

Research Article

Antimetastatic Potential of Rhodomyrtone on Human Chondrosarcoma SW1353 Cells

Malatee Tayeh¹ and Ramida Watanapokasin ²

¹Department of Medical Technology, School of Allied Health Sciences, Walailak University, Nakhon Si Thammarat 80160, Thailand

²Department of Biochemistry, Faculty of Medicine, Srinakharinwirot University, Bangkok 10110, Thailand

Correspondence should be addressed to Ramida Watanapokasin; ramidabc@gmail.com

Received 24 February 2020; Revised 9 June 2020; Accepted 8 July 2020; Published 27 July 2020

Academic Editor: Mario Ledda

Copyright © 2020 Malatee Tayeh and Ramida Watanapokasin. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Chondrosarcoma is primary bone cancer, with the forceful capacity to cause local invasion and distant metastasis, and has a poor prognosis. Cancer metastasis is a complication of most cancers; it is one of the leading causes of cancer-related death. Rhodomyrtone is a pure compound that has been shown to induce apoptosis and antimetastasis in skin cancer. However, the inhibitory effect of rhodomyrtone on human chondrosarcoma cell metastasis is largely unknown. Effect of rhodomyrtone on cell viability in SW1353 cell was determined by MTT assay. Antimigration, anti-invasion, and antiadhesion were carried out to investigate the antimetastatic potential of rhodomyrtone on SW1353 cells. Gelatin zymography was performed to determine matrix metalloproteinase-2 (MMP-2) and MMP-9 activities. The effect of rhodomyrtone on the underlying mechanisms was performed by Western blot analysis. The results demonstrated that rhodomyrtone reduced cell viability of SW1353 cells at the low concentration ($<3 \mu\text{g/mL}$); cell viability was $>80\%$. Rhodomyrtone at the subcytotoxic concentrations (0.5, 1.5, and $3 \mu\text{g/mL}$) significantly inhibited cell migration, invasion, and adhesion of SW1353 cells in a dose-dependent fashion. Protein expression of integrin αv , integrin β3 , and the downstream migratory proteins including focal adhesion kinase (FAK) and the phosphorylation of serine/threonine AKT, Ras, RhoA, Rac1, and Cdc42 were inhibited after treatment with rhodomyrtone. Moreover, we found that rhodomyrtone decreased the protein level of MMP-2 and MMP-9 as well as the enzyme activity in SW1353 cells. Meanwhile, tissue inhibitor of metalloproteinase-1 (TIMP-1) and TIMP-2 expression was increased in a dose-dependent fashion. Besides, rhodomyrtone dramatically inhibited the expression of growth factor receptor-bound protein-2 (GRB2) and the phosphorylated form of extracellular signal regulation kinase1/2 (ERK1/2) and c-Jun N-terminal kinase1/2 (JNK1/2). These results indicated that rhodomyrtone inhibited SW1353 cell migration, invasion, and metastasis by suppressing integrin $\alpha\text{v}\beta\text{3}$ /FAK/AKT/small Rho GTPases pathway as well as downregulation of MMP-2/9 via ERK and JNK signal inhibition. These findings indicate that rhodomyrtone possessed the antimetastasis activity that may be used for antimetastasis therapy in the future.

1. Introduction

Chondrosarcoma, a malignant bone cancer, is the most frequent type of primary bone cancer [1]. Conventional chondrosarcoma is the most frequent all chondrosarcoma (80–90%). It can arise in any bone, but the most common sites are femur, humerus, tibia, pelvis, and ribs. It is more commonly seen in adults aged 20 to 60 years [2]. The metastatic capacity of conventional chondrosarcoma correlates with histological grading, about 15% of all metastatic

disease-related deaths occur more than 5 years after diagnosis [3]. High-grade chondrosarcomas comprise grades II and III chondrosarcoma that are very aggressive and frequently metastasize to the lung [4]. They are highly insensitive to radiotherapy or chemotherapy; thereby the surgical resection remains the primary method of therapy [5]. Cancer metastasis is the principal cause of mortality in cancer patients. Cancer metastasis consists of multiple steps involving the spread of tumor cells from the original site, migration and invasion through the basement membrane,

survival in the circulatory system, and proliferation into a secondary site [6]. Integrins are primary receptors for cellular adhesion, mediating dynamic cell-cell and cell-extracellular matrix interactions [7]. They are a heterodimeric transmembrane glycoprotein which formed from different combinations of 18 α -subunits and 8 β -subunits that can associate to form 24 unique integrin heterodimers [8]. They have been implicated in the metastasis of ovarian, pancreatic, lung, bladder, osteosarcoma, and chondrosarcomas [9–14]. Overexpression of $\alpha 2\beta 1$, $\alpha v\beta 1$, $\alpha v\beta 3$, $\beta 1$, and $\beta 1$ integrin in human chondrosarcoma is associated with tumor progression, metastasis, and poor prognosis [5, 8, 15, 16]. Integrins initiate transmembrane signaling through activation of the downstream effectors such as FAK, PI3K/AKT, and Rho GTPases family for cancer cell migration [8, 17–19]. Small Rho GTPases including RhoA, Rac1, and Cdc42 are well known to regulate cell migration [20], and overexpression of these molecules in human cancer correlates with cancer progression [21]. There are many reports demonstrating that overexpression of RhoA associated with various cancers' progression such as testicular cancer, breast cancer, colon, lung cancers, and head and neck squamous cell carcinoma [22–24]. Rac1 is overexpressed in various cancers such as testicular cancer [22], breast cancer [23], and leukemia [25]. Furthermore, overexpression of Cdc42 has been reported to be in several human cancers, such as breast, non-small cell lung cancer, and melanoma [23, 26, 27]. Besides, several studies indicated that the expression of matrix metalloproteinases (MMPs) is regulated by integrin signaling [28, 29]. MMP-2 and MMP-9 are the important proteases involved in cancer cell migration, spreading, invasion, and metastasis [30]. They are highly expressed in various malignant tumors, and activation of these enzymes enables the degradation of ECM by cancer cells allowing their access to the vasculature as well as migration and invasion into the target organ and development of cancer metastasis [31–33].

Rhodymyrtus tomentosa, a medicinal plant, is native to Southern and Southeast Asia, from India, Sri Lanka, east to southern China, Hong Kong, Taiwan, Philippines, Thailand, south to Malaysia and Sulawesi and USA [34]. It is used as a traditional medicine for the treatment of many diseases such as anti-inflammation, antidiarrheal, laxative, carminative, and urinary tract infections [35, 36]. Rhodymyrtone is a pure compound isolated from the leaves of this medicinal plant species. It exhibited antibacterial activities, immunomodulatory activity, antiproliferation, delayed wound closure, and induced apoptosis in HaCaT cells [37–42]. Our recent studies demonstrated that rhodymyrtone induced apoptosis in skin cancer [43]. It was also reported as antimetastasis of human skin cancer cells [44]. However, the effect of rhodymyrtone on the metastasis in chondrosarcoma cells is largely unknown. In this study, molecular mechanisms underlying the antimetastasis effect of rhodymyrtone in SW1353 cells were investigated in vitro to determine the potential of rhodymyrtone as a novel chemotherapeutic drug.

2. Materials and Methods

2.1. Chemicals and Reagents. Rhodymyrtone was provided by Dr. Wilawan Mahabusarakum, Department of Chemistry,

Faculty of Science, Prince of Songkla University. Dimethyl sulfoxide (DMSO), ethylenediaminetetraacetic acid (EDTA), gelatin A, β -mercaptoethanol, and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Matrigel Matrix was obtained from Corning Incorporated (Bedford, MA, USA). Coomassie Brilliant Blue R-250 and protein assay kit were purchased from Bio-Rad Labs (Hercules, CA, USA). Rabbit monoclonal anti-MMP-2, -MMP-9, -TIMP-1, -TIMP-2, -RAS, -GRB2, ERK1/2, p-ERK1/2, p38, p-p38, JNK1/2, pJNK1/2, FAK, AKT, pAKT (Ser473), pAKT (Thr308), NF- κ B, RhoA, Rac1, CDC42, Integrin αv , integrin $\beta 3$ and β -actin were purchased from Cell Signaling Technology (Danvers, MA, USA).

2.2. Cell Culture. Chondrosarcoma cell line SW1353 (ATCC, Manassas, VA, USA) initiated from a primary grade II chondrosarcoma of the right humerus from a 72-year-old Caucasian female. SW1353 was cultured in DMEM (Gibco, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (FBS) (GE, Healthcare, Life Science, Little Chalfont, UK) and antibiotic (100 U/mL of penicillin and 100 μ g/mL of streptomycin) (GE, Healthcare, Life Science, Little Chalfont, UK). The cells were incubated at 37°C in a humidified atmosphere of 5% (v/v) CO₂.

2.3. Cell Viability Assay. Cell viability was analyzed using MTT assay as previously described [43]. Briefly, 3×10^3 cells of SW1353 were seeded in 96-well plates and incubated with or without rhodymyrtone (0–5 μ g/mL) for 0, 6, 12, and 24 h. DMSO was added for dissolving the formazan crystals. Absorbance was detected at a wavelength of 570 nm by an Epoch™ Microplate spectrophotometer. Percentages of cell viability (%) were determined by comparing with the untreated control.

2.4. Apoptotic Cells Assay. SW1353 cells were seeded in a 6-well plate at a density of 1.5×10^5 cells/well. After treatment with or without rhodymyrtone (0–5 μ g/mL) for 24 h, the treated cells were washed with phosphate-buffered saline (PBS), fixed with 3.7% paraformaldehyde, and stained with Hoechst 33342 for 15 min. The nuclear morphological changes were observed under a fluorescence microscope IX73 model (Olympus) with an ultraviolet filter.

2.5. Scratch Assay. Scratch assay was performed for detecting the effect of rhodymyrtone on SW1353 cell migration, as described in the previous report [45]. Briefly, cells were grown to 90% confluence in 6-well plates and the wound areas were created with a sterile pipette tip. The cellular debris was washed with PBS, and cells in each well were treated with or without rhodymyrtone (0–3 μ g/mL) for 24 h. The wound closure was observed and captured with a phase-contrast inverted microscope (Olympus, Tokyo, Japan). The wound area was measured by NIH ImageJ software, version 1.46r. The experiments were repeated three times.

2.6. Transwell Migration Assay. The antimigration potential of rhodomyrtone was assayed by the transwell chamber in 24-well plates. Cells were treated with or without rhodomyrtone (0–3 $\mu\text{g/mL}$) for 24 h. After incubation, the migrated cells were fixed and stained. The percentage of the migratory cells for each treatment was calculated as previously described [43].

2.7. Cell Invasion Assay. The ability of cancer cells to invade through the membrane coated with 30 μg of Matrigel (Corning incorporated, Bedford, MA, USA) was determined by the Boyden chamber assay as previously described [43]. Briefly, SW1353 cells were pretreated with or without rhodomyrtone (0–3 $\mu\text{g/mL}$) for 24 h. Cells in serum-free medium were plated onto the upper side of the chamber, while the serum-containing medium (10% FBS) was added into the lower side of the chamber. After 24 h of incubation, the invaded cells were fixed, stained, and photographed under an inverted microscope. The data are presented as percentage of cell invasion from five random fields. The experiments were repeated three times and each experiment was carried out in duplicate.

2.8. Cell Adhesion Assay. The adhesion ability of SW1353 cells on the Matrigel-coated 96-well plate was performed according to the previous method [43]. Percentage (%) of cell adhesion was calculated by comparing with the untreated control. The experiments were carried out in triplicate and three independent experiments were performed.

2.9. Gelatin Zymography. Gelatin zymography was performed for determining the effect of rhodomyrtone on MMP-2/9 activities in culture media. The conditioned media were collected and subjected to gelatin containing SDS-PAGE (0.1% gelatin). The gelatinolytic activity was analyzed as in our previous research [43].

2.10. Western Blot Analysis. The Western blotting analysis was performed to investigate the metastasis-related proteins. Proteins were mixed with sample buffer and boiled for 10 min. Denatured proteins were separated in the 8–12% SDS-PAGE and blotted onto a PVDF membrane (Millipore Corporation, Billerica, MA, USA). After that, the membranes were immunoblotted with an appropriate dilution of the specific primary antibody, followed by incubation with the appropriate secondary antibody conjugated with horseradish peroxidase (Cell Signaling Technology, Danvers, MA). The protein bands were detected by using Immobilon™ Western Chemiluminescent HRP Substrate (Merck KGA, Darmstadt, DE) and exposed to Chemiluminescent Imaging System (GeneGenome Gel Documentation, Synoptics Ltd., Cambridge, UK). The relative density was quantified as in our previous research [43].

2.11. Statistical Analysis. Data were reported as mean \pm standard deviation (SD) of three independent experiments. Statistical significance was analyzed by a one-way

ANOVA test. The differences between treatment groups and the untreated groups were considered statistically significant at $p < 0.05$. SPSS 17.0 software was carried out for statistical analyses.

3. Results and Discussion

3.1. Rhodomyrtone at Low Concentrations Did Not Affect Cell Viability in SW1353 Cells. Our previous study demonstrated that rhodomyrtone inhibited cell growth and induced apoptosis in skin cancer cells [44]. In this study, we investigated whether rhodomyrtone suppressed cell viability in human chondrosarcoma SW1353 cells. MTT assay was performed to determine the cell viability and cell proliferation effect of rhodomyrtone on SW1353 cells. Figure 1(a) shows that rhodomyrtone suppressed SW1353 cell viability in a dose- and time-dependent fashion. Rhodomyrtone reduced cell viability of SW1353 cells at the high concentration ($>3 \mu\text{g/mL}$) but at the lower concentration (0–3 $\mu\text{g/mL}$) cell viability was $>80\%$. Furthermore, we investigated the nuclear morphological changes of SW1353 cells after treatment with rhodomyrtone (0–5 $\mu\text{g/mL}$). Figure 1(b) shows that no apoptotic cells and nuclear morphological changes were detected after treatment with rhodomyrtone at concentration ranges 0–3 $\mu\text{g/mL}$. Likewise, our previous study found that rhodomyrtone could inhibit A431 cell proliferation at high concentrations while at the lower concentration it did not [44]. Therefore, rhodomyrtone at concentration range (0–3 $\mu\text{g/mL}$) was selected for subsequent experiments. This result indicates that rhodomyrtone at higher concentrations inhibited cell viability, but at the concentrations ranging from 0–3 $\mu\text{g/mL}$ it did not.

3.2. Rhodomyrtone at Subcytotoxic Concentrations Inhibited SW1353 Cell Migration. Cancer metastasis is the predominant cause of cancer-related death. Cell migration, invasion, and adhesion are key steps in tumor progression [6]. Disruption of any metastatic steps is a target for preventing the development of cancer metastases. In our previous study, rhodomyrtone has been reported to suppress skin cancer metastasis by inhibiting cell migration [43]. Thus, we determined the effect of rhodomyrtone on cancer cell migration in SW1353 cells. Wound healing and the transwell chamber assay were used to investigate the antimigration effect of rhodomyrtone in SW1353 cells. Rhodomyrtone has been shown to significantly reduce cell migration to the wound area after treatment with rhodomyrtone at the subcytotoxic concentrations (0.5, 1.5, and 3 $\mu\text{g/mL}$) as shown in Figures 2(a) and 2(b). Cell migration rate was reduced to $41.4 \pm 6\%$, $35.8 \pm 9\%$, and $17.3 \pm 6\%$, respectively. To confirm the antimigration potential of rhodomyrtone in SW1353 cells, a transwell chamber assay was performed. The result showed cell migration was significantly suppressed by rhodomyrtone in a concentration-dependent manner as shown in Figures 2(c) and 2(d). The inhibition of SW1353 cell migration by rhodomyrtone was independent of the direct cytotoxic effects on cancer cells.

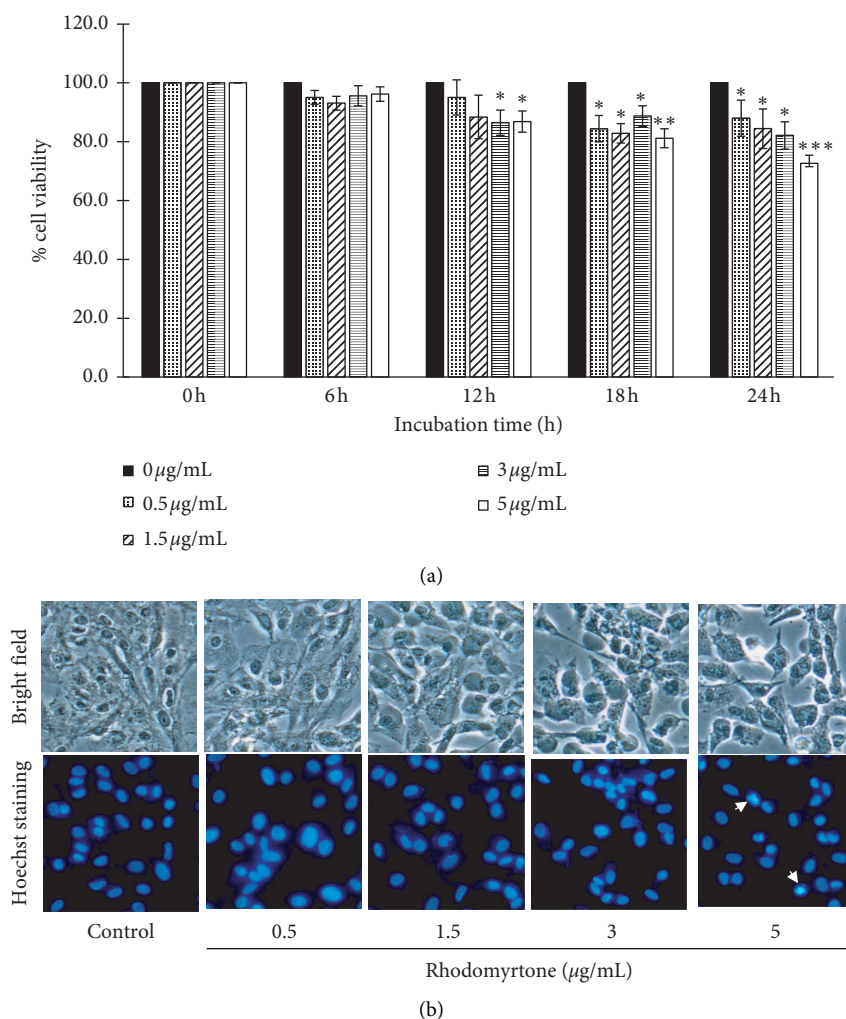


FIGURE 1: Effect of SW1353 cell viability by rhodomyrtone. (a) Cell viability was analyzed by MTT assay for 24 h. Rhodomyrtone inhibited SW1353 cell growth in a time- and concentration-dependent manner. (b) Nuclear morphology changes were detected by Hoeschst 33342 staining. Rhodomyrtone at 0–3 µg/mL (cell viability >80%) was selected for subsequent experiments. The arrows indicate apoptotic cells. Data were expressed as mean \pm standard deviation (SD) from three independent experiments.

3.3. Rhodomyrtone at the Subcytotoxic Concentrations Inhibited SW1353 Cell Invasion and Adhesion. To explore the effect of rhodomyrtone on cancer cell metastasis in SW1353 cell, we investigated the inhibition of SW1353 cell invasion by rhodomyrtone using Matrigel-coated Boyden chamber assay. Figures 3(a) and 3(b) show that rhodomyrtone reduced the SW1353 cell invasion in a concentration-dependent manner ($***p < 0.001$). The percentage of invaded cells was $48.2 \pm 4.4\%$, $46.4 \pm 10.1\%$, and $43.9 \pm 2.9\%$ when treated with 0.5, 1.5, and 3 µg/mL of rhodomyrtone, respectively. Also, the inhibitory effect of rhodomyrtone on cell adhesion of SW1353 cells was performed by MTT assay. The result demonstrated that cell adhesion ability of SW1353 cells to Matrigel-coated wells could reduce at $84.8 \pm 9.8\%$, $76.0 \pm 8.2\%$, and $64.0 \pm 7.1\%$ after treatment by 0.5, 1.5, and 3 µg/mL of rhodomyrtone (Figure 3(c)), respectively. These results indicated that rhodomyrtone inhibits chondrosarcoma cells metastasis without apparent cytotoxicity. Likewise, our previous study has also reported that

rhodomyrtone at the nontoxic concentrations exhibited strong inhibition on human skin cancer cell metastasis by reducing cell migration, cell invasion, and cell adhesion ability [43].

3.4. Rhodomyrtone at the Subcytotoxic Concentrations Inhibited the Expression and Activity of MMP-2 and MMP-9. Previous reports showed MMP-2 and MMP-9 expression was correlated with cancer invasion and the upregulation of MMPs was observed in invasive cancer cells [46–48]. The inhibition of MMP-2 and MMP-9 enzyme activity and protein expression has been shown to inhibit cancer cell migration and invasion in many types of tumor cells [49–52]. In this study, we investigated the expression and activity of MMP-2 and MMP-9 after treatment with rhodomyrtone at low concentrations. Gelatin zymography was performed to determine the activity of MMP-2 and MMP-9. The result demonstrated that rhodomyrtone significantly

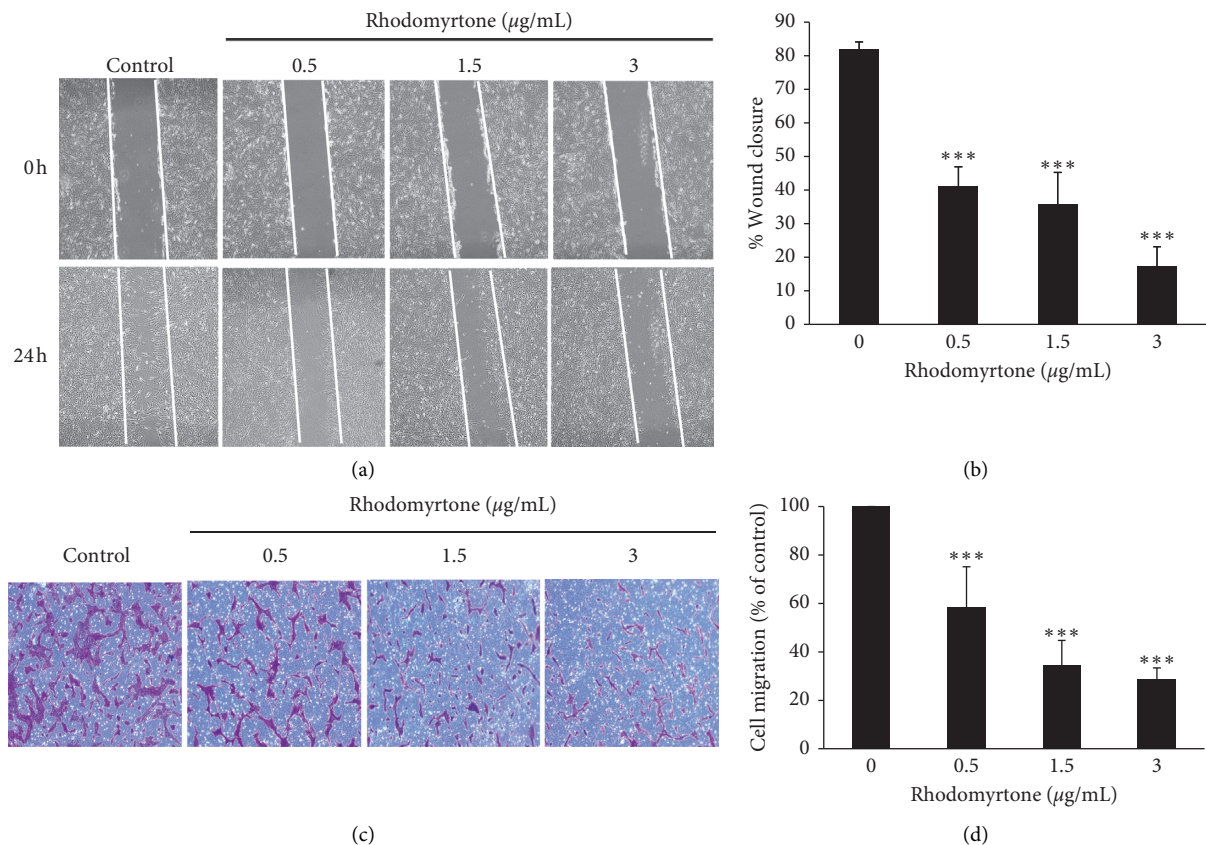


FIGURE 2: Antimigration of rhodomyrtone was determined by wound healing and transwell chamber assay. (a) The wound areas were created by a sterile micropipette tip and treated with or without the subcytotoxic concentrations of rhodomyrtone for 24 h. Cell migration of SW1353 cells was captured under a 40x magnification of the microscope. (b) The wound closure was analyzed by measuring the area between the edges (white lines indicated the wound edge). (c) Rhodomyrtone also significantly suppressed cell migration in a dose-dependent manner as detected with the transwell chamber. (d) Quantitative analysis of the migratory cells was calculated by NIH ImageJ. Data represent mean \pm standard deviation (SD) from three independent experiments. *** $p < 0.001$ vs. untreated control.

reduced the activity of MMP-2 and MMP-9 in a concentration-dependent manner as shown in Figures 4(a) and 4(b). The protein expression of MMP-2 and MMP-9 was determined by Western blot analysis. The result showed MMP-2 and MMP-9 protein expression was significantly suppressed by rhodomyrtone as compared to the untreated control as shown in Figures 4(c) and 4(d). These results revealed that the rhodomyrtone inhibited both MMP-2 and MMP-9 activities and protein expression in SW1353 cells. Thus, inhibition of MMPs activities and protein expression is the target for preventing cancer metastases. This is consistent with previous reports, showing that resveratrol attenuated MMP-9 and MMP-2 regulated differentiation of HTB94 cells [52]. Some studies demonstrated that curcumin and curcumin derivative inhibited cancer cell invasion through the downregulation of MMPs in human A549 lung cancer cells [53], MDA-MB-231 human breast cancer cells [54], MCF-7 cells [55], and hepatocellular carcinoma [56].

3.5. Rhodomyrtone at the Subcytotoxic Concentrations Induced the Expression Endogenous Inhibitor of MMP-2 and MMP-9. In this study, we found that the activities of MMP-2 and MMP-9 were inhibited by rhodomyrtone. The activities of

MMPs are specifically inhibited by a group of tissue inhibitors of metalloproteinases (TIMPs); TIMP-1 and TIMP-2 have been known to interact with MMP-9 and MMP-2, respectively. Several studies reported that overproduction of TIMPs can reduce metastasis whereas a low level of TIMPs correlates with tumor progression [17, 57, 58]. In this research, the expression of TIMP-1 and TIMP-2 was analyzed by Western blot analysis. We found that rhodomyrtone significantly increased TIMP-1 and TIMP-2 protein expression in a concentration-dependent manner (Figures 5(a) and 5(b)). This is consistent with our previous study which showed that rhodomyrtone reduced A431 cell metastasis by suppressing MMP-2/9 activities and increasing the expression of TIMP-1 and TIMP-2 [43]. Similarly, Ferrari and colleagues showed that the upregulation of TIMP-1 by genipin could inhibit MMP-2 activity and suppressed the metastasis of HepG2 and MHCC97 L cells [57]. Likewise, the inhibition of A549 cell metastasis by increasing TIMP-2 expression [58]. This result indicated that the enzyme activities of MMP-2 and MMP-9 were inhibited by their endogenous inhibitors.

3.6. Rhodomyrtone at the Subtoxic Concentrations Down-regulated Integrin α v, Integrin β 3, FAK, AKT, and Rho GTPase Signaling Pathway. Western blot analysis was performed

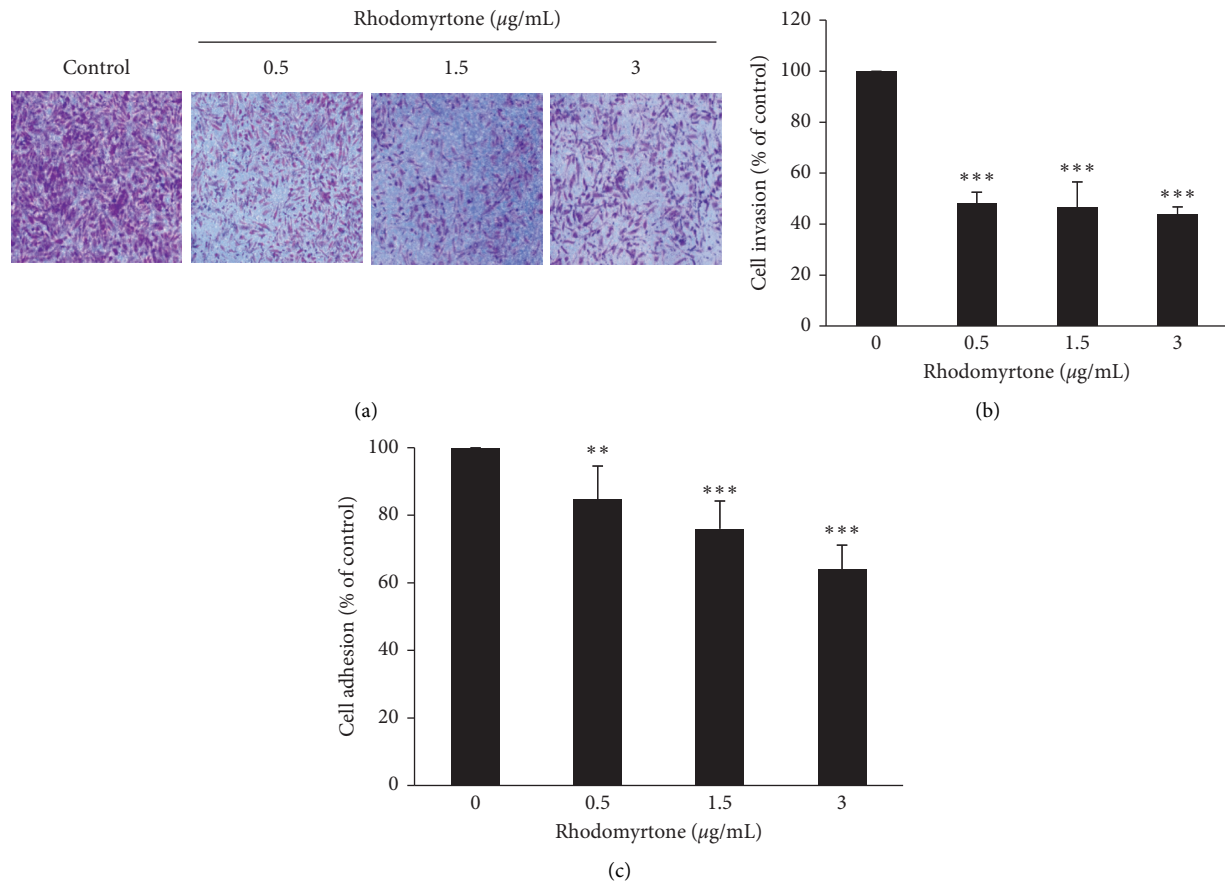


FIGURE 3: Cell invasion and adhesion assay. (a) Anti-invasion activity of rhodomymrtone was analyzed by the Matrigel-coated Boyden chamber assay. The cells invaded to the lower side of the chamber were stained and photographed under a 40x magnification of the microscope. (b) Percentage of invaded cells showed significant decrease after treatment with rhodomymrtone in a concentration-dependent manner. (c) Cells adhesion ability of treated cells was markedly inhibited after treatment with rhodomymrtone. The values are presented as mean \pm standard deviation (SD) from three independent experiments. ** $p < 0.01$ and *** $p < 0.001$ vs. untreated control.

to clarify the molecular mechanism of rhodomymrtone on human chondrosarcoma cell metastasis. Integrins are cellular receptors that associate the extracellular matrix to intracellular signaling molecules and regulate many cellular processes, including cell migration, adhesion, survival, growth, and differentiation [59, 60]. Integrin $\alpha\beta3$ has been reported to encourage cancer cell survival, migration, invasion, and angiogenesis in human chondrosarcoma [16]. Therefore, inhibition of integrin $\alpha\upsilon$ and integrin $\beta3$ expression is the good target for the metastasis treatment in human chondrosarcoma. We investigated whether rhodomymrtone could inhibit the expression of these metastasis-associated integrins. We found that rhodomymrtone at the subcytotoxic concentrations (0.5, 1.5, and 3 $\mu\text{g/mL}$) significantly reduced integrin $\alpha\upsilon$ expression in a dose-dependent manner but not much on integrin $\beta3$ (Figure 6(a)). This finding indicates that rhodomymrtone inhibited SW1353 cell metastasis by modulating integrin $\alpha\upsilon$ and integrin $\beta3$. This is consistent with that of Wu and colleagues who showed berberine suppressed chondrosarcoma cell migration by inhibiting the $\alpha\beta3$ integrin expression [61]. Moreover, we further evaluated the downstream effectors of integrin including

focal adhesion kinase (FAK), AKT, and p-AKT and small Rho GTPases family. There are many reports indicating that integrins initiate transmembrane signaling through activation of FAK and activate the downstream effectors such as AKT and Rho GTPases families for cancer cell migration [8, 17–19]. Rho GTPases family including Ras, RhoA, Rac1, and Cdc42 are well known to regulate cytoskeleton remodeling, cell migration, malignant transformation, cell polarity, invasion, and metastasis [20]. Ras and Rac-1 regulate lamellipodia formation, Cdc42 stimulates plasma membrane protrusion filopodia, and RhoA promotes stress fiber formation [19, 55, 62, 63]. Western blot analysis showed that rhodomymrtone significantly suppressed FAK expression and the downstream effectors including total AKT, phosphorylated AKT, Ras, RhoA, Rac1, and Cdc42 in SW1353 cell (Figures 6(c) and 6(f)).

Consistent with the effect of artonin E on migration and invasion inhibition of lung cancer cells via suppression of activated FAK, activated AKT, and CDC42 [62], *Coptidis Rhizoma* inhibits hepatic carcinoma cell migration by inactivation of Rho signaling [64]. Likewise, inhibition of Rac1 in MDA-MB-231 cells was shown to inhibit lamellipodia formation and, subsequently, cellular

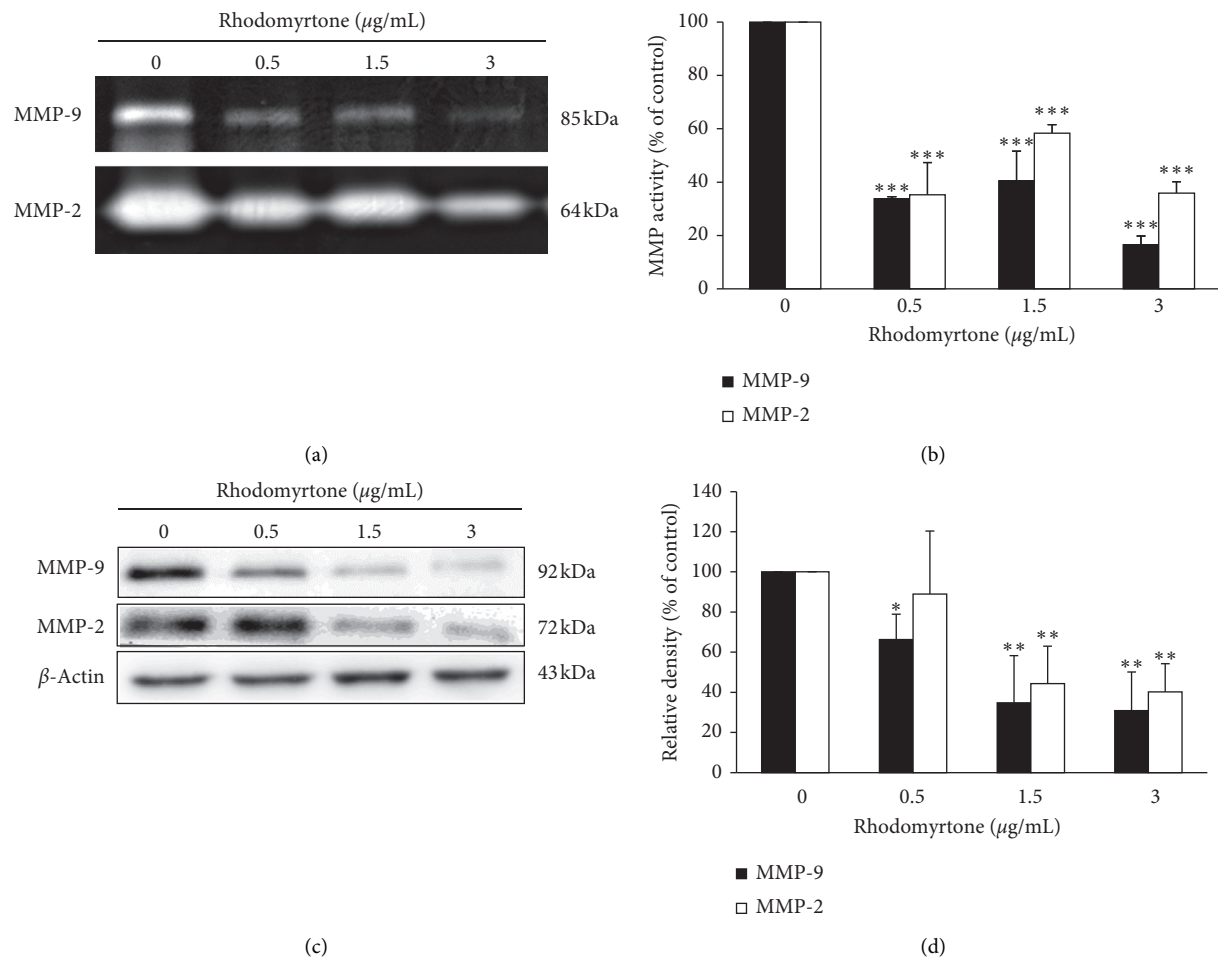


FIGURE 4: Effect of rhodomyrtone on MMP-2 and MMP-9 activities and protein expression. (a) Photograph presented the gelatinolytic activity of MMP-2 and MMP-9. (b) Quantitative analysis of MMP-2 and MMP-9 activities was calculated using NIH ImageJ. (c) Expression of MMP-2 and MMP-9 proteins was detected by using the specific antibodies. (d) Protein levels of MMP-2 and MMP-9 were significantly suppressed by rhodomyrtone in a concentration-dependent manner. Data are presented as mean \pm standard deviation (SD) from three independent experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. untreated control.

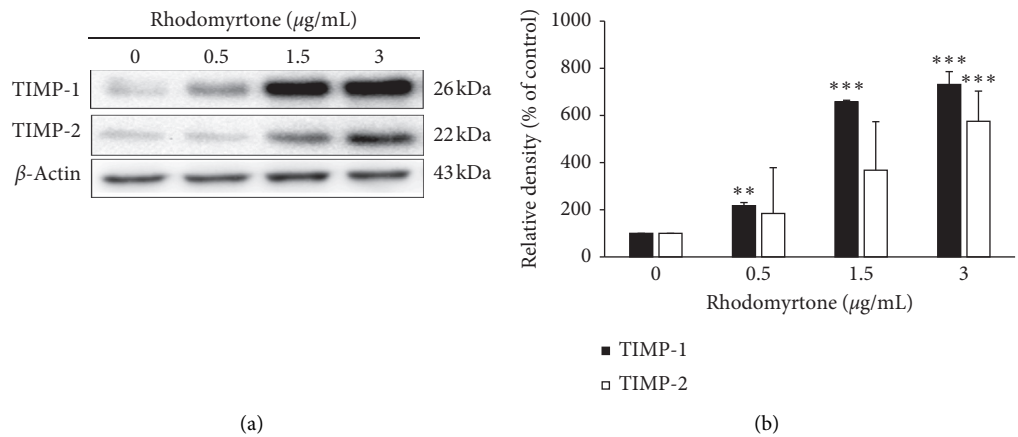


FIGURE 5: Western blot analysis of TIMPs protein level in SW1353 cells. (a) Expression of TIMP-1 and TIMP-2 protein was detected. (b) Rhodomyrtone significantly increased the expression of TIMP-1 and TIMP-2 protein levels. Data are presented as mean \pm standard deviation (SD) from three independent experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. untreated control.

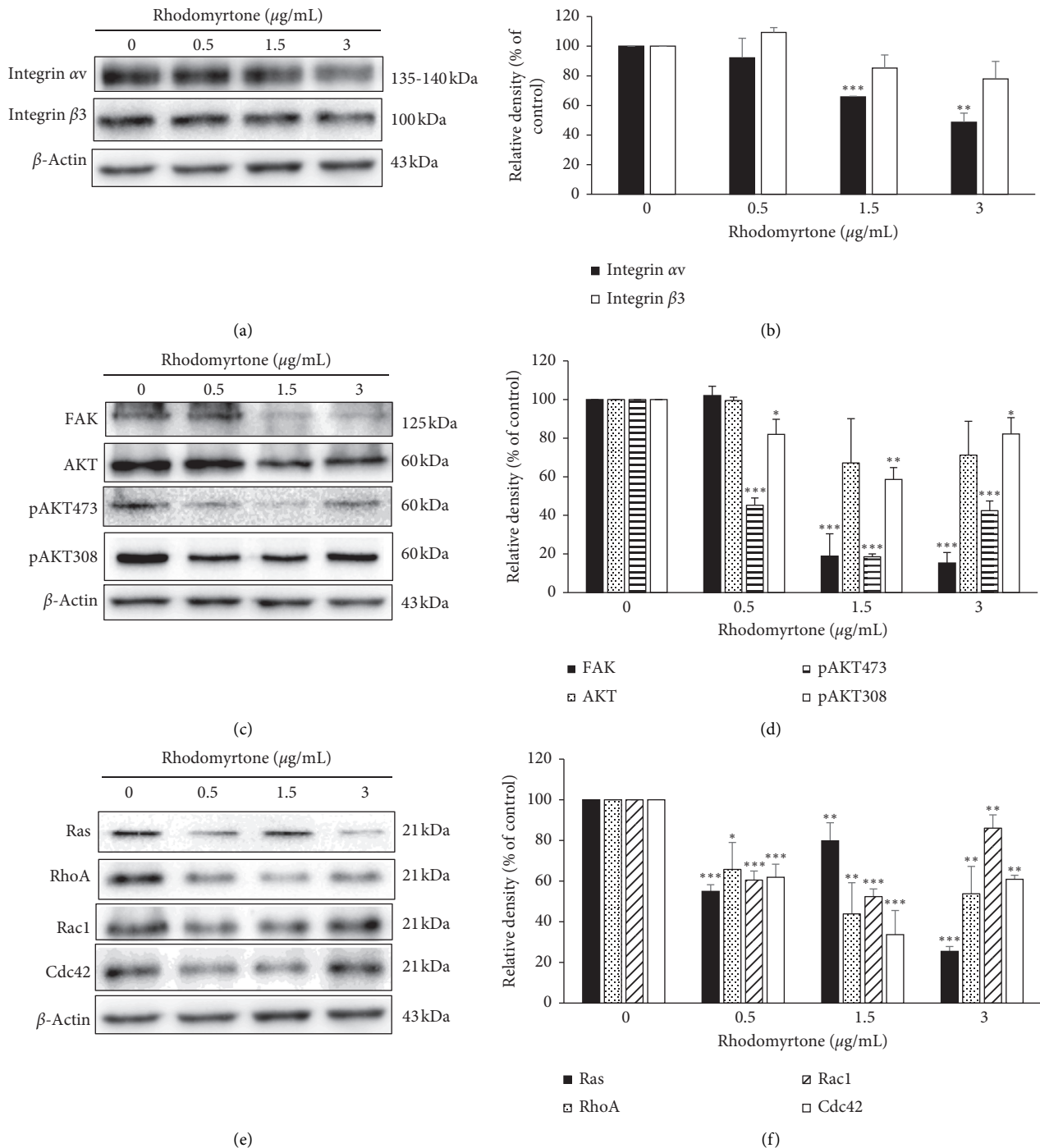


FIGURE 6: Western blot analysis of integrin αv , integrin $\beta 3$ protein levels, and the effector proteins. (a) Western blotting analysis of $\alpha v\beta 3$ integrin detected by using specific antibodies. β -Actin was used for equal loading. (b) Relative density of proteins was significantly inhibited after treatment with rhodomlyrtone. (c) FAK, AKT, pAKT (Ser473), and pAKT (Thr308) were detected by using specific antibodies and β -actin is used as the loading control. (d) The relative density of proteins was significantly inhibited after treatment with rhodomlyrtone in a concentration-dependent manner. (e) Specific antibodies were used for detecting Ras, RhoA, Rac1, and cdc42. β -Actin was used for equal loading. (f) The relative density of proteins was significantly inhibited after treatment with rhodomlyrtone. Data are represented as mean \pm standard deviation (SD) from three independent experiments. ** $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. the untreated control.

migration [65]. These results suggested that rhodomlyrtone inhibited cell migration by significantly suppressing integrin αv , but not much on integrin $\beta 3$. The

downstream migratory proteins of FAK, AKT, and Rho GTPase signaling pathways were also suppressed by rhodomlyrtone.

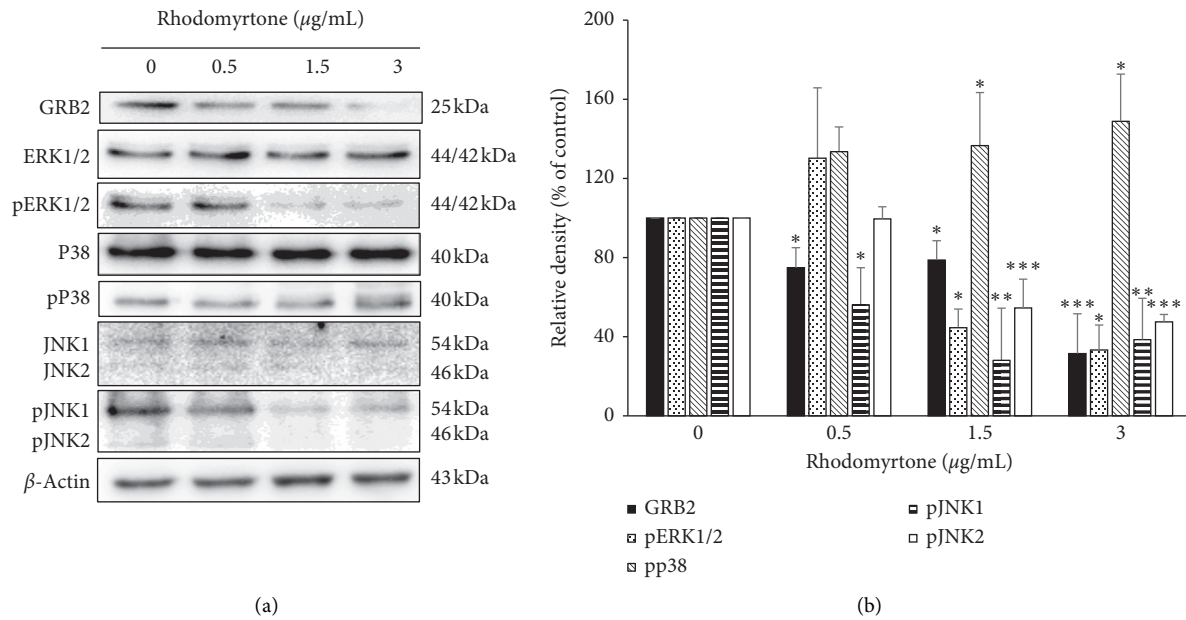


FIGURE 7: Western blot analysis of the protein level of GRB2 and MAPK pathway in SW1353 cells. (a) Expression of GRB2 and MAPK protein members was determined by Western blot analysis after treatment with or without rhodomyrtone and β -Actin is used as a loading control. (b) Relative density of proteins was significantly inhibited after treatment with rhodomyrtone. Data are presented as mean \pm standard deviation (SD) from three independent experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. the untreated control.

3.7. Rhodomyrtone at the Subtoxic Concentrations Suppressed the Expression of ERK/JNK and Induced the Expression of p38 Signaling Pathway. In this study, we showed MMP-2 and MMP-9 expression was suppressed by rhodomyrtone at the subcytotoxic concentrations. MMP-2 and MMP-9 are the important proteases involved in cancer cell migration, spreading, invasion, and metastasis [30]. They are highly expressed in various malignant tumors. The activation of these enzymes enables the degradation of ECM by cancer cells allowing their access to the vasculature as well as their migration and invasion into the target organ and development of cancer metastasis [31, 33]. Mitogen-activated protein kinase (MAPK) signaling pathway especially has been reported to be involved in regulation of MMP-2 and MMP-9 expression [16, 66–69]. Reduction of MAPK protein members may have the potential to interrupt cancer cell migration, invasion, and progression. Here, we investigated the effect of rhodomyrtone on MAPK protein members in SW1353 cells. The results showed that GRB2, phosphorylated ERK1/2, and JNK1/2 were significantly inhibited after treatment with rhodomyrtone in a concentration-dependent manner (Figure 7). Many previous studies showed that the expression of MMPs such as MMP-1, MMP-2, MMP-9, and MMP-13 in human chondrosarcoma is regulated by integrin $\alpha\beta3$ /FAK/ERK pathway [17, 70–72]. Likewise, the previous report demonstrated that expression of MMP-2 and MMP-9 in cancer cells was regulated by JNK/MAPK pathway [73]. Our previous study showed rhodomyrtone inhibited skin cancer cell metastasis by suppression of MMP-2/9 expression through inhibiting ERK1/2 and p38 signaling pathway [43]. Moreover, we found that rhodomyrtone

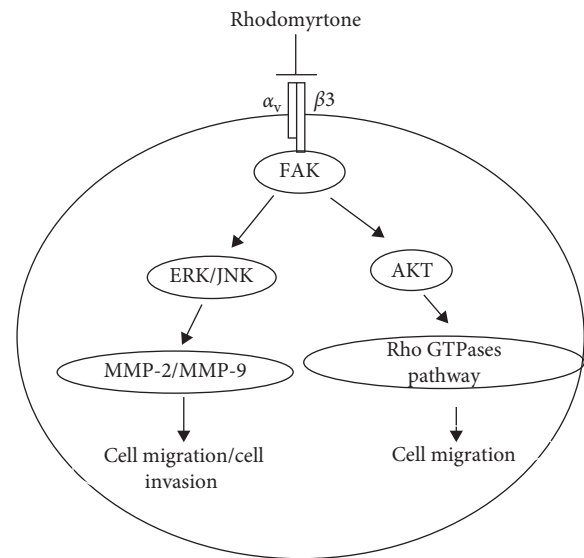


FIGURE 8: Rhodomyrtone inhibited SW1353 cell metastasis by suppressing integrin $\alpha\beta3$ /FAK/AKT/small Rho GTPases pathway as well as downregulation of MMP-2/9 via ERK and JNK signaling pathway inhibition.

induced expression of phosphorylated p38. Activation of p38 MAPK signaling has been reported to be involved in the overexpression of TIMP-1, the endogenous inhibitor of MMP-2, and it may be responsible for MMP-2 inhibition [56]. These findings suggest that rhodomyrtone at the subcytotoxic concentrations inhibited human chondrosarcoma cell invasion by suppressing MMP-2 and MMP-9 expression via ERK and JNK signal inhibition.

4. Conclusions

Our present study demonstrated that rhodomyrtone could inhibit SW1353 cell motility, migration, invasion, and adhesion by suppressing integrin $\alpha v\beta 3$ /FAK/AKT/small Rho GTPases pathway as well as downregulation of MMP-2/9 via ERK and JNK signaling pathway inhibition (Figure 8). These findings indicate that rhodomyrtone is a new antimetastasis agent for the treatment of cancer metastasis. In vivo studies need to be performed in the next phase to determine the therapeutic efficacy.

Data Availability

The data and materials used in the study are available from authors upon reasonable request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

All authors equally contributed to this study.

Acknowledgments

This work was funded by the Strategic Wisdom and Research Institute, Srinakharinwirot University, Bangkok, Thailand. The authors would like to thank Strategic Wisdom and Research Institute, Srinakharinwirot University, Bangkok, Thailand.

References

- [1] Z. Dai, P. Lei, J. Xie, and Y. Hu, "Antitumor effect of resveratrol on chondrosarcoma cells via phosphoinositide 3-kinase/AKT and p38 mitogen-activated protein kinase pathways," *Molecular Medicine Reports*, vol. 12, no. 2, pp. 3151–3155, 2015.
- [2] X. B. Yan, T. Xie, S. D. Wang, Z. Wang, H. Y. Li, and Z. M. Ye, "Apigenin inhibits proliferation of human chondrosarcoma cells via cell cycle arrest and mitochondrial apoptosis induced by ROS generation-an in vitro and in vivo study," *International Journal Clinical Experimental Medicine*, vol. 11, no. 3, pp. 1615–1631, 2018.
- [3] Y.-C. Fong, W.-H. Yang, S.-F. Hsu et al., "2-methoxyestradiol induces apoptosis and cell cycle arrest in human chondrosarcoma cells," *Journal of Orthopaedic Research*, vol. 25, no. 8, pp. 1106–1114, 2007.
- [4] G. Polychronidou, V. Karavasilis, S. M. Pollack, P. H. Huang, A. Lee, and R. L. Jones, "Novel therapeutic approaches in chondrosarcoma," *Future Oncology*, vol. 13, no. 7, pp. 637–648, 2017.
- [5] H. E. Tzeng, C. H. Tang, S. H. Wu et al., "CCN6-mediated MMP-9 activation enhances metastatic potential of human chondrosarcoma," *Cell Death and Disease*, vol. 9, no. 10, Article ID 955, 13 pages, 2018.
- [6] J. J. Bravo-Cordero, L. Hodgson, and J. Condeelis, "Directed cell invasion and migration during metastasis," *Current Opinion in Cell Biology*, vol. 24, no. 2, pp. 277–283, 2012.
- [7] R. Rathinam and S. K. Alahari, "Important role of integrins in the cancer biology," *Cancer and Metastasis Reviews*, vol. 29, no. 1, pp. 223–237, 2010.
- [8] J. C. Chen, Y. C. Fong, and C. H. Tang, "Novel strategies for the treatment of chondrosarcomas: targeting integrins," *BioMed Research International*, vol. 2013, Article ID 396839, 11 pages, 2013.
- [9] B. Xue, W. Wu, K. Huang et al., "Stromal cell-derived factor-1 (SDF-1) enhances cells invasion by $\alpha v\beta 6$ integrin-mediated signaling in ovarian cancer," *Molecular and Cellular Biochemistry*, vol. 380, no. 1–2, pp. 177–184, 2013.
- [10] H. Sawai, H. Funahashi, M. Yamamoto et al., "Interleukin-1 α enhances integrin α ," *Oncology*, vol. 65, no. 2, pp. 167–173, 2003.
- [11] K. Takenaka, M. Shibuya, Y. Takeda et al., "Altered expression and function of beta1 integrins in a highly metastatic human lung adenocarcinoma cell line," *International Journal of Oncology*, vol. 17, no. 6, pp. 1187–1194, 2000.
- [12] C. Heyder, E. Gloria-Maercker, W. Hatzmann, B. Niggemann, K. S. Zänker, and T. Dittmar, "Role of the beta1-integrin subunit in the adhesion, extravasation and migration of T24 human bladder carcinoma cells," *Clinical and Experimental Metastasis*, vol. 22, no. 2, pp. 99–106, 2005.
- [13] S. W. Wang, H. H. Wu, S. C. Liu et al., "CCL5 and CCR5 interaction promotes cell motility in human osteosarcoma," *PLoS One*, vol. 7, no. 4, Article ID 35101, 11 pages, 2012.
- [14] C. H. Tang, Y. T. Keng, and J. F. Liu, "HMGB-1 induces cell motility and $\alpha 5\beta 1$ integrin expression in human chondrosarcoma cells," *Cancer Letters*, vol. 322, no. 1, pp. 98–106, 2012.
- [15] C. M. Wu, T. M. Li, S. F. Hsu et al., "IGF-I enhances $\alpha 5\beta 1$ integrin expression and cell motility in human chondrosarcoma cells," *Journal of Cellular Physiology*, vol. 226, no. 12, pp. 3270–3277, 2011.
- [16] C. Y. Lee, M. J. Su, C. Y. Huang et al., "Macrophage migration inhibitory factor increases cell motility and up-regulates $\alpha v\beta 3$ integrin in human chondrosarcoma cells," *Journal of Cellular Biochemistry*, vol. 113, no. 5, p. 1590, 2011.
- [17] C. H. Hou, Y. C. Chiang, Y. C. Fong, and C. H. Tang, "WISP-1 increases MMP-2 expression and cell motility in human chondrosarcoma cells," *Biochemical Pharmacology*, vol. 81, no. 11, pp. 1286–1295, 2011.
- [18] P. Chanvorachote, S. Chamni, C. Ninsontia, and P. P. Phiboonchaiyanan, "Potential anti-metastasis natural compounds for lung cancer," *Anticancer Research*, vol. 36, no. 11, pp. 5707–5718, 2016.
- [19] N. Petpiroon, B. Sritularak, and P. Chanvorachote, "Phoyunnanin E inhibits migration of non-small cell lung cancer cells via suppression of epithelial-to-mesenchymal transition and integrin αv and integrin $\beta 3$," *BMC Complementary and Alternative Medicine*, vol. 17, no. 1, Article ID 553, 2017.
- [20] J. L. Orgaz, C. Herraiz, and V. Sanz-Moreno, "Rho GTPases modulate malignant transformation of tumor cell," *Small GTPases*, vol. 5, no. 4, Article ID 29019, 2014.
- [21] E. Sahai and C. J. Marshall, "RHO-GTPases and cancer," *Nature Review Cancer*, vol. 2, no. 2, pp. 133–142, 2002.
- [22] T. Kamai, T. Yamanishi, H. Shirataki et al., "Overexpression of RhoA, Rac1 and Cdc42 GTPases is associated with progression in testicular cancer," *Clinical Cancer Research*, vol. 10, no. 14, pp. 4799–4805, 2004.
- [23] G. Fritz, C. Brachetti, F. Bahlmann, M. Schmidt, and B. Kaina, "Rho GTPases in human breast tumours: expression and mutation analyses and correlation with clinical parameters," *British Journal of Cancer*, vol. 87, no. 6, pp. 635–644, 2002.

- [24] G. Fritz, I. Just, and B. Kaina, "Rho GTPases are over-expressed in human tumors," *International Journal of Cancer*, vol. 81, no. 5, pp. 682–687, 1999.
- [25] J. Wang, Q. Rao, M. Wang et al., "Overexpression of Rac1 in leukemia patients and its role in leukemia cell migration and growth," *Biochemical and Biophysical Research Communications*, vol. 386, no. 4, pp. 769–774, 2009.
- [26] Y. Liu, Y. Wang, Y. Zhang et al., "Abnormal expression of p120-catenin, E-cadherin, and small GTPases is significantly associated with malignant phenotype of human lung cancer," *Lung Cancer*, vol. 63, no. 3, pp. 375–382, 2009.
- [27] M. G. Tucci, G. Lucarini, D. Brancorsini et al., "Involvement of E-cadherin, beta-catenin, Cdc42 and CXCR4 in the progression and prognosis of cutaneous melanoma," *British Journal of Dermatology*, vol. 157, no. 6, pp. 1212–1216, 2007.
- [28] S. N. Yang, H. T. Chen, H. K. Tsou et al., "Leptin enhances cell migration in human chondrosarcoma cells through OBRL leptin receptor," *Carcinogenesis*, vol. 30, no. 4, pp. 566–574, 2009.
- [29] E. I. Deryugina, M. A. Bourdon, G. X. Luo, R. A. Reisfeld, and A. Strongin, "Matrix metalloproteinase-2 activation modulates glioma cell migration," *Journal of Cell Science*, vol. 110, pp. 2473–2482.
- [30] L. J. McCawley and L. M. Matrisian, "Matrix metalloproteinases: multifunctional contributors to tumor progression," *Molecular Medicine Today*, vol. 6, no. 4, pp. 149–156, 2000.
- [31] Y. Itoh and H. Nagase, "Matrix metalloproteinases in cancer," *Essays in Biochemistry*, vol. 38, pp. 21–36, 2002.
- [32] L. M. Coussens, B. Fingleton, and L. M. Matrisian, "Matrix metalloproteinase inhibitors and cancer: trials and tribulations," *Science*, vol. 295, no. 5564, pp. 2387–2392, 2002.
- [33] M. Egeblad and Z. Werb, "New functions for the matrix metalloproteinases in cancer progression," *Nature Review Cancer*, vol. 2, no. 3, pp. 161–174, 2002.
- [34] A. Winotai, T. Wright, and J. A. Goolsby, "Herbivores in Thailand on *Rhodomirtus tomentosa* (Myrtaceae), an invasive weed in Florida," *Florida Entomologist*, vol. 88, no. 1, pp. 104–105, 2005.
- [35] A. Panthong, D. Kanjanapothi, and W. C. Taylor, "Ethnobotanical review of medicinal plants from Thai traditional books, part I: plants with anti-inflammatory, antiasthmatic and antihypertensive properties," *Journal of Ethnopharmacology*, vol. 18, no. 3, pp. 213–228, 1986.
- [36] A. Panthong, D. Kanjanapothi, T. Taesotikul, and W. C. Taylor, "Ethnobotanical review of medicinal plants from Thai traditional books, part II: plants with antidiarrheal, laxative and carminative properties," *Journal of Ethnopharmacology*, vol. 31, no. 2, pp. 121–156, 1991.
- [37] S. Dachriyanus, M. V. Sargent, B. W. Skelton et al., "Rhodomirtone, an antibiotic from *Rhodomirtus tomentosa*," *Australian Journal of Chemistry*, vol. 22, pp. 229–232, 2002.
- [38] J. Saising, A. Hiranrat, W. Mahabusarakam, M. Ongsakul, and S. P. Varavathikunchai, "Rhodomirtone from *Rhodomirtus tomentosa* (Aiton) Hassk. as a natural antibiotic for staphylococcal cutaneous infections," *Journal of Health Science*, vol. 54, no. 5, pp. 589–595, 2008.
- [39] S. Limsuwan and S. P. Voravuthikunchai, "*Boesenbergia pandurata* (Roxb.) Schltr., *Eleutherine americana* Merr. and *Rhodomirtus tomentosa* (Aiton) Hassk. as antibiofilm producing and antiquorum sensing in *Streptococcus pyogenes*," *FEMS Immunology & Medical Microbiology*, vol. 53, no. 3, pp. 429–436, 2008.
- [40] S. Limsuwan, E. N. Trip, T. R. Kouwen et al., "Rhodomirtone: a new candidate as natural antibacterial drug from *Rhodomirtus tomentosa*," *Phytomedicine*, vol. 16, no. 6–7, pp. 645–651, 2009.
- [41] W. Sianglum, P. Srimanote, W. Wonglumsom, K. Kittiniyom, and S. P. Voravuthikunchai, "Proteome analyses of cellular proteins in methicillin-resistant *Staphylococcus aureus* treated with rhodomirtone, a novel antibiotic candidate," *PLoS One*, vol. 6, no. 2, Article ID 16628, 10 pages, 2011.
- [42] J. Chorachoo, D. Saeloh, T. Srichana et al., "Rhodomirtone as a potential anti-proliferative and apoptosis inducing agent in HaCaT keratinocyte cells," *European Journal of Pharmacology*, vol. 772, pp. 144–151, 2016.
- [43] M. Tayeh, S. Nilwarangoon, W. Mahabusaraku, and R. Watanapokasin, "Anti-metastatic effect of rhodomirtone from *Rhodomirtus tomentosa* on human skin cancer cells," *International Journal of Oncology*, vol. 50, no. 3, pp. 1035–1043, 2017.
- [44] M. Tayeh, S. Nilwarangkoon, C. Tanunyutthawongse, W. Mahabusarakum, and R. Watanapokasin, "Apoptosis and antimigration induction in human skin cancer cells by rhodomirtone," *Experimental and Therapeutic Medicine*, vol. 15, no. 6, pp. 5035–5040, 2018.
- [45] M. K. Lu, Y. W. Shih, T. T. Chang Chien, L. H. Fang, H. C. Huang, and P. S. Chen, "α-Solanine inhibits human melanoma cell migration and invasion by reducing matrix metalloproteinase-2/9 activities," *Biological & Pharmaceutical Bulletin*, vol. 33, no. 10, pp. 1685–1691, 2010.
- [46] C. L. Liao, K. C. Lai, A. C. Huang et al., "Gallic acid inhibits migration and invasion in human osteosarcoma U-2 OS cells through suppressing the matrix metalloproteinase-2/-9, protein kinase B (PKB) and PKC signaling pathways," *Food and Chemical Toxicology*, vol. 50, no. 5, pp. 1734–1740, 2012.
- [47] S. Yodkeeree, C. Ampasavate, B. Sung, B. B. Aggarwal, and P. Limtrakul, "Demethoxycurcumin suppresses migration and invasion of MDA-MB-231 human breast cancer cell line," *European Journal of Pharmacology*, vol. 627, no. 1–3, pp. 8–15, 2010.
- [48] M. Takahashi, N. Oka, T. Naroda et al., "Prognostic significance of matrix metalloproteinases-2 activation ratio in renal cell carcinoma," *International Journal of Urology*, vol. 9, no. 10, pp. 531–518, 2002.
- [49] O. Jung, J. Lee, Y. J. Lee et al., "Timosaponin AIII inhibits migration and invasion of A549 human non-small-cell lung cancer cells via attenuations of MMP-2 and MMP-9 by inhibitions of ERK1/2, Src/FAK and β-catenin signaling pathways," *Bioorganic & Medicinal Chemistry Letters*, vol. 26, no. 16, pp. 3963–3967, 2016.
- [50] C. Yan and D. D. Boyd, "Regulation of matrix metalloproteinase gene expression," *Journal of Cellular Physiology*, vol. 211, no. 1, pp. 19–26, 2007.
- [51] X. Cheng, J. Gu, M. Zhang et al., "Astragaloside IV inhibits migration and invasion in human lung cancer A549 cells via regulating PKC-α-ERK1/2-NF-κB pathway," *International Immunopharmacology*, vol. 23, no. 1, pp. 304–313, 2014.
- [52] E. J. Gweon and S. J. Kim, "Resveratrol attenuates matrix metalloproteinase-9 and -2-regulated differentiation of HTB94 chondrosarcoma cells through the p38 kinase and JNK pathways," *Oncology Reports*, vol. 32, no. 1, pp. 71–78, 2014.
- [53] Y. C. Liao, Y. W. Shih, C. H. Chao, X. Y. Lee, and T. A. Chiang, "Involvement of the ERK signaling pathway in fisetin reduces invasion and migration in the human lung cancer cell line A549," *Journal of Agricultural and Food Chemistry*, vol. 57, no. 19, pp. 8933–8941, 2009.
- [54] K. Liew, P. V. Yong, Y. M. Lim, V. Navaratnam, and A. S. Ho, "2-Methoxy-1,4-Naphthoquinone (MNQ) suppresses the

- invasion and migration of a human metastatic breast cancer cell line (MDA-MB-231)," *Toxicology in Vitro*, vol. 28, no. 3, pp. 335–339, 2014.
- [55] Y. W. Shih, S. T. Chien, and M. D. Shi, "Apigenin regulates matrix metalloproteinase-2/9 and Rho GTPase family through FAK signal to reduce breast cancer MCF-7 cells metastasis," *International Journal of Biotechnology and Bioengineering*, vol. 3, no. 7, pp. 249–258, 2017.
- [56] N. Wang, M. Zhu, S. W. Tsao, K. Man, Z. Zhang, and Y. Feng, "Up-regulation of TIMP-1 by genipin inhibits MMP-2 activities and suppresses the metastatic potential of human hepatocellular carcinoma," *PLoS One*, vol. 7, no. 9, Article ID 46318, 11 pages, 2012.
- [57] C. Ferrari, S. Benassi, F. Ponticelli et al., "Role of MMP-9 and its tissue inhibitor TIMP-1 in human osteosarcoma," *Acta Orthopaedica Scandinavica*, vol. 75, no. 4, pp. 487–491, 2004.
- [58] M. Gao, J. H. Zhang, F. X. Zhou et al., "*Angelica sinensis* suppresses human lung adenocarcinoma A549 cell metastasis by regulating MMPs/TIMPs and TGF- β 1," *Oncology Reports*, vol. 27, no. 2, pp. 585–593, 2012.
- [59] J. Park, K. Singha, S. Son et al., "A review of RGD-functionalized nonviral gene delivery vectors for cancer therapy," *Cancer Gene Therapy*, vol. 19, no. 11, pp. 741–748, 2012.
- [60] M. Deb, D. Sengupta, and S. K. Patra, "Integrin-epigenetics: a system with imperative impact on cancer," *Cancer and Metastasis Review*, vol. 31, no. 1-2, pp. 221–234, 2012.
- [61] C. M. Wu, T. M. Li, T. W. Tan, Y. C. Fong, and C. H. Tang, "Berberine reduces the metastasis of chondrosarcoma by modulating the α v β 3 integrin and the PKC δ , c-Src, and AP-1 signaling pathways," *Evidence-based Complementary and Alternative Medicine*, vol. 2013, Article ID 423164, 10 pages, 2013.
- [62] K. Plaibua, V. Pongrakhananon, P. Chunhacha, B. Sritularak, and P. Chanvorachote, "Effects of artonin e on migration and invasion capabilities of human lung cancer cells," *Anticancer Research*, vol. 33, no. 8, pp. 3079–3088, 2013.
- [63] A. Maiuthed and P. Chanvorachote, "Cisplatin at sub-toxic levels mediates integrin switch in lung cancer cells," *Anticancer Research*, vol. 34, no. 12, pp. 7111–7117, 2014.
- [64] N. Wang, Y. Feng, E. P. Lau et al., "F-actin reorganization and inactivation of rho signaling pathway involved in the inhibitory effect of *Coptidis Rhizoma* on hepatoma cell migration," *Integrative Cancer Therapies*, vol. 9, no. 4, pp. 354–364, 2010.
- [65] B. L. Montalvo-Ortiz, L. Castillo-Pichardo, E. Hernández et al., "Characterization of EHOp-016, novel small molecule inhibitor of Rac GTPase," *Journal of Biological Chemistry*, vol. 287, no. 16, pp. 13228–13238, 2012.
- [66] C. Y. Lee, C. Y. Huang, M. Y. Chen, C. Y. Lin, H. C. Hsu, and C. H. Tang, "IL-8 increases integrin expression and cell motility in human chondrosarcoma cells," *Journal of Cellular Biochemistry*, vol. 112, no. 9, pp. 2549–2557, 2011.
- [67] P. C. Brooks, S. Strömblad, L. C. Sanders et al., "Localization of matrix metalloproteinase MMP-2 to the surface of invasive cells by interaction with integrin α v β 3," *Cell*, vol. 85, no. 5, pp. 683–69, 1996.
- [68] C. H. Hou, R. S. Yang, S. M. Hou, and C. H. Tang, "TNF- α increases α v β 3 integrin expression and migration in human chondrosarcoma cells," *Journal of Cellular Physiology*, vol. 226, no. 3, pp. 792–799, 2011.
- [69] K. B. Reddy, S. M. Nabha, and N. Atanaskova, "Role of MAP kinase in tumor progression and invasion," *Cancer and Metastasis Reviews*, vol. 22, no. 4, pp. 395–403, 2003.
- [70] Y.-J. Chen, Y.-Y. Wei, H.-T. Chen et al., "Osteopontin increases migration and MMP-9 up-regulation via α v β 3 integrin, FAK, ERK, and NF- κ B-dependent pathway in human chondrosarcoma cells," *Journal of Cellular Physiology*, vol. 221, no. 1, pp. 98–108, 2009.
- [71] S. C. Hsu, J. H. Lin, S. W. Weng et al., "Crude extract of *Rheum palmatum* inhibits migration and invasion of U-2 OS human osteosarcoma cells by suppression of matrix metalloproteinase-2 and -9," *BioMedicine*, vol. 3, no. 3, pp. 120–129, 2013.
- [72] Y.-C. Fong, C.-Y. Lin, Y.-C. Su et al., "CCN6 enhances ICAM-1 expression and cell motility in human chondrosarcoma cells," *Journal of Cellular Physiology*, vol. 227, no. 1, pp. 223–232, 2012.
- [73] Y. Fong, K. H. Shen, T. A. Chiang, and Y. W. Shih, "Acacetin inhibits TPA-induced MMP-2 and u-PA expressions of human lung cancer cells through inactivating JNK signaling pathway and reducing binding activities of NF-kappaB and AP-1," *Journal of Food Science*, vol. 75, no. 1, pp. 30–38, 2010.