

Chronic oral supplementation with sepiapterin prevents endothelial dysfunction and oxidative stress in small mesenteric arteries from diabetic (db/db) mice

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1 We previously reported that acute incubation with tetrahydrobiopterin (BH₄) or sepiapterin, a cofactor for endothelial nitric oxide synthase and a stable precursor of BH₄, respectively, enhanced the acetylcholine (ACh)-induced relaxation of isolated small mesenteric arteries (SMA) from diabetic (db/db) mice. In this study, we investigated the effect of chronic oral supplementation of sepiapterin (10 mg kg⁻¹ day⁻¹) to db/db mice on endothelium function, biopterin levels and lipid peroxidation in SMA.

2 Oral dietary supplementation with sepiapterin had no effect on glucose, triglyceride, cholesterol levels and body weight. SMA from db/db mice showed enhanced vascular reactivity to phenylephrine, which was corrected with sepiapterin supplementation. Furthermore, ACh, but not sodium nitroprusside-induced relaxation, was improved with sepiapterin supplementation in db/db mice.

3 BH₄ levels and guanosine triphosphate cyclohydrolase I activity in SMA were similar in db/+ and db/db mice. Sepiapterin treatment had no effects on BH₄ or guanosine triphosphate cyclohydrolase I activity. However, the level of dihydrobiopterin + biopterin was higher in SMA from db/db mice, which was corrected following sepiapterin treatment.

4 Thiobarbituric acid reactive substance, malondialdehyde, a marker of lipid peroxidation, was higher in SMA from db/db mice, and was normalized by sepiapterin treatment.

5 These results indicate that sepiapterin improves endothelial dysfunction in SMA from db/db mice by reducing oxidative stress. Furthermore, these results suggest that decreased biosynthesis of BH₄ may not be the basis for endothelial dysfunction in SMA from db/db mice.

British Journal of Pharmacology (2003) **140**, 701–706. doi:10.1038/sj.bjp.0705476

Keywords: Endothelial dysfunction; type II diabetes; tetrahydrobiopterin; sepiapterin; oxidative stress; db/db mice

Abbreviations: ACh, acetylcholine; BB, biobreeding; BH₂, dihydrobiopterin; BH₃^{*}, trihydrobiopterin radical; BH₄, (6R)-5,6,7,8-tetrahydrobiopterin; DAHP, 2,4-diamino-6-hydroxypyrimidine; DHPR, dihydropteridine reductase; eNOS, endothelial nitric oxide synthase; GTP, guanosine triphosphate; NO, nitric oxide; NOS, nitric oxide synthase; PE, phenylephrine; PEG-SOD, polyethylene glycol-superoxide dismutase; ROS, reactive oxygen species; SMA, small mesenteric artery; SNP, sodium nitroprusside; SHR, spontaneous hypertensive rats

Introduction

(6R)-5,6,7,8-tetrahydro-L-biopterin (BH₄) is an essential cofactor required for the biosynthesis of nitric oxide (NO) by nitric oxide synthase (NOS). Although the exact mechanism whereby BH₄ regulates NOS activity is not known, several different hypotheses have been proposed viz., BH₄ exerts an allosteric action to stabilize the active dimeric state of NOS, it plays a redox active role in stimulating NOS, it increases the binding of L-arginine to NOS and scavenges reactive free radicals or it plays a role in the electron transfer to the ferrous-dioxy intermediate by forming a pterin radical, thus enabling the formation of a heme-based oxidant that rapidly hydroxylates L-arginine (Klatt *et al.*, 1994; Kojima *et al.*, 1995;

Mayer & Werner, 1995; Hurshman *et al.*, 1999; Wei *et al.*, 2001).

In situations where there are suboptimal concentrations of BH₄, endothelial NOS (eNOS) will generate superoxide in addition to NO. For instance, biochemical studies with purified constitutive NOS have demonstrated that in the presence of suboptimal concentrations of BH₄, there is an uncoupling of NOS and subsequent formation of superoxide and hydrogen peroxide (Pou *et al.*, 1992; Wever *et al.*, 1997; Vasquez-Vivar & Kalyanaraman, 2000). Blockade of BH₄ synthesis with 2,4-diamino-6-hydroxypyrimidine (DAHP), a selective guanosine triphosphate (GTP) cyclohydrolase I inhibitor, results in impaired endothelial function in isolated aorta, coronary, mesenteric and cerebral arteries (Cosentino & Katusic, 1995; Kinoshita *et al.*, 1997; Tiefenbacher *et al.*, 2000; Pannirselvam *et al.*, 2002). Compared to control values,

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levels of BH₄ have been shown to be decreased in aorta from insulin-resistant rats, in coronary endothelial cells from biobreeding (BB) diabetic rats and in aorta from spontaneously hypertensive (SHR) rats (Shinozaki *et al.*, 1999; Meininger *et al.*, 2000; Hong *et al.*, 2001). Thus, supplementation with BH₄ would be expected to restore impaired endothelium function and improve cardiovascular function in diabetes and other cardiovascular pathological conditions.

BH₄ has been shown to improve endothelium-dependent vasorelaxation in isolated aorta from streptozotocin-induced diabetic rat, fructose-fed insulin-resistant rats and coronary arteries following reperfusion injury (Tiefenbacher *et al.*, 1996; Pieper, 1997; Shinozaki *et al.*, 1999). BH₄ has also been shown to improve relaxation in blood vessels from patients with coronary artery disease, type II diabetes, smokers and atherosclerosis (Heitzer *et al.*, 2000; Maier *et al.*, 2000; Tiefenbacher *et al.*, 2000; Heitzer *et al.*, 2001). Chronic treatment with BH₄ has been shown to improve endothelial dysfunction in aorta from fructose-fed insulin-resistant rats and decrease the elevated blood pressure in SHR rats (Shinozaki *et al.*, 2000; Hong *et al.*, 2001). We have reported endothelial dysfunction in isolated small mesenteric arteries (SMA) from diabetic (db/db) mice and also shown that acute incubation with BH₄, sepiapterin or with a combination of polyethylene glycol superoxide dismutase (PEG-SOD) and catalase significantly improved acetylcholine (ACh)-induced relaxation (Pannirselvam *et al.*, 2002). The objective of the present study was to investigate the effects of the chronic oral supplementation of sepiapterin on endothelial function, biopterin contents and lipid peroxidation in SMA from diabetic (db/db) mice.

Methods

Animals

Male C57BL/KsJ diabetic mice (db/db) and nondiabetic controls (db/+), 6-week-old, were purchased from The Jackson Laboratory (Bar Harbour, ME, U.S.A.). At 8 weeks of age, animals were divided into four groups – group 1: db/+ mice receiving powder chow for 8 weeks, group 2: db/+ mice receiving sepiapterin (10 mg kg⁻¹ day⁻¹) in powder chow for 8 weeks, group 3: db/db mice receiving powder chow for 8 weeks and group 4: db/db mice receiving sepiapterin (10 mg kg⁻¹ day⁻¹) in powder chow for 8 weeks. In accordance with a protocol approved by the University of Calgary animal care committee, mice were killed by cervical dislocation. Heparinized blood samples were collected for blood biochemistry. The mesenteric arcade was removed and first-order branches of the mesenteric artery (measuring approximately 150–200 µm in diameter) were dissected in cold Krebs' solution of the following composition (in mM): NaCl 120, NaHCO₃ 25, KCl 4.8, NaH₂PO₄ 1.2, MgSO₄ 1.2, dextrose 11.0, CaCl₂ 1.8, aerated with 95% O₂ and 5% CO₂. The rest of the arterial bed was immediately stored at -70°C for biochemical measurements.

Wire myograph studies

Isometric tension studies using wire myograph were performed as described previously (Pannirselvam *et al.*, 2002). SMA were cut into 2 mm ring and mounted on a Mulvany–Halpern

myograph. The passive tension-internal circumference was determined by stretching to achieve an internal circumference equivalent to 90% of that of the blood vessel under a transmural pressure of 100 mmHg. All the experiments were performed at 37°C. Concentration–response curves to phenylephrine (PE) were constructed and normalized relative to the contraction induced by 120 mM KCl. Subsequently, endothelium-dependent and independent relaxations to ACh and sodium nitroprusside (SNP), respectively, were studied in tissues precontracted with a submaximal concentration of PE.

Determination of GTP cyclohydrolase I activity and biopterin content in SMA

GTP cyclohydrolase I activity in SMA was measured as previously described (Vann *et al.*, 2000). Homogenized SMA was incubated with Tris-HCl buffer (pH 7.4), 10 mM dithiothreitol, 10 mg ml⁻¹ bovine serum albumin, 10 mM GTP for 2 h at 37°C. The reactions were terminated with 1 M HCl and oxidized with iodine reagent (1% I₂/2% KI, 1:1, wv⁻¹). Neopterin triphosphate was dephosphorylated by incubating with 10 U of alkaline phosphatase for 30 min at 37°C. The reactions were terminated by adding 1 M phosphoric acid. Samples were centrifuged and the amount of neopterin formed was measured by reverse-phase high-performance liquid chromatography with fluorometric detection as described earlier. Neopterin levels were normalized to the amount (mg) of protein.

The biopterin contents of the SMA were measured by reverse-phase high-performance liquid chromatography as described previously (Howells *et al.*, 1986). SMA were homogenized in 25 mM triethanolamine-HCl (pH 7.4) containing 1 mg ml⁻¹ dithioerythritol and diethylenetriaminepentaacetic acid and separated using ultrafree-MC tubes (Millipore Co., U.S.A.) by centrifuging at 5000 r.p.m. for 15 min at 4°C. The filtrates were immediately used for BH₄ and biopterin assays. BH₄ was measured directly using an ESA Coulochem electrochemical detector, dihydrobiopterin (BH₂) was measured by fluorescence (358/447 nm) after oxidation on the conditioning cell electrode and biopterin was measured by its natural fluorescence (358/447 nm). BH₄ and biopterin measured were normalized relative to the amount (mg) of protein.

Determination of lipid peroxidation in plasma and SMA

Thiobarbituric acid reactive substance, malondialdehyde, a marker for lipid peroxidation, was measured in plasma and SMA, using a commercial kit (OXI-TEK, ZeptoMetrix Co. U.S.A.) according to the manufacturer's instructions. The levels of malondialdehyde were normalized to milligrams of protein.

Protein assay

Protein content was determined by the method of bicinchoninic acid using a commercial kit from Pierce Co., U.S.A.

Drugs

L-Sepsiapterin, a precursor of BH₄, was obtained from Schireks Laboratories (Switzerland). All other chemicals were purchased from Sigma Chemicals (St Louis, MO, U.S.A.).

Biochemical measurement of plasma glucose, triglyceride and cholesterol were performed using commercial kit from Sigma Diagnostics (St Louis, MO, U.S.A.).

Statistics

All values are expressed as mean \pm s.e.m.. The relaxation is expressed as mean percentage (\pm s.e.m.). In all experiments, n equals the number of animals used in the protocol. The concentration–response curves among the four groups were compared using a repeated measures ANOVA test. Statistical significance of difference in biochemical parameters among four groups was performed using one-way ANOVA. Multiple comparisons of the paired groups were performed using Student–Newman–Keuls method. A P -value of less than 0.05 was considered statistically significant.

Results

Metabolic characteristics in db/db mice

The db/db mice showed higher body weight compared to db/+ mice. Oral treatment with sepiapterin had no effect on body weight in either db/db or db/+ mice (Table 1). The db/db mice showed elevated levels of plasma glucose, cholesterol and triglyceride compared to db/+ mice. However, sepiapterin treatment had no effect on the biochemical parameters in either db/+ or db/db mice (Table 1).

Effect of sepiapterin treatment on vascular reactivity

The PE-induced maximum contraction was significantly higher in SMA from db/db mice compared to db/+ mice. Treatment with sepiapterin significantly reduced the maximum contraction of SMA to PE in db/db mice, but not in SMA from db/+ mice (Figure 1). PE-induced maximum contractions were 124 ± 4 , 125 ± 9 , 150 ± 4 and 134 ± 6 (% 120 mM KCl) for db/+, db/+ treated with sepiapterin, db/db and db/db treated with sepiapterin, respectively.

Ach induced a concentration-dependent relaxation of SMA from db/+ and db/db mice; however, maximum relaxations were significantly reduced in SMA from db/db mice compared to db/+ mice. Sepiapterin treatment significantly improved the relaxation to Ach in SMA from db/db mice but not in SMA from db/+ mice (Figure 2). Sensitivity (expressed as pD_2) and maximum relaxation (expressed as %) to Ach were 7.5 ± 0.1 and 95 ± 2 , 7.3 ± 0.1 and 96 ± 1 , 6.4 ± 0.1 and 66 ± 6 and 7.2 ± 0.2 and 77 ± 2 for db/+, db/+ treated with sepiapterin, db/db and db/db treated with sepiapterin, respectively. SNP induced a concentration-dependent relaxation of SMA from db/+ and db/db mice, which were

comparable and not altered in sepiapterin-treated groups (Figure 3). Sensitivity (expressed as pD_2) and maximum relaxation (expressed as %) to SNP were 8.1 ± 0.2 and 98 ± 1 , 8.0 ± 0.1 and 95 ± 1 , 7.6 ± 0.1 and 96 ± 0.5 and 8.0 ± 0.1 and 95 ± 1 for db/+, db/+ treated with sepiapterin, db/db and db/db treated with sepiapterin, respectively.

Effect of sepiapterin treatment on GTP cyclohydrolase I activity and biopterin contents

The level of neopterin, as a measure of GTP cyclohydrolase I activity, was not significantly different in SMA from db/+ and db/db mice. Sepiapterin treatment had no effect on GTP cyclohydrolase I activity in SMA from either db/+ or db/db mice (Figure 4).

The level of BH_4 in SMA was not significantly different between db/+ and db/db mice (Figure 5). Sepiapterin treatment had no effect on BH_4 levels in SMA from either db/+ or db/db (Figure 5). The BH_2 + biopterin content in mesenteric arteries was significantly higher in db/db mice compared to db/+ mice (Figure 6). Sepiapterin treatment reduced the content of BH_2 + biopterin in SMA from db/db mice to control values (Figure 6). When the biopterin contents were expressed as a ratio of BH_4 to BH_2 + biopterin, the value was significantly lower in SMA from db/db mice compared to db/+ mice. Sepiapterin treatment had no effect on the ratio of BH_4 to BH_2 /biopterin in either db/db or db/+ mice (Figure 7).

Effect of sepiapterin treatment on lipid peroxidation levels in plasma and SMA

Malondialdehyde, a marker for lipid peroxidation, was significantly elevated in plasma and SMA from db/db mice compared to db/+ mice (Figure 8a and b). Sepiapterin treatment had no effect on malondialdehyde levels in plasma from db/db and db/+ mice (Figure 8a). Sepiapterin treatment significantly reduced the elevated levels of malondialdehyde in SMA from db/db mice, but not in db/+ mice (Figure 8b).

Discussion

Chronic oral supplementation of sepiapterin prevented the enhanced vascular contractility to PE and improved relaxation to Ach in SMA from db/db mice. Sepiapterin treatment lowered the levels of BH_2 + biopterin and malondialdehyde in SMA from db/db mice, but did not affect the levels of either BH_4 or GTP cyclohydrolase activity. These results suggest that increased oxidative stress and oxidized products of BH_4 (BH_2 + biopterin) contribute to endothelial dysfunction in SMA from db/db mice. Sepiapterin treatment restores

Table 1 Effect of sepiapterin on biochemical characteristics in db/db mice

Parameters	db/+	db/+ + sepiapterin	db/db	db/db + sepiapterin
Body weight (g)	30 ± 0.3	30 ± 0.7	$50 \pm 1^{*\dagger}$	$49 \pm 0.4^{*\dagger}$
Glucose (mg dl ⁻¹)	168 ± 19	179 ± 24	$349 \pm 21^{*\dagger}$	$345 \pm 27^{*\dagger}$
Cholesterol (mg dl ⁻¹)	70 ± 11	84 ± 5	$143 \pm 7^{*\dagger}$	$142 \pm 8^{*\dagger}$
Triglycerides (mg dl ⁻¹)	69 ± 16	72 ± 3	$119 \pm 13^{*\dagger}$	$115 \pm 8^{*\dagger}$

db/+ + sepiapterin, db/db + sepiapterin are the groups that received sepiapterin 10 mg kg⁻¹ day⁻¹ for 8 weeks in diet. * P < 0.01 compared to db/+ group and $^\dagger P$ < 0.01 compared to db/+ + sepiapterin group. The values represent mean \pm s.e.m.

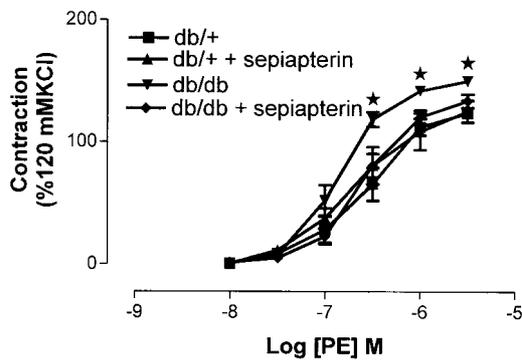


Figure 1 Effect of oral supplementation of sespiapterin on vascular reactivity to PE of SMA from db/+ and db/db mice. The PE-induced contraction was normalized to 120 mM KCl. The data are expressed as mean \pm s.e.m. based on seven to eight experiments. * $P < 0.05$ compared to db/+ and db/+ treated with sespiapterin group.

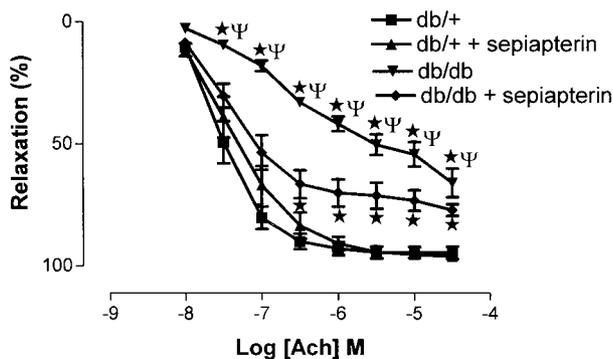


Figure 2 Effect of oral supplementation of sespiapterin on endothelium-dependent relaxation to Ach of SMA from db/+ and db/db mice. Ach-induced relaxations are expressed as percentage. The data are expressed as mean \pm s.e.m. based on seven to eight experiments. * $P < 0.05$ compared to db/+ and db/+ treated with sespiapterin group, $\psi P < 0.05$ to db/db treated with sespiapterin group.

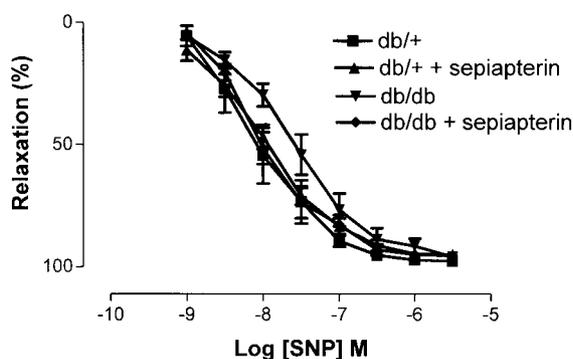


Figure 3 Effect of oral supplementation of sespiapterin on endothelium-independent relaxation to SNP of SMA from db/+ and db/db mice. SNP-induced relaxations are expressed as percentage. The data are expressed as mean \pm s.e.m. based on seven to eight experiments.

endothelial function in SMA from db/db mice by decreasing the levels of lipid peroxidation and BH₂ + biopterin contents.

We previously reported improved relaxation to Ach following acute treatment with BH₄ or sespiapterin in SMA from db/db mice (Pannirselvam *et al.*, 2002). In the current study,

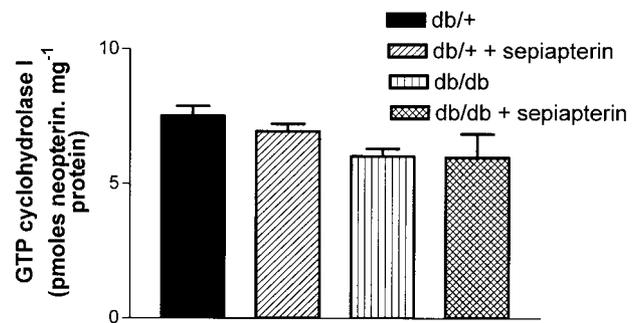


Figure 4 Effect of oral supplementation of sespiapterin on GTP cyclohydrolase activity in SMA from db/+ and db/db mice. The values are mean \pm s.e.m. neopterin formed in pmol mg⁻¹ of protein based on seven to eight experiments.

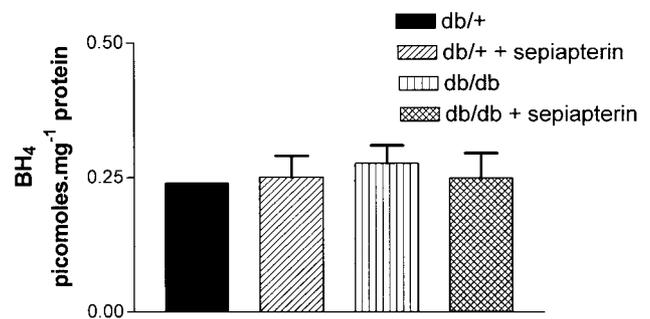


Figure 5 Effect of oral supplementation with sespiapterin on BH₄ levels in SMA from db/+ and db/db mice. The values are mean \pm s.e.m. in pmol mg⁻¹ of protein based on five to six experiments.

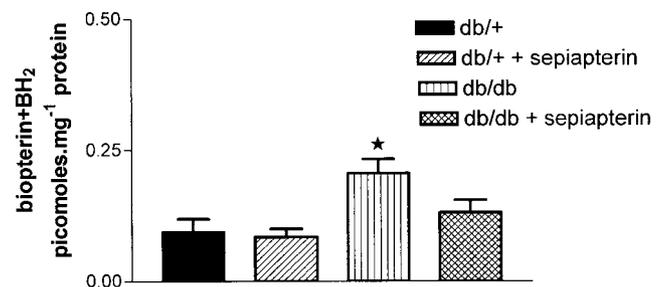


Figure 6 Effect of oral supplementation with sespiapterin on BH₂ + biopterin levels in SMA from db/+ and db/db mice. The values are mean \pm s.e.m. pmol mg⁻¹ of protein based on five to six experiments. * $P < 0.05$ compared to db/+ and db/+ treated with sespiapterin group.

we showed that chronic oral supplementation with sespiapterin improved endothelium-dependent relaxation of SMA to Ach and reduced vascular reactivity to PE. Treatment with sespiapterin, however, did not alter SNP-induced endothelium-independent relaxation. These findings are in agreement with previous reports in fructose-fed insulin-resistant rats and spontaneously hypertensive rats (Shinozaki *et al.*, 2000; Hong *et al.*, 2001). These data also support the hypothesis that a decreased bioavailability of BH₄, either due to a decreased synthesis or increased oxidation of BH₄, contributes to endothelial dysfunction in SMA from db/db mice.

BH₄ is synthesized by two pathways. The *de novo* synthesis pathway that uses GTP as a precursor where GTP cyclohydrolase I is the rate-limiting enzyme, and by a salvage pathway

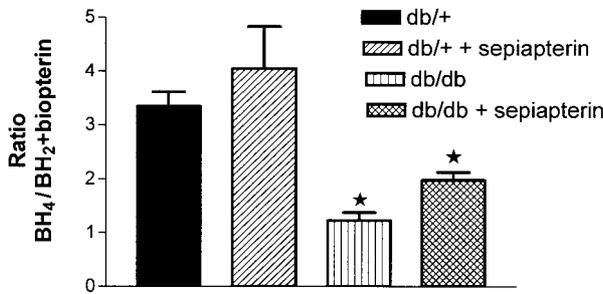


Figure 7 Effect of oral supplementation with sepiapterin on the ratio of BH₄ to BH₂ + biopterin in SMA from db/+ and db/db mice. The values are mean \pm s.e.m. pmoles mg⁻¹ of protein based on five to six experiments. * $P < 0.05$ compared to db/+ and db/+ + treated with sepiapterin group.

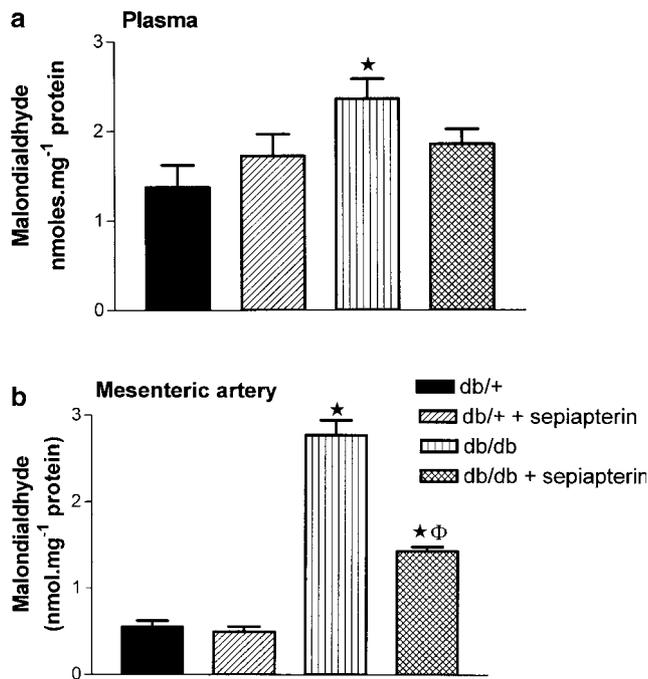


Figure 8 Effect of oral supplementation of sepiapterin on lipid peroxidation in plasma (a) and SMA (b) from db/+ and db/db mice. The values are malondialdehyde in nmol mg⁻¹ protein and expressed as mean \pm s.e.m. based on five to six experiments. * $P < 0.05$ compared to db/+ and db/+ + treated with sepiapterin, and § $P < 0.05$ compared to db/db mice.

where BH₄ is regenerated from quinonoid form of the BH₂ using dihydropteridine reductase (DHPR, Mayer & Werner, 1995). GTP cyclohydrolase I and DHPR activity were reduced in aorta from the fructose-fed insulin-resistant rat model and exhibited endothelial dysfunction with decreased levels of BH₄, decreased production of NO and an increased production of superoxide anion (Shinozaki *et al.*, 2000). Similar results were reported in coronary artery endothelial cells from BB diabetic rats (Meininger *et al.*, 2000). However, in the current study, we did not observe any change in the BH₄ level or GTP cyclohydrolase I activity in SMA from db/db mice. These data indicate that an impaired biosynthetic pathway for BH₄ is not the cellular basis for endothelial dysfunction in the db/db mouse model. The level of BH₂ + biopterin was elevated in SMA from db/db mice, thus decreasing the ratio of BH₄ to

BH₂ and biopterin. Increased oxidation of BH₄ to BH₂ and biopterin would result in decreased levels of BH₄. Surprisingly, we did not observe a change in the BH₄ levels in SMA from db/db mice. Based on the recent report, we speculate that BH₄ is oxidized to trihydrobiopterin (BH₃^{*}) radical as well as BH₂ and biopterin (Kuzkaya *et al.*, 2003). BH₃^{*} radical is converted back to BH₄ by ascorbate, maintaining levels of BH₄, whereas BH₂ and biopterin accumulates in the system probably because BH₂ cannot be converted to BH₄ due to decreased expression or activity of DHPR. Vasquez-Vivar *et al.* (2002b) reported that the relative proportion of the oxidized to the reduced BH₄ metabolites regulate eNOS activity and the generation of superoxide. Thus, in the present study, increased levels of BH₂ + biopterin may competitively antagonize the binding of BH₄ to eNOS and thus uncouple eNOS. However, further experiments are required to investigate this hypothesis.

Increased oxidation of BH₄ in SMA from diabetic mice may result from an increased reactive oxygen species (ROS) and oxidative stress (Giugliano *et al.*, 1996). ROS may impair endothelium-dependent relaxation by rapidly inactivating NO resulting in the formation of peroxynitrite. BH₄ is a powerful reducing agent and it is possible that excess peroxynitrite formation may lead to oxidation and depletion of BH₄ (Milstien & Katusic, 1999). In the present study, malondialdehyde levels were elevated in plasma and SMA from db/db mice and were restored to within the control range by sepiapterin treatment, suggesting that the restoration of the endothelial function by sepiapterin could in part be explained by an antioxidant action of sepiapterin. Direct antioxidant properties of BH₄ have also been documented by Kojima *et al.* in xanthine/xanthine oxidase free-radical generating system and phorbol myristate acetate stimulated rat macrophage generated free radicals (Kojima *et al.*, 1995). Oral supplementation with BH₄ normalized vascular superoxide production, membrane lipid peroxidation and prevented the activation of nuclear factor kappa B, activating protein-1 in insulin-resistant rats (Shinozaki *et al.*, 2000). Patel *et al.* (2002) reported that the scavenging efficiency of BH₄ was comparable to ascorbate and could be a biologically viable antioxidant (Patel *et al.*, 2002). This will also explain the effect of sepiapterin treatment on enhanced vascular reactivity to PE in SMA from db/db mice. Free radicals are known to impair endothelium integrity, enhance alpha-adrenergic receptor-mediated turnover of phosphoinositol and augment contractility through calcium channels (Chang *et al.*, 1993). Paradoxical negative reports, however, have been reported on the effects of sepiapterin on endothelial dysfunction. Sepiapterin attenuated Ach- and A23187-induced relaxation of isolated aorta from cholesterol-fed rabbits although increased the levels of BH₄ (Vasquez-Vivar *et al.*, 2002a). It is possible that prolonged exposure to high concentration of sepiapterin has a direct effect to uncoupled eNOS, resulting in the production of superoxide instead of NO.

In conclusion, increased oxidative stress through oxidation of BH₄ uncouples eNOS leading to endothelial dysfunction in SMA from db/db mice, and oral treatment with sepiapterin improves endothelial dysfunction by reducing oxidative stress.

We acknowledge the financial support of an operating grant from Norman D. MacDougall Canadian Diabetes Association (to TJA & C.R.T.) and a graduate fellowship award from Pfizer/CHS/CIHR and AHFMR (to M.P.).

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(Received May 20, 2003

Revised July 2, 2003

Accepted July 25, 2003)