Original Research Communication

Action of Pyrroloquinolinequinol As an Antioxidant Against Lipid Peroxidation in Solution

KAZUHIRO MIYAUCHI, TEIJI URAKAMI, HIROYUKI ABETA, HONGLIAN SHI, NORIKO NOGUCHI, and ETSUO NIKI

ABSTRACT

The activities of pyrroloquinoline quinone (PQQ), a coenzyme of methanol dehydrogenase and amine oxidase, and its reduced form pyrroloquinoline quinol (PQQH₂) as an antioxidant have been measured in solution. PQQH₂ was stable in the absence of oxygen but rapidly auto-oxidized to PQQ in the presence of oxygen in water. PQQH₂ was stable in an aprotic solvent such as acetonitrile, even in air. PQQ did not exert appreciable antioxidant activity, whereas PQQH₂ exerted higher reactivity than α -tocopherol toward galvinoxyl radical and peroxyl radical. PQQH₂ acted as a potent antioxidant against the oxidation of methyl linoleate in acetonitrile induced by azo compound and produced a clear induction period, from which the apparent stoichiometric number was obtained as 1.1. PQQH₂ reduced the α -tocopheroxyl radical and spared α -tocopherol in the oxidation of methyl linoleate. These results suggest that PQQH₂ may act as a potent antioxidant, particularly in combination with α -tocopherol. Antiox. Redox Signal. 1, 547–554.

INTRODUCTION

There is now increasing experimental, clinical, and epidemiological evidence suggesting that the oxidative damage of biological molecules plays a pivotal role in the pathogenesis of various diseases, as well as cancer and aging (Papas, 1999). Lipids are vulnerable to oxidation, and lipid peroxidation has been accepted to be an important event in the development of several diseases, including atherosclerosis (Steinberg et al., 1989). Consequently, the role of antioxidants, especially monophenolic and polyphenolic compounds such as vitamin E and flavonoids, has received much attention recently (Papas, 1999).

Pyrroloquinoline quinone (PQQ), 4,5-dihy-dro-4,5-dioxo-1*H*-pyrrolo[2,3-f]quinoline-2,7,9-tricarboxylic acid (Fig. 1), is a coenzyme of methanol dehydrogenase and bovine serum

amine oxidase (Duine et al., 1979; Salisbury et al., 1979). Trace amounts of free PQQ have been detected in several organs and the plasma and urine of humans (Kumazawa et al., 1992). Various physiological or pharmacological effects of PQQ like vitamin or growth factor activity (Killgore et al., 1989), enhancing effect of DNA synthesis in human fibroblasts (Naito et al., 1993) and inhibitory effect on the formation of hydrocortisone-induced cataract in embryos (Nishigori et al., 1989) have been reported. Its reduced form pyrroloquinoline quinol (PQQH2; Fig. 1) is a catechol derivative and expected to exert an antioxidant function. In fact, the effect of PQQ as an antioxidant has been reported against the lipid peroxidation of rat brain homogenate (Hamagishi et al., 1990). A protective effect of PQQ against experimental liver injury induced by hepatotoxins such as carbon tetrachloride, D-galactosamine, thioac-

FIG. 1. Chemical structure of PQQ and PQQH2.

etamide, and allyl formate has been observed (Watanabe et al., 1988). The reduction-oxidation cycle of PQQ-PQQH₂ has been also studied (Itoh et al., 1986; Sugioka et al., 1988). Thus, the pharmacological application of PQQ (Davidson, 1992) and its esters (Urakami et al., 1995/1996a, 1997) has received considerable attention. However, the basic kinetic data for PQQ and PQQH₂ as an antioxidant have been lacking. The present study was undertaken to obtain kinetic data on the antioxidant action of PQQ and PQQH₂ against lipid peroxidation in solution.

MATERIALS AND METHODS

Materials

PQQ was prepared by a method described previously (Urakami et al., 1993, 1995/1996a,b) that uses Hyphomicrobium denitrificans TK 0411 (Urakami et al., 1995). PQQH₂ was prepared by reducing PQQ with sodium borohydride (Itoh et al., 1986). 2R,4′R,8′R-α-Tocopherol was supplied from Eisai Co. (Tokyo, Japan). Methyl linoleate obtained from Sigma (St. Louis, MO) was purified before use as described previously (Gotoh et al., 1992). 2,2′-Azobis(2,4-dimethylvaleronitrile) (AMVN) was purchased from Wako Pure Chemical Inc. (Osaka, Japan) and used as received. Other chemicals were those of the highest grade available commercially.

Methods

The absorption spectra were measured with a spectrophotometer (Shimadzu UV-2200, Kyoto, Japan) at either 37°C or room temperature

under air, unless otherwise specified. In some experiments, the absorption spectrum was taken under vacuum by mixing PQQH₂ powder and solvent after degassing the system using a vacuum line. The rates of interaction of antioxidants with galvinoxyl were measured by following the decrease in the maximum absorption of galvinoxyl at 429 nm using a rapid-mixing stopped-flow appratus (RX1000, Applied Photophysics, Letherhead, UK). The reactivity of PQQH₂ toward the peroxyl radical was estimated by a competition method using *N*,*N*′-diphenyl-*p*-phenylenediamine (DPPD) (Noguchi *et al.*, 1998).

The oxidation of methyl linoleate was carried out at 37°C in air in the presence and absence of the radical initiator AMVN and antioxidant. The formation of methyl linoleate hydroperoxides was measured with high-performance liquid chromatography (HPLC) apparatus equipped with a UV detector monitored at 234 nm. Samples were injected onto a reverse-phase column LC18 (particle size 5 μ m; 4.6 × 250 mm; Supelco, Tokyo), methanol/H₂O (95/5, vol/vol) being used as an eluent with a flow rate of 1.0 ml/min.

The electron spin resonance (ESR) spectrum of the radicals was recorded on X-band ESR spectrometer (JEOL, Tokyo, Japan).

RESULTS

Reduction of PQQ to PQQH₂ and stability of PQQH₂

It is conceivable that PQQ exerts antioxidant capacity not *per sv* but only when it is reduced to PQQH₂. PQQ was readily reduced *in vitro* to PQQH₂ by sodium borohydride, NADPH, and NADH, and the characteristic absorption spectrum of PQQH₂ was observed (Fig. 2). PQQ was stable at 37°C under air but PQQH₂ was unstable in water and in the presence of oxygen. In the absence of oxygen, PQQH₂ was stable even in water and the absorption spectrum of PQQH₂ did not change. However, the introduction of air or 2% oxygen–98% nitrogen gas induced a rapid change in absorption spectrum as shown in Fig. 2B, which suggests the autoxidation of PQQH₂ to PQQ. On the other hand,

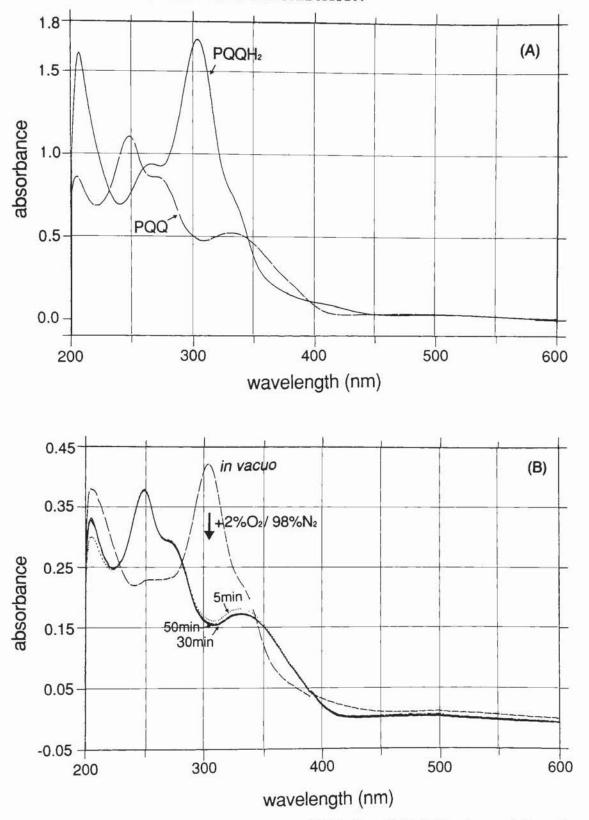


FIG. 2. A. Absorption spectra of PQQ and PQQH₂ in Tris-HCl buffer, pH 7.4. B. The change of absorption spectrum observed when 2% oxygen–98% nitrogen gas was introduced to an aqueous solution of PQQH₂ in vacuo.

PQQH₂ was found to be stable in organic aprotic solvent, even under air atmosphere.

Interaction of PQQ and PQQH2 with galvinoxyl

To estimate the reactivities of PQQ and POOH₂ toward radicals, the interaction of POO and PQQH2 with galvinoxyl radical was studied spectrophotometrically. It is well known that the hydrogen-donating antioxidant reacts rapidly with galvinoxyl radical and that the rate correlates with the reactivity of the antioxidant toward peroxyl radicals, the chaincarrying species in lipid peroxidation. The rate of interaction between PQQH₂ and galvinoxyl was measured at 37°C in air by following the decrease in the absorption of galvinoxyl in acetonitrile-DMSO (93:2 by volume) solution. PQQ did not react with galvinoxyl at appreciable rate, whereas PQQH₂ did rapidly. Fig. 3 shows the decay of absorption at 429 nm of galvinoxyl caused by the reaction with PQQH₂ and also with α -tocopherol, a major lipophilic antioxidant in vivo, for comparison. It can be seen that PQQH2 reacts with galvinoxyl radical faster than α -tocopherol.

The stoichiometric number of hydrogen of PQQH₂ that reacts with galvinoxyl was measured from a decrease in absorption at 429 nm

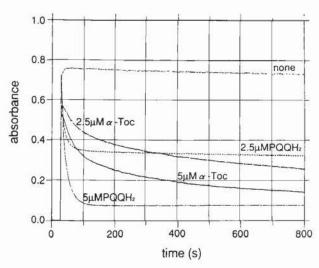


FIG. 3. Interaction of PQQH₂ with galvinoxyl. The rate of interaction between PQQH₂ and galvinoxyl (5.9 μ M) was measured at 37°C in acetonitrile–DMSO (93:2 by vol) by following a decrease in the absorption at 429 nm by galvinoxyl as described in Materials and Methods. The results with α -tocopherol (α -Toc) are also included for comparison.

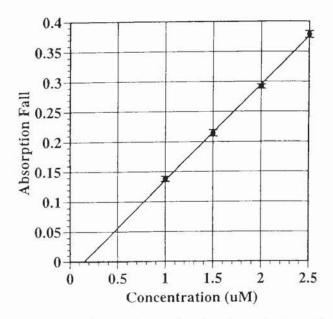


FIG. 4. Stoichiometric number for the reduction of galvinoxyl by PQQH₂. PQQH₂ (1–2.5 μ M) was reacted with 5 μ M galvinoxyl in acetonitrile at 37°C in air and the fall in the absorption at 429 nm was measured. The data points represent mean \pm SD of five independent experiments. The slope of the plot of this absorption fall against PQQH₂ concentration gives the stoichiometric number as n=1.07.

in the presence of excess galvinoxyl. As shown in Fig. 4, the absorption fall was proportional to PQQH₂ concentration and from the slope and extinction coefficient for galvinoxyl, $\epsilon = 1.50 \times 10^5 \, \mathrm{M}^{-1} \mathrm{cm}^{-1}$, the stoichiometric number was obtained as n = 1.07.

Reaction of PQQH2 with the peroxyl radical

To follow the reaction of PQQH₂ with the peroxyl radical, the change in the absorption spectrum during the incubation of PQQH₂ with AMVN was followed under air in acetonitrile–dimethylsulfoxide (DMSO) (93/2 by vol/vol) solvent. PQQH₂ was stable under air in this solvent, but the addition of AMVN induced the change in absorption spectrum as shown in Fig. 5. The absorption at 305 nm due to PQQH₂ decreased, while that at 330 nm by PQQ increased with time, suggesting that PQQH₂ was oxidized to PQQ by a reaction with the peroxyl radicals.

The reactivity of PQQH₂ toward peroxyl radicals was estimated by the competition method

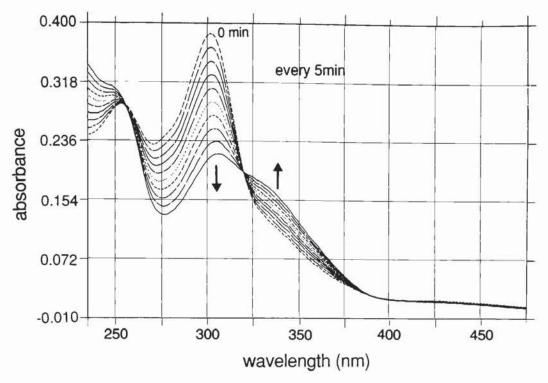


FIG. 5. Change in absorption spectrum during the oxidation of PQQH₂ by peroxyl radical. PQQH₂ (20 μ M) was incubated in acetonitrile–DMSO (93:2 vol/vol) at 37°C under air in the presence of AMVN (1.0 mM).

using DPPD (Noguchi *et al.* 1998). It is known that DPPD reacts with two molecules of peroxyl radicals to give *N,N'*-diphenyl-*p*-benzo-quinone diimine (DPBQ) (reaction 1).

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DPBQ has a strong absorption at 440 nm and the rate of reaction 1 can be followed from its increase with time. When another antioxidant IH is added to this system, it scavenges peroxyl radicals in competition with DPPD (reaction 2),

$$AO_2^{\bullet} + IH \xrightarrow{k_1} AOOH + I^{\bullet}$$
 (2)

It was found as observed previously (Noguchi *et al.*, 1998) that the rate of increase in the absorption at 440 nm was directly proportional to AMVN concentration but independent of DPPD concentration. Because DPPD reacts with two radicals to give DPBQ, the rate of free radical flux from AMVN is obtained by multi-

plying the rate of DPBQ formation by 2. The rate of free radical generation from AMVN, Ri, is given in Equation (3),

$$Ri = 2ek_d[AMVN]$$
 (3)

where e and $k_{\rm d}$ are the efficiency of free radical formation and the rate constant for unimolecular decomposition of AMVN, respectively. The rate of DPBQ formation induced by 5.0, 10, and 20 mM AMVN was obtained as 0.840, 1.76, and 3.45 nM/sec, which gives $ek_{\rm d}$ as $1.72 \times 10^{-6}~{\rm s}^{-1}$ as an average under the reaction conditions employed here. The effects of PQQH₂ and α -tocopherol on the formation DPBQ from DPPD (Fig. 6) show that PQQH₂ competes with DPPD more strongly than α -tocopherol, suggesting that PQQH₂ is more reactive toward peroxyl radicals than α -tocopherol, as observed toward galvinoxyl.

Inhibition of oxidation of methyl linoleate by PQQH₂

Methyl linoleate has been often used as a model substrate for assessing the antioxidant

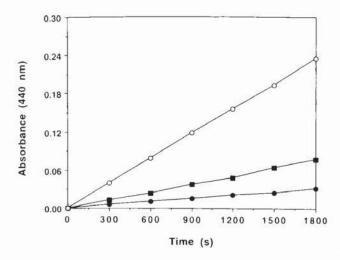


FIG. 6. Effect of PQQH₂ and a-tocopherol on the formation of DPBQ from DPPD. DPPD (0.1 m*M*) was incubated with AMVN (10 m*M*) at 37°C in acetonitrile–DMSO (93:2 vol/vol) in the absence (\bigcirc) and presence of PQQH₂ (0.1 m*M*, \blacksquare) or α -tocopherol (0.1 m*M*, \blacksquare), and the formation of DPBQ was followed by an absorption at 440 nm.

activity against lipid peroxidation, as its oxidation mechanism and products have been well established and its oxidation rate can be easily and quantitatively measured. As shown in Fig. 7, PQQH₂ suppressed the oxidation of methyl linoleate and produced a distinct in-

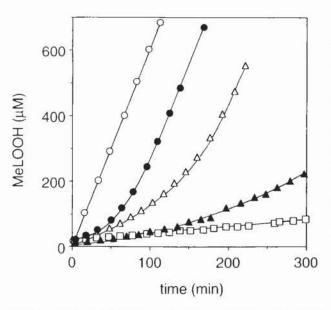


FIG. 7. Effect of PQQH₂ against the oxidation of methyl linoleate. Methyl linoleate (151 mM) was oxidized in acetonitrile–DMSO (93:2, vol/vol) in the presence of AMVN (0.10 mM) and PQQH₂ at 37°C in air and the formation of methyl linoleate hydroperoxide (MeLOOH) was followed with an HPLC. The concentrations of PQQH₂ were 0 (\bigcirc), 2 (\blacksquare), 4 (\triangle), 10 (\blacksquare), and 20 μ M (\square).

duction period in a dose-dependent manner. It has been known that the induction period t_{inh} is proportional to the antioxidant concentration and inversely proportional to the rate of chain initiation, Ri,

$$t_{\rm inh} = \frac{n \, [\rm antioxidant]}{\rm Ri}$$
 (4)

where n is the stoichiometric number of peroxyl radicals scavenged by each antioxidant molecule (Niki *et al.*, 1984). It was found in the present study that the induction period was proportional to the PQQH₂ concentration and the stoichiometric number n was obtained as 1.1.

The antioxidant activity of PQQH₂ against the oxidation of methyl linoleate was also studied in the presence of α -tocopherol. The results are shown in Fig. 8. It shows that PQQH₂ extended the induction period and, more importantly, spared α -tocopherol. It was found by ESR study that PQQH₂ reduced α -tocopheroxyl radical (data not shown).

DISCUSSION

The antioxidant action of phenolic compounds has been well documented. Above all, monophenolic compounds such as tocopherol and butylated hydroxytoluene and polyphenols such as flavonoids have been studied extensively (Papas et al., 1999). Dihydroxybenzenes are also known to act as a potent antioxidant. Ubiquinol and α-tocopheryl hydroquinone have been shown to act as a potent antioxidant in combination with α -tocopherol (Shi et al., 1999a and papers cited therein). Catechols also act as an antioxidant by either chelating metal ions or scavenging radicals. For example, catecholestrogens such as 2-hydroxyestradiol have high antioxidant capacity (Lacort et al., 1995). PQQH2 is another type of catechol compound, but the activity of PQQH2 as a radical-scavenging antioxidant has not been studied. The present study shows that $PQQH_2$ acts as a potent radical-scavenging antioxidant. As described above, PQQH2 exerted higher reactivites than α -tocopherol toward galvinoxyl and peroxyl radicals and, more importantly, it

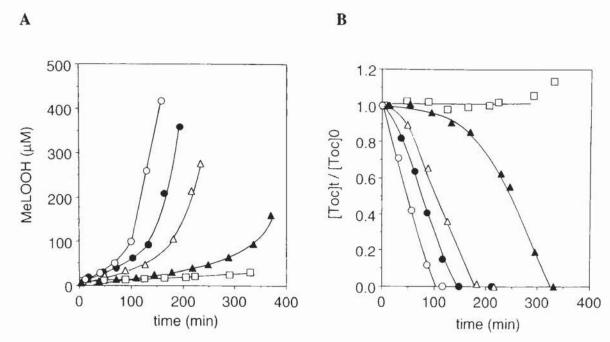


FIG. 8. Effect of PQQH₂ on the oxidation of methyl linoleate in the presence of α -tocopherol. Methyl linoleate (151 mM) was oxidized in the presence of AMVN (0.10 mM), α -tocopherol (2.0 μ M), and PQQH₂ of various concentrations at 37°C in air in acetonitrile–DMSO (93:2, vol/vol). The formation of methyl linoleate hydroperxide (MeLOOH) (A) and consumption of α -tocopherol (B) were followed. [Toc]t/[Toc]o denotes remaining fraction of α -tocopherol. The concentrations of PQQH₂ were 0 (\bigcirc), 2 (\bigcirc), 4 (\triangle), 10 (\triangle), and 20 μ M (\square).

is capable of reducing α -tocopheroxyl radical to regenerate α -tocopherol. This makes PQQH₂ particularly a potent antioxidant in combination with α -tocopherol. This may be also important under such conditions where α -tocopherol-mediated-peroxidation (Bowry and Stocker, 1993) is important.

PQQH₂ like ubiquinol and α -tocopheryl hydroquinone (Shi et al., 1999a), is unstable, especially in the presence of water and air, and readily undergoes auto-oxidation to give PQQ, which makes the apparent stoichiometric number smaller. The instability of PQQH₂ in water implies the involvement of transition metal ions. The auto-oxidation of PQQH2 is mediated by hydroperoxyl radical and/or superoxide, which may attack lipids as well and reduce the antioxidant efficacy of PQQH₂ as observed with ubiquinol and α -tocopherylhydroquinone (Shi et al., 1999a,b). PQQ, like unbiquinone and α-tocopherylquinone, does not act as an antioxidant per se, but it may be reduced to PQQH₂ by a reducing system in vivo. In fact, it has been reported that the ratio of ubiquinol/ ubiquinone is much higher than 1 in vivo (Aberg et al., 1992), suggesting an efficient reducing capacity *in vivo*. In conclusion, PQQH₂ is a potent antioxidant and may play a role in the defense system *in vivo* against oxidative stress in combination with other antioxidants such as vitamin E.

ABBREVIATIONS

AMVN, 2,2'-azobis(2,4-dimethylvaleronitrile); DMSO, dimethyl sulfoxide; DPBQ, *N*,*N*'-diphenyl-*p*-benzoquinone diimine; DPPD, *N*,*N*'-diphenyl-*p*-phenylene diamine; ESR, electron spin resonance; PQQ, pyrroloquinoline quinone; PQQH₂, pyrroloquinoline quinol.

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Address reprint requests to: Dr. Etsuo Niki RCAST, University of Tokyo 4-6-1 Komaba, Meguro Tokyo 153-8904, Japan

E-mail: eniki@oxygen.rcast.u-tokyo.ac.jp

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