

Research article

GPR99, a new G protein-coupled receptor with homology to a new subgroup of nucleotide receptors

Timo Wittenberger†¹, Susanne Hellebrand†¹, Antonia Munck¹, Hans-Jürgen Kreienkamp², H Chica Schaller¹ and Wolfgang Hampe*¹

Address: ¹Zentrum für Molekulare Neurobiologie, Universität Hamburg, Martinistr. 52, D-20246 Hamburg, Germany and ²Institut für Zellbiochemie und Klinische Neurobiologie, Universität Hamburg, Martinistr.52, D-20246 Hamburg, Germany

E-mail: Timo Wittenberger - timo.wittenberger@altanapharma.com; Susanne Hellebrand - susanne.hellebrand@roche.com; Antonia Munck - munck@uke.uni-hamburg.de; Hans-Jürgen Kreienkamp - kreienkamp@uke.uni-hamburg.de; H Chica Schaller - schaller@zmnh.uni-hamburg.de; Wolfgang Hampe* - hampe@uke.uni-hamburg.de

*Corresponding author †Equal contributors

Published: 5 July 2002

Received: 12 March 2002

BMC Genomics 2002, 3:17

Accepted: 5 July 2002

This article is available from: <http://www.biomedcentral.com/1471-2164/3/17>

© 2002 Wittenberger et al; licensee BioMed Central Ltd. Verbatim copying and redistribution of this article are permitted in any medium for any purpose, provided this notice is preserved along with the article's original URL.

Abstract

Background: Based on sequence similarity, the superfamily of G protein-coupled receptors (GPRs) can be subdivided into several subfamilies, the members of which often share similar ligands. The sequence data provided by the human genome project allows us to identify new GPRs by in silico homology screening, and to predict their ligands.

Results: By searching the human genomic database with known nucleotide receptors we discovered the gene for GPR99, a new orphan GPR. The mRNA of GPR99 was found in kidney and placenta. Phylogenetic analysis groups GPR99 into the P2Y subfamily of GPRs. Based on the phylogenetic tree we propose a new classification of P2Y nucleotide receptors into two subgroups predicting a nucleotide ligand for GPR99. By assaying known nucleotide ligands on heterologously expressed GPR99, we could not identify specifically activating substances, indicating that either they are not agonists of GPR99 or that GPR99 was not expressed at the cell surface. Analysis of the chromosomal localization of all genes of the P2Y subfamily revealed that all members of subgroup "a" are encoded by less than 370 kb on chromosome 3q24, and that the genes of subgroup "b" are clustered on one hand to chromosome 11q13.5 and on the other on chromosome 3q24-25.1 close to the subgroup "a" position. Therefore, the P2Y subfamily is a striking example for local gene amplification.

Conclusions: We identified a new orphan receptor, GPR99, with homology to the family of G protein-coupled nucleotide receptors. Phylogenetic analysis separates this family into different subgroups predicting a nucleotide ligand for GPR99.

Background

The superfamily of G protein-coupled receptors (GPRs) is one of the largest human gene families [1]. Many different approaches have been undertaken to identify new GPRs,

both biochemically and by database searches. Recently, we have identified several new GPR genes from the database of expressed sequence tags by a complex computational strategy [2]. With the availability of the human

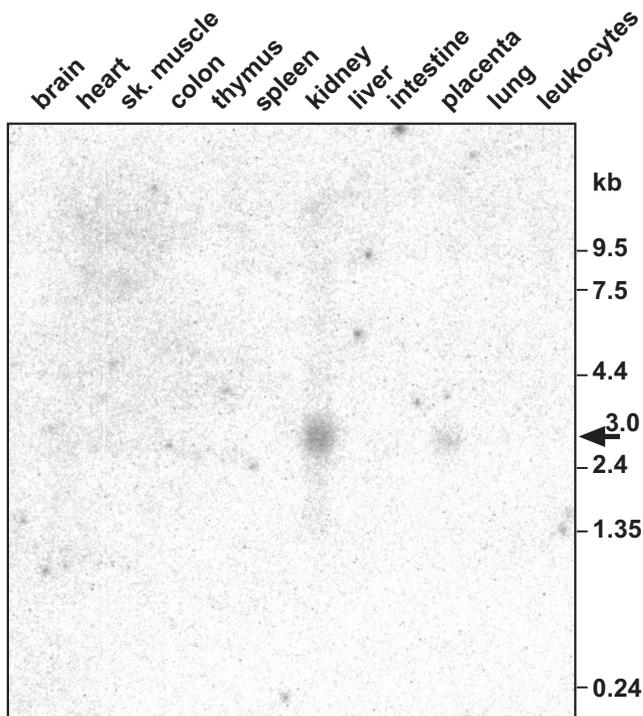


Figure 1
Northern blot analysis of human GPR99.

genomic sequence another source for data mining became accessible, which is especially valuable for GPR searches, since many GPR genes contain no or only a few introns. Nevertheless, the existence of pseudogenes, many of which are not transcribed or lead to truncated proteins, makes it necessary to prove the expression of each putative gene found in the genome. For this publication we used some of the new receptors with homology to the subfamily of P2Y nucleotide receptors [2] to search for further GPRs in the human genomic database.

Results and discussion

Identification of GPR99

Performing TBLASTN searches of the human genomic database with the known nucleotide GPRs we identified an additional ORF, which we named GPR99. The ORF reaches from bp 140188 to 141201 of the BAC clone with the accession number AC026756.15. The putative start codon in a Kozak context is located 15 bp behind an in-frame stop codon. The last 20 bp of the ORF and the 3' UTR are found on an EST clone derived from thyroid epithelium (AW827323). Sequencing the entire EST clone from the I.M.A.G.E consortium [3], which was supplied by the Resource Center of the German Human Genome Project at the Max-Planck-Institute for Molecular Genetics, confirmed the genomic sequence behind the coding region for transmembrane domain six. To show mRNA expres-

sion of full-length GPR99, we amplified the entire coding region together with the upstream stop codon from a human placenta cDNA library by PCR. Direct sequencing of the PCR product was in perfect accordance with the genomic data. The sequence was submitted to GenBank under accession number AF370886. mRNA expression of GPR99 was proven by northern blot analysis. A 3.0 kb mRNA was detected in kidney and, to a lower extent, in placenta (Fig. 1).

Sequence analysis

GPR99 shares 36% identical amino acids both with the P2Y₁ receptor (57% similarity) and with GPR91 (55% similarity) as its closest homologs (Fig. 2). "Fingerprint" analysis for GPR subtypes [4] also groups GPR99 into the P2 purinoceptor subfamily. No receptors from other species with a similar degree of relatedness were identified in the databases.

Phylogenetic analysis of all human members of the P2Y subfamily of GPRs results in three subgroups designated "a", "b", and "n" (Fig. 3A). Subgroup "n" (for non-nucleotide receptors) contains P2Y₅, P2Y₇, P2Y₉, and P2Y₁₀, all of which, for homology reasons, were designated P2Y receptors. Closer analysis revealed that none of these is activated by nucleotides, but one of them, P2Y₇, binds leukotriene B4 [5]. In contrast, GPR99 belongs to subgroup "b" which, in addition to the new orphan receptor GPR91, consists of five receptors with proven nucleotide agonists, hinting at a similar ligand for GPR91 and GPR99. Subgroup "a" consists of the orphan receptors GPR87 and H963, and of GPR86/P2Y₁₃[6], P2Y₁₂ and KIAA0001, which bind the nucleotide ligands ADP, ADP, and UDP-glucose, respectively.

A closer analysis of residues implicated in direct nucleotide interactions confirms the existence of two distinct nucleotide receptor subgroups. At the beginning of transmembrane domain seven, all receptors of subgroup "b", including GPR99, share the motif Y-X-V-T-R-P-L, which is not found in the other GPRs (Fig. 3B). The highly specific positively charged arginine residue of this motif was shown to play an important role for binding of negatively charged nucleotides [7]. In the same position the subgroup "a" members share the motif K-E-X-T-L-X-L. P2Y₅, P2Y₇, P2Y₉, P2Y₁₀ and other non-nucleotide receptors lack both of these motifs (Fig. 3B). These motifs might, therefore, be good diagnostic tools to predict ligands for additional orphan receptors.

Towards the end of transmembrane domain seven many GPRs share the motif D/N-P-X-X-Y. GPR99 lacks the highly conserved proline residue of this motif (Fig. 3B) and shares a leucine residue at this position with two GPRs encoded by Herpes simplex virus six. Since also some mel-

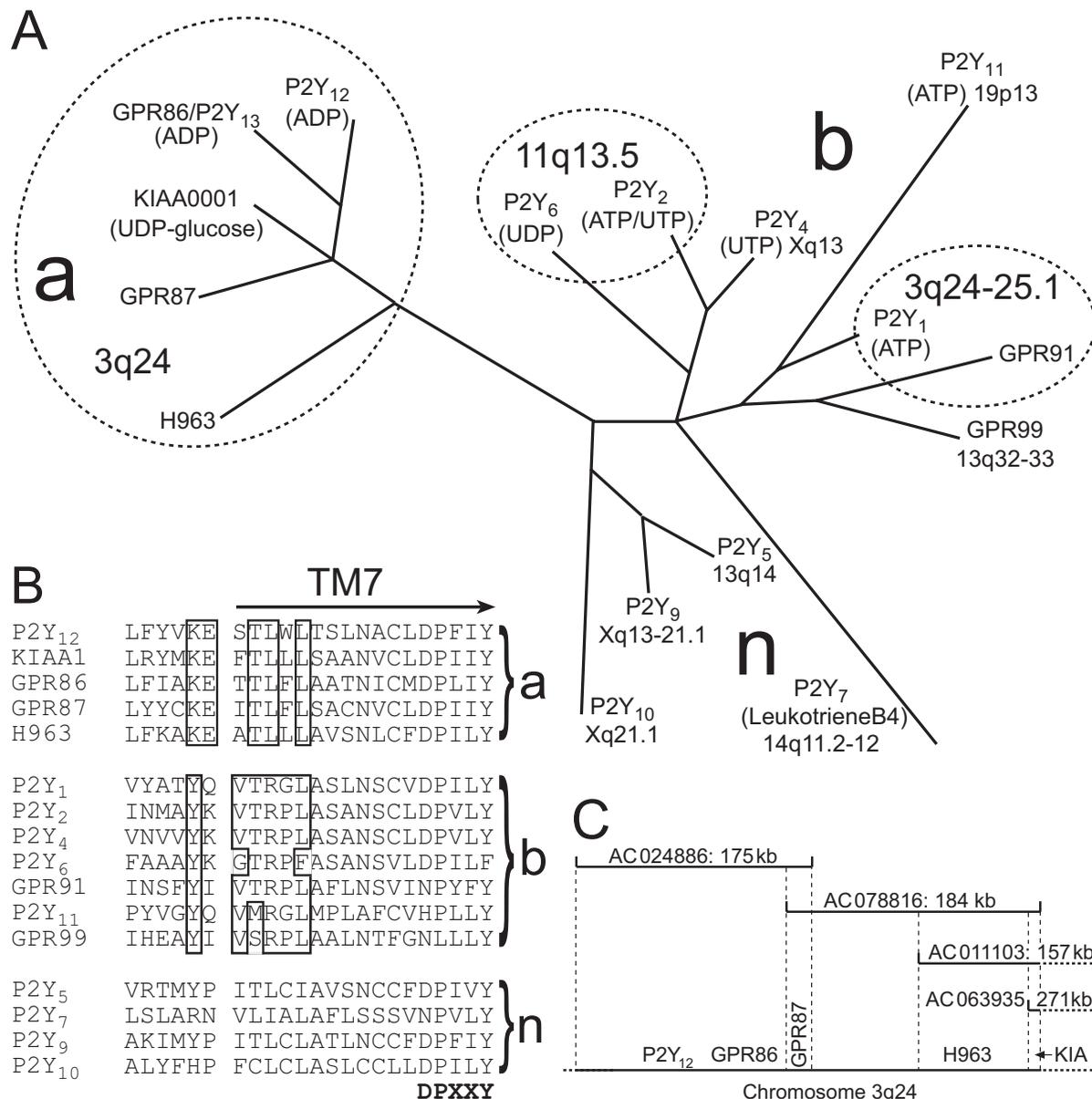


Figure 3
Phylogenetic analysis of human nucleotide GPRs. (A) Phylogenetic tree. The two nucleotide-receptor subgroups "a" and "b" and the related non-nucleotide receptors (n) are indicated. Chromosomal localizations and known endogenous ligands with highest affinities (in brackets) are indicated for each GPR. Phylogenetic analysis assigned a support value of at least 53% to each internal branch. (B) Alignment of putative nucleotide-binding motifs at the beginning of transmembrane domain seven. The proline-containing motif towards the end of transmembrane domain seven is indicated below the alignment. The accession numbers of the receptors are: GPR86, AAK01864; GPR87, AAK01858; GPR91, AF348078; H963, AAC51846; KIAA0001, BAA02791; P2Y₁, CAA07339; P2Y₂, XP_006367; P2Y₄, P51582; P2Y₅, P43657; P2Y₆, Q15077; P2Y₇, Q15722; P2Y₉, Q99677; P2Y₁₀, AAB57836; P2Y₁₁, AAB88674; P2Y₁₂, AAG48944. (C) Schematic diagram of the subgroup "a" gene cluster on chromosome 3q24. In the upper part the overlapping BAC clones, coding for the receptors indicated below, are given. The genes of P2Y₁₂, GPR86, and GPR87 are located in a head-to-tail configuration within a distance of 46.5 kb on one contig of AC024886. The GPR87 gene is also present on AC078816, which overlaps with AC024886 in 15 kb of its sequence. The H963 gene is encoded by AC078816 and AC011103, the KIAA0001 gene by AC078816, AC011103, and AC063935. From these data the order of the genes can be deduced. Gaps in the known sequence of the BAC clones coding for H963 and KIAA0001 make it impossible to determine the orientation and exact size of the intergenic regions.

In contrast to other nucleotide GPR genes, which are often found in clusters, we did not recognize further GPR genes close to GPR99. The chromosomal localization of the other nucleotide GPRs supports the proposed subgroup classification, since all genes of the subgroup "a" receptors are located within 355 kb on two overlapping BAC clones (AC024886 and AC078816) on chromosome 3q24 (Fig. 3C). The five subgroup "a" genes are, therefore, excellent examples for gene duplication events and divergent evolution of the resulting genes. The subgroup "b" genes are more scattered throughout the genome with clusters of P2Y₁ and GPR91 on chromosome 3q24-25.1, not far away from the subgroup "a" cluster, and of P2Y₂ and P2Y₆ on chromosome 11q13.5 [9] on the same BAC clone consisting of 191 kb (AP002761).

Functional assays

A full-length cDNA clone of GPR99 was constructed by ligating error-free PCR products. For expression in *Xenopus* oocytes this cDNA was cloned into pGemHE [10]. In vitro transcribed cRNA was injected together with the cRNA of a G-protein coupled inwardly rectifying potassium channel, and the oocytes were analyzed by whole cell clamp as described [11,12]. None of the applied nucleotide ligands (ATP, CTP, GTP, UTP, ADP, GDP, UDP, AMP, CMP, GMP, cAMP, cGMP, CMP-sialic acid, GDP-fucose, GDP-mannose, UDP-N-acetyl galactosamin, UDP-glucuronate, UDP-galactose, UDP-N-acetyl glucose) evoked stronger responses in GPR99-injected than in control oocytes. As an additional mammalian expression system we used CHO cells stably transfected with aequorin and the promiscuous G protein α subunit G _{α 16} [13]. These cells were transiently transfected with a GPR99-pcDNA3 construct. G-protein activation was analyzed by measuring the Ca²⁺-dependent luminescence of aequorin in a bioluminometer (Berthold). As in the oocyte system, the above mentioned nucleotides did not stimulate the emission of light in a GPR99-dependent manner. We conclude from this that the nucleotide derivatives assayed are either no agonists of GPR99 or that GPR99 can not activate the G proteins present in the oocyte and the CHO cells. Both systems are widely used for heterologous cell-surface expression of GPRs. Nevertheless, since no GPR99-specific antiserum was available, and since we did not add an epitope tag to GPR99, we can not prove cell-surface expression which is a prerequisite for the ligand assays used.

Conclusions

We describe the identification of GPR99 from human genomic data and its expression in kidney and placenta. With respect to a new subgroup classification for the P2Y subfamily based on sequence similarity, specific ligand-binding motifs, and gene clustering, we propose that GPR99 is a receptor for nucleotide ligands.

Materials and methods

Database searches and construction of evolutionary trees

BLASTN and TBLASTN searches of GenBank were performed using the National Center for Biotechnology Information server [14]. Amino-acid sequences were aligned with Clustal W [8]. Phylogenetic analysis was performed using the program PUZZLE [15], a support value of at least 53% was assigned to each internal branch.

Northern-blot analysis

A commercial human multiple-tissue northern blot (Clontech) was hybridized using a [³²P]-labeled 400 bp NotI-GPR99 fragment of the IMAGE clone 3014233 (AW827323) according to the manufacturer's instructions. The blot contained 1 μ g of poly-A⁺ RNA in each lane.

PCR

Based on the genomic sequence of hGPR99 an upstream primer containing the in-frame stop and start codon (AGA TGA AAG GAG ACA ACC ATG AAT G) and a downstream primer about 50 nucleotides 3' of the stop codon (CTT AGG ATG CTA GGT AAA GTA TCA GC) were designed. Full-length GPR99 was PCR amplified from placental cDNA prepared with the Marathon kit (Clontech) using a protocol with 30 cycles at an annealing temperature of 55°C and Taq polymerase.

To obtain an error-free GPR99 cDNA, the PCR products were cloned into the pGEM-T Easy vector (Promega) and a full-length clone was constructed by ligating error-free parts of two individual clones.

Ligand assays

For expression in *Xenopus* oocytes GPR99 was cloned into pGemHE [10]. In vitro transcribed cRNA was injected together with the cRNA of the G protein-coupled inwardly-rectifying potassium channel GIRK, and the oocytes were analyzed by whole cell clamp analysis [11,12].

As a mammalian expression system we used CHO cells stably transfected with aequorin and the promiscuous G protein α subunit G _{α 16} [13]. These cells were transiently transfected with GPR99 cloned into pcDNA3 (Invitrogen). Activation of G-proteins was analyzed by measuring the Ca²⁺-dependent luminescence of aequorin in a bioluminometer (Berthold).

Author's contributions

T. W. and S. H. carried out the phylogenetic and sequence analysis, A. M. performed the aequorin assay, and H.-J. K. the *Xenopus* oocyte assay. H.C.S. participated in the design of the study and W. H. initially identified and sequenced gpr99, took part in the ligand assays, and wrote the manuscript. All authors read and approved the final manuscript.

Acknowledgements

We thank P. Joost and A. Methner for discussions and S. Hempel for help with the figures. This work was supported by the Deutsche Forschungsgemeinschaft (SFB 444 and 545).

References

1. Bockaert J, Pin JP: **Molecular tinkering of G protein-coupled receptors: an evolutionary success.** *EMBO J* 1999, **18**:1723-1729
2. Wittenberger T, Schaller HC, Hellebrand S: **An expressed sequence tag (EST) data mining strategy succeeding in the discovery of new G-protein coupled receptors.** *J Mol Biol* 2001, **307**:799-813
3. Lennon GG, Auffray C, Polymeropoulos M, Soares MB: **The I.M.A.G.E. Consortium: An integrated molecular analysis of genomes and their expression.** *Genomics* 1996, **33**:151-152
4. **GPR fingerprint analysis.** [<http://www.bioinf.man.ac.uk/finger-PRINTScan/>]
5. Ralevic V, Burnstock G: **Receptors for purines and pyrimidines.** *Pharmacol Rev* 1998, **50**:413-492
6. Communi D, Gonzalez NS, Detheux M, Brezillon S, Lannoy V, Parmentier M, Boeynaems JM: **Identification of a novel human ADP receptor coupled to G(i).** *J. Biol. Chem.* 2001, **276**:41479-41485
7. Jiang Q, Guo D, Lee BX, Van Rhee AM, Kim YC, Nicholas RA, Schachter JB, Harden TK, Jacobson KA: **A mutational analysis of residues essential for ligand recognition at the human P2Y1 receptor.** *Mol Pharmacol* 1997, **52**:499-507
8. **Ensembl contig view tool.** [<http://www.ensembl.org>]
9. Somers GR, Hammet F, Woollatt E, Richards RI, Southey MC, Venter DJ: **Chromosomal localization of the human P2y6 purinoceptor gene and phylogenetic analysis of the P2y purinoceptor family.** *Genomics* 1997, **44**:127-130
10. Liman ER, Tytgat J, Hess P: **Subunit stoichiometry of a mammalian K⁺ channel determined by construction of multimeric cDNAs.** *Neuron* 1992, **9**:861-871
11. Kreienkamp H-J, Hönck H-H, Richter D: **Coupling of rat somatostatin receptor subtypes to a G-protein gated inwardly rectifying potassium channel (GIRK).** *FEBS-Lett.* 1997, **419**:92-94
12. Birgül N, Weise C, Kreienkamp H-J, Richter D: **Reverse physiology in Drosophila: identification of a novel allatostatin-like neuropeptide and its cognate receptor structurally related to the mammalian somatostatin/galanin/opioid receptor family.** *EMBO J* 1999, **18**:5892-5900
13. Stables J, Green A, Marshall F, Fraser N, Knight E, Sautel M, Milligan M, Lee M, Rees S: **A bioluminescent assay for agonist activity at potentially any G-protein-coupled receptor.** *Anal Biochem* 1997, **252**:115-126
14. **NCBI BLAST search.** [www.ncbi.nlm.nih.gov/BLAST]
15. Strimmer K, von Haeseler A: **Quartet puzzling: a quartet maximum likelihood method for reconstructing tree topologies.** *Mol Biol Evol* 1996, **13**:964-969

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMedcentral will be the most significant development for disseminating the results of biomedical research in our lifetime."

Paul Nurse, Director-General, Imperial Cancer Research Fund

Publish with **BMC** and your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours - you keep the copyright



BioMedcentral.com

Submit your manuscript here:

<http://www.biomedcentral.com/manuscript/>

editorial@biomedcentral.com