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Experimental febrile seizures are precipitated by a hyperthermiainduced respiratory alkalosis

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Abstract

Febrile seizures are frequent during early childhood, and prolonged (complex) febrile seizures are associated with an increased susceptibility to temporal lobe epilepsy. The pathophysiological consequences of febrile seizures have been extensively studied in rat pups exposed to hyperthermia. The mechanisms that trigger these seizures are unknown, however. A rise in brain pH is known to enhance neuronal excitability. Here we show that hyperthermia causes respiratory alkalosis in the immature brain, with a threshold of 0.2–0.3 pH units for seizure induction. Suppressing alkalosis with 5% ambient CO₂ abolished seizures within 20 s. CO₂ also prevented two long-term effects of hyperthermic seizures in the hippocampus: the upregulation of the I_h current and the upregulation of CB1 receptor expression. The effects of hyperthermia were closely mimicked by intraperitoneal injection of bicarbonate. Our work indicates a mechanism for triggering hyperthermic seizures and suggests new strategies in the research and therapy of fever-related epileptic syndromes.

> Febrile seizures are the most common type of convulsive events in humans between the ages of 6 months and 6 years, with a prevalence that varies from 3% to 14% between different populations worldwide^{1,2}. Although most febrile seizures are apparently benign, one-third of them are 'complex,' with a prolonged duration (>10-20 min), and are associated with a risk of subsequent epilepsy 3,4 . Febrile seizures have been extensively studied in a well-established animal model, in which prolonged seizure activity is evoked by exposing rat pups to a hyperthermic environment that raises the body temperature to a level comparable to human fever^{5,6}. Work on this model has mainly focused on the short- and long-term consequences of experimental febrile seizures, including effects on neuronal survival⁷, receptor and channel expression⁸, excitability^{9–11}, plasticity¹² and seizure susceptibility¹³. Notably, however,

COMPETING INTERESTS STATEMENT

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there is little information available on the mechanisms that are responsible for the triggering and maintenance of these hyperthermia-induced seizures.

It is known that an elevated body temperature results in an increase in the rate of breathing, especially in young children^{14–16}, which reflects the general role of ventilation in the regulation of mammalian body temperature^{17–19}. Although changes in breathing pattern do not necessarily have an effect on the partial pressure of CO₂ (*P*CO₂) in the blood, it has been reported that hyperventilation (by definition, a decrease in *P*CO₂ resulting in an alkaline shift in pH²⁰) can develop during ongoing hyperthermia¹⁸. This is interesting in the present context, as it is generally known that a rise in pH leads to an increase in neuronal excitability and often to epileptiform activity both *in vitro* and *in vivo*^{20–30}.

The present work shows that, in the rat pup model of complex experimental febrile seizures, hyperthermia leads to thermal tachypnea and to a consequent, age-dependent respiratory alkalosis that triggers and sustains convulsions. A moderate elevation of ambient CO_2 to 5% results in the abolishment of the hyperthermia-induced respiratory alkalosis and in a fast block of the seizures. Our results suggest new designs for therapies aimed at preventing or interrupting febrile seizures and related convulsive events. Our observations are also likely to be useful in devising strategies in the search for factors at the genetic and systems^{31–34} levels that enhance an individual's susceptibility for fever-related epileptic syndromes.

RESULTS

Age dependence of hyperthermia-induced tachypnea and seizures

To examine whether hyperthermia-induced seizures are associated with changes in respiration, we made parallel measurements of body temperature and breathing frequency in rat pups at postnatal days (P)8–P11 and P22–P23. In the P8–P11 rats (n = 21), breathing frequency was 163 ± 14 breaths/min under control conditions, and an increase in body temperature was closely paralleled by an increase in the breathing rate. The hyperthermia-induced seizures occurred when the breathing rate increased by about 60%, to a value of 254 ± 44 breaths/min (Fig. 1a). This rate was achieved when the body temperature increased from its control level of 33.4 ± 0.9 to 41.8 ± 0.7 °C. Both the accelerated breathing and the seizures were promptly terminated by reducing the body temperature. This was followed by a slow, transient undershoot to 140 ± 17 breaths/min in the breathing rate.

In the P22–P23 group (n = 14), exposure to hyperthermia resulted in an increase in body temperature to 44.2 ± 1.1 °C from its control level of 34.3 ± 0.7 °C, and in a 28% increase in breathing frequency from 112 ± 11 breaths/min to 144 ± 23 breaths/min (Fig. 1a). In contrast to the younger rats, the 55 min of hyperthermia experienced by the older rats did not evoke seizures⁵.

Hyperthermia induces respiratory alkalosis in the immature brain

The above experiments show that changes in body temperature produce marked changes in breathing rate in rat pups¹⁹. But this thermal tachypnea does not, as such, necessarily imply that there would be a reduction in systemic PCO_2 with a consequent increase in pH affecting the brain^{18,20}. Hence, we carried out experiments on freely moving rat pups at P8–P11 and at P22–P23 with an implanted cortical pH electrode.

At P8–P11, the accelerated breathing during hyperthermia was closely paralleled by a rise in intracortical pH (Fig. 1b). The onset of seizure activity took place when the intracortical pH was elevated by 0.23 ± 0.08 units from its control level of 7.22 ± 0.096 (n = 9). Thereafter, the pH shift showed a further slight increase to a maximum of 0.27 ± 0.04 units above control. After reduction of body temperature, the recovery of cortical pH was, in parallel with the

respiration rate, followed by a slight but consistent rebound below the initial control level (Fig. 1b). Hence, the changes in the rate of breathing and brain pH were tightly and monotonically linked to alterations in body temperature at P8–P11. In contrast to this linkage, hyperthermia produced a moderate increase in respiration rate but no detectable change in brain pH in the more mature, P22–P23 rats (pH, 7.26 ± 0.08 and 7.29 ± 0.09 in control and hyperthermia, respectively; n = 8, P = 0.52).

Brain alkalosis and seizure activity evoked by bicarbonate

To examine whether the hyperthermia-induced brain alkalosis is sufficient to trigger the seizures observed at P8–P11, we used intraperitoneal injections of bicarbonate to evoke pH shifts in the brain. Bicarbonate at 1 mmol/kg induced a small alkalosis in the cortex (0.09 \pm 0.03 units, n = 8), but no seizures were observed (Fig. 1c). But increasing the dose to 5 mmol/kg produced a fast increase in cortical pH of 0.29 \pm 0.05 units which was accompanied by seizures in seven of seven experiments (Fig. 1d). Both the duration of the alkaline cortical pH shift after injection of bicarbonate and the associated behavioral seizure activity were more brief (~5 min) than those observed under hyperthermia. This is expected, considering that the bicarbonate injection produced only a transient alkaline load. Apart from their more brief duration, the behavioral characteristics of the bicarbonate-induced seizures were similar to those induced by hyperthermia. The threshold pH change at seizure onset after administration of bicarbonate was considerably similar (0.26 \pm 0.06 units; n = 7) to what was seen under hyperthermia (Fig. 1e).

Elevation of CO₂ suppresses hyperthermia-induced ictal activity

The data described so far support the idea that the experimental febrile seizures are triggered by brain alkalosis. If this is so, a straightforward and testable prediction is that inhibition of respiratory alkalosis should suppress the hyperthermia-induced ictogenesis. We carried out simultaneous recordings with electrodes implanted into the hippocampus and into the adjacent temporal cortex in the nonanesthetized, freely moving P8–P11 pups. In 12 of 12 experiments in which movement artifacts did not preclude an accurate analysis of the exact onset times of the electrographic ictal events, a near-coincident seizure onset in the hippocampus and the adjacent temporal cortex was seen, with a mean delay of 1.3 ± 0.8 s in the cortical versus hippocampal activity (Fig. 2a). The frequency of field potential discharges during the ictal activity was 3.0-3.5 Hz in both the hippocampus and cortex (n = 26).

An inhibition of hyperventilation-induced physiological effects can be effectively achieved by increasing the ambient CO₂ level while keeping the oxygen level constant³⁵. In agreement with this, application of 5% CO₂ during fully developed hyperthermia-induced seizures completely blocked the electrographic ictal activity within approximately 15–25 s in both the hippocampus and the cortex (Fig. 2a,b). The markedly fast block of seizure activity was seen in all 22 experiments of this kind. In addition, we found that the electrographic ictal activity and the associated behavioral manifestations were completely and invariably blocked when 5% CO₂ was preapplied shortly before, and maintained throughout, the exposure to hyperthermia (Fig. 2c). Exposure of the pups to 5% CO₂ had no effect on body temperature during hyperthermia (41.8 ± 0.9 °C and 42.1 ± 1.1 °C in the absence versus presence of 5% CO₂; n = 8, P = 0.78).

As expected on the basis of the behavioral observations, the bicarbonate-induced electrographic seizures at P8–P11 were characterized by a rapid onset and brief duration of about 4.4 ± 1.8 min. Exposure to 5% CO₂ at the time of bicarbonate injection completely blocked the hippocampal and cortical ictal activity as well as their behavioral correlates (n = 11; Fig. 2d).

Finally, direct measurements of cortical pH in the P8–P11 pups during the exposure to 5% CO₂ showed that the cortical alkalosis in response to hyperthermia was completely abolished. There was a slight, delayed acidosis that achieved its maximum value of 0.10 ± 0.02 units (n = 5) with a delay of 15–20 min (Fig. 2e). The brain alkalosis seen in response to injection of 5 mmol/kg bicarbonate was also blocked by 5% CO₂ and, again, a small and slowly developing acid shift took place (0.11 ± 0.03 units, n = 5; Fig. 2e).

CO₂ blocks the long-term effects of hyperthermia

Previous work has shown a long-lasting upregulation of the hyperpolarization-activated cation current (I_h) in response to hyperthermia-induced seizures¹⁰. To test whether blocking seizure activity by exposure of the pups to 5% CO₂ inhibits this effect of hyperthermia on I_h , we carried out whole-cell recordings from CA1 pyramidal cells in hippocampal slices 8–11 d after the experimental manipulations. Indeed, the hyperthermia-induced enhancement of the hyperpolarization-activated 'sag' potential and of the rebound depolarization¹⁰ was completely blocked in slices taken from pups that were exposed to hyperthermia in the presence of 5% CO₂ (Fig. 3a). Furthermore, the hyperthermia-induced increase in the average number of action potentials evoked by the rebound depolarization¹⁰ was also blocked in these slices (Fig. 3a).

The effects of the bicarbonate-induced seizures on I_h were similar to those evoked by hyperthermia (Fig. 3b). But two consecutive injections of bicarbonate were needed for effects that were quantitatively comparable to those induced by hyperthermia. This is probably attributable to the brief duration of the seizures induced by bicarbonate.

In addition to the upregulation of I_h , another salient molecular effect characteristic of complex experimental febrile seizures is a long-term change in cannabinoid signaling¹². Therefore, we carried out CB1 receptor western blot analyses from hippocampi at 1–120 d after causing the hyperthermia-induced seizures at P10 (Fig. 4a). We observed a rise in CB1 receptor protein that peaked 5 d after the hyperthermia-induced seizures to a level of about 170% versus control (Fig. 4b). This was followed by a decline at 10 d, and a full recovery to the control level was evident at 120 d. In contrast to the hippocampi from pups that had undergone hyperthermia-induced seizures, no statistically significant changes in the expression of the CB1 protein were seen in hippocampi from pups exposed to hyperthermia in the presence of 5% CO₂.

Again, the bicarbonate seizures mimicked the effects of hyperthermia-induced seizures, and a single injection was sufficient for a pronounced upregulation ($144 \pm 16\%$; n = 6) of CB1 receptor expression (Fig. 4c). This suggests a lower threshold for the seizure effects on CB1 than on I_h. The bicarbonate-induced effect on expression of CB1 was completely suppressed when seizure activity was blocked by 5% CO₂.

DISCUSSION

Here, we show that an increase in the body temperature of rat pups leads to a pronounced increase in the rate of respiration, which is followed by an increase in brain pH that triggers ictal activity. That the ictogenesis in the present model of febrile seizures^{5,6} is causally connected to the respiratory alkalosis is supported by several lines of evidence. First, there is an extensive amount of work showing that alkalosis of brain tissue leads to enhanced neuronal excitability and to epileptiform activity^{20–30}. We found that in the P8–P11 pups (i) seizure activity induced by hyperthermia had a well-defined pH threshold of about 0.25 units; (ii) seizures were provoked by brain alkalosis of a similar magnitude after injection of bicarbonate; (iii) an increase in the ambient CO₂ to 5% blocked the hyperthermia-associated brain alkalosis as well as the experimental febrile seizures, without affecting body temperature; and (iv) the bicarbonate-induced pH changes and seizures were also blocked by 5% ambient CO₂. Finally,

although hyperthermia led to a moderate increase in respiratory frequency, there was no respiratory alkalosis and, consistent with this, no seizures in rats at P22–P23. The dependence on age of experimental febrile seizures has been well characterized^{5,6} but, so far, no clear explanation has been provided for the high propensity of rats for hyperthermia-induced seizures at around P10. This topic will be discussed in detail below.

Prevention of brain alkalosis and the consequent experimental febrile seizures by 5% CO₂ blocked two types of hyperthermia-induced changes at the level of neuronal communication and long-term plasticity: the increase in the I_h current in hippocampal pyramidal neurons¹⁰ and the increase in the expression of the CB1 cannabinoid receptor¹². Our data indicate that the upregulation of both I_h and CB1 are caused by seizure activity and not by hyperthermia as such. Moreover, we found that with regard to the long-term modulation of I_h and CB1, the effects of the hyperthermia-induced seizures were closely mimicked by those caused by bicarbonate injection. This provides additional support for the key role of brain alkalosis. We also show for the first time that robust, hyperthermia-induced electrographic seizure activity can be recorded not only in the hippocampus⁶ but also in the temporal cortex. A major conclusion that can be drawn from the present work, however, is that the basic mechanisms underlying experimental febrile seizures cannot be elucidated by focusing solely on hippocampal and cortical functions. It is obvious that the temperature-sensitive mechanisms controlling respiratory activity and acid-base homeostasis have a key role in the generation of experimental febrile seizures.

Despite the abundance of data on the age dependence of experimental febrile seizures⁵, previous work has shed little light on the question of why the immature rat brain has a particularly high susceptibility for the generation of hyperthermia-induced seizures at the end of the second week of life. It should be noted that the experimental febrile seizures are evoked in normal, healthy rat pups that do not have any specific pathological susceptibility for hyperthermia-induced seizures. Our observations provide a new view into this problem.

In mammals, the neuronal mechanisms that control respiration have the dual task of playing an important role in thermoregulation and maintaining the partial pressures of blood gases (both oxygen and CO₂) constant^{18,36,37}. But the immature neuronal mechanisms that control respiration in the neonatal rat^{19,38} are apparently not able to cope with the above dual homeostatic functions when challenged by an abnormally high increase in ambient temperature. The overall result is a thermal tachypnea that strives to cool the body, but obviously does so at the expense of a net loss in CO₂ homeostasis, which thereby becomes manifest as a substantial respiratory alkalosis in brain tissue. It is of interest to note that the CO₂ chemosensitivity of rat pups reaches a minimum at around P10 (ref. 39). Our observation that hyperthermia leads to respiratory alkalosis in P8–P11 but not in P22–P23 rats are in agreement with the fact that the 'nadir' in CO₂ chemosensitivity³⁹ coincides with the developmental time window in which the hyperthermia-induced seizures have their lowest threshold^{5,6}.

It is widely assumed that experimental febrile seizures as studied in the present rat pup model have, at least to a certain degree, a mechanistic basis similar to simple and complex febrile seizures in children. The fact that hyperthermia induces respiratory alkalosis in the immature brain leads to a number of consequences that are likely to be relevant for the understanding of the basic mechanisms underlying febrile seizures and related disorders, such as febrile seizures plus (FS+) and generalized epilepsy with febrile seizures plus (GEFS+) 40,41 . In addition to abnormalities in hippocampal and cortical functions, the disease mechanisms underlying FS, FS+ and GEFS+ may involve neuronal mechanisms that control respiration and acid-base homeostasis. Obviously, homeostatic malfunctions of these types are not mutually exclusive, and various kinds of susceptibility factors may coexist in an individual. There is notably little

information available on the effects of fever on respiratory functions in children¹⁴. Hence, data from blood gas analyses (especially of CO_2) during fever-related epileptic episodes will be important in order to further examine the relevance of the present observations for the human condition.

In future studies, it will be interesting to study whether some of the ion channel gene mutations underlying febrile seizures, FS+ and GEFS+ have an influence on the pH sensitivity of the corresponding mutated channels^{31,32,42}. Furthermore, in view of the polygenic heterogeneity underlying epilepsy³², genes coding for proteins involved in the control of respiration^{36,37} provide a novel, functionally attractive class of candidate modifier or susceptibility genes for epilepsy. This, of course, does not exclude genuinely temperature-sensitive mechanisms in FS +, GEFS+ and other kinds of fever-related epileptic syndromes⁴³.

The blocking effect of ambient 5% CO₂ on the experimental febrile seizures in the rat pups was markedly fast and potent, with a delay of only ~ 20 s to a complete suppression of electrographic ictal activity. The prompt action of CO₂ raises the possibility that, also in children, ongoing ictal activity associated with fever could be quickly suppressed by a rise in ambient CO₂. Notably, CO₂ also prevented the conspicuous effects on long-term neuronal plasticity that are known to take place after hyperthermia-induced seizures^{9–12}. In conclusion, our work indicates a mechanism for triggering hyperthermia-induced seizures and suggests new strategies in the research and therapy of fever-related epileptic syndromes.

METHODS

Rats

We used pups from timed-pregnant Wistar rats which were kept together with their littermates except during experiments. The results are mainly from P8–P11 pups in which hyperthermic seizures were reliably induced, with comparisons to pups at P22–P23. All experiments were approved by the Committee for Animal Care and Use at the University of Helsinki and by the Ethic Committee of the Charité, University Medicine, Berlin.

Induction of seizures

We raised the body temperature of the pups, monitoring rectal temperature with a thermocouple (K101; Voltcraft) in a chamber with an ambient temperature of 48 ± 2 °C. The exposure to hyperthermia lasted 55 min and, at P8–P11, the latency to onset of hyperthermia-induced seizures (31.2 ± 3.7 min, n = 32 pups) showed little variation. The seizures had a broad spectrum of behavioral characteristics^{5,34} with a duration of ~ 25 min (24.6 ± 1.7 min). Initially, there was a sudden interruption of movements followed by oral automatisms. Thereafter, clonic movements of the limbs and the head, chewing of an extremity and tonic flexion of the body took place, often associated with a loss of postural control.

We used a thin tube connected to a 26-gauge polyethylene cannula (Becton Dickinson) placed intraperitoneally for application of bicarbonate (100 mM NaHCO₃ and 154 mM NaCl). The seizures evoked by a single bicarbonate injection (5 mmol/kg) were much more brief, but markedly similar to those induced by hyperthermia. After a brief delay following injection (~ 2 min), an interruption of movements was seen, followed by hyperthermia-induced seizure–like oral automatisms and tonic-clonic movements of the limbs and the head. In some experiments on I_h, we applied two consecutive bicarbonate injections of 5 mmol/kg with an interval of 15 min.

We monitored the breathing patterns of the pups using a piezo crystal sensor (movement sensor 230; Siemens) and applied CO₂ at constant oxygen partial pressure using a prewarmed gas

mixture containing 5% CO₂, 19% O₂, 76% N₂ (AGA). A 95% change in the CO₂ concentration in the chamber took place in 8.7 \pm 4.2 s (n = 25; CO₂ analyzer CD-101; Datex).

Anesthesia

We used hypothermia⁴⁴ for chronic implantation of the Ag-AgCl electrodes in P5–P6 pups and for implantation of cortical pH electrodes 24–48 h before the experiments at P8–P11, and ketamine-xylazine anesthesia⁴⁵ for all pups older than P20.

Electrophysiology in vivo

We made direct-current recordings^{35,46} of cortical and hippocampal activity at P8–P11 using Teflon-coated silver wire (uncoated diameter, 0.125–0.25 mm; Advent Research Materials Ltd, with chlorided tips) implanted at P5 or P6. We sampled the signals at 0.5–3 kHz using a 12-bit data acquisition board (National Instruments).

We performed craniectomies without damaging the underlying dura using a standard miniature drill equipped with a 0.7-mm diameter carbide dental burr, and placed the electrodes at the following coordinates: 1.6–2.0 mm posterior from bregma, 1.6–2.0 mm lateral from midline, 1.8–2.2 mm below dura for hippocampal CA3 recordings and 0.3–0.7 mm below dura for recordings in the temporal cortex, with a subdural reference electrode above the cerebellum. We fixed the implanted electrodes using microconnectors (GM-4; Microtech) and dental acryl. We sutured the incision for electrode implantation using 6-0 monofilament nylon. After recovery from anesthesia, we returned pups into their original litter. To verify the electrode positions, we injected dye into the hippocampus and cortex, dissected the brain into coronal sections and subsequently examined the sections using light microscopy.

The threshold for detecting the onset of the electrographic seizure activity evoked by hyperthermia or bicarbonate was defined as the time point at which the ictal activity reached an amplitude greater than three standard deviations of the baseline activity.

Monitoring of cortical pH in vivo

We made H⁺-sensitive electrodes using plastic tubing with a tip outer diameter of 0.1–0.2 mm and a PVC-gelled membrane solution⁴⁷. This enabled the construction of short and stable membrane columns with a signal that is not sensitive to changes in temperature⁴⁸. We back-filled electrodes with a solution containing 150 mM NaCl, 20 mM HEPES and 10 mM NaOH.

We placed the H^+ -sensitive microelectrodes 0.5–1 mm into the cortex using a guiding plastic tubing (conical; tip outer diameter, 0.9 mm), which was implanted above the dura (2.0 mm posterior from bregma, 2.0 mm lateral from midline) and fixed with dental acryl. As the reference, we implanted an Ag-AgCl electrode close to the H^+ microelectrode.

Electrophysiology in vitro.

We performed whole-cell recordings in hippocampal slices in a submerged-type chamber that was perfused with physiological solution at $21-22 \degree C^{10,49}$ (Supplementary Methods online).

Western blotting

We performed western blot analyses from hippocampi using a CB1-specific rabbit antibody (1:5,000) raised against the C terminus of the rat CB1 protein⁵⁰. Optical densities of the bands were analyzed with the AIDA imaging software (Raytest; **Supplementary Methods**).

Statistical analysis

All data are presented as mean \pm s.d. One-way analysis of variance (ANOVA) was followed by the Bonferroni-Dunn post hoc test.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Figure 1.

Hyperthermia-induced behavioral seizures are associated with brain alkalosis. (a) Hyperthermia (HT)-induced changes in body temperature (lower panels) and breathing rate (upper panels) in P8–P11 (n = 21; left panels) and P22–P23 (right panels) rat pups. The green bar marks the onset and end of the hyperthermia-induced seizures; the red bar indicates exposure to hyperthermia. Data here and below are mean \pm (or +) s.d. Seizures did not occur in P22–P23 rats. (b) Simultaneous recording of hyperthermia-induced changes in breathing rate and intracortical pH in a P9 (left) and a P23 (right) rat. The green bar indicates hyperthermia-induced seizures. (c) Intraperitoneal application (arrow) of 1 mmol/kg bicarbonate induces a small (~0.1 pH units) alkaline shift in pH in the brain of a P9 rat, but no seizure activity. (d) Application of 5 mmol/kg bicarbonate in a P9 rat induces an intracortical alkalosis of 0.24 pH units. The green bar indicates seizure activity. (e) Summary of cortical pH changes in P8–P11 rat pups at threshold for seizure initiation during hyperthermia (n = 9) and upon injection of 5 mmol/kg bicarbonate (n = 7). The two sets of data are not statistically different (P = 0.424).



Figure 2.

Exposure of rat pups to 5% ambient CO₂ blocks hyperthermia- and bicarbonate-induced brain alkalosis and associated ictal activity. (**a**) Recording in a P9 rat showing simultaneous hyperthermia (HT)-induced ictal activity in the hippocampus and in the temporal cortex, and the fast antiepileptic action of ambient 5% CO₂. (**b**) Time delay between exposure of the pups to 5% CO₂ to the block of ongoing ictal activity in the cortex and hippocampus in the P8–P11 rats (n = 29). (**c**) Cortical recording during a continuous 25-min hyperthermia-induced seizure (upper panel) in a P9 rat. Under identical hyperthermic conditions, but in the presence of 5% CO₂, no ictal activity is observed (lower panel). The insets show simultaneous cortical and hippocampal recordings at an expanded scale. (**d**) Cortical recording immediately after the onset and before the end of ictal activity evoked by intraperitoneal application of 5 mmol/kg bicarbonate (upper panel) in a P10 rat. The bicarbonate-induced ictal activity is completely

suppressed by exposure to 5% CO₂ (lower panel). The insets show simultaneous cortical and hippocampal recordings at an expanded scale. (e) The alkalosis induced by hyperthermia or intraperitoneal injection of 5 mmol/kg bicarbonate (P9 rats) is completely abolished in the presence of 5% CO₂. During prolonged exposure to 5% CO₂, a slow acid shift is seen.



Figure 3.

Ambient 5% CO₂ applied during hyperthermia blocks the long-term upregulation of the I_h current. (a) The amplitudes of the I_h -generated voltage sag and rebound depolarization evoked by hyperpolarizing current pulses in CA1 pyramidal neurons in hippocampal slices are enhanced in slices from pups previously exposed to hyperthermia compared to control slices¹⁰, but there are no significant differences between neurons from control and hyperthermia + 5% CO_2 rats (control, *n* (slices) = 6; hyperthermia, *n* = 7; hyperthermia + 5% CO_2 , n = 5). The inset shows recordings performed 10 d after the induction of hyperthermiainduced seizures at P9. The number of action potentials evoked by the rebound depolarization after hyperpolarizing currents ≥ 0.3 nA is increased in slices from pups previously exposed to hyperthermia¹⁰. No significant differences were found between neurons from hyperthermia +5% CO₂ and control rats (bottom left panel; control, n (slices) = 5; hyperthermia, n = 6; hyperthermia + 5% CO_2 , n = 4). Five to ten current pulses of each amplitude shown on the xaxis were applied. Specimen traces are shown on the bottom right. (b) After seizures caused by one or two bicarbonate injections, the I_h-generated sag (top left panel), the rebound depolarization (top right panel) and the number of action potentials evoked by the rebound depolarization after hyperpolarizing currents ≥ 0.3 nA (bottom panel) are enhanced (control, n (slices) = 8; one bicarbonate injection, n = 8; two bicarbonate injections, n = 6). *P < 0.05, **P < 0.01, ***P < 0.001 for test versus control (ANOVA).



Figure 4.

Ambient 5% CO₂ applied during hyperthermia blocks the long-term upregulation of the CB1 receptors. (**a**) Western blots of CB1 receptor protein from hippocampi 1, 2, 3, 5, 10 and 120 d after exposing P10 rats to hyperthermia (red) or to the hyperthermia + 5% CO₂ treatment (blue), and from control littermates (white). β -tubulin was used as a reference protein for the quantification shown in **b**. (**b**) Quantitative analysis of western blots. An upregulation of the CB1 receptor protein is evident after 2 d following hyperthermia, and a maximum is seen after 5 d. No significant change in CB1 receptor protein expression was found in the hyperthermia + 5% CO2 rats compared to controls (all values represent percentage changes normalized to β -tubulin levels). The value at each time point is based on data from 12 hippocampi. (**c**) After bicarbonate injection, an upregulation in CB1 receptor is seen at 5 d after seizures (*n* = 6). This effect is fully blocked in rats in which the seizures were suppressed by 5% CO₂. ****P* < 0.001, control versus hyperthermia, or control versus bicarbonate injection.