

Polymorphism at the self-incompatibility locus in Solanaceae predates speciation

(S locus/molecular evolution/shared polymorphism)

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ABSTRACT Sequences of 11 alleles of the gametophytic self-incompatibility locus (S locus) from three species of the Solanaceae family have recently been determined. Pairwise comparisons of these alleles reveal two unexpected observations: (i) amino acid sequence similarity can be as low as 40% within species and (ii) some interspecific similarities are higher than intraspecific similarities. The gene genealogy clearly illustrates this unusual pattern of relationships. The data suggest that some of the polymorphism at the S locus existed prior to the divergence of these species and has been maintained to the present. In support of this hypothesis, the number of shared polymorphic sites was found to exceed the number found in simulations with independent accumulation of mutations. Strictly neutral evolution is exceedingly unlikely to maintain the polymorphism for such a long time. The allele multiplicity and extreme age of the alleles is consistent with Wright's classic one-locus population genetic model of gametophytic self-incompatibility. Similarities between the plant S locus and the mammalian major histocompatibility complex are discussed.

Self-incompatibility is a natural, genetically determined barrier to inbreeding. In many plant species, self-incompatibility is controlled by a single locus, the S locus, which elicits recognition and rejection of self-pollen (1). In gametophytic self-incompatibility, inhibition of pollen germination or pollen-tube growth results when the S allele expressed by the pollen matches one of the two S alleles expressed in the pistil. A highly expressed, pistil-specific protein, the S protein, has been identified as a product of the S locus. The cDNA sequences of several alleles of the S protein from three species in the Solanaceae family have been determined, and extraordinary divergence, even between alleles from the same species, is apparent (T.R.I. and T.-h.K., unpublished work).

The self-incompatibility systems of many plant species have long been of interest to evolutionary biologists, and the extreme levels of polymorphism in S-protein sequences suggest that an unusual array of evolutionary forces operate at the S locus. Early appreciation of the allelic diversity of *Oenothera organensis*, with 37 alleles identified in a population of 1000 plants (2, 3), prompted Wright (4) to examine population genetic models of self-incompatibility loci. A consequence of gametophytic self-incompatibility is that all individuals in a population are heterozygotes. Wright noticed that rare alleles have a reproductive advantage because pollen bearing a rare allele is less likely to land on a stigma with the same allele, so that rare alleles suffer fewer aborted pollinations. This frequency dependence made it less likely for a new mutation to be rapidly lost compared with a neutral allele, and Wright's model produced a balance between

mutation and random genetic drift that could maintain many alleles in the population. Additionally, this model predicts that the alleles could be far older than neutral alleles. The availability of molecular sequence data provides us with unprecedented power to test these models and allows us to explore the evolutionary past of the locus by reconstructing the genealogy of S alleles.

MATERIALS AND METHODS

Sequences and Alignment. The 11 inferred S-protein sequences examined in this study are S_{F11} , S_z , S_{1nic} , S_{2nic} , S_{3nic} , and S_{6nic} from *Nicotiana glauca* (5, 6), an ornamental tobacco species; S_{1pet} , S_{2pet} , and S_{3pet} from *Petunia inflata* (7), a species of wild petunia; and S_{2sol} and S_{3sol} from *Solanum chacoense* (8), a wild potato species. The alignment of amino acid sequences was taken from unpublished work (T.R.I. and T.-h.K.). The percentage of amino acid identity among pairs of alleles was calculated from the aligned sequences.

Gene Genealogy. A distance matrix based on amino acid similarities was calculated from the amino acid alignment, and the neighbor-joining algorithm (9) was used to generate a gene genealogy. The aligned amino acid sequences were also used to generate maximum parsimony trees (10) whose robustness was tested by "bootstrapping."

Shared Polymorphism Test. A test of the common ancestry of polymorphisms was performed based on nucleotide or amino acid sequences. A site that has the same two or more nucleotides segregating in two species is defined as a "shared polymorphic site." Shared polymorphic sites can occur either by chance or by common ancestry, so the test hinges on whether the observed number of shared polymorphic sites exceeds the number of shared polymorphic sites that would occur by independent accumulation of polymorphisms. The null distribution of numbers of shared polymorphisms was generated with a computer by randomly drawing one of the observed sequences and introducing as many random polymorphisms as were observed in the real data. After these randomly mutated sequences were constructed, the number of shared polymorphic sites was tallied exactly as for the real data. If the number of observed shared polymorphisms is greater than some critical fraction of the distribution with random mutations, then the null hypothesis is rejected and the test indicates that there is an excess of shared polymorphism. The most reasonable conclusion in such a case is that the polymorphisms have common ancestry. The same approach was also applied to the amino acid sequences.

RESULTS

The matrix of amino acid similarities indicates an extraordinary degree of divergence among alleles within each species (Table 1). The degree of sequence divergence within species

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Table 1. Pairwise amino acid similarity

	<i>N. alata</i>						<i>P. inflata</i>			<i>S. chacoense</i>	
	<i>S_{F11}</i>	<i>S_z</i>	<i>S_{1nic}</i>	<i>S_{2nic}</i>	<i>S_{3nic}</i>	<i>S_{6nic}</i>	<i>S_{1pet}</i>	<i>S_{2pet}</i>	<i>S_{3pet}</i>	<i>S_{2sol}</i>	<i>S_{3sol}</i>
<i>S_{F11}</i>	—										
<i>S_z</i>	59.7	—									
<i>S_{1nic}</i>	44.7	45.0	—								
<i>S_{2nic}</i>	44.7	43.6	67.7	—							
<i>S_{3nic}</i>	44.8	45.4	68.9	63.3	—						
<i>S_{6nic}</i>	43.1	45.1	61.0	60.0	69.9	—					
<i>S_{1pet}</i>	53.5	62.4	42.5	39.5	43.3	42.6	—				
<i>S_{2pet}</i>	53.2	61.7	40.8	40.3	41.7	39.9	73.1	—			
<i>S_{3pet}</i>	52.5	66.3	41.5	41.0	42.4	41.1	73.5	80.1	—		
<i>S_{2sol}</i>	40.1	41.6	46.7	44.7	46.0	45.3	38.8	38.8	39.3	—	
<i>S_{3sol}</i>	42.4	44.6	50.0	48.5	49.0	47.5	47.0	43.8	42.6	41.5	—

These values represent percent amino acid identity. Note the unusually low similarity, even between alleles from the same species. For example, *S_z* and *S_{1nic}* are only 45.0% similar. Also note that some alleles from the same species are even less similar than alleles from different species (e.g., 45.0% between *S_z* and *S_{1nic}* vs. 62.4% between *S_z* and *S_{1pet}*).

is on the same order as the sequence divergence between species. In fact, some alleles appear to be more similar to alleles in other species than to other alleles in the same species.

A phylogenetic tree based on neighbor-joining analysis (9) of the distance matrix calculated from the amino acid alignment reveals two major evolutionary branches (Fig. 1). One lineage contains *S_{F11}* and *S_z* of *N. alata* along with three alleles of *P. inflata*, and the other lineage includes *S_{1nic}*, *S_{2nic}*, *S_{3nic}*, and *S_{6nic}* of *N. alata* along with two alleles of *S. chacoense*. A genealogy of nearly identical topology was obtained by the parsimony method (10). The unrooted par-

simony tree shows that the smallest cluster with all six *N. alata* alleles necessarily contains the two *S. chacoense* alleles. The significance of this pattern was assessed by bootstrap resampling (12), which showed no cases out of 100 randomly generated trees in which all six *N. alata* alleles clustered together. Thus, the phylogenetic analyses indicate that some *N. alata* alleles are more closely related to alleles in other species than to the other *N. alata* alleles.

Based on the results of the phylogenetic analysis, we hypothesize that the extant polymorphism at the S locus arose prior to the time of divergence of the three solanaceous species. According to this hypothesis, one of the ancestral alleles was the most recent common ancestor of the *P. inflata* alleles as well as of *S_{F11}* and *S_z* from *N. alata*, and another was the most recent common ancestor of the two *S. chacoense* alleles and the remaining four *N. alata* alleles. Further support comes from a comparison of the average number of synonymous substitutions per site [*d_s*; calculated by the method of Nei and Gojobori (13) and corrected for underestimation by the Jukes-Cantor formula (14)] between S alleles from the same species and between sequences of the small subunit of ribulose-1,5-bisphosphate carboxylase (*RuBCs*) (15–17) from each pair of these three genera. The average number of synonymous substitutions per site among S alleles from the same species exceeds that from interspecific comparisons of the *RuBCs* sequences (0.82 vs. 0.47, respectively). Under the assumption of equal rates of synonymous substitution, this result suggests that the S alleles within a species diverged before speciation. *RuBCs* has an estimated synonymous substitution rate of 6.6×10^{-9} substitutions per site per year (18), which is typical of plant nuclear genes (19). With this rate of synonymous substitution, *RuBCs* sequences data indicate a *Petunia-Nicotiana* split at 27 million years ago, a *Nicotiana-Solanum* split at 28 million years, and a *Petunia-Solanum* split at 36 million years, making the shared polymorphisms in the S locus extraordinarily old.

The shared polymorphism test provides a means of statistically testing whether common ancestry of polymorphisms is an appropriate conclusion to draw from these data. Table 2 gives a count of the number of shared polymorphic sites among all species pairs and the count of polymorphic sites common to all three species. In all cases, the observed number of shared polymorphic sites is significantly greater than the number that would be expected had the polymorphisms occurred independently. As Fig. 2 shows, in some cases none of the randomly generated sequences had as many shared polymorphisms as the observed number.

A more conservative test generates random mutations only at those sites that are observed to be polymorphic in the

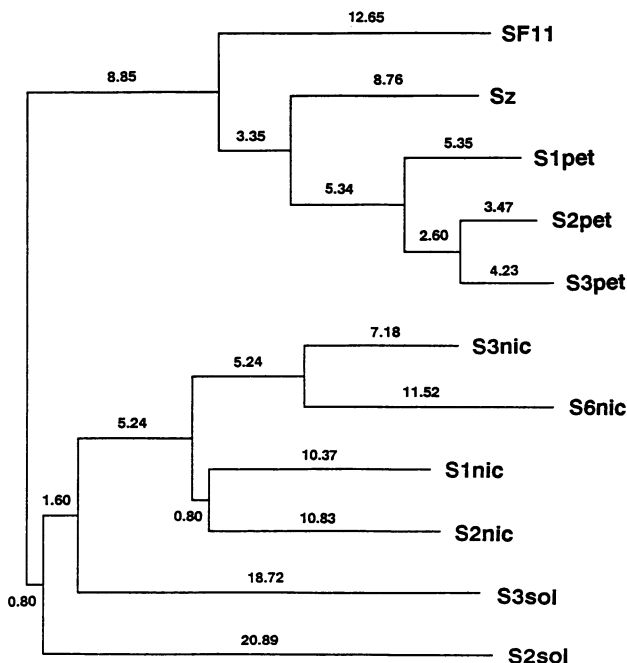


FIG. 1. S-locus gene genealogy. Note that the tree indicates that some alleles are more related to alleles in other species than to other alleles in the same species. For example, *S_z* and *S_{F11}* are clustered with the *P. inflata* alleles rather than with the other four *N. alata* alleles. Similarly, the other four *N. alata* alleles cluster with the two *S. chacoense* alleles rather than with *S_z* and *S_{F11}*. The tree was constructed from a distance matrix by the neighbor-joining algorithm (9), using software provided by J. H. Ferguson, N. Saitou, and M. Nei. Amino acid similarity values were taken from the 250 PAM table (11), appropriate for highly diverged sequences and summed over aligned residues. The distances, labeled as branch lengths, were calculated by subtracting each pairwise similarity value from 100.

Table 2. Significance tests of the numbers of shared nucleotide polymorphisms

	Sample	Random pairs of alleles
No. of polymorphic sites		
<i>N. alata</i>	360	208
<i>P. inflata</i>	154	114
<i>S. chacoense</i>	235	235
No. of shared polymorphisms		
<i>N. alata</i> and <i>P. inflata</i>	75	16.6
<i>N. alata</i> and <i>S. chacoense</i>	102	35.5
<i>P. inflata</i> and <i>S. chacoense</i>	25	16.7
All three species	16	3.11
No. of randomly mutated sequences with greater than the observed number of shared polymorphisms		
<i>N. alata</i> and <i>P. inflata</i>	0/1000	0/1000
<i>N. alata</i> and <i>S. chacoense</i>	0/1000	0/1000
<i>P. inflata</i> and <i>S. chacoense</i>	5/1000	2/1000
All three species	0/1000	25/1000

This analysis used the alignment of the 11 nucleotide sequences without the leader peptide (a total of 618 base pairs). The first block of the table reports the number of polymorphic sites and the average number of differences between pairs of alleles within each species. The second block reports the number of polymorphisms shared among all observed alleles for each pair of species and for pairs of alleles drawn from each species. A site is considered to have a shared polymorphism if, among the alleles in the sample, two species share the same pair of distinct nucleotides. The third block reports the significance test ascertained by a computer simulation that generated random, independent mutations. One allele from each species was chosen and was randomly assigned as many mutations as were observed in the first block. The number of shared polymorphisms in this randomly generated sample was compared to the observed number. The third block of the table reports the number of such random samples that have more shared polymorphisms than did the actual data.

sample. This test is appropriate if monomorphic sites are strongly constrained, and results in a larger number of shared polymorphic sites by chance. Even by this more conservative approach, all of the tests reported in Table 2 remain significant. Comparable results were obtained from an analysis of shared amino acid polymorphisms (Fig. 3). We conclude that the excess shared polymorphism is due to common ancestry and that the polymorphisms must have predated speciation.

DISCUSSION

The findings of the molecular analyses are consistent with predictions of Wright's classic one-locus population genetic model of gametophytic self-incompatibility (4). In this model, functionally new alleles generated by newly arisen mutations are initially favored over more frequent alleles. Opposing this spontaneous introduction of new variation is the loss of alleles by random genetic drift. The model attains an equilibrium in which many more alleles are maintained in a finite population than could be maintained at a strictly neutral locus. Recent theoretical analyses indicate that alleles at a gametophytic self-incompatibility locus will have very long residence times and expected coalescence times far exceeding those of neutral alleles (20, 21). The inevitable consequence of the extreme age of alleles is that they will have highly divergent sequences.

Explanations for the extraordinary allelic diversity that do not invoke the role of self-incompatibility fail to explain the observed pattern of interspecific shared polymorphism. Although a high mutation rate can also generate high levels of allelic diversity, it would not result in the large number of shared polymorphisms observed between alleles. The uniqueness of the S locus has been demonstrated by genomic

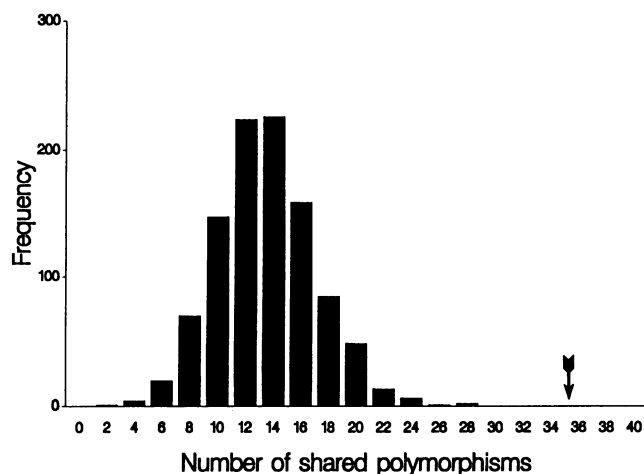


FIG. 2. The test of shared nucleotide-site polymorphism. The histogram indicates the distribution of number of shared nucleotide polymorphisms that are obtained from 1000 computer-generated samples for the tobacco-potato comparison. For each sample, one tobacco sequence and one potato sequence were drawn at random. Random pairs of tobacco sequences had an average of 208 polymorphic sites, and the potato sequences had an average of 235 polymorphic sites, so 208 mutations were introduced to the tobacco sequence and 235 mutations to the potato sequence. The number of shared polymorphic sites was then tallied from this sample, and the entire process was repeated 1000 times. The arrow indicates the average observed number of shared polymorphisms between tobacco and potato alleles. Because the observed number is greater than the randomly generated sample, we conclude that the shared polymorphisms are not due to chance coaccumulation of mutations, but rather that they represent common ancestry.

Southern blots of *N. alata* (6) and *P. hybrida* (22), which excludes the possibility of intergenic exchange as a mechanism for generating allelic diversity. Because there is only one gene, analysis of gametophytic S alleles is much simpler than that of the multigenic sporophytic self-incompatibility system of *Brassica* (23).

The clearest case of shared polymorphism would have alleles from each of two different species on each of two sides of a deep branch in the gene genealogy. The pattern observed here has only *Nicotiana* alleles on two sides of a deep branch. A tree constructed from linked shared polymorphic sites would have alleles from each species in both of the major clusters. However, hybridization and introgression or recombination/gene conversion (24) among these ancient alleles could have broken down associations among polymorphic sites, resulting in *P. inflata* alleles that cluster together while still exhibiting an excess of shared polymorphisms with *N. alata*. Genomic sequence data, which show extreme intraspecific sequence diversity for ≈ 5 kilobases downstream from the S locus, suggest that there is a mechanism for restricting intragenic recombination in this region (unpublished results). The extraordinary age of alleles and sequence divergence among alleles could only be found for a neutral locus in an implausibly large population.

Comparable evidence for ancient polymorphism that predates speciation has previously been found in the mammalian major histocompatibility complex (MHC) class I locus (24–27). Both the MHC locus and the S locus are highly polymorphic and have highly divergent alleles. The extraordinary age of the polymorphism in both systems appears to have been maintained by natural selection through a mechanism involving self-/non-self-recognition. In the case of the human MHC, evidence suggests that either overdominance or frequency-dependent selection maintains the high allelic diversity (28). The operation of diversifying natural selection at the S locus should result in an excess of nonsynonymous sub-

	10	20	30	40	50
Sf11	DFEYLQVLTPASFCY...	AMH.CERIA	PNFTIHGLWPDNVKTRLHNC		
Sz	DFDYMQVLTPASFCY...	PKNFCSRIA	PNFTIHGLWPDNVKTRLHNC		
S1nic	NFKYMQVLTPITAFG...	VMN.CERT	PTNFTIHGLWPDNVSTELNYC		
S2nic	AFKYMQVLTPITAFG...	IKH.CERT	PTNFTIHGLWPDNVSTELNYC		
S3nic	AFKYMQVLTPITAFG...	HTTSP.CERT	PNFTIHGLWPDNVSTELNYC		
S6nic	AFKYMQVLTPITAFG...	CKNI.PSNFTIHGLWPDNVSTELNYC			
S1pet	NFKYMQVLTPASFCFR...	PKNICRPA	KNFTIHGLWPEITGFRLEFC		
S2pet	NFDYQVLTPASFCY...	PKNFCKRK	SNFTIHGLWPKMKHFRLEFC		
S3pet	NFDYQVLTPASFCYR...	PKNICRRI	PNFTIHGLWPEKHFRLFC		
S2sol	TFDYMKVLQWPMYC...	HKFCERI	PNFTIHGLWPDNKKYLLNFC		
S3sol	TFEHLQVLTPITSPFC...	KERCIRS	SNFTIHGLWPDNTSTRNFC		
	60	70	80	90	100
Sf11	KPKPTYSYFT.GKMLMDLKHWMKLFQDYGRTQPSWKYQYIKHGSCC				
Sz	TSEKYVNFADQSPILDDLDHWMELKYHDFGLQFLWRQYQKHGTCC				
S1nic	DRQKQKLFDDDKKQKQDLDRWPDLTLDKDDCKKQGFWSYKYKHGTCC				
S2nic	DRSKPTMFTDQKQKQDLDRWPDLTTKTKFDSLDKQAFWKDVEVYKHGTCC				
S3nic	SGEDYEKLDLDDKQKQDLDRWPDLTIAADCKEIQVFKKHYKHGTCC				
S6nic	GKEDDYNIIMDGPEKQKGLYVRWPDLTIREKADCKEIQVFKKHYKHGTCC				
S1pet	TGDPKYEITFDNNVYDLERHWQMKFDENYAKYBQPLWSYKYKHGTCC				
S2pet	TGDK.YSRFKEDNIIMVLERHWQMKFDENYAKYBQPLWSYKYKHGTCC				
S3pet	DGDKFVSFLKDRIVEDLERHWQMKFDEKFAKIQPLWTEYKHGTCC				
S2sol	.RSYAYNALTNVREKSKLDLDRWPDLTSSKMSMTKKEQKFWYKYKHGTCC				
S3sol	.KIVKYNKIKDEHKIDALEYGVNPLTTTEAVSKEDQVFWGKYKHGTCC				
	110	120	130	140	150
Sf11	QKRYMQNTYFGLALRLKDKFDLLRLTLQTHRIIPGSSY.TFQDIFDAIKTV				
Sz	IFRYMQNTYFGLALRLKDKFDLLRLTLQTHRIIPGSSY.TFQDIFDAIKTV				
S1nic	LFQYMQNTYFGLALRLKDKFDLLRLTLQTHRIIPGSSY.TVQKYNNTIKAI				
S2nic	SDKFDREQYFDLALRLKDKFDLLRLTLQTHRIIPGSSY.TVQKYNNTIKAI				
S3nic	SESYMLTQYFDLALRLKDKFDLLRLTLQTHRIIPGSSY.TVQKYNNTIKAI				
S6nic	SEIYMQNTYFGLALRLKDKFDLLRLTLQTHRIIPGSSY.TVQKYNNTIKAI				
S1pet	SKIYMQKAYFLLALRLKDKFDLLRLTLQTHRIIPGSSY.TFQDIFDAIKTV				
S2pet	KNLYDQAYFLLALRLKDKFDLLRLTLQTHRIIPGSSY.TFQDIFDAIKTV				
S3pet	SNLYDQAYFLLALRLKDKFDLLRLTLQTHRIIPGSSY.TFQDIFDAIKTV				
S2sol	EKLYMQAQYFNLTMNLKDKFDLLRLTLQTHRIIPGSSY.LLSNGRPLRLQ				
S3sol	TDLYDKDAYFDLALRLKDKFDLLRLTLQTHRIIPGSSY.SSNIQNAVSKV				
	160	170	180	190	200
Sf11	SQE.NPDIKCAEV.TKGTPELVEIGICFTFNADSMFRCPQSDTCOKT.AKVLFR				
Sz	TNQVDPDLKCVH.IKGVRELVEIGICFTFNADSMFRCPQSDTCOKT.AKVLFR				
S1nic	TGK.FPNLTCKNQ....MELKEIGICFDQKVKWVIDCPNPKICKPT.NKGVMPF				
S2nic	TGG.FPNLTCKNQ....RELKEIGICFDQKVKWVIDCPNPKICKPT.NKGVMPF				
S3nic	TQG.YPNLSCTKQ....MELKEIGICFDQKVKWVIDCPNPKICKPT.NKGVMPF				
S6nic	TGK.YPNLSCTKQ....QELNEVGICFDQKVKWVIDCPNPKICKPT.NKGVMPF				
S1pet	TNQVDPDLKCVH.IKGVRELVEIGICFTFNADSMFRCPQSDTCOKT.AKVLFR				
S2pet	TNNKDPDLKCVH.IKGVRELVEIGICFTFNADSMFRCPQSDTCOKT.AKVLFR				
S3pet	TNNKDPDLKCVH.IKGVRELVEIGICFTFNADSMFRCPQSDTCOKT.AKVLFR				
S2sol	TMKVFPFLKCIDN.NGIMELLEVGICFDPAATKVPICHRPWICHADENTRIELVK				
S3sol	TQGV.PHVTGPNRFRKTSSELLEALCFDPAQNVICHRPCKTNSKGTGKIFTP				

FIG. 3. Amino acid alignment of 11 S-allele proteins. The bold residues are sites at which two species share an amino acid polymorphism. Simulations were performed as in Table 2 to test the significance of the number of shared polymorphisms. One allele of each of two species was randomly drawn, and random mutations in nucleotide sequences were generated, based on the number of observed polymorphisms in each species. These sequences were then translated, and the amino acid sequences were compared to ascertain the number of shared amino acid polymorphisms. The numbers of shared amino acid polymorphisms observed in the data (and the count of cases in the random simulations having a greater number of shared polymorphisms) were as follows: *N. alata*-*P. inflata*, 17 (8/1000); *N. alata*-*S. chacoense*, 23 (0/1000); *P. inflata*-*S. chacoense*, 6 (0/1000); polymorphisms shared by all three species, 4 (12/1000).

stitutions over synonymous substitutions. Further analyses of the rates of synonymous and nonsynonymous substitution and of the coalescence properties of self-incompatibility alleles will be presented elsewhere. Because of its wide

distribution over nearly half of the families of flowering plants (1), gametophytic self-incompatibility may prove to be one of the most general mechanisms for generating trans-specific polymorphism.

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