

NIH Public Access Author Manuscript

Gut. Author manuscript; available in PMC 2014 June 20.

Published in final edited form as:

Gut. 2013 March ; 62(3): 452–460. doi:10.1136/gutjnl-2011-301146.

Transcriptome analysis identifies TNF superfamily receptors as potential therapeutic targets in alcoholic hepatitis

Silvia Affò¹, Marlene Dominguez¹, Juan José Lozano¹, Pau Sancho-Bru¹, Daniel Rodrigo-Torres¹, Oriol Morales-Ibanez¹, Montserrat Moreno¹, Cristina Millán¹, Aurora Loaeza-del-Castillo¹, José Altamirano¹, Juan Carlos García-Pagán¹, Vicente Arroyo¹, Pere Ginès¹, Juan Caballería¹, Robert F Schwabe², and Ramon Bataller¹

¹Liver Unit, Hospital Clínic, Institut d'Investigacions Biomèdiques August Pi i Sunyer (IDIBAPS), Centre Esther Koplowitz, CIBER de Enfermedades Hepáticas y Digestivas (CIBERehd), Barcelona, Spain

²Department of Medicine, Columbia University, College of Physicians and Surgeons, New York, New York, USA

Abstract

Objective—Alcoholic hepatitis (AH) is a severe clinical condition that needs novel therapies. The identification of targets for therapy is hampered by the lack of animal models of advanced AH. The authors performed a translational study through a transcriptome analysis in patients with AH to identify new molecular targets.

Design—Hepatic gene expression profiling was assessed by DNA microarray in patients with AH (n=15) and normal livers (n=7). Functional analysis was assessed by gene set enrichment analysis. Quantitative PCR was performed in patients with AH (n=40), hepatitis C (n=18), non-alcoholic steatohepatitis (n=20) and in mouse models of acute and chronic liver injury. Protein expression was assessed by immunohistochemistry and western blotting.

Results—Gene expression analysis showed 207 genes >5-fold differentially expressed in patients with AH and revealed seven pathways differentially regulated including 'cytokine— cytokine receptor interaction'. Several tumour necrosis factor (TNF) superfamily receptors, but not ligands, were overexpressed in AH. Importantly, Fn14 was the only TNF superfamily receptor exclusively upregulated in AH compared with other liver diseases and correlated with both 90-day

Competing interests None.

Correspondence to: Dr. Ramon Bataller, Liver Unit, Hospital Clínic, Villarroel 170, Barcelona 08036, Spain; bataller@clinic.ub.es. Additional materials are published online only. To view these files please visit the journal online (http://dx.doi.org/10.1136/gutjnl-2011-301146).

Transcript profiling: Microarray data have been deposited in NCBI's Gene Expression Omnibus (GEO; accession number GSE28619).

Contributors SA and MD performed the experiments, collected and analysed data, interpreted the data and wrote the manuscript. JJL: analysed data and interpreted the data of the microarray. PS-B, DR-T, OM-I, MM, CM, ALC, JA: collected and analysed data, interpreted the data and contributed in the revision of the manuscript. JCG-P, VA, PG, JC, RFS: interpreted the data and contributed to the revision of the manuscript. RB: designed the study, interpreted the data and wrote the manuscript. SA and MD contributed equally to this work.

Ethics approval Approval provided by the Ethics Committee of the Hospital Clinic of Barcelona, Spain.

Provenance and peer review Not commissioned; externally peer reviewed.

mortality and severity of portal hypertension. Fn14 protein expression was detected in areas of fibrogenesis and in a population of hepatocytes. Fn14 expression was increased in experimental models of liver injury and was detected in progenitor cells.

Conclusion—Translational research revealed that TNF superfamily receptors are overexpressed in AH. Fn14, the receptor for TNF-like weak inducer of apoptosis, is selectively upregulated in patients with AH. TNF superfamily receptors could represent a potential target for therapy.

INTRODUCTION

Alcoholic hepatitis (AH) is the most severe form of alcoholic liver disease (ALD).¹ It is characterised by hepatocellular damage, steatosis and pericellular fibrosis. Patients with severe AH have a poor short-term prognosis.² Current therapies are not fully effective and novel targeted therapies are needed. The development of such therapies is hampered by a poor knowledge of the molecular mechanisms. Based on animal models,^{3–6} tumour necrosis factor (TNF) α was proposed to play a pivotal role in AH.^{3–9} Consequently, drugs interfering TNF α were tested in these patients¹⁰¹¹ but the results were disappointing due to an increased incidence of severe bacterial infections.¹² There are no experimental models that mimic the main findings of AH in humans. To overcome this limitation, translational studies with human samples are required.¹³

Microarray analysis using high-throughput screening technology has emerged as an important tool to study gene expression patterns and molecular events in complex diseases.¹⁴ Functional interpretation of microarray data is currently performed using different softwares.¹⁵ Gene profiling analysis has been performed in different types of chronic liver diseases.^{16–18} Here, we use modern informatics tools to perform a functional analysis capable of identifying the pathways implicated in the pathogenesis of AH. We studied a series of biopsy-proven AH and fragments of normal livers and identified different pathways that may play a pathogenic role, including 'cytokine-cytokine receptor interaction'. A detailed analysis allowed us the identification of the TNF superfamily as a potential disease driver. The TNF superfamily contains a variety of cytokines and receptors that activate signalling pathways regulating cell survival, death and differentiation. The most relevant agonists include $TNF\alpha$, TNF-like weak inducer of apoptosis (TWEAK) and Fas ligand. Studies from experimental models of alcoholic liver injury have implicated TNFa, Fas ligand and their receptors in liver damage and remodelling.⁴⁵¹⁹²⁰ Little is known on the role of TWEAK and its specific receptor Fn14. Activation of Fn14 controls many cellular activities including proliferation, migration, differentiation, apoptosis, angiogenesis and inflammation.²¹ Moreover, experimental evidence indicates that the TWEAK-Fn14 axis is implicated in progenitor cell expansion and liver regeneration.²²²³

MATERIALS AND METHODS

Patients

Patients admitted to the Liver Unit of the Hospital Clínic, Barcelona (2007–2009), with clinical, analytical and histological features of AH were prospectively included. The inclusion criteria have been previously described.¹³²⁴²⁵ All patients had histological

diagnosis of AH. Patients with malignancies or any other potential cause of liver disease were excluded from the study. Liver biopsies were obtained using a transjugular approach. As controls, we included patients with chronic hepatitis C-induced liver disease (HCV) (n=18). All patients had HCV genotype 1 and did not receive previous antiviral treatment. We also included a cohort of patients with morbid obesity and associated non-alcoholic steatohepatitis (NASH) (n=20) according to Kleiner's criteria (supplementary table 1).²⁶ A laparoscopic liver biopsy was obtained in these patients during bariatric surgery. In all patients, liver specimens were analysed by an expert liver pathologist and a part of the biopsy was submerged into a RNA stabilisation solution (RNAlater, Ambion, Austin, Texas, USA). The protocol was conformed to the ethical guidelines of the 1975 Declaration of Helsinki and was approved by the Ethics Committee of the Hospital Clinic of Barcelona and only patients with signed informed consent were included.

Selection of fragments of normal human livers

Fragments of normal livers were obtained from optimal cadaveric liver donors (n=3) or resection of liver metastases (n=4). All controls had normal serum aminotransferase levels and normal liver histology (supplementary table 2). Criteria to obtain normal livers were: (1) no past history of liver disease, alcohol abuse or metabolic syndrome; (2) absence of maintained arterial hypotension before laparotomy; (3) normal serum levels of aminotransferases; (4) time of brain death to laparotomy less than 12 h; (5) normal liver histology; (6) no use of vasodilator drugs; and (7) liver specimens obtained immediately after laparotomy and before vascular clamp. Samples obtained during resection of liver metastasis were obtained at a minimum distance of 5 cm from the tumour.

Microarray studies and functional analysis of microarray data

A subset of patients with severe AH based on clinical criteria (Maddrey's discriminant function >32) (n=15) was randomly selected to perform DNA microarray analysis. The epidemiological, clinical and analytical characteristics of this subset of patients were representative of the whole series of patients (n=40, table 1). High quality RNA samples were hybridised to GeneChips (Affymetrix *Hgu133plus*, Affymetrix, Santa Clara, California, USA) and a functional analysis was performed with the resulting data (see details in supplementary Materials and methods).

Real-time PCR

Quantitative real-time PCR (qPCR) was performed as previously described using commercial primer-probe pairs (Applied Biosystems, Foster City, California, USA) for CXCL3, CXCL4, CXCL5, CXCL6, CXCL10, Fas, TNFRSF12A (Fn14), CXCL1 (Gro-α), IL-8, CCL2 (MCP-1), osteoprotegerin, CCL5, TNFα, TNFRSF1, TRAF1, TRAF3, TRAILR1 and TNFSF12 (TWEAK).²⁷ Data were normalised to 18s and gene expression values were calculated based on the Ct method. The results were expressed as 2^{- Ct}.

Immunohistochemistry

Paraffin-embedded liver sections were incubated with anti-Fn14 (1:40 overnight at 4°C, Abcam, Cambridge, UK), anti-epithelial cell adhesion molecule (EpCAM) (1:100 for 2 h at

room temperature, Dako, Glostrup, Denmark), anti-pancytokeratin (1:4000 for 1 h at room temperature, Dako) and NF- κ B p65 (1:50 overnight at 4°C, Cell Signalling, Beverly, Massachusetts, USA) primary antibodies.

Western blot

Western blot studies were performed using standard procedures. Membranes were incubated with antibodies against AKT, phospho-AKT (Ser473), p38 MAP Kinase, phospho-p38 MAPK (Thr180/Tyr182) (Cell Signaling) and reprobed with GAPDH (Abcam). After washes, membranes were incubated with horseradish peroxidase-conjugated secondary antibody. Proteins were detected by enhanced chemiluminescence (Biological Industries, Beit-Haemek, Israel).

High precision-cut liver slices of mice livers

Male C57BL/6 mice were used to obtain 250 μ m slices from fresh liver sections using a Vibratome VT1000S (Leica Microsystems, Wetzlar, Germany). Samples were washed in PBS, soaked in 4% agarose solution (Ultrapure LMP Agarose, Invitrogen, Carlsbad, California, USA) for 20 min, and then orientated, mounted and immobilised using cyanoacrylate glue. Tissue slices were placed on organotypic tissue culture plate inserts for up to 24 h (Millicell®-CM; Millipore, Massachusetts, USA). Tissues were maintained at 37°C in a 5% CO₂ humidified incubator using 1.1 ml of Williams'Medium E (Sigma-Aldrich, St. Louis, Missouri, USA) supplemented with 1% inactivated fetal bovine serum, 2 mM L-Glutamine, 50 U/ml penicillin, 50 μ g/ml streptomycin, 0.1 μ M insulin, 15 mM HEPES and 50 μ M β -mercaptoetanol for up to 48 h. After 24 h in culture, tissue slices were incubated with TWEAK (R&D Systems, Minneapolis, Minnesota, USA), TGF β 1 (Sigma-Aldrich), TNF α (R&D Systems) and lipopolysaccharide (LPS, Sigma-Aldrich) for 24 h. Slices were then transferred to a 1.5 ml tube and homogenised with a polypropylene pestle in 1 ml Trizol®Reagent and total RNA was obtained.

Experimental mouse models

Different models of acute and chronic liver injury were performed including carbon tetrachloride (CCl₄), 3,5-diethoxycarbonyl-1,4-dihydro-collidin (DDC) and acetaminophen administration (see supplementary Materials and methods for details). Moreover, we used a model of acute and acute-on-chronic ethanol-induced liver injury. In the acute model, male Balb/c mice (n=10) were fasted for 8 h with free access to water and then gavaged a single dose of 50% ethanol (5 g/kg body weight) or water. Animals were sacrificed 8 h after gavage. In the acute-on-chronic model, male Balb/c mice (n=10) were intra-peritoneally administered with CCl₄ (Sigma-Aldrich; diluted 1:4 in oil) or vehicle (oil) at a dose of 0.5 ml/kg body weight twice per week for a total of five injections. Two days after the last CCl₄ injection, mice were fasted for 8 h with free access to water, and then they were gavaged a single dose of 50% ethanol (5 g/kg body weight) or water and sacrificed 8 h after gavage.

In all animal models, livers were excised and collected for RNA extraction and immunohistochemistry. All animal procedures were approved by the Ethics Committee of Animal Experimentation of the University of Barcelona and were conducted in accordance with the National Institutes of Health *Guide for the Care and Use of Laboratory Animals*.

Statistical analysis

Continuous variables were described as means (\pm SE) and were compared using Student t test or Mann—Whitney U test, as appropriate, depending on their normality test. Differences between categorical variables were assessed by Fisher's exact test or the χ^2 test with Yates correction for continuity, when necessary.

Correlations between variables were evaluated using Spearman's ρ or Pearson's *r*, when appropriate. In order to identify those molecules associated with short-term mortality (90 days) we fitted a Cox univariate regression analysis. We did not perform a multivariate Cox regression analysis because of the high risk of overfitting due to the scarce numbers of events (n=11) in our sample, according to a proportion rule of variables/events of 1:10.

The area under the receiver characteristic curve (AUROC) analysis was used to determine the best cut-off value and the accuracy (sensitivity and specificity) of continuous variables associated with 90-day mortality. Finally, we performed a comparative risk analysis using the Kaplan—Meier method. Comparisons were performed by the log-rank test. All statistical analyses were performed using SPSS V.14.0 for Windows (SPSS Inc.).

RESULTS

General characteristics of patients

We prospectively included 40 patients with clinical, analytical and histological characteristics of AH. Thirty patients (75%) developed at least one major complication during hospitalisation: two-thirds presented ascites and almost a half of the patients developed bacterial infections, whereas only a minority developed renal failure, encephalopathy or variceal haemorrhage.

The majority of patients had severe sinusoidal portal hypertension and 78% were classified as severe AH (ABIC score >6.71) at admission (for ABIC score calculation see: http://www.lille-model.com/score.asp?score=abic).²⁵ The majority of patients had moderate or severe steatosis (69%) of diffuse distribution, marked hepatocyte ballooning (61%) and marked necro-inflammation (37%). The main general characteristics of patients are depicted in table 1.

Unsupervised hierarchical clustering analysis

Gene expression profile analysis was first assessed in a representative subgroup of patients with severe AH (n=15) that was obtained from the original cohort (n=40). This analysis revealed 207 genes >5-fold differentially regulated in patients with AH compared with controls. The most upregulated genes that were differentially regulated are shown in table 2. Unsupervised clustering analysis allowed a clear differentiation in gene expression between livers with AH and controls (p<0.001) (figure 1). All patients with AH clustered together and showed a homogeneous pattern of gene expression. Functional analysis assessed using the gene set enrichment analysis program and the Kyoto Encyclopedia of Genes and Genomes pathway database revealed seven pathways differentially regulated in patients with AH including the cytokine—cytokine receptor interaction pathway (table 3). Full microarray

data have been deposited in NCBI's Gene Expression Omnibus (accession number GSE28619).

Analysis of the cytokine—cytokine receptor interaction pathway in different types of chronic liver diseases

Because cytokines are currently considered potential targets of therapy in patients with AH, we focused on the cytokine—cytokine receptor interaction pathway (figure 2). First, the microarray data confirmed previous results from our group indicating that CXC chemokines are markedly overexpressed in AH.¹³²⁴ Second, we found that some CC chemokines including MCP-1 were upregulated in these patients while others remained unchanged. Interestingly, none of the agonists belonging to the TNF superfamily (including $TNF\alpha$ and Fas ligand) were differently expressed in livers with AH. In contrast, several TNF superfamily receptors (eg, FAS, TRAILR1 and Fn14) were markedly overexpressed. To confirm the results obtained in the microarray analysis, we measured the expression of selected genes by quantitative PCR in our series of patients with AH (n=40) and in patients with other liver diseases including chronic hepatitis C (n=18) and NASH (n=20). Validated genes included seven CXC and two CC chemokines and genes belonging to the TNF superfamily (Fas, Fn14, TNFa, TNFRSF1 and TRAILR1). The expression of the selected genes assessed by qPCR corroborated the results obtained with the microarray studies (p<0.01 for all). Among TNF receptor superfamily, both Fn14 and TRAILR1 were markedly overexpressed in AH compared with normal livers (p<0.001) (figure 3A). Importantly, the only TNF superfamily receptor exclusively overexpressed in AH was Fn14. Fn14 was nearly 20-fold overexpressed in AH, downregulated in chronic hepatitis C and unchanged in NASH, suggesting that it may play a specific role in AH. Moreover, we confirmed that TNFa, FAS and TNFRSF1 are not increased in our series of patients with AH compared with normal livers. While TNF α was increased in chronic hepatitis C (p=0.0002) and NASH (p=0.02), TNFRSF1 and FAS were exclusively upregulated in patients with NASH (p<0.0005). Finally, we measured the expression of TNFR-associated factor (TRAF) consensus binding motif contained in the cytoplasmic tail of most of the TNFR superfamily members including Fn14.²¹ We found that TRAF3, but not TRAF1, was increased in AH (figure 3A).

Relationship between Fn14 and short-term prognosis in patients with AH

Because Fn14 was the only TNF superfamily receptor exclusively overexpressed in AH (figure 3A), we next explored the correlation between Fn14 hepatic expression and key clinical events. We explored whether baseline hepatic expression of Fn14 correlates with short-term survival in patients with AH. Cox regression analysis showed that hepatic gene expression of Fn14 (HR: 1.05, 95% CI 1.00 to 1.11, p=0.03), IL-8 (HR: 1.14, 95% CI 1.02 to 1.26, p=0.019), CXCL-5 (HR: 1.01, 95% CI 1.004 to 1.02, p=0.006), CXCL1 (HR: 1.001, 95% 1.00 to 1.003, p=0.018) and CXCL6 (HR: 1.01, 95% CI 1.004 to 1.03, p=0.009) was associated with 90-day mortality. The best cutoff level for Fn14 gene expression was 22 $(2^{-} Ct)$ (AUROC: 0.72; sensitivity 75% and specificity 62%, data not shown). Kaplan—Meier survival analysis was performed to compare 90-day mortality according to hepatic expression of Fn14 in patients with AH. Patients with higher Fn14 gene expression (>22) had worse 90-day survival than patients with lower hepatic gene expression (p=0.02) (figure

3B). We next explored whether Fn14 expression correlated with the degree of portal hypertension, a major determinant of mortality in patients with AH. Patients with high (>22-fold-expression) Fn14 gene expression showed more severe portal hypertension than patients with low expression (<22-fold-expression) (HVPG 21.5 ± 2 vs 17.5 ± 1 mm Hg, respectively; p=0.04) (figure 3C). Importantly, Fn14 and TRAILR1 hepatic gene expression correlated with the ABIC score, a system that identifies patients with severe disease (r=0.44, p=0.01 and r=0.41, p=0.02, respectively). In addition, hepatic gene expression of Fn14 positively correlated with most of CXC chemokines gene expression (supplementary figure 1). Collectively, these results suggest that Fn14 could be implicated in the pathogenesis of disease severity in AH.

Immunohistochemistry analysis of Fn14 expression in patients with AH

We next studied Fn14 at the protein level. Fn14 staining was barely detected in normal human livers while it was mainly expressed in parenchymal cells around the fibrogenic areas in patients with AH (figure 4A, B). Because Fn14 has been reported to be expressed in progenitor cells in damaged tissues²²²³ we next explored whether Fn14 was expressed in progenitor cells or in hepatocytes that derive from these cells. For this purpose, sequential liver sections from patients with AH were stained with anti-Fn14, anti-pancytokeratin (a marker of progenitor cells) and anti-EpCAM, a surface marker of human hepatic progenitor cells that is also expressed in newly generated hepatocytes derived from progenitor cells but not in mature hepatocytes.²⁸ Fn14 was expressed in hepatocytes at the edge of regenerative nodules colocalising with EpCAM positive cells (figure 4C, D) and it was expressed only in a subpopulation of pancytokeratin positive cells (figure 4E, F). These results suggest that Fn14 is mainly expressed in a fraction of hepatocytes and in a subpopulation of progenitor cells in patients with AH.

Expression of Fn14 in experimental models and in precision-cut liver slices in mice

Because there are no animal models of severe AH, we next explored the expression of Fn14 in mouse models of acute and chronic liver injury. Well-established animal models of acute and chronic liver injury were performed. Fn14 gene expression (p<0.05) was increased in mice treated with CCl₄, DDC diet and acute acetaminophen, but not by chronic administration of CCl₄ (figure 5A). Remarkably, overexpression of Fn14 was higher in the acetaminophen and DDC treated animals, two animal models of liver injury that are characterised by the expansion of liver progenitor cells. Immunohistochemical studies showed that Fn14 was expressed in progenitor cells (figure 5B) and weakly expressed in a subpopulation of hepatocytes. These results are in accordance with previous studies describing the expression of Fn14 in liver progenitor cells.²²²³

We next studied the expression of Fn14 in an experimental model of acute and acute-onchronic alcohol-induced liver damage. Acute ethanol exposure strongly induced Fn14 gene expression (figure 5C) but not other TNFa superfamily members including TNFa, TRAIL and TWEAK (data not shown). In the acute-on-chronic model, Fn14 was also increased, but in a lesser extent than in the purely acute model (figure 5C). Moreover, we studied several signalling pathways potentially implicated in Fn14-induced biological effects. We found that p38, AKT and NF- κ B p65 were activated in the livers of animals exposed to ethanol compared with control mice (figure 5D, E).

We finally used high precision-cut liver slices from mice livers to investigate which mediators are involved in Fn14 upregulation in the injured liver. Liver slices were exposed to inflammatory mediators involved in the pathogenesis of AH (LPS and TNF α) as well as TWEAK and TGF β 1, a major fibrogenic mediator. Incubation of liver slices with LPS, TNF α and TWEAK did not increase Fn14 gene expression, while TGF β 1 increased its expression (figure 5E).

DISCUSSION

The current study investigates the hepatic gene expression profile in patients with AH using a high-throughput approach. We performed a functional analysis of the hepatic transcriptome, which allowed us to identify several pathways potentially implicated in the pathogenesis of this devastating medical condition. Besides confirming some previous data¹³²⁴ we provide evidence that other novel biological pathways such as 'TNF superfamily receptors' and 'ECM (extracellular matrix)—receptors interaction' are potentially implicated in this disease. The results of this study could be relevant, since it could provide useful human data to research groups devoted to the study of ALD. Importantly, some of the results obtained in our study differ from those obtained in rodents exposed to alcohol (ie, the lack of hepatic upregulation of TNF α and its specific receptors).⁴⁵ These discrepancies between human and experimental data reinforce the need to develop an experimental model that reproduces all the features of AH.

Because current therapies (ie, corticosteroids) are not effective in many patients with severe AH,²⁹³⁰ targeted therapies are urgently needed. In the absence of a well-accepted model of AH in rodents, studies in human samples seem mandatory. Our translational approach to identify novel therapies consists of analysing gene expression in the livers from patients with biopsy-proven AH. In all patients, liver specimens were obtained using a transjugular approach by an experienced team, which also allowed the measurement of portal pressure. We selected carefully the patients so it was not surprising that gene expression pattern was quite homogeneous throughout our series. It is important to stress that the study was performed in patients with the most severe form of ALD. Whether the results obtained in our study also apply to patients with moderate forms of ALD is unknown and deserves further investigation.

A functional analysis revealed a homogeneous pattern of altered hepatic gene expression in patients with AH. This pattern allowed the identification of several pathways implicated in AH including cytokine—cytokine receptor interaction. A detailed analysis of these pathways revealed relevant information. First, we confirmed previous data from our laboratory suggesting that CXC chemokines such as Gro- α and IL-8 may play a major role in AH.¹³²⁴ Second, we provide evidence that some selected CC chemokines such MCP-1 are also differentially regulated in these patients. This finding could be relevant since we previously found a marked lymphocytic infiltration in livers with AH, which is the hallmark biological property of CC chemokines.¹³ We are currently investigating the functional role of this type

of chemokines in our laboratory. Most importantly, data obtained in the microarray analysis and quantitative PCR studies corroborated previous results showing that TNF α is not differentially expressed in livers with AH. Therefore, we think that the current paradigm that TNF α plays a pivotal role in severe cases of ALD may be revised. These results could have important therapeutic implications, since they can explain, at least in part, why TNF α blocking agents are ineffective in these patients.¹²

The most striking finding of our study was the marked upregulation of several receptors belonging to the TNF superfamily such as Fn14, TRAILR1 and FAS in patients with AH. In contrast, none of the ligands including TWEAK, TRAIL, FAS ligand and TNF α were differentially regulated. Interestingly, we found that Fn14 was the only receptor exclusively overexpressed in patients with AH, while TRAILR1 was overexpressed in AH and NASH and TNFRSF1 and FAS were exclusively upregulated in NASH. Because of the specific overexpression of Fn14 in AH patients, we decided to further explore this TNF receptor. We first investigated whether Fn14 hepatic expression at admission predicts mortality in patients with AH. Fn14 expression was markedly associated with mortality rate at 3 months, suggesting a potential pathogenic role for this receptor in the pathogenesis of AH.

Fn14 is the receptor for TWEAK, a cytokine belonging to the TNF superfamily that has powerful biological properties.³¹ Importantly, Fn14 is also able to signal in a ligandindependent manner when it is ectopically overexpressed in vitro.²¹ We hypothesise that in AH, Fn14 could act in a TWEAK-independent manner. In fact, experimental studies in precision-cut liver slices from mice showed that TWEAK did not induce Fn14 gene expression even if capable of inducing other pro-fibrogenic and pro-inflammatory genes in the liver (supplementary figure 2). Moreover, TWEAK serum levels were not increased in patients with AH. The mechanisms leading to Fn14 upregulation in AH are actually unknown. We showed that TGF β 1, a profibrogenic cytokine markedly increased in AH, increases hepatic expression of Fn14. This effect was not induced by TNF α , TWEAK and LPS, suggesting that fibrogenic, rather than inflammatory mediators, could regulate Fn14 hepatic expression.

Recent data from our laboratory indicate that liver regeneration is probably impaired in patients with AH.²⁷ Accumulation of hepatic progenitor cells correlates with patient mortality, suggesting a defect in the maturation process that generates functional hepatocytes in these patients. Because Fn14 is expressed in progenitor cells in damaged tissues,²²²³ we explored whether Fn14 is also expressed in experimental models characterised by acute and chronic liver damage and progenitor cell expansion. In both models, we found a marked expression of Fn14. This finding, together with the colocalisation of Fn14 with progenitor cells and newly generated hepatocytes derived from progenitor cells in livers from patients with AH, suggests a potential role for this receptor in progenitor cell differentiation. Importantly, ethanol administration to mice resulted in increased Fn14 expression, suggesting that alcohol abuse stimulates Fn14 expression in the liver. Functional studies modifying Fn14 expression in an animal model of AH are required to delineate the precise role of this pathway in AH.

In conclusion, our study demonstrates that several pathways, including the cytokine cytokine receptor interaction, may play a role in AH in humans. Among genes belonging to this pathway, we found that TNFa superfamily receptors are markedly upregulated. One of these receptors, Fn14, was exclusively overexpressed in AH compared with other types of chronic liver diseases and to control livers. Fn14 hepatic expression correlated with disease severity and degree of portal hypertension. Future studies in animal models of acute AH are required to delineate the role of Fn14 in liver injury and regeneration in this severe liver disease.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

Funding This study was supported by grants from the Instituto de Salud Carlos III (FIS PI080237, FIS PS09/01164 and FIS PI080126 to RB, JC and PG, respectively). SA received a grant from IDIBAPS. PS-B was funded by the Ministerio de Ciencia e Innovación, Juan de la Cierva (JCI-2009-03849) and by the Instituto de Salud Carlos III, Miguel Servet (CP11/00071). DR-T received a grant from the Ministerio de Educación, FPU programme. MD and JA received a grant from Fundación Banco Bilbao Vizcaya Argentaria. This work was performed in Centre Esther Koplowitz (CEK).

References

- Nagata K, Suzuki H, Sakaguchi S. Common pathogenic mechanism in development progression of liver injury caused by non-alcoholic or alcoholic steatohepatitis. J Toxicol Sci. 2007; 32:453–68. [PubMed: 18198478]
- Lucey MR, Mathurin P, Morgan TR. Alcoholic hepatitis. N Engl J Med. 2009; 360:2758–69. [PubMed: 19553649]
- Tilg H, Diehl AM. Cytokines in alcoholic and non alcoholic steatohepatitis. N Engl J Med. 2000; 343:1467–76. [PubMed: 11078773]
- Iimuro Y, Gallucci RM, Luster MI, et al. Antibodies to tumour necrosis factor alpha attenuate hepatic necrosis and inflammation caused by chronic exposure to ethanol in the rat. Hepatology. 1997; 26:1530–7. [PubMed: 9397994]
- Yin M, Wheeler MD, Kono H, et al. Essential role of tumour necrosis factor alpha in alcoholinduced liver injury in mice. Gastroenterology. 1999; 117:942–5. [PubMed: 10500078]
- Neuman MG, Brenner DA, Rehermann B, et al. Mechanisms of alcoholic liver disease: cytokines. Alcohol Clin Exp Res. 2001; 25(5 Suppl ISBRA):251S–3S. [PubMed: 11391079]
- 7. Mcclain CJ, Barve S, Deaciuc I, et al. Tumor necrosis factor and alcoholic liver disease. Alcohol Clin Exp Res. 1998; 22:248S–52S. [PubMed: 9727645]
- Mcclain CJ, Barve S, Deaciuc I, et al. Cytokines in alcoholic liver disease. Semin Liver Dis. 1999; 19:205–19. [PubMed: 10422201]
- Mookerjee RP, Sen S, Davies NA, et al. Tumour necrosis factor alpha is an important mediator of portal and systemic haemodynamic derangements in alcoholic hepatitis. Gut. 2003; 52:1182–7. [PubMed: 12865279]
- Spahr L, Rubbia-Brandt L, Frossard JL, et al. Combination of steroids with infliximab or placebo in severe alcoholic hepatitis: a randomized controlled pilot study. J Hepatol. 2002; 37:448–55. [PubMed: 12217597]
- 11. Tilg H, Jalan R, Kaser A, et al. Anti-tumor necrosis factor-alpha monoclonal antobody therapy in severe alcoholic hepatitis. J Hepatol. 2003; 38:419–25. [PubMed: 12663232]
- Naveau S, Chollet-Martin S, Dharancy S, et al. A double-blind randomized controlled trial of infliximab associated with prednisolone in acute alcoholic hepatitis. Hepatology. 2004; 39:1390–7. [PubMed: 15122768]

- Dominguez M, Miquel R, Colmenero J, et al. Hepatic expression of CXC chemokines predicts portal hypertension and survival in patients with alcoholic hepatitis. Gastroenterology. 2009; 136:1639–50. [PubMed: 19208360]
- Fu S, Zhao H, Shi J, et al. Peripheral arterial occlusive disease: global gene expression analyses suggest a major role for immune and inflammatory responses. BMC Genomics. 2008; 9:369.
 [PubMed: 18673543]
- Dopazo J. Functional interpretation of microarray experiments. OMICS. 2006; 10:398–410. [PubMed: 17069516]
- 16. Seth D, Leo MA, McGuinness PH, et al. Gene expression profiling of alcoholic liver disease in the baboon (Papio hamadryas) and human liver. Am J Pathol. 2003; 163:2303–17. [PubMed: 14633604]
- Seth D, Gorrell MD, McGuinness PH, et al. SMART amplification maintains representation of relative gene expression: quantitative validation by real time PCR and application to studies of alcoholic liver disease in primates. J Biochem Biophys Methods. 2003; 55:53–66. [PubMed: 12559588]
- Seth D, Gorrell MD, Cordoba S, et al. Intrahepatic gene expression in human alcoholic hepatitis. J Hepatol. 2006; 45:306–20. [PubMed: 16797773]
- Tagami A, Ohnishi H, Moriwaki H, et al. Fas-mediated apoptosis in acute alcoholic hepatitis. Hepatogastroenterology. 2003; 50:443–8. [PubMed: 12749243]
- 20. Taïeb J, Mathurin P, Poynard T, et al. Raised plasma soluble Fas and Fas-ligand in alcoholic liver disease. Lancet. 1998; 351:1930–1. [PubMed: 9654266]
- 21. Winkles JA. The TWEAK-Fn14 cytokine-receptor axis: discovery, biology and therapeutic targeting. Nat Rev Drug Discov. 2008; 7:411–25. [PubMed: 18404150]
- 22. Jakubowski A, Ambrose C, Parr M, et al. TWEAK induces liver progenitor cell proliferation. J Clin Invest. 2005; 115:2330–40. [PubMed: 16110324]
- Tirnitz-Parker JEE, Viebahn SC, Jakubowski A, et al. Tumor necrosis factor—like weak inducer of apoptosis is a mitogen for liver progenitor cells. Hepatology. 2010; 52:291–302. [PubMed: 20578156]
- Colmenero J, Bataller R, Sancho-Bru P, et al. Hepatic expression of candidate genes in patients with alcoholic hepatitis: correlation with disease severity. Gastroenterology. 2007; 132:687–97. [PubMed: 17258719]
- Dominguez M, Rincon D, Abraldes JG, et al. A new scoring system for prognostic stratification of patients with alcoholic hepatitis. Am J Gastroenterol. 2008; 103:2747–56. [PubMed: 18721242]
- 26. Kleiner DE, Brunt EM, Van Natta M, et al. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. Hepatology. 2005; 41:1313–21. [PubMed: 15915461]
- Sancho-Bru P, Altamirano J, Rodrigo-Torres D, et al. Liver progenitor cell markers correlate with liver damage and predict short-term mortality in patients with alcoholic hepatitis. Hepatology. Published Online First: 25 January 2012. 10.1002/hep.25614
- Yoon SM, Gerasimidou D, Kuwahara R, et al. Epithelial cell adhesion molecule (EpCAM) marks hepatocytes newly derived from stem/progenitor cells in humans. Hepatology. 2011; 53:964–73. [PubMed: 21319194]
- 29. Christensen E, Gluud C. Glucocorticoids are ineffective in alcoholic hepatitis: a meta-analysis adjusting for confounding variables. Gut. 1995; 37:113–18. [PubMed: 7672658]
- Imperiale TF, Mccullough AJ. Do corticosteroids reduce mortality from alcoholic hepatitis? A meta-analysis of the randomized trials. Ann Intern Med. 1990; 113:299–307. [PubMed: 2142869]
- Wiley SR, Winkles JA. TWEAK, a member of the TNF superfamily, is a multifunctional cytokine that binds the TweakR/Fn14 receptor. Cytokine Growth Factor Rev. 2003; 14:241–9. [PubMed: 12787562]

Significance of this study

What is already known about the subject?

- Alcoholic hepatitis (AH) is a severe form of alcoholic liver diseases that carries a poor short-term prognosis.
- Current therapies to treat AH (eg, corticosteroids) are not fully effective in many patients and targeted therapies are urgently needed.
- CXC chemokines are overexpressed in livers from patients with AH and may represent novel targets for therapy.
- Tumour necrosis factor α is overexpressed in animal models of moderate alcoholic liver disease.

What are the new findings?

- There are seven pathways differentially regulated in patients with alcoholic hepatitis (AH) compared with normal livers including 'focal adhesion', 'cell cycle' and 'cytokine—cytokine receptor interaction'.
- Tumour necrosis factor (TNF) superfamily receptors, but not ligands, are markedly over-expressed in patients with AH.
- Fn14 is the only TNF superfamily receptor exclusively upregulated in AH compared with other liver diseases and correlates with both 90-day mortality and severity of portal hypertension.

How might it impact on clinical practice in the foreseeable future?

• The identification of potential novel targets for therapy will stimulate the development of new targeted therapies for this severe clinical condition and will help the design of new clinical trials.

Activation in the second secon

Figure 1.

Microarray data of patients with alcoholic hepatitis (n=15) compared with control livers (n=7). The intensity of each colour denotes the standardised ratio between each value and the average expression of each gene across all samples. Red coloured pixels correspond to an increased abundance of the transcript in the indicated sample, whereas green pixels indicate decreased transcript levels.



Figure 2.

Gene expression of the members of 'cytokine—cytokine receptor interaction' pathway in patients with alcoholic hepatitis (AH). In grey colour, ligands and receptors that were found upregulated in patients with AH.



Figure 3.

(A) Fn14, TNF α , TNFRSF1, TRAILR1 and FAS gene expression in patients with alcoholic hepatitis (AH) (n=40), HCV (n=18) and non-alcoholic steatohepatitis (NASH) (n=20) compared with normal livers (n=7) (*p<0.05; **p<0.005; #p<0.005); TRAF1 and TRAF3 gene expression in patients with AH compared with normal livers (*p<0.05). (B) Kaplan—Meier curve showing 90-day survival according to Fn14 gene expression at baseline. A value of 22-fold expression with respect to controls was identified as the cut-off value with better sensitivity and specificity to define patients with low and high Fn14 gene expression. (C) Severity of portal hypertension among patients with low (<22-fold) and high (>22-fold) Fn14 gene expression (*p=0.04).



Figure 4.

(A) Representative photomicrograph of a normal liver staining for Fn14 (200× magnification); (B) Representative photomicrograph of a liver with alcoholic hepatitis (AH) stained with anti-Fn14 (original magnification, ×400); (C, D) Immunohistochemistry of consecutive slices of livers with AH stained with anti-Fn14 and anti-EpCAM, respectively (original magnification, ×200); (E, F) Immunohistochemistry of consecutive slices of livers with AH stained with anti-Fn14 and anti-EpCAM, respectively (original magnification, ×200); (E, F) Immunohistochemistry of consecutive slices of livers with AH stained with anti-Fn14 and anti-EpCAM, respectively (original magnification, ×200).



Figure 5.

(A) Hepatic Fn14 gene expression in mice with chronic and acute CCl₄ administration and DDC diet treated and receiving acetaminophen (APAP) mice (see supplementary Materials and methods) (*p<0.05 with respect to controls); (B) Fn14 hepatic protein expression in control mice and in mice treated with APAP; Fn14 expression was detected in progenitor cells (arrows) and was weakly expressed in some hepatocytes (original magnification, ×400); (C) Hepatic Fn14 gene expression from mice with CCl₄ administration, CCl₄ plus binge ethanol (EtOH) gavage and binge EtOH gavage (see Materials and methods) (*p<0.05 with respect to controls); (D) Western blot analysis from the same groups of mice showed in

(C). Representative western blot of phospho-p38 MAPK (Thr180/Tyr182), p38 MAP Kinase, phospho-AKT (Ser473) and AKT reprobed with GAPDH; (E) Representative immunohistochemical images of control and gavage EtOH administrated mice ($200 \times$ magnification) using anti-NF- κ B p65 antibody; NF- κ B p65 was strongly activated in the liver of mice gavaged with EtOH in progenitor cells and in some hepatocytes as well as in inflammatory cells (arrows); (F) Fn14 hepatic gene expression in mice high precision-cut liver slices exposed to different mediators including TWEAK, TNF α , LPS and TGF β 1. TGF β 1 was the only mediator that increased Fn14 gene expression (*p<0.05).

Table 1

Clinical, analytical and hepatic haemodynamic characteristics of patients with alcoholic hepatitis (AH, n=40)

Variables	Mean±SE or percentage	
Age (years)	49±1	
Male (%)	65	
Maddrey's discriminant function	59±6	
MELD score	22±1	
ABIC score	7.65±0.19	
Alcohol intake (gr/d)	107±5	
90-day mortality (%)	28	
Glucose (mg/dl)	111±5	
Creatinine (mg/dl)	0.84±0.06	
AST (U/L)	155±12	
ALT (U/L)	61±5	
GGT (U/L)	600±83	
Bilirubin (mg/dl)	12.5±1.21	
Albumin (mg/dl)	26.2±0.55	
Platelet count (×10 ⁹)	139±14	
Leucocytes count (×109/l)	10.04±7.43	
Neutrophils count (%)	74±8.32	
INR	1.92±0.12	
HVPG (mm Hg)	19.5±1	
Cirrhosis (%)	60	
Severe AH $(\%)^*$	77.5	

* Severe AH was defined as an ABIC score 6.71 points.

MELD, Model for End-stage Liver Disease; ABIC, Age-Bilirubin-INR-Creatinine score; ALT, alanine aminotransferase; AST, aspartate aminotransferse; GGT, gamma glutamyl transpeptidase; INR, international normalised rate; HVPG, hepatic venous pressure gradient.

Table 2

Most relevant genes differentially expressed in patients with alcoholic hepatitis compared with controls

Accession N°	Gene name	Fold expression
	Extracellular matrix/fibrosis	
M 83248	Osteopontin	67.17
AW190565	Lysyl oxidase-like 4	39.95
NM_000089	Collagen, type I, a 2	38.23
NM_005764	PDZK1 interacting protein 1	35.08
NM_002423	Matrix metallopeptidase 7	28.70
NM_003247	Thrombospondin 2	16.69
K01228	Collagen, type I, a 1	14.97
N30339	Collagen, type V, α 1	14.49
BC001388	Annexin A2	12.72
NM_001845	Collagen, type IV, a 1	10.54
NM_003254	TIMP metallopeptidase inhibitor 1	8.88
AU146808	Collagen, type III, a 1	8.76
X05610	Collagen, type IV, a 2	7.65
AK026829	Laminin, a 2	6.70
NM_000393	Collagen, type V, α 2	6.18
	Inflammation/immunity	
NM_004591	Chemokine (C-C motif) ligand 20	90.21
NM_002993	Chemokine (C-X-C motif) ligand 6	45.70
NM_004221	Interleukin 32	15.37
NM_000584	Interleukin 8	14.90
NM_016639	Tumour necrosis factor receptor superfamily, member 12A (Fn14)	10.32
NM_001511	Chemokine CXC ligand 1 (Gro-a)	9.42
NM_001565	Chemokine CXC ligand 10	6.99
AI817041	Chemokine CXC receptor 7	5.32
	Cell communication/cell cycle/apoptosis	
NM_015515	Keratin 23 (histone deacetylase inducible)	126.81
BG327863	CD24 molecule	21.29
BC002700	Keratin 7	15.26
NM_002276	Keratin 19	6.41
NM_000224	Keratin 18	5.06
	Other functions	
NM_020299	Aldo-keto reductase family 1, member B10	484.19
NM_005564	Lipocalin 2	56.66
NM_006398	Ubiquitin D	56.64
NM_016548	Golgi membrane protein 1	31.50
J04152	Tumour-associated calcium signal transducer 2	22.40
NM_002354	Tumour-associated calcium signal transducer 1	17.49
NM 000903	NAD(P)H dehvdrogenase, guinone 1	16.91

Accession N°	Gene name	Fold expression
AL514445	Regulator of G-protein signalling 4	15.65
NM_001442	Fatty acid binding protein 4, adipocyte	15.32
NM_005980	S100 calcium binding protein P	10.50

Table 3

Biological pathways for differentially expressed genes in patients with alcoholic hepatitis revealed in the functional analysis

Pathways	ID	Involved genes	FDR
Focal adhesion	HSA04510	58	0.03
Cell cycle	HSA04110	49	0.05
Cytokine—cytokine receptor interaction	HSA04060	33	0.05
Cell communication	HSA01430	24	0.01
Extracellular matrix—receptor interaction	HSA04512	24	0.01
Aminoacyl-tRNA biosynthesis	HSA00970	23	0.17
GAP junction	HSA04540	15	0.14

FDR, false discovery rates