

Research Article

Spectrophotometric Determination of Gemifloxacin Mesylate, Moxifloxacin Hydrochloride, and Enrofloxacin in Pharmaceutical Formulations Using Acid Dyes

Ayman A. Gouda,¹ Alaa S. Amin,² Ragaa El-Sheikh,¹ and Amira G. Yousef¹

¹ Chemistry Department, Faculty of Science, Zagazig University, Zagazig 44519, Egypt ² Chemistry Department, Faculty of Science, Benha University, Benha, Egypt

Correspondence should be addressed to Ayman A. Gouda; aymangouda77@gmail.com

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Simple, rapid, and extractive spectrophotometric methods were developed for the determination of some fluoroquinolones antibiotics: gemifloxacin mesylate (GMF), moxifloxacin hydrochloride (MXF), and enrofloxacin (ENF) in pure forms and pharmaceutical formulations. These methods are based on the formation of ion-pair complexes between the basic drugs and acid dyes, namely, bromocresol green (BCG), bromocresol purple (BCP), bromophenol blue (BPB), bromothymol blue (BTB), and methyl orange (MO) in acidic buffer solutions. The formed complexes were extracted with chloroform and measured at 420, 408, 416, 415, and 422 nm for BCG, BCP, BPB, BTB, and MO, respectively, for GMF; at 410, 415, 416, and 420 nm for BCP, BTB, BPB, and MO, respectively, for MXF; and at 419 and 414 nm for BCG and BTB, respectively, in case of ENF. The analytical parameters and their effects are investigated. Beer's law was obeyed in the ranges 1.0–30, 1.0–20, and 2.0–24 μ g mL⁻¹ for GMF, MXF, and ENF, respectively. The proposed methods have been applied successfully for the analysis of the studied drugs in pure forms and pharmaceutical formulations. Statistical comparison of the results with the reference methods showed excellent agreement and indicated no significant difference in accuracy and precision.

1. Introduction

Fluoroquinolones are the second-generation members of quinolone antibiotics fluorinated in position 6 and bearing a piperazinyl moiety at position. They are considered to be the most effective Gram-positive and Gram-negative pathogens to combat infection caused by microorganisms that are resistant to other microbials, such as tetracyclines. Also, they have some activity against mycobacteria, mycoplasmas, rickettsias, and the protozoan *Plasmodium falciparum* [1–3]. There is a substantial body of literature related to both the mechanism of their action as DNA gyrase inhibitors and the influence of systematic structural modifications on their biological activity. Gemifloxacin mesylate (GMF) is (R,S)-7-[(4Z)-3-(aminomethyl)-4-(methoxyimino)-1-pyrrolidinyl]-1-cyclopropyl-6-fluoro-1,4-dihydro-4-oxo-1,8-naphthyridine-3-carboxylic acid methanesulfonate. Moxifloxacin (MXF) is {1-cyclopro-

pyl-7-[2,8-diazobicyclo (4.3.0) nonane]-6-fluoro-8-methoxy-1,4 dihydro-4-oxo-3-quinolone carboxylic acid}. Enrofloxacin (ENF) is (1 cyclopropyl-7-(4-ethyl-1-piperazinyl)-6-fluoro-1,4-dihydro-4-oxo-3-quinolone carboxylic acid) (Scheme 1). GMF and MXF are fourth-generation synthetic broad-spectrum 8-methoxy fluoroquinolone antibacterial drug derivatives. Due to their clinical advantages, GMF and MXF are receiving a great interest and there was an increase in number of their pharmaceutical dosage forms in the market in the recent past. Enrofloxacin is the first fluoroquinolone developed for veterinary application and is potentially available for the treatment of some urinary tract, respiratory tract, and skin infectious diseases in pets and livestock [2]. There are no official (pharmacopoeia) methods that have been found for the assay of GMF and MXF in their pharmaceutical formulations. ENF is official in the United States Pharmacopeia (USP) [4].



SCHEME 1: The chemical structure of the studied fluoroquinolones.

Several methods have been reported for the determination of fluoroquinolones either in pure forms, dosage forms, or biological fluids like chromatography [5–9], microchip electrophoresis [10], chiral counter-current chromatography [11], capillary zone electrophoresis [12], electrochemistry [13– 15], atomic absorption spectrometry [16, 17], and spectrofluorimetry [18–21]. However, these methods are expensive and not available at most quality control laboratories. For routine analysis of the studied drugs, a simple, rapid, and cost effective analytical method was required.

The spectrophotometric technique continues to be the most preferred method for the assay of different classes of drugs in pure, pharmaceutical formulations and in biological samples, for its simplicity and reasonable sensitivity with significant economical advantages. Spectrophotometric methods are reported for the assay of GMF [22–32], MXF [16, 33–40], and ENF [17, 41–47]. These methods were associated with some major drawbacks such as decreased selectivity due to measurement in ultraviolet region and/or decreased simplicity of the assay procedure (e.g., tedious precipitation, heating, or liquid-liquid extraction steps in the ion-pair formation-based methods). For these reasons, it was worthwhile to develop a new simple and selective spectrophotometric method for the determination of the studied drugs in their pharmaceutical dosage forms.

In the present work, we report the development of accurate and precise extractive spectrophotometric methods based on the chloroform soluble ion-pair complexes between the studied fluoroquinolone antibiotics (GMF, MXF, and ENF) and some acid dyes (BCG, BCP, BTB, BPB, or MO). The absorbance measurements were measured at optimum wavelengths. The proposed methods were applied successfully for the determination of the studied drugs in pure and dosage forms. No interference was observed from the additives. The methods provide rapid, economic procedures and more sensitive compared to the previously reported spectrophotometric methods. These methods were validated by the statistical data.

2. Experimental

2.1. Apparatus. All absorption spectra were made using Kontron Unikon 930 (UV-Visible) spectrophotometer (German) with a scanning speed of 200 nm/min and a band width of 2.0 nm, equipped with 10 mm matched quartz cells. The pH values of different buffer solutions were checked using a Hanna pH-meter instrument (pH 211) (Romania) equipped with a combined glass-calomel electrode.

2.2. Materials and Reagents. All reagents and chemicals used were of analytical or pharmaceutical grade and all solutions were prepared fresh daily.

Materials. Pharmaceutical grade gemifloxacin mesylate (GMF) was supplied by Al-Obour Pharmaceutical and Chemical Industries Company, Egypt. Moxifloxacin hydrochloride (MXF) reference standard was provided by Sabaa, Kahira Company, Egypt. Enrofloxacin (ENF) was kindly provided by Pharma Swede, Egypt (AVICO).

All pharmaceutical preparations were obtained from commercial sources in the local markets. Factive tablets were obtained from Oscient Pharmaceuticals Corporation, USA, labeled to contain 320 mg GMF per tablet; Flobiotic tablets were obtained from Hikma Pharmaceutical and Chemical Industries Company, Egypt, labeled to contain 320 mg GMF per tablet. GemiQue tablets were obtained from Obour Pharmaceutical and Chemical Industries Company, Egypt, labeled to contain 320 mg GMF per tablet. Avelox tablets were obtained from Bayer, Germany, labeled to contain 400 mg MXF per tablet. Moxiflox tablets were obtained from EVA Pharm. & Chem. Ind. Company, Egypt, labeled to contain 400 mg MXF per tablet. Moxifloxacin tablets were obtained from Sabaa International Company for Pharmaceuticals and Chemical Industries, S.A.E., labeled to contain 400 mg MXF per tablet. Enrocin 10% injectable (Alexandria Company for Pharmaceuticals and Chemical Industries, Alexandria, Egypt) was labeled to contain 10% ENR and Avitryl 20% injectable (AVICO Veterinary Pharmaceuticals) was labeled to contain 200 mg ENR.

2.3. Preparation of Stock Standard Solutions. Stock standard solutions of GMF, MXF, and ENR ($100 \mu g \text{ mL}^{-1}$ and $1.0 \times 10^{-3} \text{ M}$) were prepared by dissolving an exact weight of pure drugs in least amount of 0.1 M HCl; the mixture was warmed at 50°C in a water bath for 5.0 min, agitated by an electrical shaker for another 5.0 min, cooled to room temperature, and diluted to 100 mL with bidistilled water in a 100 mL measuring flask. The standard solutions were found stable for at least one week without alteration when kept in an amber coloured bottle and stored in a refrigerator when not in use.

2.4. Reagents. Bromocresol green (BCG), bromocresol purple (BCP), bromophenol blue (BPB), bromothymol blue (BTB), and methyl orange (MO) (BDH Chemicals Ltd., Poole, England) were used without further purification. Stock solutions $(1.0 \times 10^{-3} \text{ M})$ of reagents were prepared by dissolving the appropriate weight of each reagent in 10 mL of 96% ethanol and diluted to 100 mL with bidistilled water. These solutions are stable for at least one week if kept in the refrigerator.

Series of buffer solutions of KCl-HCl (pH = 1.5-4.2), NaOAc-HCl (pH = 1.99-4.92), NaOAc-AcOH (pH = 3.0-5.6), and potassium hydrogen phthalate-HCl (pH = 2.0-7.0) were prepared by following the standard methods [48].

2.5. General Procedures

2.5.1. For GMF. Aliquots of (0.1-3.0 mL) the standard drug solution (100 μ g mL⁻¹) were transferred to 10 mL measuring flasks and added 2.0 mL of acetate buffers of pH 3.0 and 3.5 using (BCG or BCP) and (BPB, BTB or MO), respectively and then added 2.0 mL of all reagent solutions $(1.0 \times 10^{-3} \text{ M})$. The mixture was extracted twice with 10 mL chloroform by shaking for 2.0 min and then allowed to stand for clear separation of the two phases and the chloroform layer was passed through anhydrous sodium sulphate. The absorbance of the yellow colored complexes was measured at 420, 408, 416, 415, and 422 nm, using BCG, BCP, BPB, BTB, and MO, respectively, against corresponding reagent blank similarly prepared. All measurements were made at room temperature $(25 \pm 2^{\circ}C)$. The procedures were repeated for other analyte aliquots and calibration plots were drawn to calculate the amount of drugs in unknown analyte samples.

2.5.2. For MXF. Aliquots of (0.1-2.0 mL) the standard drug solution (100 μ g mL⁻¹) were transferred to 10 mL measuring flasks and added 2.0 mL of potassium hydrogen phthalate-HCl buffer of pH 3.5 and 3.0 using BCP or MO and BPB or BTB, respectively, then added to 2.0 mL of all reagent solutions $(1.0 \times 10^{-3} \text{ M})$. The mixture was extracted twice with 10 mL chloroform by shaking for 2.0 min and then allowed to stand for clear separation of the two phases and the chloroform layer was passed through anhydrous sodium sulphate. The absorbance of the yellow colored complexes was measured at 410, 415, 416, and 420 nm using BCP, BTB, BPB, and MO, respectively, against corresponding reagent blank similarly prepared. All measurements were made at room temperature $(25 \pm 2^{\circ}C)$. The procedures were repeated for other analyte aliquots and calibration plots were drawn to calculate the amount of drugs in unknown analyte samples.

2.5.3. For ENF. Aliquots of (0.2-2.4 mL) the standard drug solution $(100 \,\mu\text{g mL}^{-1})$ were transferred to 10 mL measuring flasks and added 2.0 mL of acetate buffer of pH 3.0 using BCG or BTB and then added to 2.0 mL of reagent solutions $(1.0 \times 10^{-3} \text{ M})$. The mixture was extracted twice with 10 mL chloroform by shaking for 2.0 min, then allowed to stand for clear separation of the two phases and the chloroform layer was

passed through anhydrous sodium sulphate. The absorbance of the yellow colored complexes was measured at 419 and 414 nm using BCG and BTB, respectively, against corresponding reagent blank similarly prepared. All measurements were made at room temperature ($25 \pm 2^{\circ}$ C). The procedures were repeated for other analyte aliquots and calibration plots were drawn to calculate the amount of drug in unknown analyte samples.

2.6. Applications to Pharmaceutical Formulations

2.6.1. Procedure for Tablets. The contents of ten tablets (Factive, Flobiotic, or GemiQue) labeled to contain 320 mg GMF per tablet and (Avelox or Moxiflox) labeled to contain 400 mg MXF per tablet were crushed, powdered, and weighted out and the average weight of one tablet was determined. An accurate weight equivalent to 10 mg GMF or MXF was dissolved in 20 mL of 0.5 M HCl with shaking for 5.0 min and filtered. The filtrate was diluted to 100 mL with bidistilled water in a 100 mL measuring flask to give 100 μ g mL⁻¹ stock solution. An aliquot of the diluted drug solution was treated as described previously.

2.6.2. Procedure for Injection. Accurate volumes of Enrocin 10% or Avitryl 20% of injectable quantity equivalent to 200 mg were extracted with 10 mL of 0.5 M HCl, diluted with water, and sonicated for about 5.0 min. The extracts were transferred into 100 mL volumetric flasks and then diluted to volume with bidistilled water. Aliquots of these solutions were transferred into a series of 10 mL volumetric flasks, and the analysis was completed as previously mentioned.

2.7. Stoichiometric Relationship. The stoichiometric ratios of the ion-associates formed between the drugs under investigation and the reagents were determined by applying the continuous variation [49] and the molar ratio [50] methods at the wavelengths of maximum absorbance. In continuous variation method, equimolar solutions were employed: 5.0 \times 10^{-4} M standard solutions of drug and 5.0×10^{-4} M solutions of dye were used. A series of solutions was prepared in which the total volume of the studied drugs and the dye was kept at 2.0 mL. The drug and reagent were mixed in various complementary proportions (0:2, 0.2:1.8, 0.4:1.6,...,2:0, inclusive) and completed to volume in a 10 mL calibrated flask with the appropriate solvent for extraction following the above mentioned procedure. In the molar ratio method, the concentrations of GMF, MXF, and ENF are kept constant (1.0 mL of 5.0×10^{-4} M) while that of dyes (5.0×10^{-4} M) are regularly varied (0.2-2.4 mL). The absorbance of the prepared solutions optimum is measured at optimum condition at wavelength for each complex.

3. Results and Discussion

3.1. Absorption Spectra. The nitrogenous drugs are present in positively charged protonated forms and anionic dyes of sulfonephthalein group present mainly in anionic form at $pH \ge 2.5$. So when treated with an acid dye at pH range 2.8–4.0 of

acidic buffers solutions, a yellow ion-pair complex which is extracted with chloroform is formed. The absorption spectra of the ion-pair complexes, which were formed between GMF, MXF, or ENF and reagents, were measured in the range 350– 550 nm against the blank solution. The ion-pair complexes of GMF and BCG, BCP, BPB, BTB, and MO show maximum absorbance at 420, 408, 416, 415, and 422 nm, respectively; of MXF and BCP, BTB, BPB, and MO show maximum absorbance at 410, 415, 416, and 420 nm, respectively and of ENF and BCG and BTB show maximum absorbance at 419 and 414 nm, respectively.

3.2. Optimum Reaction Conditions for Complex Formation. The optimization of the methods was carefully studied to achieve complete reaction formation, highest sensitivity, and maximum absorbance.

3.2.1. Effects of pH on Ion-Pair Formation. The effect of pH on the drug-reagent complex was studied by extracting the colored complexes in the presence of various buffers. It was noticed that the maximum color intensity and highest absorbance value were observed in NaOAc-AcOH buffer of pH 3.0 or 3.5 using BCG or BCP and BPB, BTB, or MO, respectively, for GMF (Figure 1) and pH 3.0 using BCG or BTB for ENF. Whereas for MXF, the highest absorbance value was observed in potassium hydrogen phthalate-HCl buffer of 3.0 and 3.5 using BCP or MO and BPB or BTB, respectively, in addition to the stability of the color without affecting the absorbance at the optimum pH values. Further, 2.0 mL of the buffers solutions gave maximum absorbances and reproducible results.

3.2.2. Effect of Extracting Solvents. The effect of several organic solvents, namely, chloroform, carbon tetrachloride, methanol, ethanol, acetonitrile, *n*-butanol, benzene, acetone, ethyl acetate, diethyl ether, toluene, dichloromethane, and chlorobenzene, was studied for effective extraction of the colored species from aqueous phase. Chloroform was found to be the most suitable solvent for extraction of colored ion-pair complexes for all reagents quantitatively. Experimental results indicated that double extraction with total volume 10 mL chloroform, yielding maximum absorbance intensity, stable absorbance for the studied drugs and considerably lower extraction ability for the reagent blank and the shortest time to reach the equilibrium between both phases.

3.2.3. Effects of Reagents Concentration. The effect of the reagents was studied by measuring the absorbance of solutions containing a fixed concentration of GMF, MXF, or ENF and varied amounts of the respective reagents. Maximum color intensity of the complex was achieved with 2.0 mL of 1.0×10^{-3} M of all reagents solutions, although a larger volume of the reagent had no pronounced effect on the absorbance of the formed ion-pair complex (Figure 2).

3.2.4. Effect of Time and Temperature. The optimum reaction time was investigated from 0.5 to 5.0 min by following the color development at ambient temperature $(25 \pm 2^{\circ}C)$. Complete color intensity was attained after 2.0 min of mixing for



FIGURE 1: Effect of pH of acetate buffer solution on ion-pair complex formation between GMF and $(1.0 \times 10^{-3} \text{ M})$ reagents.



FIGURE 2: Effect of volume of $(1.0 \times 10^{-3} \text{ M})$ reagent on the ion-pair complex formation with GMF.

all complexes. The effect of temperature on colored complexes was investigated by measuring the absorbance values at different temperatures. It was found that the colored complexes were stable up to 35°C. At higher temperatures, the drug concentration was found to increase due to the volatile nature of the chloroform. The absorbance remains stable for at least 12 h at room temperature for all reagents.

3.3. Stoichiometric Relationship. The stoichiometric ratio between drug and dye in the ion-pair complexes was determined by the continuous variations method (Figure 3). Job's method of continuous variation of equimolar solutions was employed: $a 5.0 \times 10^{-4}$ M standard solution of drug base and 5.0×10^{-4} M solution of BCG, BCP, BPB, BTB, or MO, respectively, were used. A series of solutions was prepared in which the total volume of drug and reagent was kept at 2.0 mL for BCG, BCP, BPB, BTB, and MO, respectively. The absorbance was measured at the optimum wavelength. The results indicate that 1:1 (drug: dye) ion-pairs are formed through the electrostatic attraction between positive protonated GMF⁺, MXF⁺, or



FIGURE 3: Job's method of continuous variation graph for the reaction of MXF with dyes BCP, BPB, BTB, and MO, $[drug] = [dye] = 5.0 \times 10^{-4}$ M.



MXF-BPB ion-pair complex

SCHEME 2: Proposed mechanism of the reaction between MXF and BPB salt.

ENF⁺ and negative BCG⁻, BCP⁻, BPB⁻, BTB⁻, and MO⁻. The extraction equilibrium can be represented as follows:

$$\mathrm{GMF}^{+}_{(\mathrm{aq})} + \mathrm{D}^{-}_{(\mathrm{aq})} \longleftrightarrow \mathrm{GMF}^{+}\mathrm{D}^{-}_{(\mathrm{aq})} \longleftrightarrow \mathrm{GMF}^{+}\mathrm{D}^{-}_{(\mathrm{org})}, \quad (1)$$

where GMF^+ and D^- represent the protonated GMF and the anion of the dye, respectively, and the subscripts (aq) and (org) refer to the aqueous and organic phases, respectively (Scheme 2).

3.4. Conditional Stability Constants (K_f) of Ion-Pair Complexes. The stability of the ion-pair complexes was evaluated. The formation of the ion-pair complexes was rapid and the yellow color extracts were stable at least for 12 h for drug-dye without any change in color intensity and with the maximum absorbance at room temperature. The conditional stability constants (K_f) of the ion-pair complexes for the studied drug were calculated from the continuous variation data using the following equation [51]:

$$K_f = \frac{A/A_m}{\left[1 - A/A_m\right]^{n+1} C_M(n)^n},$$
 (2)

where A is the observed maximum absorbance, A_m is the absorbance value corresponding to intersection of the two tangents of the curve, C_M is the mole concentration corresponding to maximum absorbance, and n is the stoichiometry with which dye ion associates with drugs. The log K_f values for drug-dye ion-pair associates were calculated in Table 1.

3.5. Method of Validation

3.5.1. Linearity. At described experimental conditions for GMF, MXF, and ENF determination, standard calibration curves with reagents were constructed by plotting absorbance versus concentration. The statistical parameters were given in the regression equation calculated from the calibration graphs. The linearity of calibration graphs was proved by the high values of the correlation coefficient (r) and the small values of the *y*-intercepts of the regression equations. The apparent molar absorptivities of the resulting colored ion-pair complexes and relative standard deviation of response factors for each proposed spectrophotometric method were also calculated and recorded in Table 1. The molar absorptivity of BCP > BCG > BTB > MO > BPB ion-pair complexes for GMF, while for MXF the molar absorptivity of BCP > BTB > BPB > MO ion-pair complexes, also, the molar absorptivity of BCG > BTB ion-pair complexes for ENF.

3.5.2. Sensitivity. The limits of detection (LOD) and quantitation (LOQ) for the proposed methods were calculated using the following equation [51, 52]:

$$LOD = \frac{3s}{k}, \qquad LOQ = \frac{10s}{k}, \tag{3}$$

where is the standard deviation of the response of the blank or the standard deviation of intercepts of regression lines and k is the sensitivity, namely, the slope of the calibration graph.

In accordance with the formula, the limits of detection for GMF were found to be 0.23, 0.26, 0.52, 0.28, and 0.87 μ g mL⁻¹ for BCG, BCP, BTB, BPB, and MO methods, respectively. Whereas, for MXF the detection limits were found to be 0.21, 0.56, 0.25, and 0.41 μ g mL⁻¹ for BCP, BTB, BPB, and MO methods, respectively. Also, for ENF the detection limits were found to be 0.48 and 0.51 μ g mL⁻¹ for BCG and BTB methods, respectively.

According to this equation, the limit of quantitation for GMF was found to be 0.77, 0.87, 1.73, 0.93, and 2.90 μ g mL⁻¹ for BCG, BCP, BTB, BPB, and MO methods, respectively. Whereas, for MXF the detection limits were found to be 0.70, 1.87, 0.83, and 1.37 μ g mL⁻¹ for BCP, BTB, BPB, and MO methods, respectively. Also, for ENF the detection limits were found to be 1.6 and 1.70 μ g mL⁻¹ for BCG and BTB methods, respectively.

3.5.3. Accuracy and Precision. Specificity of ion-pair reaction and selective determination of GMF, MXF, and ENF which were the basic nitrogenous compounds with acid dyes could be possible. Percentage relative standard deviation (RSD%) as precision and percentage relative error (RE%) as accuracy of the suggested methods were calculated. Precision was carried out by six determinations at four different concentrations in these spectrophotometric methods. The percentage relative error was calculated using the following equation:

$$RE\% = \left[\frac{founded - added}{added}\right] \times 100.$$
(4)

The interday and intraday precision and accuracy results are shown in Tables 2, 3, and 4. These results of accuracy and precision show that the proposed methods have good repeatability and reproducibility.

3.5.4. Robustness and Ruggedness. For the evaluation of the method robustness, some parameters were interchanged: pH, dye concentration, wavelength range, and shaking time. The capacity remains unaffected by small deliberate variations. Method ruggedness was expressed as RSD% of the same procedure applied by two analysts and with two different instruments on different days. The results showed no statistical differences between procedures done with different analysts and instruments suggesting that the developed methods were robust and rugged.

3.6. Effects of Interference. To assess the usefulness of the method, the effect of diluents, excipients, and additives which often accompany GMF, MXF, and ENF in their dosage forms (starch, lactose, glucose, sucrose, talc, sodium chloride, titanium dioxide, and magnesium stearate) was studied. The results indicated that there is no interference from excipients and additives, indicating a high selectivity for determining the studied GMF, MXF, and ENF in their dosage forms.

3.7. Analysis of Pharmaceutical Formulations. The proposed methods have been successfully applied to the determination of GMF, MXF, and ENF in pharmaceutical dosage forms. Six

			GMF				ŚW	CF		EN	н
Farameters	BCG	BCP	BTB	BPB	MO	BCP	BTB	BPB	MO	BCG	BTB
Wavelengths $\lambda_{\rm max}$ (nm)	420	408	415	416	422	410	415	416	420	419	414
ЬН	3.0	3.0	3.5	3.5	3.5	3.0	3.5	3.0	3.5	3.0	3.0
Beer's law limits ($\mu g m L^{-1}$)	1.0 - 16	1.0-12	2.0-16	1.0 - 16	3.0-30	1.0-12	2.0 - 18	1.0 - 10	2.0-20	2.0-20	2.0-24
Molar absorptivity ε (L/mol ⁻¹ cm ⁻¹) × 10 ⁴	2.1787	3.9244	1.8904	2.4457	0.9386	3.3572	1.9365	4.1976	1.2876	1.4126	1.198
Sandell's sensitivity (ng cm ⁻²)	22.3	12.4	25.7	19.9	51.7	13.0	22.6	10.4	34.0	25.4	30.0
$\log K_f$	5.25 ± 0.13	4.90 ± 0.10	4.95 ± 0.08	5.36 ± 0.12	4.76 ± 0.09	4.86 ± 0.07	4.98 ± 0.11	5.12 ± 0.09	5.20 ± 0.07	4.82 ± 0.12	5.14 ± 0.09
Regression equation"											
Intercept (a)	0.0016	0.0042	0.0087	0.0064	-0.0006	-0.0091	-0.0058	-0.0137	0.0299	0.0066	0.0005
Slope (b)	0.0447	0.0805	0.0382	0.0498	0.0196	0.0764	0.0441	0.0953	-0.0023	0.0393	0.0334
Correlation coefficient (r)	0.9998	0.9999	0.9993	7666.0	0.9996	0.9991	0.9997	0.9994	0.9995	0.9998	0.9995
$LOD \ (\mu g \ m L^{-1})^b$	0.23	0.26	0.52	0.28	0.87	0.21	0.56	0.25	0.41	0.48	0.51
$LOQ (\mu g m L^{-1})^b$	0.77	0.87	1.73	0.93	2.90	0.70	1.87	0.83	1.37	1.60	1.70
Mean ± SD	99.80 ± 1.14	99.60 ± 0.74	99.90 ± 0.90	99.75 ± 1.05	99.65 ± 0.86	99.95 ± 0.74	100.10 ± 1.07	99.60 ± 0.82	99.70 ± 0.79	100.05 ± 0.98	99.80 ± 0.87
RSD %	1.14	0.74	0.90	1.05	0.86	0.74	1.07	0.82	0.79	0.98	0.87
RE %	1.20	0.77	0.94	1.10	0.90	0.78	1.12	0.86	0.83	1.03	0.91
t-test ^c	0.23	0.50	0.19	0.29	0.42	0.01	0.11	0.26	0.19	0.24	0.07
F-test ^c	3.49	1.47	2.18	2.96	1.98	1.35	1.54	1.10	1.19	1.24	1.02
${}^{a}A = a + bC$, where <i>C</i> is the conce ^b LOD: limit of detection; LOQ: lir ^c The theoretical values of <i>t</i> and <i>F</i> i	entration in $\mu g r$ mit of quantifica at $P = 0.05$ are 2	mL^{-1} and y is thation; ε : molar al 2.571 and 5.05, r_1	e absorbance ur bsorptivity. espectively.	uits.							

Mathod	A ddad (I -1)		Int	raday			In	terday	
TATENTION	Autor (Mg IIII)	Recovery %	Precision RSD % ^a	Accuracy RE %	Confidence limit ^b	Recovery %	Precision RSD % ^a	Accuracy RE %	Confidence limit ^b
	5.0	99.30	0.53	-0.80	4.996 ± 0.026	100.60	0.43	0.60	5.03 ± 0.022
BCG	10	99.50	0.65	-0.50	9.950 ± 0.065	99.30	0.59	-0.70	9.930 ± 0.059
	15	99.10	0.82	-0.90	14.865 ± 0.122	99.90	0.83	-0.10	14.985 ± 0.124
	4.0	100.60	0.47	0.60	4.024 ± 0.019	99.30	0.51	-0.70	3.988 ± 0.02
BCP	8.0	99.20	0.72	-0.80	7.936 ± 0.057	99.70	0.70	-0.30	7.976 ± 0.056
	12	100.20	1.17	0.20	12.024 ± 0.141	100.30	1.10	0.30	12.036 ± 0.132
	5.0	100.10	0.49	0.10	5.005 ± 0.025	99.60	0.62	-0.40	4.98 ± 0.031
BPB	10	99.80	0.79	-0.20	9.98 ± 0.079	99.40	0.87	-0.60	9.94 ± 0.086
	15	99.70	1.01	-0.30	14.955 ± 0.151	100.20	1.24	0.20	15.03 ± 0.23
	5.0	99.60	0.65	-0.40	4.98 ± 0.032	99.70	0.54	-0.30	4.985 ± 0.027
BTB	10	100.10	0.88	0.10	10.01 ± 0.088	99.30	0.75	-0.70	9.93 ± 0.074
	15	100.40	0.93	0.40	15.06 ± 0.140	100.20	1.08	0.20	15.03 ± 0.162
MO	10	99.60	0.44	-0.40	9.96 ± 0.044	99.50	0.52	-0.50	9.95 ± 0.052
	20	99.80	0.68	-0.20	19.96 ± 0.136	99.70	0.83	-0.30	19.94 ± 0.166
	30	100.30	0.98	0.30	30.09 ± 0.295	100.20	1.20	0.20	30.06 ± 0.361
^a Mean of s ^b Confiden	ix determinations; RSD ce limit at 95% confiden	%: percentage rela ce level and five de	tive standard deviation;] egrees of freedom $(t = 2.^{10})$	RE %: percentage relat 571).	tive error.				

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MICHION	Auucu (Mg IIII	Recovery %	Precision RSD % ^a	Accuracy RE %	Confidence limit ^b	Recovery %	Precision RSD % ^a	Accuracy RE %	Confidence limit ^b
	2.0	99.50	0.36	-0.50	1.99 ± 0.007	99.00	0.38	-1.00	1.98 ± 0.008
BCP	6.0	100.20	0.57	0.20	6.012 ± 0.034	99.60	0.70	-0.40	5.976 ± 0.042
	10	100.50	0.81	0.50	10.05 ± 0.081	99.80	0.92	-0.20	9.98 ± 0.092
	5.0	99.20	0.42	-0.80	4.96 ± 0.021	100.30	0.50	0.30	5.015 ± 0.025
BPB	10	99.50	0.76	-0.50	9.95 ± 0.076	99.20	0.79	-0.80	9.92 ± 0.078
	15	99.70	0.90	-0.30	14.955 ± 0.135	99.60	1.16	-0.40	14.94 ± 0.173
	2.0	99.40	0.50	-0.60	1.988 ± 0.01	100.10	0.58	0.10	2.002 ± 0.012
BTB	6.0	99.20	0.79	-0.80	5.952 ± 0.047	99.70	0.76	-0.30	5.982 ± 0.045
	10	99.50	1.05	-0.50	9.95 ± 0.104	99.30	1.07	-0.70	9.93 ± 0.106
	5.0	100.30	0.42	0.30	5.015 ± 0.021	100.40	0.47	0.40	5.02 ± 0.024
MO	10	100.60	0.85	0.60	10.06 ± 0.086	99.80	0.83	-0.20	9.98 ± 0.083
	15	99.90	1.19	-0.10	14.985 ± 0.178	99.20	1.23	-0.80	14.88 ± 0.183
^a Mean of si ^b Confidenc	x determinations; RSD :e limit at 95% confiden	%: percentage rels ice level and five d	ative standard deviation;] legrees of freedom $(t = 2.1)$	RE %: percentage relat 571).	iive error.				

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TABLE 3: Intraday and interday precision and accuracy data for MXF obtained by the proposed methods.

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Mathad	Added		Int	raday			Inte	erday	
Meniou	$(\mu \mathrm{gmL}^{-1})$	Recovery	Precision	Accuracy	Confidence	Recovery	Precision	Accuracy	Confidence
	0	%	$RSD \%^{a}$	RE %	limit ^b	%	$RSD \%^{a}$	RE%	limit ^b
	5.0	99.10	0.42	-0.90	4.955 ± 0.021	09.60	0.51	-0.40	4.98 ± 0.025
BCG	10	100.20	0.78	0.20	10.02 ± 0.078	99.80	0.75	0.20	9.98 ± 0.075
	15	99.40	1.16	-0.60	14.91 ± 0.173	99.20	1.02	-0.80	14.88 ± 0.152
	5.0	99.30	0.53	-0.70	4.965 ± 0.026	100.60	0.40	0.60	5.03 ± 0.020
BTB	10	99.50	06.0	-0.50	9.95 ± 0.09	99.40	0.68	-0.60	9.94 ± 0.068
	20	100.50	1.15	0.50	20.10 ± 0.231	99.60	0.96	-0.40	19.92 ± 0.191
^a Mean of six d	eterminations; RSD%	6: percentage relative	standard deviation; Rl	E%: percentage relativ	ve error.				
^b Confidence li	mit at 95% confidenc	ce level and five degree	es of freedom ($t = 2.5$	71).					

TABLE 4: Intraday and interday precision and accuracy data for ENF obtained by the proposed methods.

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Doccut Talan (1 -1 /	D d	Factive t	tablets	Flobiotic 1	tablet	GemiQue 1	ablets
reagent taken (pg mit)	rure arag adaea (pg IIIL)	Total found ($\mu g m L^{-1}$)	Recovery $\%^a \pm SD$	Total found ($\mu g m L^{-1}$)	Recovery $\% \pm SD$	Total found ($\mu g m L^{-1}$)	Recovery % ± SD
2.0	4.0	5.96	99.30 ± 0.78	5.98	99.60 ± 0.67	6.02	100.30 ± 0.51
BCG	8.0	9.91	99.10 ± 1.02	10.02	100.20 ± 0.82	9.92	99.20 ± 0.70
	12	13.97	99.80 ± 1.24	13.96	99.70 ± 1.09	13.99	99.90 ± 0.94
2.0	2.0	3.996	99.90 ± 0.66	3.96	99.10 ± 0.42	3.97	99.30 ± 0.38
BCP	6.0	8.02	100.20 ± 0.90	7.98	99.80 ± 0.69	7.99	99.90 ± 0.84
	10	11.92	99.30 ± 0.95	12.05	100.40 ± 1.03	11.95	99.60 ± 1.17
2.0	4.0	5.97	99.50 ± 0.57	5.99	99.80 ± 0.36	6.01	100.15 ± 0.76
BPB	8.0	9.97	99.70 ± 0.76	9.96	99.60 ± 0.49	9.97	99.70 ± 0.96
	12	13.87	99.10 ± 1.14	13.97	99.90 ± 0.78	13.89	99.20 ± 1.21
2.0	4.0	6.03	100.50 ± 0.48	6.01	100.10 ± 0.53	6.01	100.10 ± 0.54
BTB	8.0	9.90	99.00 ± 0.72	9.96	99.60 ± 0.85	9.96	99.60 ± 1.03
	12	13.93	99.50 ± 0.97	13.96	99.70 ± 1.31	14.04	100.30 ± 1.15
5.0	5.0	9.97	99.70 ± 0.32	9.91	99.10 ± 0.70	9.99	99.90 ± 0.65
MO	10	14.94	99.60 ± 0.56	15.06	100.40 ± 1.05	14.97	99.80 ± 0.85
	20	24.95	99.80 ± 0.90	24.88	99.50 ± 1.25	24.90	99.60 ± 1.10
^a Average of six determination	S.						

TABLE 5: Determination of GMF in its pharmaceutical dosage forms applying the standard addition technique.

1	T-1//	/- 1	Avelox ta	iblets	Moxiflox	tablets	Moxifloxaci	n tablets
reagent	iaken ($\mu g m L$)	Fure arug added (µg mL)	Total found ($\mu g m L^{-1}$)	Recovery $\%^a \pm SD$	Total found ($\mu g m L^{-1}$)	Recovery % ± SD	Total found ($\mu g m L^{-1}$)	Recovery $\% \pm SD$
	2.0	2.0	3.98	99.40 ± 0.56	3.96	99.10 ± 0.62	3.98	99.60 ± 0.48
BCP		6.0	7.98	99.70 ± 0.83	8.06	100.70 ± 0.89	7.98	99.80 ± 1.10
		10	11.89	99.10 ± 1.20	12.02	100.20 ± 1.26	11.92	99.30 ± 1.17
	2.0	4.0	5.98	99.60 ± 0.61	5.97	99.50 ± 0.35	5.96	99.30 ± 0.36
BPB		8.0	9.99	99.90 ± 0.84	9.96	99.60 ± 0.60	9.99	99.90 ± 0.63
		12	14.07	100.50 ± 1.14	13.99	99.90 ± 0.85	14.08	100.60 ± 1.36
	2.0	2.0	3.98	99.50 ± 0.48	3.97	99.20 ± 0.33	4.01	100.20 ± 0.61
BTB		4.0	5.95	99.20 ± 0.92	5.99	99.80 ± 0.65	5.96	99.30 ± 0.84
		8.0	10.04	100.40 ± 1.17	10.01	100.10 ± 0.93	9.95	99.50 ± 1.07
	5.0	5.0	9.91	99.10 ± 0.42	9.94	99.40 ± 0.37	9.95	99.50 ± 0.45
MO		10	15.08	100.50 ± 0.76	15.03	100.20 ± 0.75	14.97	99.80 ± 0.85
		15	19.90	99.50 ± 1.16	19.92	99.60 ± 1.28	19.83	99.15 ± 0.90
^a Average of	f six determination:	in the second						

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DoggontT	halton (ug mI ⁻¹)	Duro drug oddod ($ug m I^{-1}$)	Enrocin 10%	injectable	Avitryl 20% i	njectable
Reagent 1	aken (µg mL)	Pute drug added (µg IIIL)	Total found ($\mu g m L^{-1}$)	Recovery $\%^a \pm SD$	Total found ($\mu g m L^{-1}$)	Recovery % ± SD
	5.0	5.0	9.92	99.20 ± 0.48	9.94	99.40 ± 0.46
BCG		10	14.94	99.60 ± 0.66	15.20	100.10 ± 0.72
		15	19.98	99.90 ± 0.82	19.90	99.50 ± 1.05
	5.0	5.0	10.05	100.50 ± 0.56	9.97	99.70 ± 0.52
BBTB		10	14.91	99.40 ± 0.83	15.05	100.30 ± 0.69
		15	19.84	99.20 ± 1.20	19.82	99.10 ± 0.95

TABLE 7: Determination of ENF in its pharmaceutical dosage forms applying the standard addition technique.

^aAverage of six determinations.

TABLE 8: Application of the proposed methods for the determination of GMF, MXF, and ENF in their pharmaceutical preparations.

Camulaa	Domonto dan otho do ^c			Proposed method	ds	
Samples	Reported methods	BCG	BCP	BPB	BTB	МО
Factive tablets						
$X \pm SD^a$	100.08 ± 0.56	99.90 ± 0.62	100.15 ± 0.74	99.75 ± 0.53	99.80 ± 0.71	100.20 ± 0.59
<i>t</i> -value ^b		0.20	0.07	0.39	0.28	0.13
<i>F</i> -value ^b		1.23	1.75	1.12	1.61	1.11
Flobiotic tablets						
$X \pm SD^a$	99.94 ± 0.68	99.68 ± 0.80	99.79 ± 0.57	99.90 ± 0.73	100.10 ± 0.84	100.20 ± 0.77
<i>t</i> -value ^b		0.23	0.15	0.04	0.14	0.23
<i>F</i> -value ^b		1.38	1.42	1.15	1.53	1.67
GemiQue tablets						
$X \pm SD^a$	99.85 ± 0.49	99.70 ± 0.60	100.05 ± 0.57	99.60 ± 0.38	99.96 ± 0.55	99.55 ± 0.63
<i>t</i> -value ^b		0.18	0.24	0.37	0.14	0.34
<i>F</i> -value ^b		1.50	1.35	1.66	1.26	1.65
Avelox tablets						
$X \pm SD^a$	99.03 ± 0.97		99.60 ± 0.74	99.35 ± 0.96	99.10 ± 1.20	99.50 ± 0.82
<i>t</i> -value ^b			0.47	0.21	0.04	0.34
<i>F</i> -value ^b			1.72	1.02	1.53	1.40
Moxiflox tablets						
$X \pm SD^a$	99.34 ± 0.34		99.15 ± 0.52	99.50 ± 0.46	99.62 ± 0.43	99.55 ± 0.60
<i>t</i> -value ^b			0.28	0.26	0.47	0.28
<i>F</i> -value ^b			2.34	1.83	1.60	3.11
Moxifloxacin tablets						
$X \pm SD^a$	99.94 ± 0.92		99.70 ± 1.05	99.85 ± 0.80	100.15 ± 0.98	99.90 ± 0.84
<i>t</i> -value ^b			0.16	0.07	0.14	0.03
<i>F</i> -value ^b			1.30	1.32	1.13	1.20
Enrocxin 10% injectable	2					
$X \pm SD^a$	99.85 ± 0.43	99.70 ± 0.68			100.10 ± 0.32	
<i>t</i> -value ^b		0.17			0.43	
F-value ^b		2.50			1.81	
Avitryl 20% injectable						
$X \pm SD^a$	99.78 ± 0.64	99.50 ± 0.48			99.46 ± 0.47	
<i>t</i> -value ^b		0.32			0.37	
<i>F</i> -value ^b		1.78			1.85	

^aAverage of six determinations.

^bTheoretical values for *t*- and *F*-values at five degrees of freedom and 95% confidence limit are t = 2.57 and F = 5.05.

^cReported spectrophotometric methods for GMF [29], MXF [40], and ENF [44].

replicate determinations were made. Moreover, to check the validity of the proposed methods, dosage forms were tested for possible interference with standard addition method (Tables 5, 6, and 7). There was no significant difference between slopes of calibration curves and standard addition methods. Therefore it is concluded that the excipients in pharmaceutical dosage forms of GMF, MXF, and ENF were not found any interference in the analysis of GMF, MXF, and ENF. At 95% confidence level the calculated *t*- and *F*-values did not exceed the theoretical *F*-value indicating no significant difference between the proposed methods and the reported methods for GMF [29], MXF [40], and ENF [44] (Table 8) [52]. The results show that satisfactory recovery data were obtained and the assay results were in good agreement with the reported methods.

4. Conclusion

This paper describes the application of extractive ion-pair complexation reaction with acid dyes for the quantification of some fluoroquinolones antibiotics (GMF, MXF, and ENF) in pure forms and pharmaceutical formulations. Compared with the existing visible spectrophotometric methods, the proposed methods have the advantages of being relatively simple, rapid, cost-effective, free from auxiliary reagents, and more sensitive for determination of the studied drugs in pure form and pharmaceutical formulations. Moreover, the proposed methods are free from tedious experimental steps such as heating unlike the previously reported spectrophotometric methods cited earlier. The most attractive feature of these methods is their relative freedom from interference by the usual diluents and excipients in amounts far in excess of their normal occurrence in pharmaceutical formulations. The statistical parameters and the recovery data reveal high precision and accuracy of the methods besides being robust and rugged. Therefore, the validated method could be useful for routine quality control assay of the studied drugs in pure forms and pharmaceutical formulations.

Conflicts of Interests

There is no other conflict of interests related to this paper.

Authors' Contribution

All the authors contributed to the concept and design, making and analysis of data, drafting, revising, and final approval. Ayman A. Gouda is responsible for the study registration. Ayman A. Gouda and Amira G. Yousef have done the experiments. Alaa S. Amin provided test samples, reference material, and data analysis. Ayman A. Gouda and Ragaa El-Sheikh are responsible for interpretation, paper writing, and administrative support. All authors read and approved the final paper.

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