

Masashi Arai, MD

 itochondria occupy approximately 30% cell volume of cardiac myocytes. They are key organelles that orchestrate cardiac function under physiological and pathological conditions. Their principal function is ATP production via their electron transport system. However, recent studies indicate that mitochondria have additional roles in oxidative stress production,1 intracellular Ca2+ handling,2 and apoptosis induction,3 all of which interplay to create the single functional machinery of the mitochondria. For example, intracellular Ca²⁺ concentration is a trigger that activates pyruvate dehydrogenase, isocitrate dehydrogenase, α -ketoglutarate dehydrogenase and ATP synthase, enzymes of ATP production. Intracellular Ca²⁺ concentration is controlled by the sarcoplasmic reticulum Ca2+-ATPase (SERCA2) protein, whose function depends on the amount of ATP supplied by the mitochondria. In the failing heart, impaired SERCA2 function decreases intracellular Ca²⁺, which leads to further decreases in ATP production, resulting in a vicious cycle of impaired ATP production and Ca2+ cycling.4

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Recent models using genetically engineered mice have clarified the importance of the mitochondria in securing proper cardiac function. Mitochondria possess their own DNA that encodes 13 key proteins in their electron transport system. mRNA for these proteins are transcribed by the mitochondrial DNA-specific transcription factor, Tfam, a regulator of mitochondrial DNA replication. Heart-specific Tfam knockout mice develop hearts with a dilated left ventricular chamber, low ejection fraction and atrioventricular conduction delay, resembling dilated cardiomyopathy,⁵ and have a lifespan that is significantly shorter than that of wild-type mice. In contrast, Tfam transgenic mice develop hearts that cannot undergo cardiac remodeling after myocardial infarction and show better survival than wild type mice.⁶

In this issue of the *Circulation Journal*, Mizuno et al⁷ provide experimental evidence that antenatal administration of dexamethasone enhances ATP production after birth by activating the mRNA expression of creatine kinases (CKs), key enzymes for energy transfer from sites of ATP production to those of consumption.⁸ Antenatal corticosteroid hormone treatment is recommended by the National Institutes of Health for maturation of the fetal lungs in the case of premature de-livery.⁹ The serum cortisol concentration is lower in preterm

infants than in mature infants, and cortisol accelerates the maturation of the lung, liver, gut and kidney.¹⁰ However, the effect of glucocorticoid on heart development, especially on the ATP production and transfer system, has not been fully elucidated. Cardiac myocytes express 3 different isoforms of CK: myofibrillar M-type CK (M-CK), myofibrillar B-type CK and mitochondrial CK (miCK). miCK transforms ATP produced in mitochondria into phosphocreatine (PCr), an energy "currency" in the cell, and then PCr is re-converted into ATP by myofibrillar CK located in the vicinity of ATP-consuming sites such as myofibrils and the sarcoplasmic reticulum. Models using genetically engineered mice reveal that two-thirds of the energy needed for ATPases is provided by PCr, while the remaining energy comes from ATP itself.¹¹

This study demonstrates quite low mRNA expression and M-CK concentration during the fetal stage, but which gradually increase after birth. Thus, babies are not born with a fully functional heart. Furthermore, induction of miCK mRNA expression is relatively late compared with that of M-CK, which confirms a previous report on the time difference in the development of M-CK and miCK function.¹² Because of the low PO₂ environment in the fetus, the fetal heart does not produce energy by β -oxidation of fatty acids in the mitochondria, which requires large amounts of oxygen, but by glycolysis in the cytosol, where M-CK is expressed. High concentration of oxygen after birth may thus activate both M-CK and miCK function and mRNA expression.

This study also found that antenatal glucocorticoid administration significantly increased mRNA and protein levels of M-CK and miCK, even in the late fetal, anaerobic stage. Because M-CK and miCK mRNA failed to be induced by glucocorticoid in the presence of a glucocorticoid receptor antagonist, CKs are most likely induced by signaling through such glucocorticoid receptors. The glucocorticoid receptor is a nuclear receptor that complexes with glucocorticoids to initiate transcription of many genes by binding with the glucocorticoid responsive element (GRE) of target genes.¹³ Unlike genes that have GRE, such as insulin-like growth factor II, angiotensinogen and erythropoietin, the CK genes appear to lack apparent GRE in their regulatory region. The precise mechanism for post-transcriptional control of CKs requires further investigation.

Antenatal glucocorticoid administration was also demonstrated to increase ATP levels after birth. The principal function of the CK system is energy transport from the mitochon-

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Mailing address: Masashi Arai, MD, Department of Medicine and Biological Science, Gunma University Graduate School of Medicine, 3-39-22 Showa-machi, Maebashi 371-8511, Japan. E-mail: araim@showa.gunma-u.ac.jp

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Department of Medicine and Biological Science, Gunma University Graduate School of Medicine, Maebashi, Japan

dria to ATPase, known as CK shuttling.⁸ However, the CK system is capable of increasing ATP production by enhancing the conversion of PCr to ATP when mitochondrial function is impaired¹¹ or visa versa when CK function is impaired. Thus, the CK system works with the mitochondria in both a competitive and compensatory manner. Notably, ATP production can be enhanced by manipulating the CK system by antenatal glucocorticoid administration.

Glucocorticoid induces myocyte differentiation rather than myocyte proliferation. Cardiac myocytes are able to proliferate through cell division, even after birth. Therefore, antenatal glucocorticoid therapy may decrease the number of cardiac myocytes and induce cell hypertrophy. In fact, a decrease in cell number has been reported after glucocorticoid administration.¹⁴ It is unknown whether glucocorticoid-treated cardiac myocytes have the potential to divide after birth. In this case, antenatal glucocorticoid therapy may be able to improve heart development temporarily but not to a level sufficient for adulthood. Further investigation and thorough follow-up of currently treated babies¹⁵ are required to establish this treatment as a standard therapy.

References

- Tsutsui H, Kinugawa S, Matsushima S. Oxidative stress and mitochondrial DNA damage in heart failure. *Circ J* 2008; **72**(Suppl A): A-31–A-37.
- Belkacemi L, Bedard I, Simoneau L, Lafond J. Calcium channels, transporters and exchangers in placenta: A review. *Cell Calcium* 2005; 37: 1–8.
- Feissner RF, Skalska J, Gaum WE, Sheu SS. Crosstalk signaling between mitochondrial Ca²⁺ and ROS. *Front Biosci* 2009; 14: 1197–1218.
- 4. Stanley WC, Chandler MP. Energy metabolism in the normal and failing heart: Potential for therapeutic interventions. *Heart Fail Rev*

2002; **7:** 115–130.

- Li H, Wang J, Wilhelmsson H, Hansson A, Thoren P, Duffy J, et al. Genetic modification of survival in tissue-specific knockout mice with mitochondrial cardiomyopathy. *Proc Natl Acad Sci USA* 2000; 97: 3467–3472.
- Ikeuchi M, Matsusaka H, Kang D, Matsushima S, Ide T, Kubota T, et al. Overexpression of mitochondrial transcription factor a ameliorates mitochondrial deficiencies and cardiac failure after myocardial infarction. *Circulation* 2005; **112**: 683–690.
- Mizuno M, Takeba Y, Matsumoto N, Tsuzuki Y, Asoh K, Takagi M, et al. Antenatal glucocorticoid therapy accelerates ATP production with creatine kinase increase in the growth-enhanced fetal rat heart. *Circ J* 2010; **74:** 171–180.
- Joubert F, Mazet JL, Mateo P, Hoerter JA. 31P NMR detection of subcellular creatine kinase fluxes in the perfused rat heart: Contractility modifies energy transfer pathways. *J Biol Chem* 2002; 277: 18469–18476.
- NIH Consensus Development Panel on the Effect of Corticosteroids for Fetal Maturation on Perinatal Outcomes. Effect of corticosteroids for fetal maturation on perinatal outcomes. JAMA 1995; 273: 413–418.
- Fowden AL, Li J, Forhead AJ. Glucocorticoids and the preparation for life after birth: Are there long-term consequences of the life insurance? *Proc Nutr Soc* 1998; 57: 113–122.
- Kaasik A, Veksler V, Boehm E, Novotova M, Minajeva A, Ventura-Clapier R. Energetic crosstalk between organelles: Architectural integration of energy production and utilization. *Circ Res* 2001; 89: 153–159.
- Hoerter JA, Kuznetsov A, Ventura-Clapier R. Functional development of the creatine kinase system in perinatal rabbit heart. *Circ Res* 1991; 69: 665–676.
- 13. Rhodes C, Yamada Y. Characterization of a glucocorticoid responsive element and identification of an AT-rich element that regulate the link protein gene. *Nucleic Acids Res* 1995; **23**: 2305–2313.
- Rudolph AM. Myocardial growth before and after birth: Clinical implications. Acta Paediatr 2000; 89: 129–133.
- Mulder EJ, de Heus R, Visser GH. Antenatal corticosteroid therapy: Short-term effects on fetal behaviour and haemodynamics. *Semin Fetal Neonatal Med* 2009; 14: 151–156.