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Recent Development of Antitumor Agents from Chinese Herbal Medicines; Part I. Low Molecular Compounds

Abstract

A series of low molecular compounds from Chinese herbal medicines which have proved to be, in some cases, highly effective especially in tumor therapy, is listed here (part II will deal with high molecular compounds, to be published in the next issue). In contrast to synthetic agents used in cancer chemotherapy, these natural compounds have relatively low toxicities. Many of the clinical studies referred to in this paper have been carried out on Asians. Because genetic factors influence enzyme levels, sometimes leading to striking differences in metabolism and pharmacokinetics of drugs, results obtained in

clinical studies carried out in China are not 100% transferable to the European population. The mechanisms of action of these compounds are manifold, consisting of reactions with DNA bases, intercalation in DNA, inhibition of topoisomerases, inhibition of protein kinases, induction of apoptosis etc. Some of the compounds have interesting structural features, that may be used as lead structures for the development of further antitumor agents.

Key words

Antitumor agents · Chinese herbal medicines · low molecular natural compounds · traditional Chinese medicine

Introduction

As a consequence of the experience gathered over thousands of years, Chinese herbal medicines are considered as a rich source for the discovery of new drugs. In the search of new therapeutic agents, many compounds with new structural features and mechanisms of action have been isolated from Chinese herbal medicines. In recent years, a number of Chinese herbs and their active principles were reported to be antineoplastic and cytotoxic both in experimental and clinical studies. The cytotoxic and antineoplastic mechanisms of these compounds involve DNA intercalation, inhibition of DNA topoisomerases and protein kinases, induction of apoptosis, covalent binding to enzymes of biological importance and some unknown mechanisms. The natural compounds are relatively non-toxic as compared to some syn-

thetic agents. In the present paper, a summary of the most important findings on cytotoxic and antineoplastic compounds of low molecular weight from Chinese herbal medicines is given in order to contribute to the further development of antitumor drugs.

Alkaloids

Phenanthridine Alkaloids

Lycobetaine (ungeremine, AT-1840), a quaternary phenanthridinium alkaloid from some Amaryllidaceous plants, such as *Lycoris radiata* Herb. [1] was reported to be antineoplastic. In reviews and in meeting reports [2], [3], lycobetaine was mentioned to be active in the treatment of cervical, ovarian, gastric and other can-

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Dedication

In memory of Prof. Dr. Dietrich Schmähl, Deutsches Krebsforschungszentrum, Heidelberg

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Received May 27, 2002 · Accepted November 16, 2002

Bibliography

Planta Med 2003; 69: 97–108 · © Georg Thieme Verlag Stuttgart · New York · ISSN 0032-0943

cers in clinical trials. The overall response in 233 cases of different cancers was reported to be about 35% [3]. No significant myelotoxic, cardiotoxic and hepatotoxic side effects have been observed [2].

In experimental studies, lycobetaine was found to inhibit the growth of S180 [4] and KB [5] tumor cell lines *in vitro* and to be significantly active against Ehrlich ascites carcinoma, ascites hepatoma, leukemias L1210 and P388, Lewis lung carcinoma and Yoshida ascites sarcoma in mice or in rats by *i.p.* injection. In nude mice bearing human gastric cancer xenografts, lycobetaine extended the survival time and decreased the tumor size. Moreover, it was shown to have a direct cytotoxic effect on stomach cancer cells *in vitro* even at low concentrations and to arrest carcinoma cells in the G₂/M phase as demonstrated by flow cytometry [6].

A study on the interaction of lycobetaine with calf thymus DNA revealed that lycobetaine intercalates with DNA base pairs, especially the GC-pair [7]. Lycobetaine did not bind covalently to DNA [8]. Studies on structure-activity relationships revealed that the calculated interaction energy of lycobetaine analogues with double-stranded oligonucleotides correlated with their anticancer potency [9], [10]. According to the 3-dimensional structure patterns of the drug-oligonucleotide complex, the quaternary nitrogen atom in lycobetaine plays an important role in the formation of hydrogen bonds between the compound and the oligonucleotide [11]. The betaine and a methylenedioxy group in lycobetaine are supposed to be critical for its antitumor activity [2].

Recently, we have found that lycobetaine is toxic for topoisomerases I and II, and that it strongly inhibits the growth of human tumor xenografts *in vitro* and *in vivo*. The IC₅₀ values of lycobetaine in the clonogenic assay against 21 human tumor xenografts of various tumor types range from 0.002 to 27.5 μ M with a mean IC₅₀ of 0.8 μ M. Intraperitoneal administration of lycobetaine at a dose of 30 mg/kg on days 1–5 and 8–12 to nude mice bearing the large cell lung carcinoma LXFL529 resulted in a significant growth delay of the tumors [12], corresponding to about 40% of the LD₅₀ by single *i.p.* injection [13]. Lycobetaine was localized predominately in the nucleus, it competed with ethidium bromide for intercalation into calf thymus DNA and displaced the DNA minor groove binder Hoechst 33258. At growth inhibitory concentrations, lycobetaine inhibited topoisomerases I and II, stabilized the covalent DNA-topoisomerase I intermediate, the so-called cleavable complex, and induced apoptosis. Dose-dependent induction of DNA strand breaks was detected by single cell gel electrophoresis (comet assay) [12].

Lycobetaine is structurally related to the phenanthridine alkaloid lycorine as a major Amaryllidaceous constituent and can be easi-

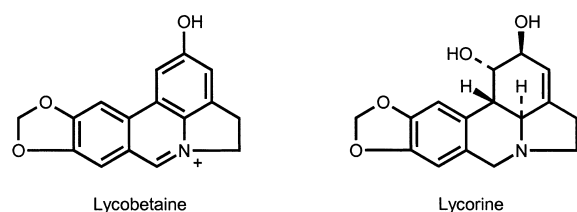


Fig. 1 Lycobetaine and lycorine.

ly obtained in high yield from this constituent by oxidation with SeO₂ [2]. Lycorine was also reported to be cytotoxic [14]. Recently, lycorine has been found to be highly active in inhibiting a number of human tumor cell lines *in vitro*, including LXFL529L, Molt4, HL60, K562, U937 [15], GXF251L, and CFX94L [16]. In contrast to lycobetaine, lycorine did not compete with ethidium bromide for intercalation into calf thymus DNA and did not displace the DNA minor groove binder Hoechst 33258, indicating that its cytotoxic mechanism is different from that of lycobetaine [15], [16]. In addition, lycorine was reported to induce flat morphology in K-Ras-transformed fibroblasts *in vitro* [17].

Benzo[c]phenanthridine Alkaloids

Among the benzo[c]phenanthridine alkaloids, nitidine, fagaronine and ethoxychelerythrine are the major components in *Radix Zanthoxyli* (Liangmianzhen), the root of *Zanthoxylum nitidum* (Roxb.) DC (Rutaceae) [18] which is listed in the Chinese Pharmacopoeia [19]. Chelerythrine, chelidonine and the related benzo[c]phenanthridine alkaloid sanguinarine are known to be the major components in the whole plant of *Chelidonium majus* L. (Papaveraceae), a medicinal herb used in European countries and in China [20].

Nitidine was reported to possess antineoplastic activity against both L1210 and P388 leukemias, Lewis lung carcinoma, and B16 melanoma in mice [21]. It increased the life span of mice inoculated with Ehrlich ascites tumor, caused a decrease in the mitotic index and size of the tumor cells, and inhibited DNA and RNA synthesis in tumors [22]. Ethoxychelerythrine also showed inhibitory activity against Ehrlich ascites carcinoma cells [23]. Nitidine chloride was noted to be effectively used in the clinical treatment of chronic myelocytic leukemia [24].

Nitidine and fagaronine were reported to bind to calf thymus DNA by intercalation [25] and to be toxic to topoisomerases I and II [26]. Nitidine exhibited strong stabilization of the covalent binary complex formed between topoisomerase I and DNA [18]. Yeast cells expressing human DNA topoisomerase I were specifi-

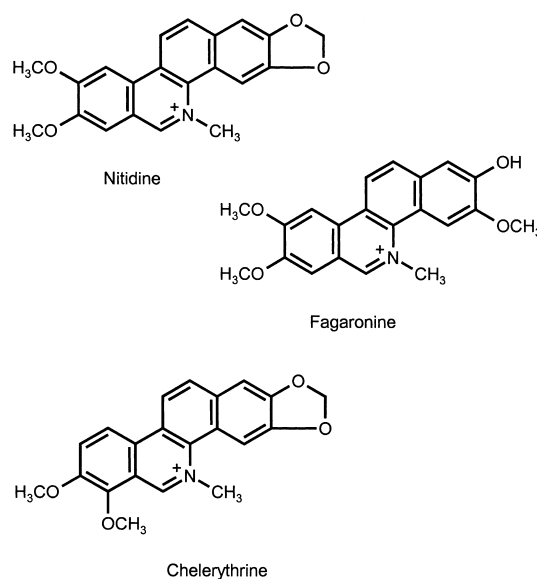


Fig. 2 Nitidine, fagaronine, and chelerythrine.

cally sensitive to nitidine [27]. Unlike camptothecin as reference compound, nitidine and fagaronine bound directly to and mediated the unwinding of B form DNA. Inhibition of topoisomerase II by nitidine was observed at the high concentration of 40 μM in comparison to the inhibition of topoisomerase I at 0.15–0.3 μM [28].

The antitumor mechanism of fagaronine is also believed to be due to inhibition of topoisomerases [29], [30]. Fagaronine is a DNA major groove intercalator, does not show any sequence specificity of DNA intercalation, but its highly electronegative oxygen of the hydroxy group is shown to be an acceptor of the hydrogen bond of the amino group of guanine in DNA [31]. In contrast, *N*-demethylfagaronine was neither cytotoxic, nor effectively interacted with DNA, suggesting that the quaternary nitrogen atom is an important prerequisite for its biological efficacy [32].

Chelerythrine was reported to be a specific protein kinase C (PKC) inhibitor [33], [34]. It completely suppressed the growth of GI-101A breast tumor cells stimulated by hydroxychloroquine and prednisone [35], blocked the expression of vascular endothelial growth factor (VEGF) mRNA in GI-101A and HL-60 cells stimulated by 12-*O*-tetradecanoylphorbol 13-acetate (TPA) or diethylstilbestrol [36], and inhibited the increased proliferation of MCF-7 cells stimulated by thymeleatoxin [37]. Chelerythrine also inhibited the proliferation of the human prostate cancer cell line, PC3 [38], and gastric cancer cell line, AGS [39]. Incubation of AGS cells with chelerythrine resulted in an arrest of cells in the G_0/G_1 phase, in induction of apoptosis and in an elevation of p53, p21, c-Myc, and Bax [39]. Chelerythrine was further reported to exhibit cytotoxic activity against a series of radioresistant and chemoresistant human squamous cell carcinoma lines. The carcinoma cells undergo apoptosis rapidly after treatment with chelerythrine *in vitro*. The antitumor effect of chelerythrine was also demonstrated in nude mice against a radioresistant, chemoresistant and p53-deficient human head and neck squamous cell carcinoma line SQ-20B with significant tumor growth delay and minimal toxicity (40). Chelerythrine also inhibited taxol-mediated polymerization of rat brain tubulin and inhibited colchicine and podophyllotoxin binding to tubulin with IC_{50} values of about 60 μM [41].

Sanguinarine was reported to be a potent inhibitor of the catalytic subunit of rat liver protein kinase A (PKA) with an IC_{50} of 6 μM , but a relatively poor inhibitor of PKC with an IC_{50} of greater than 200 μM [42]. Sanguinarine potently inhibited the growth of human keratinocytes with an IC_{50} of 0.2 μM [43]. It decreased concentration-dependently the viability of human epidermoid carci-

noma A431 cells at lower concentrations than of normal human epidermal keratinocytes. Sanguinarine treatment of A431 cells resulted in an induction of apoptosis but did not lead to formation of a DNA ladder in normal keratinocytes, even at higher concentrations [44]. Sanguinarine interacted with calf thymus DNA and altered its secondary structure. The maximum of sanguinarine-binding to DNA in buffer of low ionic strength and acidic pH and the number of bound alkaloid molecules per base pair, at saturation, is higher in G-C rich DNA than in A-T rich DNA [44].

Protoberberine Alkaloids

Like benzo[*c*]phenanthridine, protoberberine is also a four-ring system but with one saturated bond. Protoberberine alkaloids occur in numerous plants, including several medicinal plants. Berberine is the most studied protoberberine alkaloid, widely distributed in plants used in traditional Chinese medicine. Plant items listed in the Chinese Pharmacopoeia containing berberine and related protoberberine alkaloids are *Rhizoma Coptidis* (Huanglian), the rhizome of *Coptis chinensis* Franch., *C. deltoidea* C.Y. Cheng et Hsiao, or *C. teeta* Wall. (Ranunculaceae); *Caulis Mahoniae* (Ganglaomu), the stem of *Mahonia healei* (Fort.) Carr. or *M. fortunei* (Lindl.) Fedde (Berberidaceae) and *Cortex Phellodendri* (Huangbo), the stem bark of *Phellodendron chinense* Schneid. or *P. amurense* Rupr. (Rutaceae).

Berberine was reported to possess significant cytotoxicity against some human cancer cell lines, P388 murine leukemia cells [45], and 9L rat glioma cell line [46]. Recently, Iizuka et al. have reported that the extract of *Rhizoma Coptidis* and berberine significantly inhibited the proliferation of six esophageal cancer cell lines *in vitro* in a concentration-dependent manner [47].

Chi et al. reported on a flow-cytometric study of the effect of berberine on human hepatoma HepG₂ cells. Continuous exposure of HepG₂ cells to berberine resulted in a concentration-dependent growth inhibition of the cells. Berberine treatment caused a significant reduction of the S phase fraction of HepG₂ cells [48] and an arrest of gastric cancer cells in the G_2/M phase [49]. Esophageal cancer cells treated with the extract of *Rhizoma Coptidis* showed an accumulation in the G_0/G_1 phase and a relative decrease of the S phase [47]. Concentration-dependent effects of berberine were also reported on cell cycle and apoptosis in Balb/c 3T3 cells [50]. Chang et al. reported that berberine down-regulated K-Ras2 gene expression associated with morphologic differentiation in human embryonal carcinoma cells [51]. Furthermore, berberine and some related protoberberine alkaloids induced apoptosis in murine thymocytes [52] and in promyelocytic leukemia HL60 cells [53]. Berberine was also found to be a

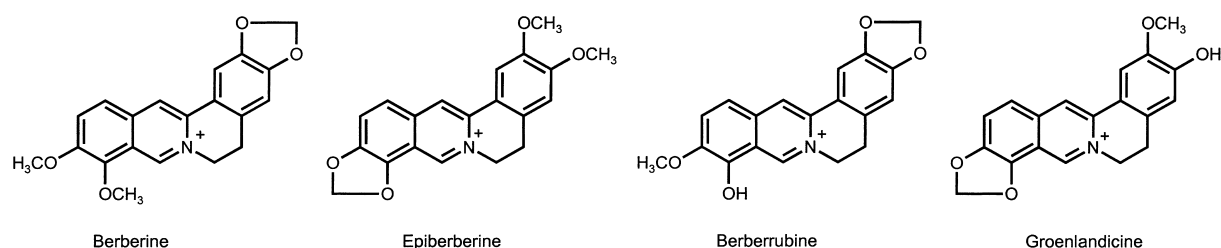


Fig. 3 Berberine, epiberberine, berberrubine, and groenlandicine.

DNA triplex binder [54] and to enhance the cytotoxicity of clinically used antitumor agents, such as nucleotides [55] and nitrosoureas [56] *in vitro*. It was also reported that berberine up-regulated the multidrug-resistant transporter (pgp-170) expression in two oral (KB, OC2), two gastric (SC-M1, NUGC-3) and two colon (COLO 205, CT 26) cancer cell lines [57], [58].

The artifact berberrubine, formed by demethylation of berberine, was found to be highly cytotoxic against P388 leukemia, L1210 leukemia, B16 melanoma and some human cancer cell lines [59]. Makhey et al. reported that berberrubine functions as a potent inhibitor of topoisomerase II *in vitro* [60]. The protoberberine alkaloids epiberberine and groenlandicine were identified as topoisomerase I inhibitors *in vitro* [61]. Topoisomerase II-mediated DNA cleavage assays showed that berberrubine poisons the enzyme by stabilizing the enzyme-DNA complexes. Berberrubine induced DNA cleavage in a site-specific and concentration-dependent manner. Comparison of the cleavage pattern of berberrubine with that of etoposide revealed that berberrubine represents a new class of antitumor agent with topoisomerase II poisoning activity as well as catalytic inhibition. The step at which berberrubine induces a cleavable complex is possibly different from that of etoposide [62].

1,4-Benzoquinone Derivatives

From the seed of *Iris pallasii* var. *chinensis* (Iridaceae), a 1,4-benzoquinone derivative with an unsaturated aliphatic side chain (irisquinone A), has been isolated [63]. The seed of *I. pallasii* var. *chinensis* is used in Chinese folk medicine as a fertility regulating agent and for the treatment of malignant diseases.

Irisquinone A was found to be cytotoxic and antitumorogenic. In mice tumor xenografts, it showed growth inhibitory activity against cervical cancer U₁₄ and Ehrlich carcinoma by *i.p.* application and against lymphosarcoma by *i.p.* and oral administration [64]. Oral treatment of mice bearing U₁₄ tumor with irisquinone A at a dosage of 100 mg/kg or *i.v.* treatment at a dosage of 5 mg/kg once every other day for 5 cycles, starting 24 h after implantation of the tumor, caused a tumor inhibition rate of 35–55% [65]. The LD₅₀ of irisquinone A in mice was about 28 mg/kg after *i.p.* and about 2.8 g/kg after oral administration. The chemotherapeutic indices (LD₅₀/ED₅₀) of irisquinone A were estimated to be 5 by *i.p.* administration and 14 by oral administration.

Radiosensitizing effects of irisquinone A were found *in vitro* against the tumor cell lines U₁₄ [64], S-180V [65], HeLa [66], against Ma 7373 breast cancer cells in mice, and against human intestinal muc adenocarcinoma in nude mice [66]. The mecha-

nism of this effect was considered to be an inhibition of oxygen consumption and depletion of glutathione in tumor cells [66]. In a clinical trial, irisquinone A, given orally to 558 patients with cancer of the lung and esophagus, or with superficial metastatic cancer during radiotherapy, significantly contributed to the reduction of tumor size and to the prolongation of survival time of the patients [64].

Related 1,4-benzoquinone derivatives were isolated from the Chinese medicinal plant *Ardisia japonica* (Thunb.) Bl. (Myrsinaceae). One of them is embelin, a 2,5-dihydroxy-1,4-benzoquinone with a saturated side chain. Embelin was found to exhibit significant antitumor activity in rats against autochthonous fibrosarcomas induced by methylcholanthrene and it prolonged the survival time of the animals [67]. An *in vitro* study with embelin using a fibrosarcoma cell line showed a concentration-dependent decrease in thymidine uptake and glutathione levels of the tumor cells [68].

Diterpenes

Rabdosia diterpenes

Several *Rabdosia* species (Lamiaceae) native to China are used in folk medicine as antitumor or anti-inflammatory agents. Studies on the chemical constituents of the leaves and stems and on their biological activities have been carried out with various *Rabdosia* species, especially *Rabdosia rubescens* (Hemsl.) Hara. Diterpene compounds, such as oridonin and ponicedin, are derived from kaurane. The diterpenes were found to be the cytotoxic principles in *Rabdosia* species [69].

115 patients suffering from inoperable esophageal carcinoma were treated with chemotherapy alone (group A) or with chemotherapy plus *R. rubescens* (group B); in group A, 10 out of 31 patients (32.3%) treated with chemotherapy alone, responded to the treatment, including 2 partial responses (greater than 50% tumor regression) and 8 minimal responses. In group B, 59 out of 84 patients (70.2%) responded to the treatment, including 10 complete responses (100% tumor regression), 16 partial and 33 minimal responses. The one-year survival rates of group A and group B were 13.6% and 41.3%, respectively. No significant differences were observed between the two groups regarding the side effects alopecia, anorexia, nausea and hyperpyrexia which occurred in more than 30% of patients [70]. Between August 1974 and January 1987, 650 patients with moderate and advanced esophageal carcinoma were treated with a combination of chemotherapy and *R. rubescens* or *R. rubescens* plus different traditional Chinese patent medicines. Forty patients survived for over 5 years (5-year survival rate 6.15%); 32 for over 6 years; 23

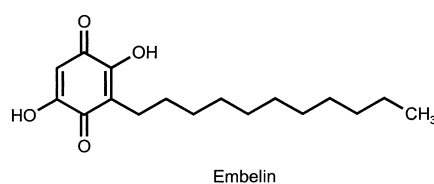
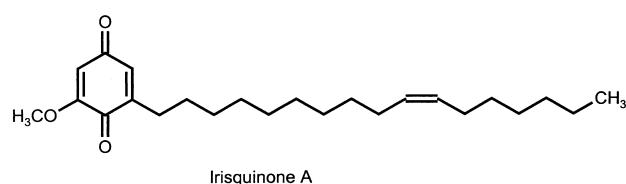


Fig. 4 Irisquinone A and embelin.

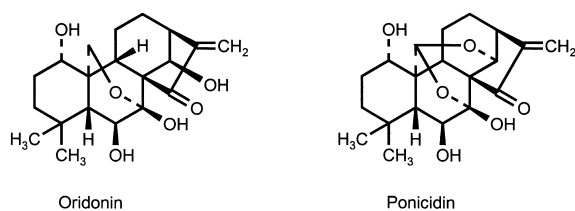


Fig. 5 Oridonin and ponocidin.

for more than 10 years, and 5 for more than 15 years [71]. PC-SPES, a combination of eight herbs including *R. rubescens* has been tested for the treatment of prostate cancer showing significantly decreased serum prostate specific antigen (PSA) of the patients [72], [73]. In a review, it was mentioned that oridonin and ponocidin were tested in clinical trials for the treatment of esophageal cancer [74].

Oridonin was reported to be cytotoxic against Ehrlich ascites carcinoma [75] and L1210 leukemia [76] in mice after *i. p.* injection. The cell killing rates of oridonin (15 mg/kg *i. p.*) on day 5 and day 8 in L1210 cells were 73% and 39%. The G_2 and S phases of L1210 cells were prolonged, while the G_1 phase was unchanged [76]. In *in vitro* investigations, oridonin exhibited inhibitory effects on the proliferation of the human gastric adenocarcinoma cell line (MGC80-3) and esophageal cancer cell line (CaEs-17) at concentrations below 15 $\mu\text{g/ml}$ [77]. An extract of *Rabdosia rubescens* exerted a cytotoxic effect on cisplatin-sensitive human ovarian cancer A2780 cells with an IC_{50} of about 0.6 mg/mL [78]. Oridonin was found to inhibit DNA, RNA, and protein syntheses in L1210 cells *in vitro* in a concentration-dependent manner. Inhibition of DNA and RNA synthesis was fast but reversible, whereas the inhibition of protein synthesis was strong and long lasting [79]. Oridonin inhibited DNA synthesis also in a cell-free system [80].

Oridonin (7.5 mg/kg for 7 days) and cisplatin (0.4 mg/kg for 4 days) showed synergistic antineoplastic effects in mice bearing Ehrlich ascites carcinoma, S180 or P388 leukemia and there was a distinct growth inhibition of Ehrlich ascites carcinoma cells *in vitro*. In the S180 cell culture, the IC_{50} values of cisplatin were reduced to 1/3.4 and 1/6.7 by cotreatment with oridonin at concentrations of 0.5 and 1 $\mu\text{g/ml}$. A greater amount of DNA cross-links and DNA-protein cross-links in S180 cells was detected when the cells were treated with cisplatin plus oridonin instead of cisplatin alone [81]. Synergistic antitumor effects of oridonin with bleomycin A_5 were also reported [82]. Several diterpenes which are structurally related to oridonin, were also reported to exert cytotoxic and antitumor activities [83], [84].

The mechanism of oridonin activity was postulated to be due to covalent binding of oridonin to a specific site of enzymes in tumor cells [85]. Oridonin and related diterpenes bearing an α -methylenecyclopentanone structure are electrophilic and can be considered as weak alkylating agents [86]. The reactivity of the α -methylene group was examined using the reaction of oridonin with adenosine and cytidine as nucleic acid model compounds and with thiols, L-cysteine, L-lysine and L-serine as model compounds representing relevant constituents of active centers of enzymes. The reaction with thiols proceeded easily under mild

conditions to give the corresponding thioether adducts, the adduct with L-cysteine being formed nearly quantitatively. However, oridonin did not react with adenosine and cytidine under the same conditions [87].

Quassinoids

Fructus Bruceae (Yadanza), the mature fruit of *Brucea javanica* (L.) Merr (Simaroubaceae), is listed in the Chinese Pharmacopoeia and is used as an antimalarial and antidiarrhetic agent and for the treatment of warts and corns by external application. A number of diterpene quassinoids characterized by an epoxy binding between positions 13 and 20 and a hydroxy function at position 5, were isolated from the fruits of *B. javanica*. Bruceantin and brusatol are two important representatives of these constituents possessing antileukemic activity. Structurally, bruceantin and brusatol differ from each other only in the acid moiety [88].

Kupchan et al. have discovered the antileukemic activity of bruceantin [89]. Bruceantin exerted growth inhibitory effects *in vitro* and *in vivo* against a series of tumor cell lines but did not show significant effects in clinical studies against solid tumors [90], [91], [92].

In contrast, an oil emulsion of the fruits of *B. javanica* was recently reported to show clinical efficacy. From 35 patients with recurrent metastatic gastrointestinal cancers treated with the oil emulsion, 24 patients (46%) showed a reduction of tumor size and improvement of clinical symptoms [93]. A clinical trial on 68 patients with brain metastasis in lung cancer treated with 10% oil emulsion combined with radiotherapy resulted in an improvement of living quality and in prolongation of median survival time (15 months) compared to radiotherapy alone (10 months) [94]. Clinical *i. v.* application of the oil emulsion was reported to improve the intracranial hypertension caused by brain metastasis from lung cancer [95].

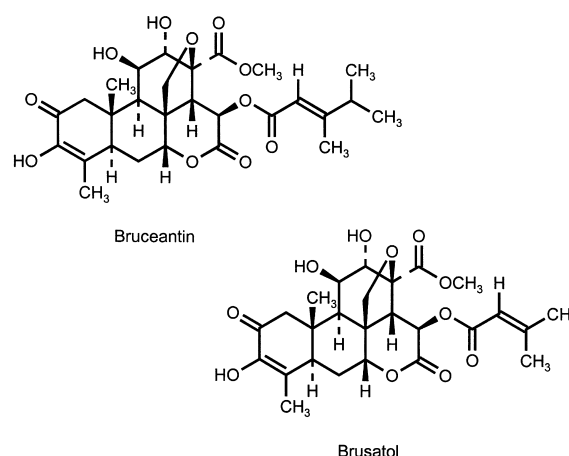


Fig. 6 Bruceantin and brusatol.

Flavones

Scutellaria Flavones

Radix Scutellariae (Huangqin), the root of *Scutellaria baicalensis* Georgi (Lamiaceae), is known to contain a number of flavones,

namely baicalein, wogonin and the corresponding 7-*O*- β -D-glucopyranosiduronides baicalin and wogonosides [96], [97]. It is listed in the Chinese Pharmacopeia and is a well known traditional Chinese medicine.

Baicalin and baicalein were reported to exhibit growth inhibitory activity towards the human hepatoma cell lines PLC/PRF/5 and Hep-G2 as well as the human pancreatic cancer cell line BxPC-3 with an IC_{50} of 20 μ g/mL for baicalin and 50 μ g/mL for baicalein [98]. Cytotoxic effects of baicalein, baicalin, and wogonin were also observed on the human bladder cancer cell lines KU-1 and EJ-1, and on a murine bladder cancer cell line (MBT-2). All three substances inhibited cell proliferation *in vitro* in a dose-dependent manner with baicalin as the most active compound. An *in vivo* study showed that the root extract of *S. baicalensis* at an oral daily dose of 10 mg per mouse for 10 days significantly inhibited the growth of MBT-2 cells implanted to C3H/HeN mice [99]. We have also found that baicalein, baicalin, wogonin, wogonoside and skullcapflavone II (neobaicalein) inhibited the growth of the human tumor cell lines LXFL 529L (large cell lung carcinoma) and HL60 at a micromolar range [100].

Baicalein strongly inhibited DNA topoisomerase II, probably by stabilizing the covalent enzyme-DNA intermediate in a ternary complex. It also inhibited the proliferation of three human hepatocellular carcinoma cell lines [101]. A structure-activity analysis revealed that flavones with hydroxy groups at the 5, 7, 3' and 4' positions favored topoisomerase II-mediated DNA cleavage. Recently, we have found that baicalein also inhibits topoisomerase I by stabilizing the otherwise cleavable complex [100]. Formation of the cleavable complex might contribute to the growth inhibitory activity of flavones on human tumor cell lines [102]. Baicalin was also found to inhibit cell proliferation and to induce apoptosis in several human prostate cancer cell lines [103].

Baicalin, wogonin, skullcapflavone II and wogonoside exhibited only weak inhibitory properties on tyrosine kinase activity of the epidermal growth factor receptor (EGFR) with IC_{50} values exceeding 60 μ M. In contrast, we found that baicalein is a potent inhibitor of EGFR with an IC_{50} of 1.1 μ M. In A431 cells overexpress-

sing EGFR, baicalein was significantly more active than in other cell lines [100]. Baicalein strongly inhibited human T-lymphoid leukemia cell proliferation with an IC_{50} of about 5 μ M. Protein tyrosine kinase activity in human T-lymphoid leukemia cells was significantly reduced by baicalein [104].

Moreover, baicalein has been reported to inhibit the activity of cAMP phosphodiesterases (PDE). We confirmed the PDE inhibitory activity of baicalein and further showed that baicalein is able to inhibit the cAMP-specific isoenzyme family PDE4 with an IC_{50} of 10 μ M. The respective glucuronide baicalin is only a weak PDE4 inhibitor. However, when LXFL 529L whole cells were incubated with baicalein, no inhibition of the intracellular PDE activity was observed and the intracellular cAMP level remained unchanged [100].

Baicalein potently inhibited the growth of a human breast carcinoma cell line, MDA-MB-435 with an IC_{50} of about 6 μ g/mL and was more active than flavones isolated from citrus fruits including hesperetin and naringenin [105]. Baicalein and related flavones also inhibited the proliferation of estrogen receptor-positive MCF-7 human breast cancer cells; the inhibition was not reversible by an addition of estrogen [106]. Baicalein and wogonin strongly inhibited xanthine oxidase, indicating that they might be useful for the remission of brain tumors, since xanthine oxidase serum levels are increased in tissues of brain tumors [107]. Baicalein as an α -glucosidase inhibitor suppressed *in vitro* invasion and *in vivo* metastasis of mouse melanoma cells [108].

Polymethoxylated Flavones

Fructus Aurantii (Zhiqiao) and Fructus Aurantii immaturus (Zhishi), the immature fruits of *Citrus aurantium* L. (Rutaceae) and its cultivated variants are listed in the Chinese Pharmacopeia. They contain polymethoxylated flavones such as tangeretin, nobiletin, sinensetin and auranetin.

The polymethoxyflavones from *C. aurantium* were found to possess a wide spectrum of biological activities. Tangeretin, nobiletin and related polymethoxyflavones were reported to exert antiproliferative activities against a number of tumor cell lines [109] and to induce differentiation of HL60 cells *in vitro* in a concentration-dependent manner [110]. HL60 cells treated with these flavones differentiated into mature monocyte/macrophage. A structure-activity study revealed that the *ortho*-catechol moiety in the 2-phenyl nucleus of flavones is a prerequisite for their antiproliferative activity and for the induction of differentiation in HL60 cells. A hydroxy group at C3 and a methoxyl group at C8 enhanced the activity [111]. Tangeretin and nobiletin also inhibited the invasion of mouse MO4 cells into embryonic chick heart fragments *in vitro* [112].

An ethyl acetate extract of orange juice did not affect the initial uptake rate of [3H]vinblastine by Caco-2 cells but significantly increased the steady-state uptake of cyclosporin A as a P-glycoprotein inhibitor. 3,3',4',5,6,7,8-Heptamethoxyflavone, tangeretin and nobiletin were found to be active compounds. They all increased the steady-state uptake of [3H]vinblastine by Caco-2 cells in a concentration-dependent manner. The ethyl acetate extract and the polymethoxyflavones also increased steady-state [3H]vinblastine uptake by LLC-GA5-COL300 cells, a cell line

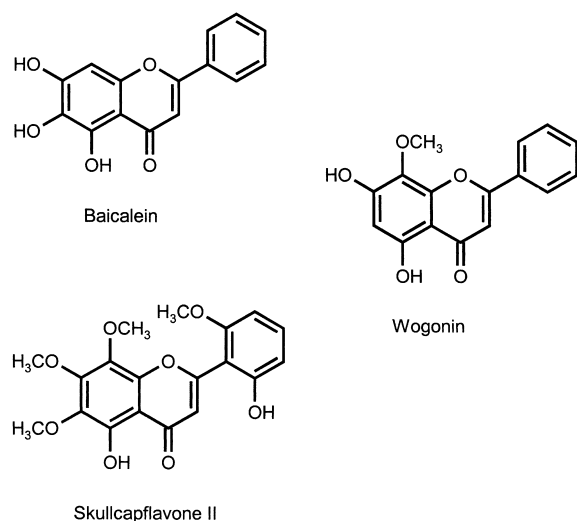


Fig. 7 Baicalein, wogonin, and skullcapflavone II.

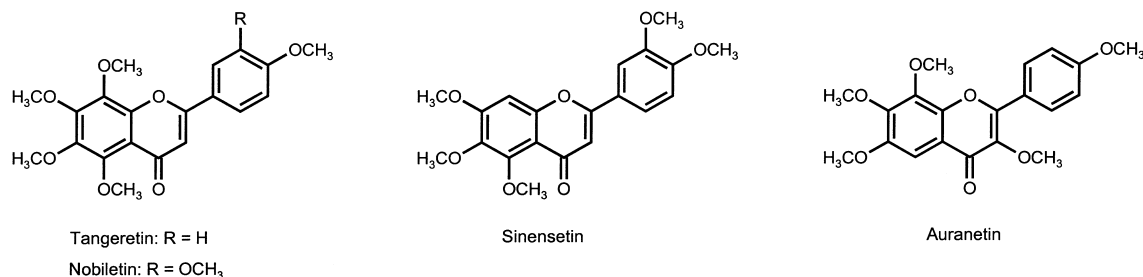


Fig. 8 Tangeretin, nobiletin, sinensetin, and auranetin.

transfected with human MDR1 cDNA [113]. The ethyl acetate extract of grapefruit juice, orange juice and the polymethoxyflavones was also reported to increase the uptake of [³H]vincristine into adriamycin-resistant human myelogenous leukemia K562/ADM cells [114].

Indigoid Bisindoles

In the 1960's, Chinese scientists began to study the active principle in the Chinese patent medicine "Danggui Longwei Wan" which had been effectively used as an antileukemic agent [115]. This official patent medicine contains 11 ingredients, including *Indigo naturalis*. The antileukemic activity was found to be associated with indirubin, a minor component in *Indigo naturalis* [116]. Indirubin is a bisindole derivative with a 3,2'-linkage, while indigo and isoindigo bisindoles possess 2,2'- and 3,3'-linkages, respectively. In experimental studies, indirubin inhibited Lewis lung carcinoma in mice and W256 sarcoma in rats [117].

Treatment of 314 patients suffering from chronic myelocytic leukemia (CML) with indirubin at daily oral doses of 300–450 mg per patient resulted in 82 complete remissions (26%), 38 partial remissions (12%) and 87 beneficial effects (28%). The overall response was 87% [118]. In a further study, treatment of CML patients with indirubin for 1.5–6 months markedly increased 5'-

nucleotidase activity of leukocytes in some cases with a palliative effect [119]. In another clinical trial including 57 cases of CML, treatment with indirubin resulted in a median survival of 31.5 months without obvious side effects [120].

Because of the manifold findings on antileukemic effects of indirubin, different studies were carried out on its mechanisms of action. Indirubin turned out to inhibit cell-free DNA synthesis catalyzed by partially purified DNA-dependent DNA polymerase from Ehrlich ascites tumor cells in a concentration-dependent manner. This inhibition could not be reversed by increasing the DNA concentration, but could be abolished by increasing the enzyme concentration. The strongest inhibition resulted after preincubation with the enzyme and DNA, suggesting that indirubin, DNA and the enzyme formed a tertiary complex [121]. The DNA polymerase 1 activities of chronic myelocytic leukemia cells from the peripheral blood of patients decreased significantly after treatment with indirubin. Similar results have been observed with *Escherichia coli* DNA polymerase 1 *in vitro* [122].

Intraperitoneal or s.c. administrations of indirubin at a daily dose of 200 mg/kg partially inhibited the incorporation of [³H]thymidine into the DNA of W256 sarcoma cells but not the incorporation of [³H]uridine into RNA and of [¹⁴C]phenylalanine into protein [123]. [³H]Thymidine incorporation into DNA of liver and spleen of healthy control mice was not affected by *i.p.* injection of indirubin, whereas hepatic and splenic [³H]thymidine incorporation was inhibited in mice bearing L7212 leukemia [124]. After incubation of indirubin with calf thymus DNA, the λ_{max} 207 nm of indirubin shifted towards a longer wavelength with decreasing absorbance. The binding of indirubin to DNA was rather weak, since indirubin molecules were easily released during precipitation with alcohol or gel filtration. The amount of bound [³H]indirubin was concentration dependent. Calf thymus DNA bound 46 indirubin molecules per 1000 nucleotides [125].

Electron microscopic examination of peripheral blood samples and leukocytes of bone marrow from patients with chronic granulocytic leukemia treated with indirubin showed a swelling of the cell nucleus membrane, swelling and degeneration of the rough endoplasmic reticulum, and condensation of chromosomes [126]. Measuring the labelling index of bone marrow cells from patients with chronic myelocytic leukemia *in vitro* using [³H]thymidine before and after indirubin treatment revealed a decrease of labelling index, the decrease being most significant in myelocytes and polychromatophilic erythroblasts [127].

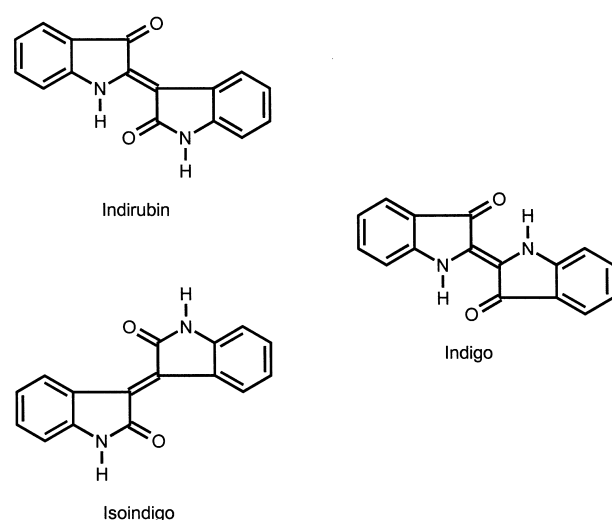


Fig. 9 Indirubin, indigo, and isoindigo.

Electron spin resonance spectrometry demonstrated that the C:C double bond obviously plays an important role in the anticancer activity of indirubin and indigo [128]. Recently, we have found that indirubin and its analogues are potent inhibitors of cyclin-dependent kinases (CDKs). The crystal structure of CDK₂ in a complex with indirubin derivatives showed that indirubin interacts with the kinase's ATP-binding site through van der Waals interactions and three hydrogen bonds [129], [130], [131]. In fact, a number of structures closely related to indirubin were reported to be cyclin-dependent kinase inhibitors [132]. Cell cycle molecular targets are being used for the development of new anticancer drugs [133].

Subacute toxicity testing of indirubin administered orally at 100–400 mg/kg per day to rats for 30 days showed no effects on leukocytes or on liver or renal function. Oral application of 80 mg/kg of indirubin per day for 1–3 months to rats caused anorexia and diarrhea, but no effects on bone marrow. Long-term administration of high doses of indirubin did not affect the hematopoietic stem cell production or DNA formation [134]. A subacute toxicity study of indirubin in dogs receiving three different doses showed no adverse effects at a low daily dosage (20 mg/kg). At a middle daily dosage of 100 mg/kg, mild diarrhea occurred within 10–30 days. Serum glutamic-pyruvic transaminase (GPT) levels were slightly elevated after 5 months in one of three dogs treated. At a high daily dosage (200 mg/kg), serious diarrhea and hematochezia occurred during 40–60 days and serum GPT levels were elevated after 3 months of treatment. Pathological changes were also observed in all tissue sections obtained after 6 months [135]. Bone marrow and blood indices, renal function, and electrocardiogram were not affected by any of the three dosages [136]. Therefore, indirubin was believed to be safe at clinical daily doses of 150–200 mg. The slight liver toxicity observed in dogs receiving 100 mg/kg corresponds to a daily dose of 2240 mg in humans [135]. In human peripheral lymphocytes [137] and in bone marrow cells of patients with chronic myelocytic leukemia [138], [139], indirubin did not increase sister chromatid exchange (SCE).

Several *N*-substituted indirubin derivatives [140], [141], indirubine acyl derivatives, indirubin oximes [142], and halogenated indirubins [143] were synthesized and tested as potentially antineoplastic agents. Some halogen-substituted indirubine derivatives were reported to exhibit higher antitumor activity against L7212 in mice and W256 sarcoma in rats than native indirubin. The most active compound was found to be 5-iodoindirubine which was active against W256 sarcoma, L7212, L7712, L759 leukemias, and Lewis lung carcinoma. It showed increases in life-span of mice bearing L7212 leukemia by 41–73%, and of rats bearing W256 sarcoma by 48–83%. The antitumor activity of 5-iodoindirubin against W256 sarcoma and L7212 leukemia was significantly higher than that of indirubin at the same dosage. *In vitro*, the IC₅₀ value of 5-iodoindirubin on [³H]thymidine incorporation into DNA of P388 cells was 6.4 µg/mL compared to 17 µg/mL of indirubin [143], [144].

Bisindole derivatives with 2,2', 3,3' or 3,2'-linkages were compared for their effects on nucleic acid and protein synthesis. They all formed complexes with calf thymus DNA as measured by UV and visible spectrometry and inhibited nucleic acid and protein

synthesis on cultured W256 sarcoma cells and *in vivo*. Isoindigo derivatives showed the highest potency in inhibiting DNA synthesis in cancer cells and in cell-free systems, whereas indigo derivatives had the lowest activity [145], [146]. *N*-Methylisoindigo (meisoindigo) was more potent than indirubin against W256 sarcoma in rats [143]. It was suggested that the improved absorption of meisoindigo is one of the major reasons for the enhanced antitumor activity as compared to indirubin. Studies on the mechanism of action of *N*-methylisoindigo indicated that it strongly inhibited DNA biosynthesis in L1210 cells and caused an arrest of S phase cells [147]. At a non-toxic concentration of 0.7 µg/mL *N*-methylisoindigo induced differentiation of ML-1 human myeloblastic leukemic cells as the most pronounced effect accompanied by the down-regulation of *c-Myb* gene expression, suggesting that the antitumor mechanisms of *N*-methylisoindigo involve these effects. *N*-Methylisoindigo was reported to be a second generation derivative, and was chosen for clinical treatment of chronic myeloid leukemia [148].

Conclusion

A series of low molecular compounds from Chinese herbal medicines, which proved to be in some cases highly effective especially in tumor therapy, has been listed here (part II will deal with high molecular compounds, to be published the next issue). These medicinal plants have been used since hundreds of years in China. In contrast to synthetic agents used in cancer chemotherapy, natural compounds have relatively low toxicities. Obviously, because of genetic factors that influence enzyme levels accounting for sometimes striking differences in metabolism and pharmacokinetics of drugs, results obtained in clinical studies carried out in China are not transferable to 100% to the European population. Since many of the clinical studies referred to in this paper have been carried out on Asians; the outcome of such studies in Caucasians or Africans might not be the same.

The mechanisms of action of these compounds are manifold, consisting in reactions with DNA bases, intercalation in DNA, inhibition of topoisomerases, inhibition of protein kinases, induction of apoptosis, etc. Some of the compounds presented here have interesting structural features, which have been or can be used as lead structures for the development of further antitumor agents.

References

- Owen TY, Wang SY, Chang SY, Lu FL, Yang CL, Hsu B. A new antitumor substance - lycobetaine (AT-1840). Ko Hsueh Tung Pao 1976; 21: 285–7
- He HM, Weng ZY. Structure-activity relationship study of the new anticancer drug lycobetaine (AT-1840). Acta Pharmaceutica Sinica 1989; 24: 302–4
- Xu B. Pharmacologic study of several anticancer agents of natural origin. Proceedings of Intern Summit on Drugs from Natural Products. October 9–12, 1995 Beijing, China: Vol 1
- Ghosal S, Singh SK, Kumar Y, Unnikrishnan S, Chattopadhyay S. Chemical constituents of Amaryllidaceae. XXVI. The role of ungeremine in the growth-inhibiting and cytotoxic effects of lycorine: evidence and speculation. Planta Medica 1988; 54: 114–6

- 5 Wang XW, Yu WJ, Shen ZM, Yang JL, Xu B. Cytotoxicity of hydroxycamptothecin and four other antineoplastic agents on KB cells. *Acta Pharmacologica Sinica* 1987; 8: 86–90
- 6 Wu YL, Wu YX, Yu CS, Zhang SY, Su ZC, Jiang SJ. The cytotoxic effect of AT-1840 and parvovirus H-1 on gastric cancer cells. *Shanghai Medical Journal* 1988; 11: 683–8
- 7 Liu J, Yang SL, Xu B. Characteristics of the interaction of lycobetaine with DNA. *Acta Pharmacologica Sinica* 1989; 10: 437–42
- 8 Liu J, Yang SL, Xu B. Effects of lycobetaine on chromatin structure and activity of murine hepatoma cells. *Science in China Series B* 1990; 33: 1459–65
- 9 Chen JZ, Chen KX, Jiang HL, Lin MW, Ji RY. Theoretical investigation on interaction binding of analogs of AT-1840 to double-stranded polynucleotide. *Progress in Natural Science* 1997; 7: 329–35
- 10 Wang P, Shi GB, Song GQ, Chen KX, Ji RY. Assignment of proton resonances and conformational characterization of oligodeoxyribonucleic acid d(CCGTACGG) in solution. *Acta Biochimica et Biophysica Sinica* 1996; 28: 703–5
- 11 Gan L, Liu XJ, Chen KL, Ji YY. Computer simulation on the interaction between antitumor agent lycobetaine and DNA. *Gaojishu Tongxun* 1992; 2: 30–3
- 12 Barthelems HU, Niederberger E, Roth T, Schulte K, Tang WC, Zankl H, Fiebig HH, Eisenbrand G, Marko D. Lycobetaine acts as a selective topoisomerase II β poison and inhibits the growth of human tumor cells. *British Journal of Cancer* 2001; 85: 1585–91
- 13 Zhang SY, Lu FL, Yang JL, Wang LJ, Xu B. Effect on animal tumors and toxicity of lycobetaine acetate. *Acta Pharmacologica Sinica* 1981; 2: 41–5
- 14 Hua DH, Saha S, Takemoto DJ. Anticancer activities of 2,5,8,9-substituted 6-oxo-1,2,3,4,5,6-hexahydrophenanthridines on multi-drug-resistant phenotype cells. *Anticancer Research* 1997; 17A: 2435–41
- 15 Niederberger E. Mechanismus-orientierte Untersuchungen zur antineoplastischen Wirkung von Naturstoffen und Naturstoff-Derivaten. Thesis, University of Kaiserslautern, Germany: 1998
- 16 Schulte K. Synthese von Phenanthridin- und Benzo[c]phenanthridin-Derivaten und Untersuchungen auf ihre biologische Wirkung. Thesis, University of Kaiserslautern, Germany: 2000
- 17 Kushida N, Atsumi S, Koyano T, Umezawa K. Induction of flat morphology in K-ras-transformed fibroblasts by lycorine, an alkaloid isolated from the tropical plant *Eucharis grandiflora*. *Drugs under Experimental and Clinical Research* 1997; 23: 151–5
- 18 Fang SD, Wang LK, Hecht SM. Inhibitors of DNA topoisomerase I isolated from the roots of *Zanthoxylum nitidum*. *Journal of Organic Chemistry* 1993; 58: 5025–7
- 19 The Pharmacopoeia of the People's Republic of China.; Vol I Ed 2000 Chemical Industrial Press, Beijing, PR China
- 20 Colombo ML, Bosio E. Pharmacological activities of *Chelidonium majus* L. (Papaveraceae). *Pharmacological Research* 1996; 33: 127–34
- 21 Stermitz FR, Gillespie JP, Amoros LG, Romero R, Stermitz TA, Larson KA, Earl S, Ogg JE. Synthesis and biological activity of some antitumor benzophenanthridinium salts. *Journal of Medicinal Chemistry* 1975; 18: 708–13
- 22 Fan YJ, Zhou J, Li M. Effect of nitidine chloride on the life cycle of Ehrlich ascites carcinoma cells in mice. *Acta Pharmacologica Sinica* 1981; 2: 46–9
- 23 Wang MH. Isolation of antitumor alkaloids from *Zanthoxylum nitidum* and structural study of its alkaloid C. *Chinese Pharmaceutical Bulletin* 1981; 16: 48
- 24 Zhu DY. Recent advances on the active components in Chinese medicines. Abstracts of Chinese Medicines (Hong Kong) 1987; 1: 251–286
- 25 Kubova N, Smekal E, Kleinwachter V, Cushman M. Binding properties of nitidine and its indenoisoquinoline analog with DNA. *Studia Biophysica* 1986; 114: 251–6
- 26 Gatto B, Sanders MM, Yu C, Wu HY, Makhey D, LaVoie EJ, Liu LF. Identification of topoisomerase I as the cytotoxic target of the protoberberine alkaloid coralyne. *Cancer Research* 1996; 56: 2795–800
- 27 Del Poeta M, Chen SF, Von Hoff D, Dykstra CC, Wani MC, Manikumar G, Heitman J, Wall ME, Perfect JR. Comparison of *in vitro* activities of camptothecin and nitidine derivatives against fungal and cancer cells. *Antimicrobial Agents and Chemotherapy* 1999; 43: 2862–78
- 28 Wang X, Henningfeld KA, Hecht SM. DNA topoisomerase I-mediated formation of structurally modified DNA duplexes. Effects of metal ions and topoisomerase I inhibitors. *Biochemistry* 1998; 37: 2691–700
- 29 Chen YZ, Tang GY, Xu BJ, Wu QJ, Lu CZ, Li JQ, Huang ZX. The formation and crystal structure of dihydronitidine and discussion of anticancer mechanism of nitidine cation. *Science in China Series B* 1992; 35: 1101–9
- 30 Larsen AK, Grondard L, Couprie J, Desoize B, Comoe L, Jardillier JC, Riou JF. The antileukemic alkaloid fagaronine is an inhibitor of DNA topoisomerases I and II. *Biochemical Pharmacology* 1993; 46: 1403–12
- 31 Fleury F, Sukhanova A, Ianoul A, Devy J, Kudelina I, Duval O, Alix AJ, Jardillier JC, Nabiev I. Molecular determinants of site-specific inhibition of human DNA topoisomerase I by fagaronine and ethoxidine. Relation to DNA binding. *Journal of Biological Chemistry* 2000; 275: 3501–9
- 32 Pezzuto JM, Antosiak SK, Messmer WM, Slaytor MB, Honig GR. Interaction of the antileukemic alkaloid, 2-hydroxy-3,8,9-trimethoxy-5-methylbenzo[c]phenanthridine (fagaronine), with nucleic acids. *Chemico-Biological Interactions* 1983; 43: 323–39
- 33 Herbert JM, Augereau JM, Gleye J, Maffrand JP. Chelerythrine is a potent and specific inhibitor of protein kinase C. *Biochemical and Biophysical Research Communications* 1990; 172: 993–9
- 34 Sampaio-Maia B, Soares-da-Silva P. Ca²⁺/calmodulin mediated pathways regulate the uptake of L-DOPA in mouse neuroblastoma neuro 2A cells. *Life Sciences* 2000; 67: 3209–20
- 35 Fernandez Y, Ramakrishnan R, Rathinavelu A. GI-101A breast tumor cell growth. *Life Sciences* 2000; 67: 567–75
- 36 Ramakrishnan R, Zell JA, Malave A, Rathinavelu A. Expression of vascular endothelial growth factor mRNA in GI-101A and HL-60 cell lines. *Biochemical and Biophysical Research Communications* 2000; 270: 709–13
- 37 Mueller H, Liu R, David F, Eppenberger U. Selective modulation of protein kinase A and protein kinase C activities in epidermal growth factor (EGF)-stimulated MCF-7 breast cancer cells. *Biological Chemistry* 1997; 378: 1023–9
- 38 Lamm ML, Long DD, Goodwin SM, Lee C. Transforming growth factor- β 1 inhibits membrane association of protein kinase C α in a human prostate cancer cell line, PC3. *Endocrinology* 1997; 138: 4657–64
- 39 Zhu GH, Wong BC, Eggo MC, Yuen ST, Lai KC, Lam SK. Pharmacological inhibition of protein kinase C activity could induce apoptosis in gastric cancer cells by differential regulation of apoptosis-related genes. *Digestive Diseases and Sciences* 1999; 44: 2020–6
- 40 Chmura SJ, Dolan ME, Cha A, Mauceri HJ, Kufe DW, Weichselbaum RR. *In vitro* and *in vivo* activity of protein kinase C inhibitor chelerythrine chloride induces tumor cell toxicity and growth delay *in vivo*. *Clinical Cancer Research* 2000; 6: 737–42
- 41 Wolff J, Knipling L. Antimicrotubule properties of benzophenanthridine alkaloids. *Biochemistry* 1993; 32: 13334–9
- 42 Wang BH, Lu ZX, Polya GM. Inhibition of eukaryote protein kinases by isoquinoline and oxazine alkaloids. *Planta Medica* 1997; 63: 494–8
- 43 Vavreckova C, Gawlik I, Muller K. Benzophenanthridine alkaloids of *Chelidonium majus*. II. Potent inhibitory action against the growth of human keratinocytes. *Planta Medica* 1996; 62: 491–4
- 44 Ahmad N, Gupta S, Husain MM, Heiskanen KM, Mukhtar H. Differential antiproliferative and apoptotic response of sanguinarine for cancer cells versus normal cells. *Clinical Cancer Research* 2000; 6: 1524–8
- 45 Dai JR, Chai H, Pezzuto JM, Kinghorn AD, Tsauri S, Padmawinata K. Studies on Indonesian Plants. V. Cytotoxic constituents of the roots of the Indonesian medicinal plant *Fibraurea chloroleuca*. *Phytotherapy Research* 1993; 7: 290–4
- 46 Chen KT, Hao DM, Liu ZX, Chen YC, You ZS. Effect of berberine alone or in combination with argon ion laser treatment on the 9L rat glioma cell line. *Chinese Medicinal Journal (Engl)* 1994; 107: 808–12
- 47 Iizuka N, Miyamoto K, Okita K, Tangoku A, Hayashi H, Yosino S, Abe T, Morioka T, Hazama S, Oka M. Inhibitory effect of Coptidis Rhizoma and berberine on the proliferation of human esophageal cancer cell lines. *Cancer Letters* 2000; 148: 19–25
- 48 Chi CW, Chang YF, Chao TW, Chiang SH, Peng FK, Lui WY, Liu TY. Flow-cytometric analysis of the effect of berberine on the expression of glucocorticoid receptors in human hepatoma HepG2 cells. *Life Sciences* 1994; 54: 2099–2107
- 49 Lin HL, Chang YF, Liu TY, Wu CW, Chi CW. Submicromolar paclitaxel induces apoptosis in human gastric cancer cells at early G1 phase. *Anticancer Research* 1998; 18: 3443–9
- 50 Yang IW, Chou CC, Yung BY. Dose-dependent effects of berberine on cell cycle pause and apoptosis in Balb/c 3T3 cells. *Naunyn-Schmiedeberg's Archives of Pharmacology* 1996; 354: 102–8

- 51 Chang KS, Gao C, Wang LC. Berberine-induced morphologic differentiation and down-regulation of c-Ki-ras2 protooncogene expression in human teratocarcinoma cells. *Cancer Letters* 1990; 55: 103–8
- 52 Miura N, Yamamoto M, Ueki T, Kitani T, Fukuda K, Komatsu Y. Inhibition of thymocyte apoptosis by berberine. *Biochemical Pharmacology* 1997; 53: 1315–22
- 53 Kuo CL, Chou CC, Yung BY. Berberine complexes with DNA in the berberine-induced apoptosis in human leukemic HL-60 cells. *Cancer Letters* 1995; 93: 193–200
- 54 Ren J, Chaires JB. Sequence and structural selectivity of nucleic acid binding ligands. *Biochemistry* 1999; 38: 16067–75
- 55 Lee SJ, Kim JB, Lee SW, Kim JH. Enhanced cytotoxicity of berberine and some anticancer nucleotides against tumor cell-lines. *Archives of Pharmacal Research* 1995; 18: 138–9
- 56 Zhang RX, Dougherty DV, Rosenblum ML. Laboratory studies of berberine used alone and in combination with 1,3-bis(2-chloroethyl)-1-nitrosourea to treat malignant brain tumors. *Chinese Medicinal Journal (English Edition)* 1990; 103: 658–65
- 57 Lin HL, Liu TY, Wu CW, Chi CW. Berberine modulates expression of mdrl gene product and the responses of digestive track cancer cells to Paclitaxel. *British Journal of Cancer* 1999; 81: 416–22
- 58 Lin HL, Liu TY, Lui WY, Chi CW. Up-regulation of multidrug resistance transporter expression by berberine in human and murine hepatoma cells. *Cancer* 1999; 85: 1937–42
- 59 Dai JR, Chai H, Pezzuto JM, Kinghorn AD, Tsauri S, Padmawinata K. Studies on Indonesian Plants. V. Cytotoxic constituents of the roots of the Indonesian medicinal plant *Fibraurea chloroleuca*. *Phytotherapy Research* 1993; 7: 290–4
- 60 Makhey D, Gatto B, Yu C, Liu A, Liu LF, LaVoie EJ. Protoberberine alkaloids and related compounds as dual inhibitors of mammalian topoisomerase I and II. *Medicinal Chemistry Research* 1995; 5: 1–12
- 61 Kobayashi Y, Yamashita Y, Fuji N, Takaboshi K, Kawakami T, Kawamura M, Mizukami T, Nakano H. Inhibitors of DNA topoisomerase I and II isolated from the *Coptis rhizomes*. *Planta Medica* 1995; 61: 414–8
- 62 Kim SA, Kwon Y, Kim JH, Muller MT, Chung IK. Induction of topoisomerase II-mediated DNA cleavage by a protoberberine alkaloid, berberrubine. *Biochemistry* 1998; 37: 16316–24
- 63 Xia GC, Zang JY, Lu XM, Xiao PG. Resource utilization and herbal study of “malinzi” (*Iris lactea* Pall. var. *chinensis*). *Acta Pharmaceutica Sinica* 1985; 20: 316–9
- 64 Li DH, Hao XG, Zhang SK, Wang SX, Liu RY, Ma KS, Yu SP, Jiang H, Guan JF. Antitumor effect and toxicity of irisquinone. *Acta Pharmacologica Sinica* 1981; 2: 131–4
- 65 Wang SX, Liu MF, Yuan L, Zhang WL, Li DH. Effects of irisquinone on cyclic nucleotides in plasma, cancer and other tissues of mice bearing U₁₄ tumor. *Chinese Journal of Clinical Oncology* 1986; 13: 241–3
- 66 Li WM, Wang SH, Kuang P, Qu BX, Song HL, Zhu LX. Radiosensitizing effect of an active principle Iq7611 from the seed of *Iris lactea* var. *chinensis*. *Tumor* 1987; 7: 97–9
- 67 Chitra M, Sukumar E, Suja V, Devi CS. Antitumor, anti-inflammatory and analgesic property of embelin, a plant product. *Chemotherapy* 1994; 40: 109–113
- 68 Chitra M, Sukumar E, Devi CS. [³H]-Thymidine uptake and lipid peroxidation by tumor cells on embelin treatment: an *in vitro* study. *Oncology* 1995; 52: 66–8
- 69 Yuan K, Hu R, Ji C, Yin M. New method for preparing oridonin by column chromatography. *China Journal of Chinese Materia Medica* 1997; 22: 478–80, 511
- 70 Wang RL, Gao BL, Xiong ML, Mei QD, Fan KS, Zuo ZK, Lang TL, Gao GQ, Ji ZC, Wei DC. Potentiation by *Rabdosia rubescens* on chemotherapy of advanced esophageal carcinoma. *Chinese Journal of Oncology* 1986; 8: 297–9
- 71 Wang RL. A report of 40 cases of esophageal carcinoma surviving for more than 5 years after treatment with drugs. *Chinese Journal of Oncology* 1993; 15: 300–2
- 72 Porterfield H. UsToo PC-SPES surveys: review of studies and update of previous survey results. *Molecular Urology* 2000; 4: 289–91
- 73 de la Taille A, Hayek OR, Burchardt M, Burchardt T, Katz AE. Role of herbal compounds (PC-SPES) in hormone-refractory prostate cancer: two case reports. *Journal of Alternative and Complementary Medicine* 2000; 6: 449–51
- 74 Lou FC, Ding LS, Ma QY, Du FL. Natural antineoplastic compounds and their structure-activity relationships. *Journal of Nanjing College of Pharmacy* 1986; 17: 152–9
- 75 Fujita T, Takeda Y, Sun HD, Minami Y, Marunaka T, Takeda S, Yamada Y, Togo T. Cytotoxic and antitumor activities of *Rabdosia* diterpenoids. *Planta Medica* 1988; 54: 414–7
- 76 Wang MY, Lin C, Zhang TM. Cytokinetic effects of oridonin on leukemia L1210 cells. *Acta Pharmacologica Sinica* 1985; 6: 195–8
- 77 Li XT, Lin C, Li PY. Comparison of *in vitro* assays for the cytotoxic effect of anticancer drugs. *Chinese Journal of Oncology* 1986; 8: 184–6
- 78 Yu JJ, Reed E. Preliminary study of the effect of selected Chinese natural drugs on human ovarian cancer cells. *Oncology Reporter* 1995; 2: 571–5
- 79 Wang MY, Lin C, Zhang TM. Autoradiographic study on the effects of oridonin on DNA, RNA and protein synthesis of leukemia L 1210 cells. *Acta Pharmacologica Sinica* 1987; 8: 164–5
- 80 Li Y, Zhang TM. Effect of oridonin on cell-free DNA synthesis *in vitro*. *Acta Pharmacologica Sinica* 1988; 9: 465–7
- 81 Gao ZG, Ye QX, Zhang TM. Synergistic effect of oridonin and cisplatin on cytotoxicity and DNA cross-link against mouse sarcoma S180 cells in culture. *Acta Pharmacologica Sinica* 1993; 14: 561–4
- 82 Zhang TM, Shou MG, Wang MY. Antitumor effects of different combinations of oridonin, bleomycin A₅ and nitrocapheane. *Acta Pharmacologica Sinica* 1986; 7: 457–60
- 83 Jiang B, Lu ZQ, Hou AJ, Zhao QS, Sun HD. *ent*-Kaurane diterpenoids from *Isodon lungshengensis*. *Journal of Natural Products* 1999; 62: 941–5
- 84 Sun HD, Lin ZW, Niu FD, Lin LZ, Chai HB, Pezzuto JM, Cordell GA. Cytotoxic *ent*-kaurene diterpenoids from three *Isodon* species. *Phytochemistry* 1995; 38: 437–42
- 85 Fujita E, Nagao Y, Node M, Kaneko K, Nakazawa S, Kuroda H. Antitumor activity of the *Isodon* diterpenoids; structural requirements for the activity. *Experientia* 1976; 32: 203–6
- 86 Zhai JK, Han WC, Ju XH. Crystal structure of nervosin and the electronic structure of its anticancer-active zone. *Acta Chimica Sinica* 1993; 51: 854–9
- 87 Fujita E, Nagao Y, Kaneko K, Nakazawa S, Kuroda H. The antitumor and antibacterial activity of the *Isodon* diterpenoids. *Chemical and Pharmaceutical Bulletin (Tokyo)* 1976; 24: 2118–27
- 88 Hall IH, Lee KH, Okano M, Sims D, Ibuka T, Liou YF, Imakura Y. Antitumor agents. XLII. Comparison of antileukemic activity of helenalin, brusatol and bruceantin and their esters on different strains of P388 lymphocytic leukemic cells. *Journal of Pharmaceutical Sciences* 1981; 70: 1147–50
- 89 Liao LL, Kupchan SM, Horwith SB. Mode of action of the antitumor compound bruceantin, an inhibitor of protein synthesis. *Molecular Pharmacology* 1976; 12: 167–76
- 90 Garnick MB, Blum RH, Canellos GP, Mayer RJ, Parker L, Skarin AT, Li FP, Henderson IC, Frei E. Phase I trial of bruceantin. *Cancer Treatment Reports* 1979; 63: 1929–32
- 91 Amato DA, Borden EC, Shiraki M, Enterline HAT, Rosenbaum C, Davis HL, Paul AR, Stevens CM, Lerner HJ. Evaluation of bleomycin, chlorozotocin, MGBG, and bruceantin in patients with advanced soft tissue sarcoma, bone sarcoma, or mesothelioma. *Investigational New Drugs* 1985; 3: 397–401
- 92 Wiseman CL, Yap HY, Bedikian AY, Bodey GP, Blumenschein GR. Phase II trial of bruceantin in metastatic breast carcinoma. *American Journal of Clinical Oncology* 1982; 5: 389–91
- 93 Cheng JH, Long H, Zu JL. Therapeutic analysis of treatment of recurrent metastatic gastrointestinal carcinomas in 35 patients with *Brucea javanica* emulsion “Yandanziyou”. *Chinese Traditional Patent Medicine* 1991; 13: 21–2
- 94 Wang ZQ. Combined therapy of brain metastasis in lung cancer. *Chinese Journal of Integrated Traditional and Western Medicine* 1992; 12: 609–10
- 95 Lu JB, Shu SY, Cai JQ. Experimental study on effect of *Brucea javanica* oil emulsion on rabbit intracranial pressure. *Chinese Journal of Integrated Traditional and Western Medicine* 1994; 14: 610–11
- 96 Wang JZ, Chen DY, Su YY. Analytical study on processing of *Scutellaria baicalensis* Georgi by HPLC. *China Journal of Chinese Materia Medica* 1994; 19: 340–1
- 97 Tomimori T, Miyaichi Y, Kizu H. On the flavonoid constituents from the roots of *Scutellaria baicalensis* Georgi. I. Yakugaku Zasshi 1982; 102: 388–91
- 98 Motoo Y, Sawabu N. Antitumor effects of saikosaponins, baicalin and baicalein on human hepatoma cell lines. *Cancer Letters* 1994; 86: 91–5

- 99 Ikemoto S, Sugimura K, Yoshida N, Yasumoto R, Wada S, Yamamoto K, Kishimoto T. Antitumor effects of scutellariae radix and its components baicalein, baicalin, and wogonin on bladder cancer cell lines. *Urology* 2000; 55: 951–5
- 100 Niederberger E, Meiers S, Genzlinger A, Zankl H, Tang WC, Eisenbrand G, Marko D. Flavones and inhibition of tumor cell growth: new aspects on the mechanism of action. In: Eisenbrand G, Dayan AD, Elias PS, Grunow W, Schlatter J, editors. *Carcinogenic and anticarcinogenic factors in food* Wiley-VCH, Weinheim: 1998: 512–3
- 101 Matsuzaki Y, Kurokawa N, Terai S, Matsumura Y, Kobayashi N, Okita K. Cell death induced by baicalein in human hepatocellular carcinoma cell lines. *Japanese Journal of Cancer Research* 1996; 87: 170–7
- 102 Austin CA, Patel S, Ono K, Nakane H, Fisher LM. Site-specific DNA cleavage by mammalian DNA topoisomerase II induced by novel flavone and catechin derivatives. *Biochemical Journal* 1992; 282: 883–9
- 103 Chan FL, Choi HL, Chen ZY, Chan PS, Huang Y. Induction of apoptosis in prostate cancer cell lines by a flavonoid, baicalin. *Cancer Letters* 2000; 160: 219–28
- 104 Huang HC, Hsieh LM, Chen HW, Lin YS, Chen JS. Effects of baicalein and esculetin on transduction signals and growth factors expression in T-lymphoid leukemia cells. *European Journal of Pharmacology* 1994; 268: 73–8
- 105 So FV, Guthrie N, Chambers AF, Moussa M, Carroll KK. Inhibition of human breast cancer cell proliferation and delay of mammary tumorigenesis by flavonoids and *Citrus* juices. *Nutrition and Cancer* 1996; 26: 167–81
- 106 So FV, Guthrie N, Chambers AF, Carroll KK. Inhibition of proliferation of estrogen receptor-positive MCF-7 human breast cancer cells by flavonoids in the presence and absence of excess estrogen. *Cancer Letters* 1997; 112: 127–33
- 107 Chang WS, Lee YJ, Lu FJ, Chiang HC. Inhibitory effects of flavonoids on xanthine oxidase. *Anticancer Research* 1993; 13: 2165–70
- 108 Umezawa K. Inhibition of experimental metastasis by enzyme inhibitors from microorganisms and plants. *Advances in Enzyme Regulation* 1996; 36: 267–81
- 109 Kawai S, Tomono Y, Katase E, Ogawa K, Yano M. Antiproliferative activity of flavonoids on several cancer cell lines. *Bioscience, Biotechnology and Biochemistry* 1999; 63: 896–9
- 110 Kawai S, Tomono Y, Katase E, Ogawa K, Yano M. Effect of *Citrus* flavonoids on HL-60 cell differentiation. *Anticancer Research* 1999; 19(A): 1261–9
- 111 Kawai S, Tomono Y, Katase E, Ogawa K, Yano M. HL-60 differentiating activity and flavonoid content of the readily extractable fraction prepared from *Citrus* juices. *Journal of Agricultural and Food Chemistry* 1999; 47: 128–35
- 112 Bracke M, Vyncke B, Opdenakker G, Foidart JM, De Pestel G, Mareel M. Effect of catechins and citrus flavonoids on invasion *in vitro*. *Clinical and Experimental Metastasis* 1991; 9: 13–25
- 113 Takanaga H, Ohnishi A, Yamada S, Matsuo H, Morimoto S, Shoyama Y, Ohtani H, Sawada Y. Polymethoxylated flavones in orange juice are inhibitors of P-glycoprotein but not cytochrome P450 3A4. *Journal of Pharmacology and Experimental Therapeutics* 2000; 293: 230–6
- 114 Ikegawa T, Ushigome F, Koyabu N, Morimoto S, Shoyama Y, Naito M, Tsuruo T, Ohtani H, Sawada Y. Inhibition of P-glycoprotein by orange juice components, polymethoxyflavones in adriamycin-resistant human myelogenous leukemia (K562/ADM) cells. *Cancer Letters* 2000; 160: 21–8
- 115 Institute of Haematology, Chinese Academy of Medical Sciences. Clinical studies of Dang Gui Lu Hui Wan in the treatment of CML. *Chinese Journal of Internal Medicine* 1979; 15: 86–8
- 116 Wu LM, Yang YP, Zhu ZH. Studies on the active principles of *Indigofera tinctoria* in the treatment of CML. *Comm Chinese Herb Med* 1979; 9: 6–8
- 117 Ji XJ, Zhang FR, Lei JL, Xu YT. Studies on the antineoplastic effect and toxicity of synthetic indirubin. *Acta Pharmaceutica Sinica* 1981; 16: 146–8
- 118 Indirubin cooperative group. Clinical study of indirubin in the treatment of 314 patients with chronic granulocytic leukemia. *Chinese Journal of Hematology* 1980; 1: 132
- 119 Gan WJ, Yang TY, Wen SD, Liu YY, Tan Z, Deng CA, Wu JX, Liu MP. Studies on the mechanism of indirubin action in treatment of chronic myelocytic leukemia (CML). II. 5'-Nucleotidase in the peripheral white blood cells of CML. *Chinese Journal of Hematology* 1985; 6: 611–3
- 120 Qian LS, Xue YP, Zhang XM, Yang TY, Yan WW, Wang ZC. Observation on the long period effect of indirubin in the treatment of 57 cases of chronic myelogenous leukemia. *Chinese Journal of Hematology* 1991; 12: 125–7
- 121 Zhang L, Wu GY, Qiu CC. Effect of indirubin on DNA synthesis *in vitro*. *Acta Academiae Medicinae Sinicae* 1985; 7: 112–6
- 122 Gan WJ, Yang TY, Wang ZC, Qian LS, Ma J, Ge YQ, Cheng BJ, Li ZM, Bo HQ. Studies on the antitumor mechanism of indirubin by treatment of chronic myelocytic leukemia (CML). *Chinese Biochemical Journal* 1987; 3: 225–30
- 123 Du DJ, Ceng QT. Effect of indirubin on the incorporation of isotope labeled precursors into nucleic acid and protein of tumor tissues. *Chinese Traditional and Herbal Drugs* 1981; 12: 406–9
- 124 Du DG, Ceng QT, Wen ZS, Wan XP. Study on the incorporation of ³H-TdR into DNA of liver and spleen of L7212, a lymphatic leukemia in mice. *Chinese Journal of Hematology* 1982; 3: 277–80
- 125 Wu GY, Liu JZ, Fang FD, Zuo J. Studies on the mechanism of indirubin action in the treatment of chronic granulocytic leukemia. V. Binding between indirubin and DNA and identification of the type of binding. *Scientia Sinica* 1982; 25B: 1071–9
- 126 Lee K, Shih CY, Yang TY, Chen LS, Chao WM, Sun CS, Wang TC, Pien SK, Sing KH. Ultratherapeutic effect of indirubin for human chronic granulocytic leukemia. *National Medical Journal of China* 1979; 59: 129–32
- 127 You YC, Mi JX, Wan JH, Yang TY, Wang ZC, Qian LS. Indirubin in the treatment of chronic myelocytic leukemia (CML) estimation of labeling index of bone marrow cells by ³H-TdR. *Chinese Journal of Oncology* 1987; 9: 418–20
- 128 Chen KY, Wang ZF, Li CY, Li WX. Study on indirubin and indigo by ESR spectrometry. *Chemical Journal of Chinese Universities* 1989; 10: 869–71
- 129 Hoessel R, Leclerc S, Endicott JA, Nobel ME, Lawrie A, Tunnah P, Leost M, Damiens E, Marie D, Marko D, Niederberger E, Tang W, Eisenbrand G, Meijer L. Indirubin, the active constituent of a Chinese antileukemia medicine, inhibits cyclin-dependent kinases. *Nature Cell Biology* 1999; 1: 60–7
- 130 Marko D, Schatzle S, Friedel A, Genzlinger A, Zankl H, Meijer L, Eisenbrand G. Inhibition of cyclin-dependent kinase 1 (CDK₁) by indirubin derivatives in human tumour cells. *British Journal of Cancer* 2001; 84: 283–9
- 131 Leclerc S, Garnier M, Hoessel R, Marko D, Bibb JA, Snyder GL, Greengard P, Biernat J, Wu YZ, Mandelkow EM, Eisenbrand G, Meijer L. Indirubins inhibit glycogen synthase kinase-3 β and CDK₅/p25, two protein kinases involved in abnormal tau phosphorylation in Alzheimer's disease. A property common to most cyclin-dependent kinase inhibitors? *Journal of Biological Chemistry* 2001; 276: 251–60
- 132 Sielecki TM, Boylan JF, Benfield PA, Trainor GL. Cyclin-dependent kinase inhibitors: useful targets in cell cycle regulation. *Journal of Medicinal Chemistry* 2000; 43: 1–18
- 133 Buolamwini JK. Cell cycle molecular targets in novel anticancer drug discovery. *Current Pharmaceutical Design* 2000; 6: 379–92
- 134 Wang JH, You YC, Mi JX, Ying HG. Effect of indirubin on hematopoietic cell production. *Acta Pharmacologica Sinica* 1981; 2: 241–4
- 135 Wen ZJ, Liu Y, Yi ZP, Yang YS. Effects of indirubin on the histology and histochemistry of canine and rat livers. *Bulletin of Chinese Materia Medica* 1988; 13: 306–7
- 136 Sichuan Institute of Traditional Chinese Medicine. Subacute toxicity of indirubin in dogs. *Chinese Traditional and Herbal Drugs* 1981; 12: 27–9
- 137 Cai YY, Xu CL, Li SH, Liu Y. Studies on sister chromatid exchanges induced by harringtonine, indirubin and pyquon before or after activation with microsome enzyme. *Acta Academiae Medicinae Sinicae* 1983; 5: 161–4
- 138 Feng BZ, Zhang YH, Qian LS, Chu YL. Effect of indirubin on SCE frequencies of BM cells in chronic myeloid leukemia. *Acta Academiae Medicinae Sinicae* 1984; 6: 308–10
- 139 Feng BZ. Sister chromatid exchange frequency of bone marrow cells and its response to indirubin in chronic myeloid leukemia. *Chinese Journal of Oncology* 1984; 6: 357–60
- 140 Wu KM, Zhang MY, Fang Z, Huang L. Synthesis of N₁-substituted derivatives of indirubin, an antileukemic compound. *Acta Pharmaceutica Sinica* 1984; 19: 513–8
- 141 Ji XJ, Zhang FR. Antineoplastic effect of indirubin derivatives and their structure-activity relationship. *Acta Pharmaceutica Sinica* 1985; 20: 137–9

- ¹⁴² Zeng QT, Du DJ, Xie DC, Wang XP, Rau CQ. Antitumor activities of indirubin derivatives. *Chinese Traditional and Herbal Drugs* 1982; 13: 24–30
- ¹⁴³ Gu YC, Li GL, Yang YP, Fu JP, Li CZ. Synthesis of some halogenated indirubin derivatives. *Acta Pharmaceutica Sinica* 1989; 24: 629–32
- ¹⁴⁴ Zeng QT, Du DJ, Si XF, Ran CQ, Wu XP. Antitumor activity of 5'-iodoindirubin. *West China Pharm Sci* 1991; 6: 63–8
- ¹⁴⁵ Wu GY, Liu JZ, Zhang L. Effect of bisindole compounds with three different kinds of linkage on the synthesis of nucleic acid and protein. *Progress in Biochemistry and Biophysics* 1985; 61: 48–51
- ¹⁴⁶ Wu KM, Zhang MY, Fang Z, Huang L. Potential antileukemic agents, synthesis of derivatives of indirubin, indigo, and isoindigotin. *Acta Pharmaceutica Sinica* 1985; 20: 821–6
- ¹⁴⁷ Ji XJ, Liu XM, Li K, Chen RH, Wang LG. Pharmacological studies of meisoindigo: absorption and mechanism of action. *Biomedical and Environmental Sciences* 1991; 4: 332–7
- ¹⁴⁸ Liu XM, Wang LG, Li HY, Ji XJ. Induction of differentiation and down-regulation of c-myc gene expression in ML-1 human myeloblastic leukemia cells by the clinically effective anti-leukemia agent meisoindigo. *Biochemical Pharmacology* 1996; 51: 1545–51