Research Paper

Genetic variation for maize root architecture in response to drought stress at the seedling stage

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Although the root system is indispensable for absorption of nutrients and water, it is poorly studied in maize owing to the difficulties of direct measurement of roots. Here, 103 maize lines were used to compare root architectures under well-watered and water-stressed conditions. Significant genetic variation, with medium to high heritability and significant correlations, was observed for root traits. Total root length (TRL) and total root surface area (TSA) had high phenotypical diversity, and TRL was positively correlated with TSA, root volume, and root forks. The first two principal components explained 94.01% and 91.15% of total root variation in well-watered and water-stressed conditions, respectively. Thus, TRL and TSA, major contributors to root variation, can be used as favorable selection criteria at the seedling stage. We found that stiff stalk and non-stiff stalk groups (temperate backgrounds) showed relatively higher mean values for root morphological diversity than the TST group (tropical/subtropical background). Of the tested lines, 7, 42, 45, and 9 were classified as drought sensitive, moderately sensitive, moderately drought tolerant, and highly drought tolerant, respectively. Seven of the 9 extremely drought tolerant lines were from the TST group, suggesting that TST germplasms harbor valuable genetic resources for drought tolerance that could be used in breeding to improve abiotic stress tolerance in maize.

Key Words: maize, drought tolerance, root system, heterotic groups.

Introduction

Maize is one of the most important cultivated grain crops around the world and is widely used to provide food, forage, and industrial raw materials. Due to rapid changes in populations, society, and economies, the demand for maize is expected to be higher than for wheat or rice by 2020 (Pingali 2001). The productivity and yield of maize are frequently limited by various biotic and abiotic stress factors, such as drought, salinity, high and low temperatures, nutrient deficiencies, disease, and insect pests. Drought stress can affect yield through different mechanisms across the whole life cycle of the maize plant (Leach *et al.* 2011). Therefore, drought is one of the most serious causes of productivity loss. Many studies have pinpointed flowering as the most drought sensitive stage, although seedling establishment is also important because of its influence on plant stand establishment (Bänziger *et al.* 2000, Leach *et al.* 2011).

In addition to anthesis-silking interval (ASI), new traits and methods have been used to help identify drought tolerant genotypes at different developmental stages and to further the creation of new cultivars (Bruce et al. 2002, Meeks et al. 2013). Drought can damage a field at any time throughout the season. The fate of seedlings will determine the structure and dynamics of most plant populations according to the "stress gradient hypothesis" (De La Cruz et al. 2008, Kitajima and Fenner 2000). Thus, phenotypical evaluation at the seedling stage is regarded as an attractive approach because it is a high-throughput and low cost method that saves space and time (Meeks et al. 2013). This approach has been successfully used to develop drought tolerant varieties in cowpea (Singh et al. 1999, Singh and Matsui 2002), cotton (Longenberger et al. 2006), wheat (Tomar and Kumar 2004), and maize (Meeks et al. 2013, Pace et al. 2014, Ruta et al. 2010). Another advantage of using seedling drought screens, where young seedlings undergo cycles

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of water stress in the greenhouse, is that phenotypical variation caused by experimental errors can be controlled better because the plants are much more uniform at an early seeding stage, compared to other periods of plant development (Wang et al. 2015). In cowpea, traits related to drought tolerance at the seedling stage have been shown to be associated with productivity and yield of adult plants (Meeks et al. 2013, Singh et al. 1999, Singh and Matsui 2002). In maize, significant correlations between seminal root traits and adult field traits have been reported, and for shoot weight versus adult plant height, and lateral root length versus brace root development (Landi et al. 1998, 2001, Pace et al. 2014).

Under drought stress, plants seek to reduce the impact of the lack of water by reducing the transpiration rate and by increasing the efficiency of water acquisition from the soil (Végh 2013). Plants have developed numerous adaptive mechanisms for better growth under drought conditions such as modification of the root system, osmotic adjustments, stomatal regulation, chemical production, and accumulation. The root system not only supports the above ground organs of the plant but also plays a crucial role in obtaining water by accessing sources far down in the soil profile. The roots are the first organs to sense a water shortage (Trachsel et al. 2010). The root system is therefore generally considered as the most important organ with respect to improving crop adaptation to water stress (Vadez 2014). Maize responds to drought stress by redirecting root growth and dry matter accumulation away from the shoot to the root (Ribaut et al. 2009, Sharp et al. 2004). In maize, this shift involves an increase in root cell wall extensibility that is mediated by increased levels of xyloglucan endotransglucosylases/hydrolases and other cell wall-loosening factors at the root tip. These modifications result in sustained growth of the root and inhibited growth of the shoot in the face of decreased water potential (Ober and Sharp 2007).

Root morphology is a poorly studied maize characteristic due to the difficulties of making direct measurements under the soil and also of observing or removing roots of plants grown under agronomic conditions. Genetic improvement to produce deep-rooted plants is considered an important strategy for improving water capture and yield stability (Kondo et al. 2003). Variation in root system architecture can be explored to improve plant vigor by improving water use efficiency and nutrient extraction under difficult growing conditions (Malamy and Benfey 1997). Primary root elongation rate and ABA accumulation under different water conditions have been studied in 12 maize inbred lines to assess the relationship between root growth and hormonal conditions (Leach et al. 2011). Differences in morphological root traits have been characterized at the seedling stage in 74 maize inbred lines to quantify the phenotypic and genotypic coefficient of variation, heritability, and interrelationships between these traits (Kumar et al. 2012). Recently, 384 inbred lines from the Ames panel were genotyped with high density single nucleotide polymorphism markers using genotyping-by-sequencing technology to study phenotypic

variation of 22 seedling root architecture traits (Pace et al. 2015). This analysis identified SNP markers throughout the genome that are associated with root architecture traits and the locations of associated SNP markers for possible candidate genes or functional markers for effects on root development have been determined (Pace et al. 2015).

Currently, digital image analysis enables a more accurate and less subjective approach to the analysis of plant root systems; the technology is also quite time- and labor-saving (Bouma et al. 2000, Himmelbauer 2004). Automated phenotypic analysis by digital image software is an innovative approach and there are several software frameworks that extract root morphology traits in two-dimensions in various hierarchies that have been widely used to characterize maize root architecture (Kumar et al. 2012, Pace et al. 2014, 2015, Ruta et al. 2010). In the present study, we aimed to (i) characterize the phenotypic variation for morphological root traits at the seedling stage in 103 maize inbred lines using digital image analysis, (ii) identify root related traits accounting for most of the variation among the tested maize lines, and (iii) compare root phenotypic diversity among different heterotic groups, and evaluate drought tolerance of maize germplasms under well-watered and water-stressed conditions.

Materials and Methods

Plant materials

One hundred and three maize inbred lines from three different heterotic groups, including stiff stalk (SS), non-stiff stalk (NSS), and tropical/subtropical (TST), were selected for identification of root architecture characteristics. Thirtyfour of the lines from China were provided by Sichuan Agricultural University of China, 38 maize lines were from the International Maize and Wheat Improvement Center (CIMMYT), 30 maize inbred lines were from the USA, and 1 line was from Nigeria. This panel includes diverse maize germplasms including 45 temperate lines and 58 tropical/ subtropical lines, and 23 of the lines were used to establish the Nested Association Mapping populations developed by Buckler et al. (2009). Detailed information on each line is given in Supplemental Table 1.

Plant growth conditions

All the test lines were planted in Jinghong, Yunnan province, China, between October 11, 2011 and February 15, 2012 for seed production. Experiments were performed in a climate-controlled chamber with a 16 h light/8 h dark photoperiod, a temperature cycle of 25°C/18°C (day/night), and 65% relative humidity in June, 2012 and repeated in January, 2013. All tested lines were grown in vermiculite with normal water potential after germination on moist filter paper for 72 h. Seedlings of uniform size at the 2-leaf stage and without visible root injuries were transferred to plastic containers holding nutrient solution (10 L solution/container) with minor modifications of the previously described

Table 1. Description of the tested traits in the study

Abbreviated name	Full trait name	Description
TRL	Total root length (cm)	The average root length of six plants
RL	Root length (cm)	RL ¹⁻³ : The average root length of six plants in diameter between 0.0 and 0.5 mm, 0.5 and 2.0 mm, and greater than 2.0 mm, respectively.
TSA	Total root surface area (cm ²)	The average root surface area of six plants
SA	Root surface area (cm ²)	SA ^{1–3} : The average root surface area of six plants in diameter between 0.0 and 0.5 mm, 0.5 and 2.0 mm, and greater than 2.0 mm, respectively.
TRT	Total root tips	The average number of root tips of six plants
RT	Root tips	RT^{1-3} : The average number of root tips of six plants in diameter between 0.0 and 0.5 mm, 0.5 and 2.0 mm, and greater than 2.0 mm, respectively.
TRV	Total root volume (cm ³)	The average root volume of six plants
RV	Root volume (cm ³)	RV ^{1–3} : The average root volume of six plants in diameter between 0.0 and 0.5 mm, 0.5 and 2.0 mm, and greater than 2.0 mm, respectively.
RAD	Root average diameter (cm)	The average root diameter of six plants
RF	Root forks	The average root forks of six plants

basal composition (Hoagland and Arnon 1950). The nutrient solution was aerated continuously and renewed every two days. Distilled water was added regularly to maintain the volume. Six plants for each inbred line grown under well-watered (WW) or water-stressed (WS) conditions (induced by adding the osmolyte 20% polyethylene glycol (w/v) PEG8000; Sigma-Aldrich) were selected for measurement of their root parameters at the 5-leaf stage. The concentration of polyethylene glycol and the stress time of four days were based on a previous report (Trachsel *et al.* 2010).

Root measurements

Measurements were made on seedlings after cultivation in control or low water potential conditions for four days. The seedlings were assigned to the two water regimes using a randomized experimental design, and two replicate experiments were performed. The complete root system was isolated from each plant and placed on a tray with no overlapping of any roots. The WinRhizo Pro 2007a (Regent Instrument Inc., Quebec, Canada) root analysis system was used to investigate root morphology based on images (400 DPI) captured using the EPSON professional scanner (Bouma et al. 2000, Magalhães et al. 2011). The following root parameters were measured: root length (RL, cm), root surface area (RSA, cm²), root average diameter (RAD, cm), root volume (RV, cm³), and the total number of root tips (TRT). More details of each measured parameter are given in Table 1.

Statistical analysis

The data from all measurements of root traits were recorded and compiled in Microsoft Excel 2007. Descriptive analyses including mean, standard deviation (SD), coefficient of variation (CV), analysis of variance (ANOVA), Pearson's correlation, and heritability (H²) estimates were calculated for the tested traits under WW and WS treatments using the SAS program (v9.3). The estimate of heritability is defined by the formula $H = V_G/(V_G + V_E)$, where

V_G and V_E represent estimates of genetic and environmental variances, respectively (Smith et al. 1998). The estimated values of phenotypes for each inbred line were calculated based on best linear unbiased prediction (BLUP) (Piepho et al. 2008). The BLUP values were then used to classify lines into three different categories according to their performance: (i) low performing lines with non-desirable root characteristics $[\leq \overline{X} - SD]$, (ii) lines with medium performance $[\geq \overline{X} - SD]$ to $[\leq \overline{X} + SD]$, and (iii) high performing genotypes with desirable traits $[\geq \overline{X} + SD]$ (Abdel-Ghani et al. 2012, Zar 2010). A polymorphic diversity index, the Shannon-Weaver diversity index (H'), was calculated for each trait (Hutcheson 1970). A principal component analysis (PCA) was performed to identify the major traits accounting for most of the variation in tested maize inbred lines using the SAS program (v9.3). A comprehensive drought resistance measurement value (D value) was introduced to estimate the tolerance capability of all tested lines. The D value was calculated across traits to evaluate maize drought tolerance using the formulas described below (Xu et al. 2009, Zhang and Xu 2009). The drought resistance coefficient (DRC) represents different drought tolerance in the various maize inbred lines. We use P_i to represent the DRC of the jth trait below:

$$DRC(\%) = WS/WW \times 100\%$$

Fuzzy subordinate function analysis was used to decrease the one-sidedness of a simple individual trait to evaluate drought tolerance in various traits. $\mu(X_j)$ stands for the subordinative function value that indicates a positive correlation between trait and drought resistance. By contrast, $1 - \mu(X_j)$ represents a negative correlation between trait and drought resistance.

$$\mu(X_j) = \frac{X_j - X_{min}}{X_{max} - X_{min}} (j = 1, 2, 3, \dots, n)$$

where

 $\mu(X_i)$ is the subordinative function value of DRC of the jth

 X_i is the DRC of the jth trait;

 X_{min} is the minimum value of the DRC of the jth trait; X_{max} is the maximum value of the DRC of the jth trait; Comprehensive drought resistance measurement was made using the formula:

$$\mathbf{D} = \sum_{j=1}^{n} \left[\mu(X_j) \times |P_j| / \sum_{j=1}^{n} |P_j| \right] (j = 1, 2, 3, ..., n)$$

where

D is the comprehensive drought resistance measurement of each maize inbred line under WS condition;

P_i is the DRC of the jth trait in each maize inbred line.

Results

Genetic variation analysis

The BLUP values and descriptive statistics for each root trait for the 103 inbred lines across two repeat experiments (excluding five lines without observed values in both biological repeats) are summarized in **Table 2**. Phenotypic variation among genotypes for each trait was confirmed by the mean, range, standard deviation, and coefficient of variation. Compared to the WW condition, the mean values of most traits were substantially decreased under the WS condition. A wide range of variation was observed for root traits in the two water treatments. Root length, root surface, and root volume (root diameter greater than 2.5 mm) had

	Table 2.	Means, coefficients	of variation and heritability	y estimates for the tested traits under	r well-watered (WW) ar	nd water-stressed (WS) conditions
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Traits	М	ean	Standard de	rivation (SD)	Coefficient of v	variation (CV%)	Heritability		
	WW	WS	WW	WS	WW	WS	WW	WS	
TRL	470.05	316.04	174.16	122.76	37.05	38.84	0.41	0.42	
TSA	81.10	55.71	31.06	22.99	38.30	41.26	0.38	0.54	
RAD	0.56	0.57	0.09	0.08	15.62	14.39	0.64	0.59	
TRV	1.16	0.80	0.54	0.39	46.78	48.12	0.43	0.64	
TRT	699.67	549.14	269.13	208.23	38.46	37.92	0.38	0.41	
RF	3184.46	1911.93	1595.42	941.71	50.10	49.25	0.58	0.48	
RL^1	309.00	197.76	129.31	83.33	41.85	56.79	0.47	0.39	
RL^2	50.51	36.08	18.72	14.86	37.07	43.19	0.31	0.51	
RL^3	1.56	1.04	0.96	0.65	61.48	59.99	0.40	0.54	
SA^1	21.31	7.76	8.50	4.41	39.89	38.27	0.43	0.43	
SA^2	13.87	9.41	5.56	4.07	40.12	46.29	0.35	0.58	
SA ³	1.55	1.05	0.95	0.63	60.89	59.37	0.38	0.50	
RV^1	0.15	0.10	0.06	0.04	39.65	38.39	0.42	0.43	
RV^2	0.35	0.23	0.15	0.11	44.10	36.05	0.40	0.61	
RV ³	0.15	0.10	0.09	0.06	62.05	55.36	0.35	0.33	

TRL: total root length; TSA: total root surface area; RAD: root average diameter; TRV: total root volume; TRT: total root tips; RF: root forks; RL¹⁻³, SA¹⁻³ and RV¹⁻³ indicate average root length, root surface area and root volume in diameter between 0.0 and 0.5 mm, 0.5 and 2.0 mm and greater than 2.0 mm, respectively.

Table 3. Analysis of variance for the tested traits under two water regimes

Variables	Type III Sur	m of squares	Mean	square	F v	alue	Significance		
variables	Genotype	Treatment	Genotype	Treatment	Genotype	Treatment	Genotype	Treatment	
df	102	1	102	1	102	1	102	1	
TRL	6,305,553.64	2,280,575.16	61,218.97	2,280,575.16	1.94	72.15	***	***	
TSA	219,795.56	62,480.89	2,133.94	62,480.89	2.63	77.12	***	***	
RAD	2.35	0.00	0.02	0.00	2.73	0.45	***	NS	
TRV	67.66	12.48	0.66	12.48	3.21	60.89	***	***	
TRT	15,746,306.45	2,084,110.65	152,876.76	2,084,110.65	1.14	15.53	NS	***	
RF	481,303,981.69	154,999,703.12	4,672,854.19	154,999,703.12	3.33	110.38	***	***	
RL^1	3,312,588.26	1,176,337.89	32,161.05	1,176,337.89	1.59	58.22	***	***	
RL^2	85,641.90	20,168.48	831.47	20,168.48	2.62	63.66	***	***	
RL ³	197.11	26.94	1.91	26.94	1.68	23.61	***	***	
SA^1	14,044.77	5,048.83	136.36	5,048.83	1.70	62.79	***	***	
SA^2	7,046.01	1,918.43	68.41	1,918.43	2.68	75.23	***	***	
SA ³	187.12	25.17	1.82	25.17	1.41	19.50	*	***	
RV^1	0.66	0.23	0.01	0.23	1.86	67.85	***	***	
RV^2	5.18	1.40	0.05	1.40	2.80	77.90	***	***	
RV ³	1.68	0.19	0.02	0.19	1.12	13.36	NS	***	

df: degree of freedom; TRL: total root length; TSA: total root surface area; RAD: root average diameter; TRV: total root volume; TRT: total root tips; RF: root forks; RL¹⁻³, SA¹⁻³ and RV¹⁻³ indicate average root length, root surface area and root volume in diameter between 0.0 and 0.5 mm, 0.5 and 2.0 mm and greater than 2.0 mm, respectively. *, ** and *** significant at P < 0.05, P < 0.01 and P < 0.001, respectively. NS, not significant.

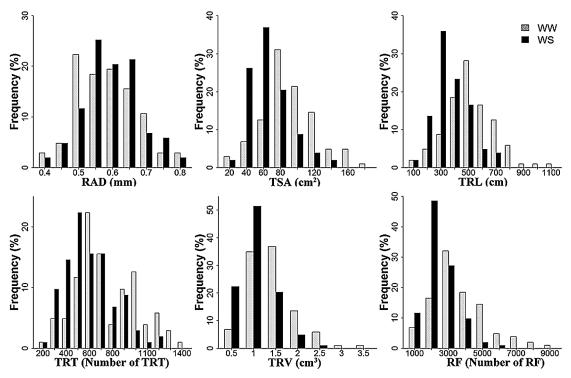


Fig. 1. Frequency distribution of variation for 9 traits in 103 maize lines. TRL: total root length; TSA: total root surface area; RAD: root average diameter; TRV: total root volume; TRT: total root tips; RF: root forks. WW: well-watered condition; WS: water-stressed condition.

relatively higher coefficients of variation (>50%) in both water treatments while average root diameter had relatively lower coefficients of variation (around 15%).

ANOVA revealed significant genetic variation among genotypes for nine traits (**Table 3**). Genotypic variation was significant for six traits at P < 0.001 and eight traits showed significant variation under different water regimes, the exception being average root diameter. The results indicate that these traits were significantly affected by water supply in the tested maize lines. The level of variation was also reflected by the distribution of traits representing different drought resistance criteria (**Fig. 1**). Histograms of frequency distribution of root traits showed approximately normal distributions. Under the WS condition, the values for seven traits (TRL, TSA, RAD, TRV, TRT, and RF) decreased significantly.

Heritability estimates

Heritability estimates for the traits are shown in **Table 2**. The traits ranged from 0.315 (RL² under WW) to 0.636 (RAD under WW). RAD showed relatively high heritability (0.636 for WW and 0.593 for WS), as did RF (0.581 for WW and 0.480 for WS) and TRV (0.429 for WW and 0.637 for WS). TRL (0.409 for WW and 0.417 for WS), TSA (0.383 for WW and 0.539 for WS), and TRT (0.376 for WW and 0.406 for WS) showed lower heritabilities. The heritability estimates for RF and RAD were higher under WW than WS conditions, but for other traits were lower under WW than WS conditions. These results indicate that heritability of root characteristics was generally lower than other

agronomic traits, and that the root morphology of maize seedlings was strongly influenced by the environment. The widely used indicator, RAD was highly heritable, suggesting that it is a reliable indicator for drought tolerance.

Genetic correlations among tested traits

Pearson correlations among the traits were calculated, and significant correlations (P < 0.01) were observed between all pairs of traits (**Table 4**). Relatively high positive correlations were found for root-related traits under the two water regimes, for example, TRL and TSA (r = 0.894 and r = 0.891 for WW and WS, respectively), and TRV and TSA (r = 0.930 and r = 0.948 for WW and WS, respectively). RAD showed significant negative correlations with TRL, TRT, RF, RL¹, RV¹, and RSA¹ under the two water regimes.

Diversity patterns with respect to heterotic groups

A comparison of the root characteristics of the different heterotic groups identified clear variation for all traits in the three groups. For most root traits, the SS and NSS groups (temperate maize backgrounds) showed higher mean values, while the TST group (tropical and subtropical backgrounds) displayed relatively lower mean values. Remarkable variation was found for total root length, and the mean values in the three heterotic groups, NSS, SS, and TST, were 501.8 cm, 526.8 cm, and 472.9 cm, respectively. Similar results were also observed for other traits including the number of root forks and tips, total root surface area and total root volume. The mean, median, and range of phenotypic variation in NSS, SS, and TST groups are shown in Fig. 2.

 Table 4. Genetic correlation among tested traits under well-watered

 (WW) and water-stressed (WS) conditions

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	TRL	TSA	RAD	TRV	TRT	RF
WW						
TRL	1	.894**	127	.670**	.764**	.941**
TSA		1	.304**	.930**	.561**	.833**
RAD			1	.606**	358**	134
TRV				1	.304**	.613**
TRT					1	.755**
RF						1
WS						
TRL	1	.891**	065	.739**	.704**	.913**
TSA		1	.287**	.948**	.605**	.849**
RAD			1	.541**	231*	.005
TRV				1	.432**	.724**
TRT					1	.587**
RF						1

TRL: total root length; TSA: total root surface area; RAD: root average diameter; TRV: total root volume; TRT: total root tips; RF: root forks; WW: well water; WS: water stress; *, ** and *** significant at P < 0.05, P < 0.01 and P < 0.001, respectively.

The maize lines in the NSS, TST, and SS groups were classified into three categories, namely high, medium, and low performance (**Table 5**). Under the WS condition, 6 (21%), 5 (9%), and 2 (11%) lines from NSS, TST, and SS groups, respectively, showed large total root volume (TRV) $[\geq \overline{X} + SD]$. Under the WW condition, 5 (18%), 11 (19%), and 2 (11%) lines from NSS, TST, and SS groups, respectively showed large TRV. For all the traits except TRV, the TST group had a lower proportion of lines with $[\geq \overline{X} + SD]$ than either the NSS or SS groups under the two water regimes (**Table 5**). The results indicated that the NSS and SS groups (temperate maize backgrounds) contained more lines

with desirable root characteristics than the TST group (tropical/subtropical germplasms) under the two water regimes.

The Shannon-Weaver diversity index (H') was calculated for all the lines and the three heterotic groups (Table 5). The H' values varied in the traits TRL, TSA, RAD, TRV, TRT, and RF with an average of 0.770, 0.784, 0.745, and 0.721 for all the maize lines, NSS, TST, and SS groups, respectively. Under the two water regimes, RAD and TRL showed relatively higher levels of variation. RF was less variable in all lines and across the three heterotic groups. The variation in root morphology among the three heterotic groups indicated that the NSS group had the highest average H' value, while the SS and TST groups had almost similar values. All the root traits except RAD showed higher H' values and higher diversity under the WS condition than the WW condition. For three traits, TRL, TRT, and RF, higher diversity was present in the TST group under the WW condition than under the WS condition.

Principal component analysis

Based on the genetic correlations among different root traits, two traits with correlation coefficients higher than 0.9 were combined in our principal components analysis. The first two principal components (PCs) explained about 94.01% and 91.15% of the total variation among the maize lines under WW and WS conditions, respectively (**Fig. 3a**, **3b**). The first PC, which explained more than 61% of the total variation, revealed that TSA and TRL, and their highly correlated traits TRV and RF, were the most important contributing traits. The most important trait in the second principal component, which contributed nearly 30% of the total variation, was RAD.

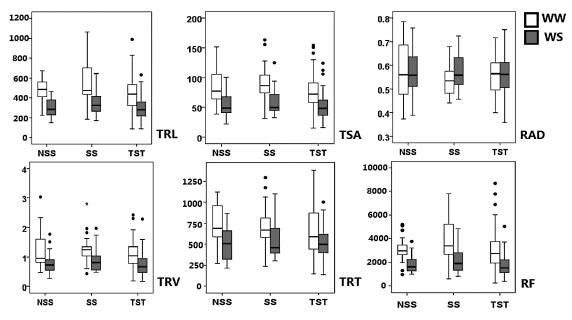
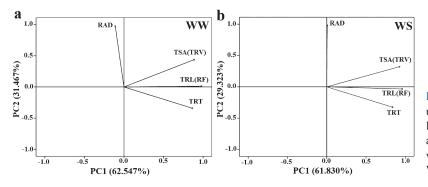


Fig. 2. Box plot showing the medians and ranges of phenotypic variation in three heterotic groups of maize. TRL: total root length; TSA: total root surface area; RAD: root average diameter; TRV: total root volume; TRT: total root tips; RF: root forks. WW: well-watered condition; WS: water-stressed condition. NSS: non-Stiff Stalk; SS: Stiff Stalk; TST: subpopulation including tropical/subtropical lines.

Table 5. The Shannon-Weaver diversity index (H') and performance categories under drought stress

Turita	Treat-	All maize lines				NSS group			TST group			SS group					
Traits	ment	Low	Medium	High	\mathbf{H}'	Low	Medium	High	\mathbf{H}'	Low	Medium	High	H'	Low	Medium	High	H^{\prime}
TRL	WW	13	83	13	0.72	3	23	2	0.59	8	43	6	0.73	1	14	3	0.65
TRL	WS	16	73	21	0.87	6	16	6	0.98	7	41	10	0.80	2	14	2	0.68
TSA	WW	12	82	15	0.73	2	22	4	0.66	8	42	7	0.76	3	11	4	0.93
TSA	WS	14	76	20	0.83	3	20	5	0.79	7	41	10	0.80	2	14	2	0.68
RAD	WW	19	71	19	0.89	5	17	6	0.94	8	41	8	0.79	5	9	4	1.04
RAD	WS	17	77	16	0.82	6	19	3	0.83	9	40	9	0.83	2	13	3	0.78
TRV	WW	10	86	13	0.66	3	19	6	0.83	4	48	5	0.54	2	14	2	0.68
TRV	WS	11	82	17	0.74	3	20	5	0.79	7	40	11	0.83	0	16	2	0.35
TRT	WW	12	80	17	0.76	2	22	4	0.66	6	42	9	0.75	1	14	3	0.65
TRT	WS	18	74	18	0.86	5	18	5	0.90	8	43	7	0.75	2	13	3	0.78
RF	WW	9	87	13	0.64	2	24	2	0.51	5	46	6	0.62	1	14	3	0.65
RF	WS	12	79	19	0.78	4	17	7	0.93	5	43	10	0.74	2	13	3	0.78

TRL: total root length; TSA: total root surface area; RAD: root average diameter; TRV: total root volume; TRT: total root tips; RF: root forks; NSS: non-Stiff Stalk; SS: Stiff Stalk; TST: subpopulation including tropical/subtropical lines; H': Shannon-Weaver diversity index; WW: well-watered condition; WS: water-stressed condition.



Germplasm resources for root morphology improvement under drought stress

The maize lines were classified into three categories, high, medium, and low performance with respect to each trait (Table 5). Most maize lines were found to fall into the medium performance category, and this was the case for all three heterotic groups. The proportion of maize lines with high performance ranged from 10 to 15%, that with medium performance ranged from 70 to 75%, and that with low performance ranged from 14 to 17%. The D value, as a synthetic index, was used to evaluate root morphology among the maize lines under drought stress (Supplemental Table 1). Based on D values, the lines could be classified into four groups. Group 1 with 7 lines, including 5003, Ji853, and Zheng22, showed sensitivity to drought with D values lower than 0.20. Group 2, with 42 lines, including Dan598 and ES40, showed moderate sensitivity to drought stress with D values between 0.20 and 0.40. Group 3 with 45 lines showed moderate tolerance with D values between 0.40 and 0.70. Group 4, with 9 lines, including CML247 and Mo17, showed drought tolerance with D values greater than 0.70. Interestingly, 7 of the 9 extremely drought tolerant lines came from the TST group (tropical genetic background). The maize lines that were extremely sensitive or extremely tolerant under drought stress are listed in **Supplemental Table 1**.

The mean values and standard deviations of the drought

Fig. 3. Principal component analysis of four traits under WW (a) and WS (b) condition. TRL: total root length; TSA: total root surface area; RAD: root average diameter; TRT: total root tips; TRV: total root volume; RF: root forks. WW: well-watered condition; WS: water-stressed condition.

resistance coefficient (DRC) for each trait in the four groups with different levels of drought tolerance are shown in **Fig. 4**. The mean values of DRC for all root traits were lowest in group 1, moderate in groups 2 and 3, and highest in group 4, except for RAD. This result indicates that drought tolerant maize lines with higher D values also had higher drought resistance coefficients, and consistent results were obtained when these two indices were calculated based on the root morphology traits used for drought tolerance screening.

Discussion

Drought stress affects many of the processes required for plant growth and development, for example, it inhibits cell elongation, reduces cell division, decreases the photosynthetic rate, and modifies root morphology (Kumar *et al.* 2004). Maize is susceptible to drought stress throughout its life cycle, and water deficits during mid to late vegetative development and flowering significantly affect root morphology, reproductive tissues, biomass production, and gain yield (Chen *et al.* 2012, Heiniger 2001, Kumar *et al.* 2004, Svačina *et al.* 2014). The root system is an indispensable organ for absorption of nutrients and water in plants and its physiological characteristics largely determine the rates of absorption of nutrients and water (Hodge *et al.* 2009).

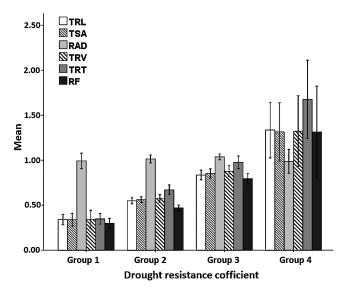


Fig. 4. The mean values and standard deviations of drought resistance coefficient for 9 traits in three groups classified for drought tolerance. Groups 1, 2, 3 and 4 represent inbred lines identified with drought sensitivity, moderate sensitivity, moderate drought tolerance and high tolerance ability under drought stress. N = 7, 42, 45 and 9 for groups 1, 2, 3 and 4, respectively. Bars are the standard deviation. TRL: total root length; TSA: total root surface area; RAD: root average diameter; TRV: total root volume; TRT: total root tips; RF: root forks.

Roots, embedded in the soil, are the first plant organs sensing water shortage (Trachsel et al. 2010). In the present investigation, the integrity of the studied root systems was maintained by growing the tested inbred lines in solution culture. It may seem counter-intuitive to conduct a waterstressed experiment in a nutrition solution. However, drought stress is manifested in multiple ways in plants grown in soil including mechanical impedance and desiccation (Trachsel et al. 2010). PEG with a molecular weight above 8000 Da is an osmolyte that does not enter the root system; it can been used to modify the osmotic potential of nutrient solutions and induce desiccation stress in a relatively controlled manner (Trachsel et al. 2010). A previous study examined the effect of growing plants under PEG8000 stress and quantified the ability of three tropical maize inbred lines to proliferate roots under conditions of adequate water supply compared to desiccation stress (Trachsel et al. 2010). Drought condition can be successfully mimicked by PEG treatment, which reduces the external free water concentration without altering the ionic composition of the cell and causes reduction in leaf water potentials (Claes et al. 1990, Ruta et al. 2010). Thus, root system responses to drought stress can be tested without root damage in solution cultures by controlling access to water.

Enhanced root growth, as evaluated by root dry weight and root-to-shoot ratios in moderate and high P-efficiency maize lines, is connected to the search for nutrients and water. Under water-stressed conditions, maize lines with different genetic backgrounds and origins displayed different drought tolerance capabilities and showed varied root architecture traits at the seedling stage (Kumar *et al.* 2012, Liang *et al.* 2013). Characterization of maize germplasm with better stress tolerance traits and screening for drought tolerant maize lines are essential to the success of breeding programs.

Traditionally, root length density and depth are considered as ideal criteria for evaluating drought tolerance of a plant root system (Kashiwagi et al. 2006). The ratio of root weight to shoot weight has been used as an index for drought resistance because large deep-rooted systems are able to extract more water while relatively smaller shoots transpire less (Srividya et al. 2011). Total root length represents the sum of the primary, crown, seminal, and lateral roots. The various components of the root system have also been selected as important traits for root morphology improvement under drought stress. In this study, we investigated the influence of water stress on the root morphology in 103 maize lines and examined various root traits, including TRL, TSA, TRT, RAD, and RV. We identified significant variation, medium to high heritability, and significant correlations for these root traits. The PCA showed that root traits such as TRL, TSA, and RF were responsible for most of the phenotypic variation at the seedling stage in the tested maize lines. In this study, H' values were calculated to compare the levels of diversity among the tested traits. A low H' value indicates an unbalanced frequency distribution for a trait with a lack of genetic diversity, while a high H' value indicates an even frequency distribution and a wide range of variation. All recorded traits showed high H' values, with TRL and RAD showing the relatively highest levels of phenotypic diversity. TRL was positively correlated with TSA, RV, and RF. In combination with the PCA analysis, we found that TRL and TSA were sufficient to explain the variation and that they could be used as favorable selection criteria for drought tolerance at the seedling stage. Moreover, TRL and TSA showed much higher drought tolerance coefficients in drought-tolerant maize lines. This result is in agreement with previous reports. For example, Kumar et al. (2012) reported that total root length and root dry weight (DW) provide the largest contribution to total phenotypic variation and might be sufficient to improve other root traits. Root surface area has been shown to have a close relationship to nutrient absorption rates (Imada et al. 2008). Thus, the larger TSA present in maize lines with high phosphorous absorbance efficiency can help the plants to overcome nutrient deficiencies (Zhang et al. 2014). However, there are some conflicting results in the literature regarding the role of root development at the seedling stage and subsequent grain yield. Manavalan et al. (2011) concluded that variation in root characteristics among parents of NAM lines is inherent but is not related to variations in kernel size. However, other researchers drew the opposite conclusion and stated that deep rooting was positively associated with seed yield and crop growth (Eghball and Maranville 1993, Pandey et al. 2000a, 2000b). There is a very weak correlation between kernel weight and TRL and

root DW, indicating a weak influence of root morphology on kernel seed size (Kumar *et al.* 2012). Therefore, a vigorous plant root system not only contributes to improving stand establishment but also enables the plant to survive in stressful conditions.

Diversity in root morphology can be exploited to improve nutrient and water use efficiency under abiotic stresses. A combination of stress resistance ability and the heterotic patterns of maize germplasm will be valuable to breeding and yield improvement. A comparison of the root characteristics of different heterotic groups indicated that SS and NSS groups (temperate maize backgrounds) showed relatively higher diversity of root morphologies than the TST group (tropical maize background). TST lines showed the highest level of diversity in TRL, TRT, and RF under the WW condition but showed lower diversity under the WS condition. Most importantly, 7 of the 9 extremely drought tolerant lines were from the TST group, indicating that the TST germplasm harbors more valuable genetic resources for drought tolerance. This result provides important information for maize hybrid breeding and improving resistance to abiotic stresses. Based on the synthetic index of D values, 7, 42, 45, and 9 maize lines were classified as drought sensitive, moderately sensitive, moderately drought tolerant, and highly drought tolerant, respectively. Drought resistance coefficients for all tested traits, except RAD, were highest in group 4, intermediate in groups 2/3, and lowest in group 1. As a result, 9 maize lines with a well-developed root system were selected for their extreme drought tolerance and could be used in maize breeding for further improvement of tolerance to abiotic stresses. Some of these lines with welldeveloped root systems have been selected and reported previously in drought tolerance screening experiments based on different selection criteria such as ASI and NDVI in different stages (Lu et al. 2011), while others are newly identified drought tolerant inbred lines and offer the possibility of new drought-responsive genes.

In conclusion, we found a range of responses to drought stress among various maize lines at the seedling stage as revealed by analysis of the root systems and physiological characteristics. We show that selection criteria based on the use of TRL and TSA at the seedling stage might be successful predictors of nutrient and water-use efficiencies in the field. The tested maize lines need be further investigated for their performance at the adult stage under WW and WS conditions. Additionally, the association of adult stage traits with seedling root traits needs to be further examined.

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