is when the number and positions of chlorines are favorable to driving a preferred polarizability vector in the molecule (8). This is the case in the coplanar PCBs with lateral substitutions, where the preferred orientation is about the longest axis (through the *para*, *para'* positions) of the

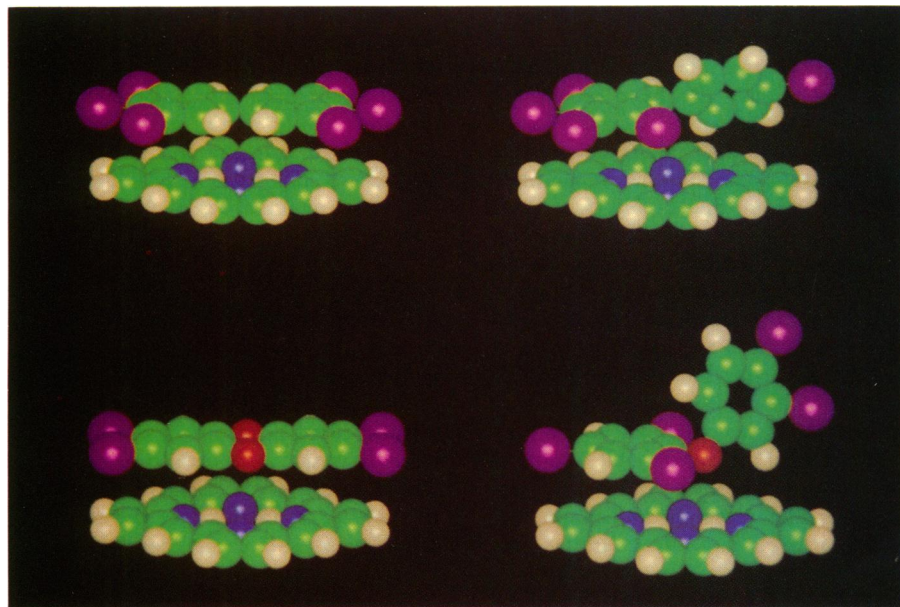
molecule. This type of preferred polarizability can facilitate the stacking interactions referred to earlier by strengthening the charge-transfer interaction or in simple, nonscientific terms, making it “stickier” (7). Both stacking and polarizability interactions might also be possible for certain stable metabolites of PCBs such as their hydroxylated metabolites.

### Areas of High Electron Density and Oxidative Metabolism

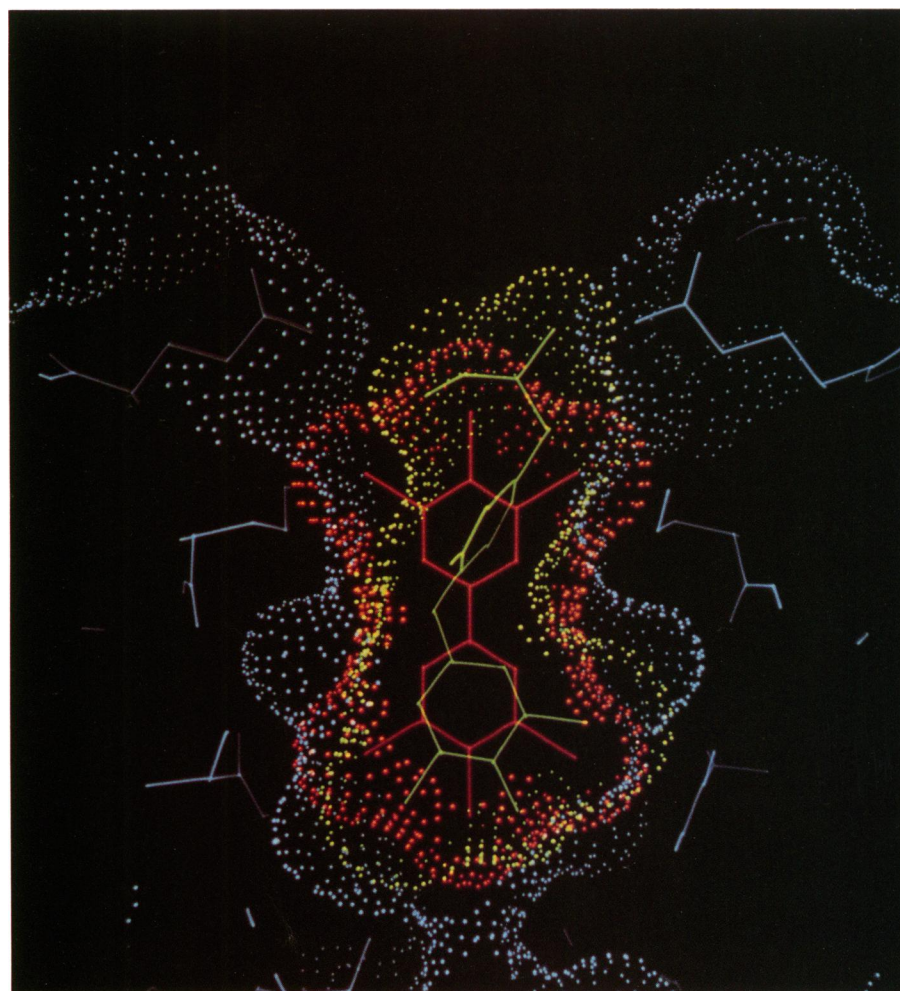
A third type of structural feature that may be important in leading to a different type of PCB toxicity is the availability of vicinal unsubstituted positions in the molecule (such as vacant *meta*, *para* positions) that can provide sites for oxidative metabolism to occur (13). In this case the chemical reactivity in the molecule that may control the rate and regiospecificity of oxidative attack involves areas of high electron density such as may occur at opposite ends of the polarization vector described above. Because reactive intermediary metabolites can be formed in such a process, it is possible that reactive metabolites can lead to covalently bound residues with biomolecules. Stable metabolites can have chemical reactivity of their own and lead to similar or different kinds of binding and toxicity as seen with the parent PCBs (14). Other types of metabolites and metabolic process-



**Figure 2.** Wire- and space-filled models comparing coplanar (3,3',4,4',5,5'-HCB, upper right) and non-coplanar (2,2',4,4',6,6'-HCB, upper left) PCBs with a chlorinated diphenyl ether (2,3',4,4',6-CDE, lower left) and thyroxine (lower right; aligned by planar face of one or more rings; note sterically inaccessible planar stacking face in 2,2',4,4',6,6'-HCB and similar condition of diphenyl ether system but with an accessible planar face). Carbon, green; hydrogen, white; oxygen, red; halogen, magenta.



**Figure 3.** Space-filled stacking models comparing coplanar (3,3',4,4',5,5'-, upper left) and non-coplanar (2,3,4,4',5,5'-, upper right) PCBs with dioxin (lower left) and chlorinated diphenyl ether (2,3',4,4',6,6'-, lower right) interactions with porphine ring system (note separation distance and planar geometrical extent of stacking interaction involving planar faces).



**Figure 4.** Molecular surface representation of 3,3',4,4',5,5'-hexachlorobiphenyl (from docking, red) and thyroxine complex (from X-ray, green) with binding domain (blue) of human protein prealbumin [compare contacts with protein surface and importance of lateral chlorines in filling binding pockets of thyroxine; see Rickenbacher et al. (17)].

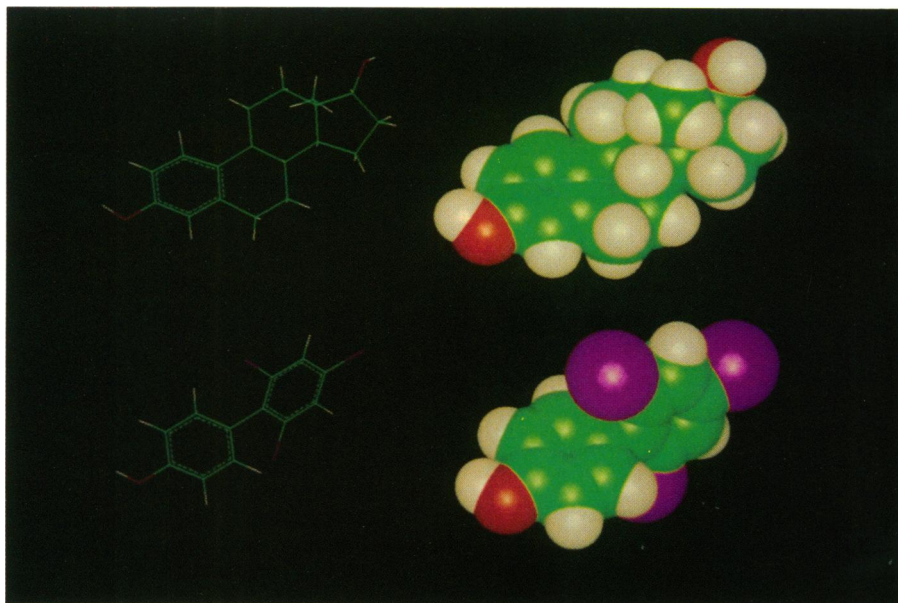
es are possible and may be important in the expression of certain forms of toxicity such as the reported association of chronic respiratory effects with methylsulfone metabolites of certain PCB congeners (3).

Although other structural features and reactivities of PCBs are possible, they have been less studied in an SAR framework. For example, certain pure phenobarbital (PB)-type inducing PCBs with *ortho* substitutions cause hepatomegaly and exhibit activity as promoters in short-term bioassays for carcinogenesis (3). The structural basis for these activities is not clear, but they may be more a function of specific substructural features than overall PCB structure consistent with the many diverse structures known to act as PB-type inducers.

## Reactivity Models and Predictions

### Stacking Models

In attempting to understand the relationship between PCB structure, chemical reactivity, and biological activity, theoretical models for relevant biological activities based on PCB molecular parameters have been sought (7). A planar (or energetically favorable coplanar) aromatic ring system is the primary structural feature that is found in all known high-affinity ligands for the Ah (or dioxin) receptor, which include both halogenated and nonhalogenated aromatic compounds. Several lines of evidence (7,15–17) now suggest that an important molecular property determining receptor binding and associated responses for these compounds is electron affinity. The dependence on this property can best be explained in terms of compound electron acceptance in a charge-transfer or stacking-type receptor model. Such a model also permits one to readily visualize steric hindrance to binding, as is the case for the *ortho*-substituted PCBs (Fig. 3), and variations in molecular size (e.g., compare PCBs with halogenated naphthalenes). The somewhat flexible but reasonably coplanar dioxin molecule shows the closest stacking followed by the non-*ortho*- and *ortho*-substituted PCBs. In general, the stacking model predicts increased binding for the class of chlorinated dioxins with increased chlorination, but this trend has not been observed in *in vitro* experimental binding studies, possibly reflecting solubility differences. There is evidence (18) for such a trend in *in vivo* absorption and tissue distribution studies in the rat. It is important to realize that a favorable, close stacking can occur in some aromatic compounds that are not totally coplanar but have a sterically accessible planar face, as in certain chlorinated diphenyl ethers (Fig. 2 and 3). This consideration bears directly on the structural relationship of PCBs to



**Figure 5.** Wire- and space-filled models comparing estrogenic PCB analog (4-OH, 2',4',6'-, bottom) with estradiol (top) (molecules aligned by common phenolic ring).

thyroid hormones, as discussed below. Other factors can play a role, such as compound solubilities and lipophilicities. Nevertheless, the receptor binding and associated biological responses are best explained in terms of changes in stereoelectronic and thermodynamic properties of ligands. As applied to PCBs, *ortho* substitution plays a major role in effecting changes in ligand binding (19,20). The relative binding affinities to the Ah receptor can be viewed as a measure of the "stacking reactivity," which can be related directly to dioxin (TCDD) and expressed as dioxin equivalents (3). This modeling work has implications in understanding the natural role and function of the Ah receptor.

### Cleft-type Models

Another important structural property in determining PCB dioxin-like toxicity is the presence of lateral chlorine substitution in the molecular structure. However, this is not always a requirement for high-affinity binding to the Ah receptor such as found for certain polynuclear aromatic hydrocarbons (PAHs). As discussed above, this structural requirement for high toxicity suggests the involvement of other binding sites with C-shaped molecular cavities or clefts with viselike properties in the molecular recognition process. A binding model of this type that has been extensively studied is the prealbumin or transthyretin interaction model. The importance of lateral substitutions in binding both thyroid hormone analogues and PCBs has been well documented (10,11) (Fig. 4). Interestingly, this binding interaction model provides another structural linkage between PCBs and thyroid hormones. In fact, there is a family of preal-

buminlike proteins apparently involved in regulating thyroid hormone action (21). As for stacking reactivity and Ah receptor binding, relative binding affinities to prealbumin can be viewed as a measure of the "lateral chlorine polarizability reactivity," which can be related directly to thyroxine and expressed as thyroxine equivalents. PCB interactions with this family of proteins have potential biological consequences as both agonists and antagonists.

### Estrogen-Active Analog Model

While the stacking and cleft-type models have both pointed to a structural relationship of PCBs to thyroid hormones, other work (22) has suggested a structural relationship to steroid hormones, particularly estrogens. The steroidal estrogens are relatively rigid, polycyclic molecules with pronounced asymmetry, and their binding to the estrogen receptor generally reflects a high degree of stereospecificity. Several classes of nonsteroidal synthetic estrogens have been based on phenyl-substituted hydrocarbons (23) because of the structural relationship to the phenolic A ring in estradiol. Hydroxylated PCBs are often found as metabolites of the parent PCBs (13). In this regard, we theorized that certain hydroxylated PCBs (Fig. 5) that were also conformationally restricted due to *ortho* substitution might be effective in binding the estrogen receptor. This proved to be the case in that the most active PCB analogues were those that combined a *para*-hydroxyphenyl ring with some degree of *ortho* substitution. However, it is not clear if there is some unique property of the hydroxyl group (such as hydrogen bonding) or if any other similar-sized

group (such as chlorine) could be substituted. It is possible that increased *ortho* chlorines in the PCB structure might also provide important hydrophobic bulk structure in binding the estrogen receptor, but there was not any direct evidence for this. In additional analysis of these data (unpublished results), the solubility/lipophilic properties of the PCB are also proving to be important.

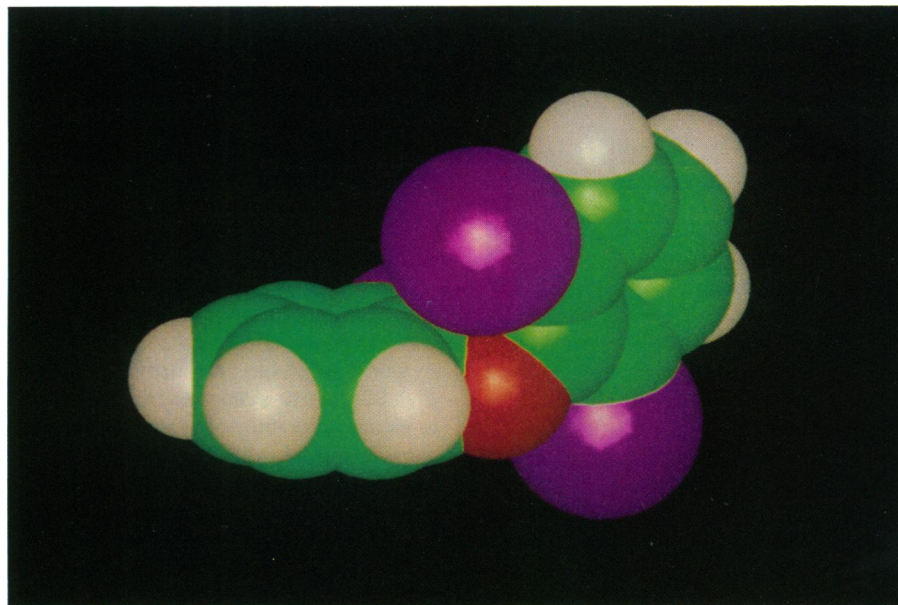
Although *ortho* substitution in PCBs can significantly lower the dioxinlike toxicity, it can potentially have the opposite effect on any estrogen-receptor-mediated toxicity. This PCB active analog modeling approach to estrogen receptor binding has implications for understanding any associated toxicity. As was suggested for other PCB reactivities and dioxin and thyroxine equivalents, it is possible to express this PCB reactivity in terms of estradiol equivalents.

## Possible Relationships to Toxic Endpoints

### Relationship to Chlorinated Diphenyl Ethers

As mentioned earlier, the chlorinated diphenyl ethers (CDEs) are an important test case for the structural requirements for dioxin like toxicity for PCBs. CDEs contain two phenyl rings as do PCBs but for energetic reasons are unlikely to achieve coplanar states in their binding interactions, even in the absence of *ortho* substitution. Any dioxinlike toxicity and associated Ah receptor binding are more likely to reflect molecular interactions involving non-coplanar states, especially with *ortho*-chlorine substituents. The main structural difference in the two classes of compounds is the ether oxygen bridge (C-O-C angle of 120°) contained in the CDEs. In addition to serving as structurally related model compounds for PCBs, the CDEs are also of interest as potential environmental contaminants, such as their association with used transformer fluid (24). For both PCBs and CDEs, chlorine substitutions provide steric constraint that defines the minimum energy conformation of the aromatic rings.

However, the effects on overall conformational properties and structure can be quite different in the context of having an accessible planar face for a stacking-type interaction to occur as postulated for the dioxin-receptor-binding interaction. In the case of PCBs, any degree of *ortho* substitution substantially raises the energy barrier to internal rotation about the pivot bond (25), and it is likely that *ortho*-PCBs interact with the dioxin receptor in their non-planar minimum energy conformations (26). Therefore, to the extent that toxicity is truly mediated by coplanar conformers, *ortho*-PCBs are likely to be relatively non-



**Figure 6.** Space-filled model showing the effects of *ortho*-chlorine substitution in a tri-*ortho*-substituted chlorinated diphenyl ether. Note the relationship (in plane or in  $\pi$ -cloud) of chlorines to stacking ring (as in Figure 3).

toxic. Lethality in guinea pigs may be an example (10) of this because the nearly isosteric 3,3',4,4',5,5'-hexachlorobiphenyl is about 1/100 as toxic as dioxin (reflecting the energy cost to achieve a coplanar state) and the structurally related 2,3,3',4,4',5,5'-heptachlorobiphenyl is relatively nontoxic at significantly larger (>10-fold) doses.

In the case of CDEs, the effects of *ortho* substitution on dioxin-receptor-binding are likely to have an enhancing effect. *Ortho* substituents provide desirable minimum-energy conformations (Fig. 3) because the rings now approach a mutually perpendicular relationship with an accessible planar face for entering into stacking interactions. An additional enhancing effect is realized because there is less energy cost (more favorable binding free energy from freezing out rotatable bonds) required to position the more rigid binding groups. A similar effect of *ortho* substituents in hydroxylated PCBs in enhancing estrogen receptor binding was previously described (22). Although other binding modes are possible, they are somewhat less favored energetically, but in general also seriously limit the accessibility of the other phenyl ring system in entering into other binding interactions such as those involving lateral substituents.

Recent work (27,28) has shown interesting differences in the structure-activity relationships for CDEs as compared to PCBs. The evidence suggests that both classes of compounds can produce dioxin-like toxic effects mediated by binding to the dioxin receptor. For example, non-*ortho*-but laterally substituted CDEs were less toxic than their mono-*ortho*-substitut-

ed analogues, and similar results were also observed for their enzyme-inducing potencies. Active congeners were also shown to include CDEs with multiple *ortho*-substitutions. These results are consistent with the anticipated enhancing effects of *ortho* substitution in CDEs on dioxin receptor binding through a stacking molecular mechanism as previously predicted (7).

In addition, an interesting result was found (28) in certain highly chlorinated CDEs that contain multiple *ortho* substitutions, especially in both rings. The potencies were shown to be similar to those found (27) for the non-*ortho*- and mono-*ortho*-substituted compounds previously studied. It is apparent that high degrees of *ortho* substitution in CDEs do not significantly diminish their Ah-receptor-mediated responses as seen for the PCBs. It is proposed that in these cases increased conformational restriction is brought about by additional steric interactions between the ring systems. The favored conformers would place the adjacent *ortho*-chlorine in the plane of the stacking ring (see Figure 3), which could influence the nature of the stacking interaction. Highly chlorinated diphenyl ethers, especially with high degrees (three or four *ortho*-chlorines) of *ortho* substitution, would increase the polarizability and steric rigidity of the system, both expected to further enhance a stacking interaction. It is anticipated that the stacking ring would be the most highly substituted ring, although this is not always clear (e.g., when both rings contain the same number of chlorines). However, high degrees of *ortho* substitution would not only lead to chlorine atoms positioned

in the stacking plane but also to close through-space interactions of *ortho*-chlorine atoms with the face of the adjacent aromatic ring system (Fig. 6). This latter effect can be relieved somewhat in the absence of *meta*-chlorine substituents that buttress the adjacent *ortho* substituent (29). Both the in-plane and through-space intramolecular effects would weaken any intermolecular stacking interaction. Nevertheless, it is anticipated that the structure-activity relationships for these types of molecules would be complex and difficult to interpret. *Ortho* substituents in CDEs can have both qualitatively and quantitatively different effects on molecular structure and reactivity as compared to the effects seen in the PCBs.

The available SAR data on CDEs support the involvement of a conformationally restricted diphenyl system with an accessible planar face that can participate in stacking interactions with protein binding sites. This type of analysis of CDEs and model for active compounds further support the proposal (20) for a stacking type interaction in dioxin receptor binding and should be helpful in guiding the selection of other congeners to test for the ability to produce dioxinlike toxicity. The potential toxicity of members of this class of chlorinated aromatic hydrocarbons needs further attention because hexa- to decachlorodiphenyl ethers, all of which are likely to have high degrees of *ortho* substitution (wider range of active congeners possible), have been identified (30) in human adipose samples at concentrations as high as 2000 pg/g, and they are known contaminants of commercial products such as technical-grade pentachlorophenol (31). The close structural relationship of active CDEs to thyroid hormones is also of considerable interest and compatible with the possible thyromimetic properties (7) of these compounds. Clearly, in the context of this stacking model for binding, the non-*ortho*-substituted congeners do not take on special significance and, other things being equal, may have reduced activity relative to their *ortho*-substituted counterparts. Preliminary quantitative structure-activity relationship (QSAR) studies (Waller C, and McKinney J, unpublished observations) with the CDEs based on the stacking-model concept look promising in understanding the dioxin-receptor-mediated responses of these chemicals. In a recent teratogenicity study (32) with nine CDEs in mice, 2,2',4,5,6'-penta- and 2,3',4',6'-tetrachlorodiphenyl ether resulted in the loss of litters at relatively low doses. Both of these CDEs contain one di-*ortho*-substituted ring, and the other has lateral (*meta*, *para*) substituents, but they are not fully *ortho* substituted to maximize the steric

interactions shown in Figure 6. These results appear to be consistent with the structural requirements of active congeners predicted by our models.

### Relationship to Thyroid Hormones

What emerges from such analysis of SAR results is that these structurally related compounds (PCBs and CDEs) can participate in similar molecular recognition processes in their potential interactions with specific molecular sites in biological systems in which the nature, accessibility, and polarizability of planar (or coplanar) face interactions can vary. The obvious next step in this type of analysis is to compare these molecular recognition processes with those that have been well documented (33,34) for thyroid hormone analogues as natural halogenated diphenyl ether systems of considerable biological importance. The SAR literature for thyroid-hormone-binding proteins points to two basic types of binding proteins: those that bind 3,5,3'-triiodo-L-thyronine ( $T_3$ ) in preference to L-thyroxine ( $T_4$ ) and those that bind  $T_4$  in preference to  $T_3$ . Interestingly, the SARs for the  $T_3$  binding proteins seem to depend more on the structural properties of the inner (tyrosyl) ring, whereas the SARs for the  $T_4$  binding proteins depend more on the outer (phenolic) ring properties (Fig. 2). In this regard, it is interesting to note that the nuclear receptors for thyroid hormones show considerable dependence of binding on a sterically constrained inner ring brought about by the 3,5- (*ortho*) substituents. We have further shown (29) that these results can be reasonably explained using a donor-acceptor or stacking aromatic ring model as previously described. In this model framework it is not surprising that biphenyl thyroid hormone analogues (which lack an ether oxygen bridge) are inactive (33) because they basically represent worse-case *ortho*-substituted PCBs. Binding ligands for the triiodothyronine ( $T_3$ ) nuclear receptor and the dioxin receptor may share common molecular recognition factors in the expression of their binding activities.

In contrast, the lateral arrangement of the *ortho*-diiodophenolic structure in the outer ring of thyroid hormones is an important feature characterizing the binding of all three major transport proteins (globulin, prealbumin, and albumin), and is the sole major binding feature for albumin. In fact, the  $T_4$  binding proteins appeared to be ideally suited to bind lateral substituted chlorinated aromatic compounds such as the lateral (*meta*, *para*) substituted PCBs. Molecular modeling and experimental studies (10,11) now provide considerable support for this hypothesis. The best binders (four- to eightfold better

than  $T_4$ ) have only lateral chlorine substituents, and additional *ortho*-chlorines do not lower binding much (two- to threefold better than  $T_4$ ). One lateral chlorine (as in the 2,4,6-pattern) is sufficient for appreciable (about half  $T_4$ ) binding activity. When no chlorines (biphenyl) or all (2,2',6,6') *ortho* positions are filled, the compounds show no binding activity. As persistence in the environment and in biological tissues usually equates to a high degree of lateral substitution, it is anticipated that most, if not all, PCB residues found in human tissues are likely to effectively bind this family of proteins. It is also known (10,11,35) that hydroxy PCBs can bind this family of proteins, which suggests that certain PCB hydroxylated metabolites could produce the same or similar responses associated with binding. Similar SAR results (11) were shown for a  $T_4$  nuclear-binding protein (12) and the microsomal deiodinase (36) in liver tissue. Other proteins that may give similar results include tyrosine hydroxylase and the dopamine receptor.

An important consideration here is that *ortho* chlorination in PCBs does not significantly affect this binding activity. One indirect effect of *ortho* substitution on this binding activity may be to effectively inhibit binding to the  $T_3$  family of binding proteins that presumably require a stacking interaction. Therefore, any biological responses associated with this "laterality" type of binding activity (as in the prealbumin interaction model) could be elicited by a much larger number of PCB congeners of environmental concern because most are *ortho* substituted to some degree. The recent finding of PCB neurotoxicity (37) with certain *ortho* but lightly substituted congeners may fall into this category. Biochemical thyroid status at the cellular level has been shown (38) to depend not only on the concentration of free  $T_4$  but also on the concentration of prealbumin. Prealbumin is synthesized in the brain (as well as the liver), where it plays a major role in transport of thyroid hormone to the central nervous system (39).

### Relationship to Retinoids

Another factor that may bear on some of the toxicological properties of PCBs relates to evidence (40) suggesting a close association between thyroid hormone and retinoid transport, metabolism, and gene transcription. For example, prealbumin binds to retinol-binding protein, and this serves as both a transport and protective mechanism for the smaller protein (41). Earlier findings (42) of prealbumin depletion in rat sera of dioxin-treated animals coupled with findings (McKinney JD et al. unpublished observations) of a parallel increase in retinol-binding protein and

retinol are consistent with this view and further support a role for thyroid-hormone-binding proteins in the mechanism of toxicity of these compounds. SAR studies (43) on the effects of PCBs on retinoid concentrations in serum and tissues indicate that there are at least two classes of PCBs (i.e., the coplanar PCBs that affect hepatic, renal, and serum retinoids and other *ortho*-substituted PCBs that only decrease serum retinol). It is not clear whether these effects are secondary to the effects of PCBs on thyroid hormone metabolism. These effects may also reflect differences in preferences for the two classes of thyroid-hormone-binding proteins described earlier.

### Relationship to Estradiol

Another class of hydroxylated PCBs (presumed metabolites) have been shown (22) to bind specifically to the mouse uterine estrogen receptor and as such represent potential agonists/antagonists ligands for certain estrogen receptor functions. As described earlier, this is believed to be associated with a close match of the phenolic ring with the A-ring in estradiol and enhanced steroidlike structure brought about by the *ortho* substituents. Developmental and reproductive toxicity are possible outcomes of interactions of this type with steroid hormone receptors. For example, reproductive impairment in marine mammals has been directly attributed to feeding on PCB-contaminated fish (44). The potential human health effects of exposure to *ortho*-substituted PCBs and their hydroxylated metabolites should be considered in this light.

### Conclusions

From a structural chemistry point of view, there are at least two distinct classes of PCBs: the non-*ortho*- and *ortho*-substituted congeners. The important distinguishing feature of the non-*ortho*-substituted PCBs is the energetic accessibility of the coplanar state that can facilitate stacking interactions such as may occur in a receptor-binding domain. The effects of *ortho* substitution on PCB reactivity are less clear, but they can include three basic types: hindrance to stacking interactions, conformational restriction and more rigid steroidlike structure, and increased carbon-chlorine bond polarizability in nonlateral positions. Lateral chlorine substituents can be important in enhancing the binding properties of both classes of PCBs through polarizability interactions.

For the non-*ortho*-substituted (coplanar) congeners, most, if not all, of the important toxic effects we know about appear to be mediated by binding to the dioxin or Ah receptor. A simple stacking

interaction model involving planar faces is offered as a way to visualize the Ah-receptor-binding process, but it must be viewed as an oversimplification of a complex binding interaction. Further support for the stacking model is derived from analysis of the important structural features of the more active dioxinlike chlorinated diphenyl ethers. The activity of the chlorinated diphenyl ethers also provides support for the thyromimetic properties of these classes of compounds. The prealbumin interaction model is offered as a way to visualize the requirement for lateral substituents in cleft-type binding interactions. The estrogen active analog modeling approach offers one way of understanding the importance of *ortho* substituents and conformational restriction in receptor binding. For the *ortho*-substituted congeners, the link to specific toxic endpoints are not well established, and it is likely that multiple mechanisms are involved. To some extent, quantitative differences in responses appear to be due to variations in the degree of coplanarity or conformational restriction (both dependent on degrees of *ortho* substitution) or accessibility of planar faces and laterality that exists among the PCB congeners. Agonist and antagonist effects may also depend on variations and combinations of the important reactivity properties (kinds and amount) among the congeners. There appear to be some areas of overlap between the two PCB classes such as the weak dioxinlike properties of certain mono-*ortho*-substituted PCBs, but the significance of this is not fully understood. As *ortho* substitution in biphenyls is expected to lower their dioxinlike binding properties in biological systems, one might anticipate differences in their toxicokinetic properties based on the degree of *ortho* substitution (45).

As suggested earlier, PCBs may interfere with endocrine function in various ways. Their ability to directly mimic natural hormones (both as potentially potent and persistent agonists and antagonists) including recognition of their specific binding sites is emerging as a guiding concept in the case of the thyroid and estrogenic steroid hormones. Indirect effects might include alterations in the receptor number or affinity of other hormones involved in multihormonal regulating systems. For example, there is evidence (46) for a direct effect of thyroid hormone on the hepatic synthesis of estrogen receptors. The antiestrogenic action of dioxin (47) and possibly coplanar PCBs (48) in down-regulating estrogen receptors could be related to their potential thyromimetic properties. Toxicity could result from over- or underexpression of normal hormonal responses that may be mediated by both

receptor and nonreceptor protein and associated DNA interactions. For example, recent work (49) suggests that the human estrogen receptor structural gene contains a DNA sequence that binds activated mouse and human Ah receptor.

The potential for reproductive and developmental toxicity (50) is of particular concern. Thyroid hormones are known to be essential for brain development (51), and many investigators (52) have provided evidence demonstrating effects of hormones on immature brain cell maturation, myelinogenesis, protein and nucleic acid metabolism, and electrical activity of the growing brain. In addition, there are a number of well-documented examples of the regulation of single and multiple clusters of genes through multiple hormonal interactions (53). Thyroidal, adrenal, and gonadal hormones are all necessary for the normal differentiation of the nervous system and involved in the ultimate expression of the target genes in several of these multihormonal regulating systems. Because humans are exposed to mixtures of PCB congeners, it is likely that there are complex interactive biological effects. Understanding the biological properties of PCBs and how they are determined by the molecular structure are important research objectives. Perhaps more direct molecular biological approaches would be helpful in elucidating the involvement of both receptor and nonreceptor protein-mediated responses. In turn, such mechanistic insight should be helpful in reducing the scientific uncertainty associated with health hazard/risk assessments associated with human exposures to PCBs and related compounds.

## REFERENCES

- Hileman B. Concerns broaden over chlorine and chlorinated hydrocarbons. *Chem Eng News* 71: 11-20(1993).
- McFarland VA, Clarke JU. Environmental occurrence, abundance, and potential toxicity of polychlorinated biphenyl congeners: considerations for congener specific analysis. *Environ Health Perspect* 81:225-239(1989).
- WHO. Environmental health criteria document for polychlorinated biphenyl and terphenyls. EHC 140. Geneva:International Programme for Chemical Safety, World Health Organization, 1993.
- Safe S. Polychlorinated biphenyls (PCBs): environmental impact, biochemical and toxic responses, and implications for risk assessment. *Crit Rev Toxicol* 24:1-63(1994).
- Weinstein H, Rabinowitz J, Liebman MN, Osman R. Determinants of molecular reactivity as criteria for predicting toxicity: problems and approaches. *Environ Health Perspect* 61:147-162(1985).
- McKinney JD, Fawkes J, Jordan S, Chae K, Oatley S, Coleman RE, Briner W. 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin (TCDD) as a potent and persistent thyroxine agonist: a mechanistic model for toxicity bases on molecular reactivity. *Environ Health Perspect* 61:41-53(1985).
- McKinney JD. Multifunctional receptor model for dioxin and related compound toxic action: possible thyroid hormone-responsive effector-linked site. *Environ Health Perspect* 82: 323-336(1989).
- McKinney JD, Singh P. Structure-activity relationships in halogenated biphenyls: unifying hypothesis for structural specificity. *Chem-Biol Interact* 33:271-283(1981).
- Ishida T, Kamiuchi K, Kuwahara A, Doi M, Inoue M. Stacking and hydrogen bonding interactions between phenylalanine and guanine nucleotide: crystal structure of L-phenylalanine-7-methylguanosine-5'-monophosphate complex. *Biochem Biophys Res Commun* 136:294-299(1986).
- McKinney JD, Chae K, McConnell EE, Birnbaum L. Structure-induction versus structure-toxicity relationships for polychlorinated biphenyls and related aromatic hydrocarbons. *Environ Health Perspect* 60:57-68(1985).
- Rickenbacher U, McKinney JD, Oatley SJ, Blake CCF. Structurally specific binding of halogenated biphenyls to thyroxine transport protein. *J Med Chem* 29:641-648(1986).
- McKinney JD, Fannin R, Jordan S, Chae K, Rickenbacher U, Darden T, Pedersen L. Polychlorinated biphenyls (PCBs) and related compound interactions with specific binding sites for thyroxine in rat liver nuclear extracts. *J Med Chem* 30:79-86(1987).
- Kato S, McKinney JD, Matthews HB. Metabolism of hexachlorobiphenyls in the rat. *Toxicol Appl Pharmacol* 53:389-398(1980).
- Yamamoto HA, Yoshimura H. Metabolic studies on polychlorinated biphenyls. III. Complete structure and acute toxicity of the metabolites 2,4,3',4'-tetrachlorobiphenyl. *Chem Pharm Bull* 21:2237-2242(1973).
- Cheney BV, Tolly T. Electronic factors affecting the receptor binding of dibenzo-*p*-dioxins and dibenzofurans. *Int J Quant Chem* 16: 87-110(1979).
- Sjoberg P, Murray JS, Brinck T, Evans P, Politzer P. The use of electrostatic potential at the molecular surface in recognition interactions: dibenzo-*p*-dioxins and related systems. *J Mol Graphics* 8:81-90(1990).
- Kafafi SA, Afeefy HY, Said HK, Hakimi JM. A new structure-activity model for the Ah receptor binding. Polychlorinated dibenzo-*p*-dioxins and dibenzofurans. *Chem Res Toxicol* 5:856-862(1992).
- Abraham K, Wiesmuller T, Brunner H, Krowke R, Hagenmaier H, Neubert D. Absorption and tissue distribution of various polychlorinated dibenzo-*p*-dioxins and dibenzofurans (PCDDs and PCDFs) in the rat. *Arch Toxicol* 63:193-202(1989).
- McKinney JD, Long GA, Pedersen L. PCB and dioxin binding to cytosol receptors: a theoretical model based on molecular parameters. *Quant Struct Act Rel Pharmacol Chem Biol* 3:99-105(1984).
- McKinney JD, Darden T, Lyerly MA, Pedersen L. PCB and related compound binding to Ah receptor(s): theoretical model based on molecular parameters and molecular mechanics. *Quant Struct Act Rel in Pharmacol Chem and Biol* 4:166-172(1985).
- Blake CCF, Oatley S. Protein-DNA and protein-hormone interactions in prealbumin: a model of the thyroid hormone nuclear receptor. *Nature* 268:115-120(1977).
- Korach KS, Sarver P, Chae K, McLachlan JA,

- McKinney JD. Estrogen receptor-binding activity of polychlorinated hydroxybiphenyls: conformationally restricted structural probes. *Mol Pharmacol* 33:120–126(1988).
23. Landvatter SW, Katzenellenbogen JA. Stereochemical considerations in the binding of nonsteroidal estrogens to the estrogen receptor. *Mol Pharmacol* 20:43–51(1981).
  24. Albro PA, Parker CE. General approach to the fractionation and class determination of complex mixtures of chlorinated aromatic compounds. *J Chromatogr* 197:155–169(1980).
  25. McKinney JD, Pedersen L. Biological activity of polychlorinated biphenyls (PCBs) related to conformationally structure. *Biochem J* 240:621–622(1986).
  26. Waller C, McKinney JD. Comparative molecular field analysis of polyhalogenated dibenzop-dioxins, dibenzofurans, and biphenyls. *J Med Chem* 35:3660–3666(1992).
  27. Howie L, Dickerson R, Davis D, Safe S. Immunosuppressive and monooxygenase induction activities of polychlorinated diphenyl ether congeners in C57BL/6N mice: quantitative structure-activity relationships. *Toxicol Appl Pharmacol* 105:254–263(1990).
  28. Harper N, Howie L, Connor K, Arellano L, Craig A, Dickerson R, Safe S. Immunosuppressive and monooxygenase induction activities of highly chlorinated diphenyl ether congeners in C57BL/6 and DBA/2 mice. *Fundam Appl Toxicol* 20:496–502(1993).
  29. Chae K, McKinney JD. Molecular complexes of thyroid hormone tyrosyl rings with aromatic donors. Possible relationship to receptor protein interactions. *J Med Chem* 31:357–362(1988).
  30. Stanley JS, Cramer PH, Ayling RE, Thornburg KR, Remmers JC, Breen JJ, Schwemberger J. Determination of the prevalence of polychlorinated diphenyl ethers (PCDPEs) in human adipose tissue samples. *Chemosphere* 20:981–985(1990).
  31. Nilsson CA, Renberg L. Further studies on impurities in chlorophenols. *J Chromatogr* 89:325–333(1974).
  32. Francis BM. Presented at the Society for Environmental Toxicology and Chemistry, Cincinnati, Ohio, 1992.
  33. Jorgensen EC. Thyroid hormones and analogs. II. Structure-activity relationships. In: *Hormonal proteins and peptides-thyroid hormones*, vol 6 (Li CH, ed). New York:Academic Press, 1978;107–204.
  34. Cody, V. Thyroid hormones: crystal structure, molecular conformation, binding, and structure-function relationships. In: *Recent progress in hormone research*, vol 34 (Greep RO, ed). New York:Academic Press, 1978, 437–447.
  35. Brouwer A, Van den Berg KJ. Binding of a metabolite of 3,4,3',4'-tetrachlorobiphenyl to transthyretin reduces serum vitamin A transport by inhibiting the formation of protein complex carrying both retinol and thyroxine. *Toxicol Appl Pharmacol* 85:301–312(1986).
  36. Rickenbacher U, Jordan S, McKinney JD. Structurally specific interaction of halogenated dioxin and biphenyl derivatives with thyroxine-5'-deiodinase in rat tissue. In: *ACS symposium series no. 413. Bioactive mechanisms: proof, SAR, and prediction*. Washington, DC:American Chemical Society, 1989;354–369.
  37. Shain W, Bush B, Seegal R. Neurotoxicity of polychlorinated biphenyls: structure-activity relationship of individual congeners. *Toxicol Appl Pharmacol* 111:33–42(1991).
  38. Ramaker J, Wood WG. Transthyretin — an explanation of anomalous serum thyroid hormone values in severe illness. *J Clin Chem Clin Biochem* 28:155–161(1990).
  39. Dickson PW, Aldred AR, Marley PD, Guo-Fen T, Howlett GJ, Schreiber G. High prealbumin and transferrin mRNA levels in the choroid plexus of rat brain. *Biochem Biophys Res Commun* 127:890–895(1985).
  40. Yen PM, Sugawara Chin WW. Triiodothyronine ( $T_3$ ) differentially affects  $T_3$ -receptor/retinoic acid receptor and  $T_3$ -receptor/retinoid X receptor heterodimer binding to DNA. *J Biol Chem* 267:23248–23252(1992).
  41. Peterson PA. Studies on the interaction between prealbumin, retinol binding protein, and vitamin A. *J Biol Chem* 246:44–49(1971).
  42. Albro PW, Corbett JT, Harris M, Lawson LD. Effects of 2,3,7,8-tetrachlorodibenzo-p-dioxin on lipid profiles in tissue of the Fisher rat. *Chem-Biol Interact* 23:315–330(1978).
  43. Chen LC, Berberian I, Koch B, Mercier M, Azais-Braesco V, Glauert HP, Chow CK, Robertson LW. Polychlorinated and polybrominated biphenyl congeners and retinoid levels in rat tissues: structure-activity relationships. *Toxicol Appl Pharmacol* 114:47–55(1992).
  44. Reijnders PJH. Reproductive failure in common seals feeding on fish from polluted coastal waters. *Nature* 33:456–457(1986).
  45. Borlakoglu JT, Wilkins JPG. Metabolism of di-, tri-, tetra-, penta-, and hexachlorobiphenyls by hepatic microsomes isolated from control animals and animals treated with Aroclor 1254, a commercial mixture of polychlorinated biphenyls (PCBs). *Comp Biochem Physiol* 105C:95–106(1993).
  46. Freyschuss B, Eriksson H. Evidence for a direct effect of thyroid hormones on the hepatic synthesis of estrogen receptors in the rat. *Steroid Biochem* 31:247–249(1988).
  47. DeVito MJ, Thomas T, Martin E, Umbreit TH, Gallo MA. Antiestrogenic action of 2,3,7,8-tetrachlorodibenzo-p-dioxin: tissue specific regulation of estrogen receptor in CD1 mice. *Toxicol Appl Pharmacol* 113:284–292(1992).
  48. Jansen HT, Cooke PS, Porcelli J, Liu T-C, Hansen LG. Estrogenic and antiestrogenic actions of PCBs in the female rat: in vitro and in vivo studies. *Reprod Toxicol* 7:237–248(1993).
  49. White TEK, Gasiewicz TA. The human estrogen receptor structural gene contains a DNA sequence that binds activated mouse and human Ah receptors: a possible mechanism of estrogen receptor regulation by 2,3,7,8-tetrachlorodibenzo-p-dioxin. *Biochem Biophys Res Commun* 193:956–962(1993).
  50. Gray LE, Ostby J, Marshall R, Andrews J. Reproductive and thyroid effects of low-level polychlorinated biphenyl (Aroclor 1254) exposure. *Fundam Appl Toxicol* 20:288–294(1993).
  51. Ford DH, Crammer EB. Developing nervous system in relation to thyroid hormones. In: *Thyroid hormones and brain development* (Grave GD, ed). New York:Raven Press, 1977; 1–18.
  52. Eayrs JT. Endocrine influence on cerebral development. *Arch Biol* 75:529–565(1964).
  53. Roy AK. Role of thyroid hormone in the expression of  $\alpha_2$ -globulin and other multihormonally regulated genes. In: *Molecular basis of thyroid hormone action* (Oppenheimer JH, Samuels HH, eds). New York:Academic Press, 1983;214–243.

## European Organization for Research and Treatment of Cancer

## U.S. National Cancer Institute

**The European Organization for Research and Treatment of Cancer (EORTC) and the U.S. National Cancer Institute (NCI) are offering an exchange program to enable cancer researchers to work at NCI or EORTC-related institutions for one to three years.**

### General Conditions

Awardees will receive an annual subsistence allowance of \$30,000. Half of this amount will be provided by U.S. sources, the remainder by European sources.

European awardees will receive the U.S. contribution either from the NCI or from their extramural host institution. The European contribution of the exchangeship will be provided either by the

scientist's home institution or by a European grant-giving agency.

For American awardees, the host institution must be affiliated with the EORTC.

### Documentation

The following documents are required, in English, from all applicants:

- Completed application form.
- Description of the research to be undertaken, not to exceed three typewritten pages.
- Letter of invitation from the prospective host.
- Agreement to release the applicant from the home institution for the duration of the exchangeship.
- Assurance of intention to return to the home institution at the end of the exchangeship.

- Statement concerning the provision of 50 percent of financial support by European sources. Non-EORTC member country candidates must continue at full salary at the home institution for the duration of the exchangeship.

- Three letters of recommendation mailed directly to the NCI Liaison Office by the recommending individuals.

### For More Information Contact:

EORTC/NCI Exchange Program  
NCI Liaison Office  
83, Avenue E. Mounier  
1200 Brussels, Belgium  
Telephone: (32) (2) 772-22-17  
Telefax: (32) (2) 770-47-54

# EXCHANGE PROGRAM