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Coibacins A and B: Total Synthesis and Stereochemical Revision

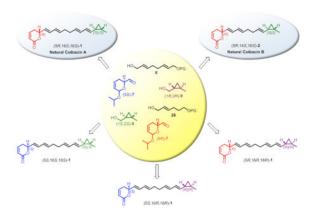
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Abstract



The interface between synthetic organic chemistry and natural products was explored in order to unravel the structure of coibacin A, a metabolite isolated from the marine cyanobacterium cf. Oscillatoria sp. that exhibits selective antileishmanial activity and potent anti-inflammatory properties. Our synthetic plan focused on a convergent strategy that allows rapid access to the desired target by coupling of three key fragments involving E-selective Wittig and modified Julia olefinations. CD measurements and comparative HPLC analyses between the natural product and four synthetic stereoisomers led to determination of its absolute configuration thus correcting the original assignment at C-5 and unambiguously establishing those at C-16 and C-18. Additionally, we have synthesized coibacin B based on the assignment of configuration for coibacin A.

Author Contributions

V. M. T. Carneiro and C. M. Avila contributed equally to this work.

The authors declare no competing financial interest.

Supporting Information

Experimental procedures, full spectroscopic data for new compounds, HPLC chromatograms and CD curves. This material is available free of charge via the Internet at http://pubs.acs.org.

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1. INTRODUCTION

Marine organisms are a rich source of diverse and biologically active natural products, which have inspired the development of novel pharmacologically relevant compounds. The α,β -unsaturated δ -lactone moiety is a privileged scaffold widely distributed among natural products that display a broad range of biological activities, such as fostriecin, $^{2-4}$ leptomycins, 5 and callystatin A. 6,7

The unequivocal assignment of the absolute configuration of the structure of a novel compound is one of the key issues in natural products chemistry. In this regard, and despite the fact that spectroscopic and crystallographic techniques are highly advanced, chemical synthesis still makes important contributions in natural products elucidation.^{8,9}

Recently, we described the isolation and structure elucidation of four unsaturated polyketides lactone derivatives, named coibacins A–D (compounds 1–4, Figure 1), from the marine cyanobacterium cf. *Oscillatoria* sp. which displayed the dihydropyran-2-one moiety as well as either a cyclopropyl ring (coibacins A and B) or a methyl vinyl chloride (coibacins C and D). ¹⁰ The methyl cyclopropyl ring and methyl vinyl chloride are similar to those observed in other marine cyanobacterial metabolites such as curacin A (5), ¹¹ and jamaicamide A (6), ¹² respectively. These co-occurring metabolites in a single organism suggest an intriguing flexibility in the biosynthetic pathway. ¹⁰

Among these cyanobacterial metabolites, coibacin A (1) displayed potent and selective activity against axenic amastigotes of *Leishmania donovani* (IC₅₀ = 2.4 μ M). Coibacin B (2) was less active as a leishmanicidal drug (IC₅₀ = 7.2 μ M); however, it exhibited higher cytotoxicity against human cancer lung cell lines (NCI-H460), with an IC₅₀ value of 17.0 μ M compared to 31.5 μ M for coibacin A (1). Evaluation of anti-inflammatory activity by cell-based nitric oxide (NO) inhibition assay¹³ revealed that coibacin B (2) was the most active coibacin representative (IC₅₀ = 5 μ M). Because coibacin A (1) was isolated in the largest amount, it was further evaluated and shown to reduce gene transcription of several inflammatory cytokines TNF- α , IL1- β , IL-6 and iNOS (1, at 10 μ g/mL).¹⁰

The absolute configuration of the dihydropyran-2-one moiety was determined to be S based on a positive Cotton effect at λ 259 nm, observed in circular dichroism (CD) measurements, while the *trans* relationship of the substituents in the methyl cyclopropyl ring was established using NOESY correlations and J-coupling constant analysis. However, because of the scarcity of the natural products, the absolute configuration of cyclopropyl portion could not be assigned. 10

Due to our interest into the synthesis and biological properties of α,β -unsaturated δ -lactones $^{14-18}$ we embarked on the total synthesis of stereoisomers of coibacin A (1) with the aim of establishing its absolute configuration and providing enough material for further exploration of its biological properties. Additionally, coibacin B (2) was synthesized based on the assignment of configuration for coibacin A (1) by a similar synthetic route.

2. RESULTS AND DISCUSSION

Assuming the *S* absolute configuration of δ -lactone moiety and the *trans* relationship between the substituents in the cyclopropyl ring, we envisioned a convergent strategy based on the coupling of three key fragments to quickly provide the two possible isomers (5*S*,16*S*, 18*S*)- and (5*S*,16*R*,18*R*)-1 (Scheme 1). This synthetic route would be carried out by *E*-selective Wittig olefination of dihydropyran aldehyde (5*S*)-7 with a tri-*n*-butyl phosphorane derived from alcohol 8. The resulting tetrahydropyran would be converted to the corresponding aldehyde for the final modified Julia olefination with a sulfone derived from

(1*S*,2*S*)- or (1*R*,2*R*)-9. The enantiomerically pure fragment (5*S*)-7 would be synthesized from commercially available (*R*)-glycidol, (*R*)-10, by similar methodology as previously reported for its enantiomer (5*R*)-7, involving as key steps ring opening of a chiral epoxide and ring-closing metathesis reaction of the corresponding diene. Preparation of fragment 8 could be performed in few steps from 1,4-butanediol (11) using tandem oxidation/Wittig olefination. 21 *trans*-Cyclopropyl fragment (1*S*,2*S*)-9 has been described using Charette asymmetric cyclopropanation of *trans*-crotyl alcohol (12) mediated by dioxaborolane-derived chiral ligand (4*R*,5*R*)-13. Similarly, the enantiomer (1*R*,2*R*)-9 could be obtained employing (4*S*,5*S*)-13.

Our synthesis began with the preparation of aldehyde (5S)-7 from (R)-10 according to a modification of the procedures described by Crimmins & King²⁰ and Boger *et al.*¹⁹ for (5R)-7. Protection of (R)-10 with *tert*-butyldiphenylsilyl chloride (TBDPSCl) and Cu(I)-mediated epoxide opening with vinylmagnesium bromide were carried out to provide allylic alcohol (S)-14 in 81% overall yield.²⁰ The dihydropyran-2-one (S)-15 was obtained in good yield after conversion of secondary alcohol (S)-14 to the corresponding ester, followed by ring-closing metathesis using Grubbs I catalyst (71% yield, 2 steps). Reduction of (S)-15 with diisobutylaluminum hydride (DIBALH) and protection of the resulting lactol as the corresponding isopropyl ketal afforded (5S)-16 in 86% yield as a single isomer.^{19,23} Deprotection of silyl ether group of (5S)-16 (83% yield) and Swern oxidation furnished the desired aldehyde (5S)-7,²⁰ which was synthesized from (R)-10 in 8 steps and 42% overall yield (Scheme 2).

The synthesis of fragment **8** started with the conversion of 1,4-butanediol (**11**) to the diester **18**. Two reaction steps were necessary to perform this transformation because treatment of 1,4-butanediol (**11**) with 20 equiv of magnesium dioxide (MnO₂) and 2.5 equiv of [(ethoxycarbonyl)methylene]triphenylphosphorane in dichloromethane or chloroform furnished the desired bis-olefinated compound **18** in very poor yield (15–18%) even after reflux for 3 d. Thus, mild conditions were employed to conduct a desymmetrization reaction *via* tandem oxidation of unactivated **11** with MnO₂ and *in situ* Wittig olefination to provide olefin (*E*)-**17** in 75% yield.²¹ The scale up of this reaction (from 2 to 30 mmol of substrate) required increased reaction time from 2 to 7 d and optimization studies indicated that 50% reduction in the amount of MnO₂ did not affect the yield. Byproducts of about 4% of diester **18** and 10% of (*Z*)-**17** were obtained from this reaction. Intermediate **17** was homologated to afford **18** in good yield (77%) after one-pot oxidation with pyridinium chlorocromate (PCC) and Wittig olefination. In this reaction the byproduct (2*E*,6*Z*)-isomer of **18** was isolated in 7% yield (Scheme 3).

Reduction of **18** to the corresponding diol with DIBALH and mono-protection with *tert*-butyldimethylsilyl chloride (TBSCl) afforded fragment **8** (PG = TBS) in 55% yield for 2 steps. Conversion to the allylic bromide **19** upon treatment with triphenylphosphine (Ph₃P) and carbon tetrabromide (CBr₄) in the presence of 2,6-lutidine was accomplished in high yield (98%);²⁴ however, in the absence of base the yield decreased to 45% and the dibromide byproduct was obtained in 28% yield. Subsequently, substitution with tri-*n*-butylphosphine (*n*-Bu₃P) afforded phosphonium salt **20** in quantitative yield. Therefore, **20** was prepared in 6 steps and 31% overall yield from readily available diol **11** (Scheme 3).

trans-Cyclopropyl fragment (1*S*,2*S*)-**9** was obtained from Charette asymmetric cyclopropanation of *trans*-crotyl alcohol (**12**) mediated by dioxaborolane-derived chiral ligand (4*R*,5*R*)-**13**.^{22,25} Treatment of (1*S*,2*S*)-**9** with 2-mercaptobenzothiazole (BTSH), Ph₃P and diisopropyl azodicarboxylate (DIAD) afforded (1*S*,2*S*)-**21** after Mitsunobu reaction.²⁵ Oxidation with ammonium molybdate/H₂O₂ provided sulfone (1*S*,2*S*)-**22** in 39% yield for 3

steps and enantiomeric ratio (er) of 94:6 (Scheme 4). Preparation of (1R,2R)-22 was accomplished from the same starting material 12 but using dioxaborolane (4S,5S)-13 and the overall yield was 31%. The enantiomeric ratio of (1S,2S)-22 and (1R,2R)-22 was determined by chiral HPLC analysis. ²⁶

Synthesis of the C-1/C-14 fragment of isomers (5*S*,16*R*,18*R*)- and (5*S*,16*S*,18*S*)-1 (Scheme 1) involved coupling of aldehyde (5*S*)-7 with tri-*n*-butyl phosphonium salt **20** by *E*-selective Wittig olefination (Scheme 5). When this reaction was performed with potassium *tert*-butoxide (*t*-BuOK) in a mixture of toluene/THF (5:1), the desired product (5*S*)-**23** was obtained as a 5:1 mixture of *E:Z* isomers. Using the DMSO derived lithium base [LiCH₂S(O)CH₃] in toluene a 7:1 *E:Z* isomeric ratio was formed. However, in both cases the yields were low (46% or less) regardless of the reaction scale and modifications in the amount of base or Wittig salt (**20**) employed. Therefore, we used sodium hexamethyldisilazide (NHMDS) as a more hindered base to prevent deprotonation α to the carbonyl group of (5*S*)-7. Fortunately, the yield was increased to about 70% (6:1 *E:Z* ratio) by using 2 equiv of NHMDS and 1.5 equiv of **20** in toluene. The moderate *E* selectivity of this reaction may be attributed to the low steric hindrance of the ylide.²⁷ The triene (5*S*)-**23** was converted uneventfully to aldehyde (5*S*)-**24** upon deprotection of the TBS ether followed by MnO₂-mediated allylic alcohol oxidation in 86% yield for 2 steps (Scheme 5).

The final coupling between aldehyde (5*S*)-24 and sulfonylbenzothiazole 22 required extensive optimization in order to improve the *E:Z* ratio of the newly formed double bond.²⁸ To avoid use of expensive enantiomerically enriched substrate, racemic 22 was employed in the modified Julia olefination reaction (Table 1). When NHMDS was used, either DMF or THF afforded (5*S*)-25 as a 1.7:1 mixture of *E:Z* isomers at C-14 in moderate to low yields (entries 1–3, Table 1). Under the same condition, reduction of the reaction time led to an increase in the yield from 28% to 65%, probably due to reduced decomposition of the product in the reaction medium (entries 2 and 3). Use of DME as solvent increased the *E:Z* ratio at C-14 to 2.7:1 and the yield (95%) (entry 4). Addition of hexamethylphosphoramide (HMPA) did not affect the selectivity (entries 5 and 6) although studies in the literature have shown improvement of the *E* selectivity in the presence of this additive.^{29,30} While LHMDS, as expected, favored the *Z* configuration of the newly formed double bond (entry 7), KHMDS displayed similar results to those reported for NHMDS (entry 8).

The isomers (5S,16S,18S)- and (5S,16R,18R)-1 were prepared as described in Scheme 6. Coupling of aldehyde (5S)-24 with sulfones (1S,2S)- or (1R,2R)-22 provided the tetraenes (5S,16S,18S)- and (5S,16R,18R)-25, respectively, as mixture of four geometric isomers with the major isomer displaying all E configuration. The mixtures of isomers (5S,16S,18S)- and (5S,16R,18R)-1 were obtained after oxidation of (5S,16S,18S)- and (5S,16R,18R)-25 with PCC and CaCO₃ in 22% and 29% yield, respectively. Separation of the major isomers by semi-preparative reverse phase HPLC³³ afforded dihydropyran-2-ones (5S,16S,18S)- and (5S,16R,18R)-1 which displayed 1 H- and 1 3C-NMR data identical to each other, as well as to that reported for natural coibacin A (1), in spite of their diastereoisomeric relationship.

Comparison of the specific optical rotation of the synthetic samples of (5S,16S,18S)-1 $([\alpha]_D^{20} - 10, c \ 0.1, \text{CHCl}_3)$ and (5S,16R,18R)-1 $([\alpha]_D^{20} - 100, c \ 0.1, \text{CHCl}_3)$ with that reported for natural coibacin A (1) $([\alpha]_D^{20} + 46, c \ 0.1, \text{CHCl}_3)$, indicated non-identity of the compounds. Based on the opposed signal of the specific optical rotation of the synthetic and natural compounds, we suspected that the original assignment of absolute configuration of lactone moiety was in error. Therefore, we conducted the total synthesis of isomers (5R,16S,18S)- and (5R,16R,18R)-1 by a similar route as described above for their enantiomers (See Experimetal Section for further details). As expected, these compounds also displayed

identical 1 H- and 13 C-NMR data to that reported for natural coibacin A (1). Despite their dextrorotatory values, the specific optical rotations of (5*R*,16*S*,18*S*)-1 ([α]_D²⁰ + 100, c 0.1, CHCl₃) and (5*R*,16*R*,18*R*)-1 ([α]_D²⁰ + 10, c 0.1, CHCl₃) proved to be significantly different from that reported for the natural coibacin A (1).

Due to the difficulty of unequivocal assignment of the absolute configuration of coibacin A (1) based only on NMR and specific optical rotation data, we carried out HPLC comparisons³⁴ of natural coibacin A (1) with all four synthetic stereoisomers displaying a *trans* configuration of the cyclopropane ring (Figure 2). When a mixture of these four compounds was co-injected with natural coibacin A (1), an enhancement in the intensity of the peak corresponding to (5R,16S,18S)-1 was observed, unequivocally establishing the absolute configuration of the natural product. Additionally, the CD curve of (5R,16S,18S)-1 was identical to the one described for coibacin A (1), with both displaying a positive Cotton effect (see CD curves in Supporting Information). Therefore these results led to the revision of the configuration at C-5 in our original publication which was previously assigned to be S, 10 possibly due to a misapplication of the rules described by Beecham. 35

It is interesting to compare the chirality established here for coibacin A (1) with that of curacin A (5, Figure 1), a methylcyclopropane-containing metabolite isolated from another marine cyanobacterium, Moorea producens (formerly Lyngbya majuscula). There, detailed biosynthetic mechanism studies have shown that S-configuration of the secondary methyl group appended to the cyclopropyl ring is established by the trajectory of hydride delivery to the β-position of an achiral enone. ³⁶ However, the adjacent cyclopropyl methine center is rendered chiral (and in the case of curacin A, cis-configured relative to the methyl substituent) after collapse of the enolate to displace chloride and form the three-membered ring. Thus, because curacin A (5) and coibacin A (1) possess secondary methyl groups of identical chirality, divergence in their biosynthetic pathways must lie within this second step. A range of possibilities exist, including the geometry of the intermediate enone formed by the enoyl CoA hydratase 1, the relative positions of the oxyanion and primary chloride substituents to the methyl group prior to cyclopropyl ring formation, and the overall orientation of the substrate in the enoyl reductase. Thus, similar to the subtle alternate functioning of β-branch forming enzymes that lead to the diverse functional groups observed in curacin A (5, Figure 1) and jamaicamide A (6, Figure 1) (cyclopropyl ring versus vinyl chloride),³⁷ the stereo-structures of the coibacins implicate additional dimensions of flexibility in this biosynthetic manifold.

A similar synthetic approach was applied to the synthesis of coibacin B (2) displaying 5R, 14S, 16S configuration, the same one found for coibacin A (1) (Scheme 7). Fragment **26** was planned to be obtained after two steps from alcohol **17**, already prepared from diol **11** for the synthesis of coibacin A (1).

Protection of alcohol **17** with TBSCl, followed by reduction of ester group with excess of DIBALH, furnished fragment **26** (PG = TBS) in high yield (94%, 2 steps). Appel reaction converted the alcohol **26** to the corresponding bromide **27** in 91% yield, which was treated with *n*-Bu₃P to give the tri-*n*-butyl phosphonium salt **28** in quantitative yield (Scheme 8).

E-selective Wittig reaction between (5R)-7 and 28 mediated by NHMDS in toluene furnished (5R)-29 as a 6:1 mixture of *E*:*Z* isomers at C-6 in 69% yield. Alcohol (5R)-30, obtained in 94% yield from deprotection of TBS ether (5R)-29, was oxidized under Swern conditions and the unstable crude product was immediately used in the modified Julia olefination with sulfone (1S,2S)-22 to afford (5R,14S,16S)-31 as a mixture of geometric isomers. While the use of NHMDS in DME provided a 1:1 mixture of geometric isomers at C-12 double bond, better *E* selectivity (2.2:1 E:Z ratio) was obtained by using of KHMDS.

The moderate yield for the Swern oxidation and Julia olefination (41%) can be explained by the instability of aldehyde derived from alcohol (5R)-30. The mixture (5R,14S,16S)-31 was treated with PCC in the presence of CaCO₃ to give a mixture of four geometric isomers (5R, 14S,16S)-2′ in 46% yield. The major isomer (5R,14S,16S)-2, with all double bonds displaying E configuration, was isolated by semi-preparative reverse phase HPLC (Scheme 9).

Synthetic (5R,14S,16S)-2 and natural coibacin B displayed identical spectra within the limits of NMR data (see Supporting Information). As for coibacin A (1), the specific optical rotation for the synthetic sample of coibacin B (2) ($[\alpha]_D^{20} + 95$, c 0.1, CHCl₃) was higher than that reported for the natural compounds ($[\alpha]_D + 59$, c 0.1, CHCl₃). Fortunately, the CD curve of synthetic (5R,14S,16S)-2 closely resembles to that of natural coibacin A (1), providing strong evidence that this represents the correct isomer of coibacin B (2).

3. CONCLUSION

In summary, total syntheses of four coibacin A (1) stereoisomers displaying a trans-configured cyclopropane ring revealed the natural substance to possess 5R,16S,18S absolute configuration. The total synthesis of coibacin A (1) was carried out in 12 steps (longest linear route) and 3.4% overall yield. Additionally, we synthesized the correct isomer of coibacin B (2) based on the assignment of configuration for coibacin A (1). Studies aimed to assess the cytotoxic and anti-inflammatory properties of these compounds are underway.

4. EXPERIMENTAL SECTION

General Information

THF and DME were freshly distilled from sodium/benzophenone. DCM, DMF, HMPA and Et₃N were freshly distilled over CaH₂. PhMe and MeCN were dried over 4 Å molecular sieves for 48 h prior to use. TLC analyses were performed in silica gel plates and the spots were revealed using UV and/or the following solutions: phosphomolybdic acid in EtOH, vanillin in EtOH/H₂SO₄ and *p*-anisaldehyde in EtOH/AcOH/H₂SO₄. Flash column chromatography was carried out using silica gel 60 (35–60 μm). $^1 H^{-1} H$ COSY (90°) and $^1 H^{-13} C$ HSQC NMR experiments were used for confirmation of NMR peak assignments. The HPLC analyses were performed at room temperature with a photodiode array detector. All melting points were measured in open capillaries and were uncorrected. HRMS analyses were performed using ESI-TOF mass spectrometer. IUPAC names of the compounds were generated using ChemBioDraw Ultra 13.0; however, the usual numbering of the carbon atoms as shown in Scheme 1 was adopted to refer to the compounds and NMR peak assignments.

(S)-1-((tert-Butyldiphenylsilyl)oxy)pent-4-en-2-ol, (S)-14

To a solution of (R)-10 (3.35 g, 45.2 mmol) and imidazole (4.00 g, 58.8 mmol) in anhyd DCM (100 mL), was added TBDPSCl (15.4 mL, 54.3 mmol) at 0 °C and under N₂. The solution was allowed to warm to rt and stirred for 2 h. The solution was quenched by the addition of water (100 mL). The layers were separated and the aqueous phase was extracted with Et₂O (2 × 50 mL). The combined organic layers were washed with brine (100 mL), dried over Na₂SO₄, filtered and concentrated. The crude product was purified by flash column chromatography (hexanes/Et₂O 9:1) to provide (S)-tert-butyl(oxiran-2-ylmethoxy)diphenylsilane (13.5 g, 43.3 mmol, 87%) as a colorless oil. [α]_D²⁰ – 2.6 (c 9.4, CHCl₃). Lit.³⁹ [α]_D²³ – 2.46 (c 9.07, CHCl₃). Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature.³⁹

To a solution of (*S*)-tert-butyl(oxiran-2-ylmethoxy)diphenylsilane (13 g, 42 mmol) in anhyd THF (19 mL) and under N_2 was added CuI (0.634 g, 3.32 mmol, 8 mol%) at rt. This suspension was stirred for 15 min. After that, the temperature was reduced to $-30\,^{\circ}$ C and vinylmagnesium bromide (100 mL, 100 mmol, 1M in THF) was added by syringe pump (over 1 h). The mixture was then stirred an additional hour at this temperature. The reaction was carefully quenched with saturated NH₄Cl solution (100 mL) at $-30\,^{\circ}$ C, the cooling bath was removed and the reaction was stirred for more 20 min. The layers were separated and the aqueous phase was extracted with Et₂O (2 × 100 mL). The combined organic layers were washed with brine (100 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude product was purified by flash column chromatography (hexanes/Et₂O 7:3) to afford alcohol (*S*)-14 (13.4 g, 39.4 mmol, 94%) as a colorless oil. $[\alpha]_D^{20} - 2.4$ (c 9.6, CHCl₃). Lit.⁴⁰ $[\alpha]_D^{20} - 2.6$ (c 9.17, CHCl₃). Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature.⁴⁰

(S)-6-(((tert-Butyldiphenylsilyl)oxy)methyl)-5,6-dihydro-2H-pyran-2-one, (S)-15

To a solution of alcohol (*S*)-**14** (12.97 g, 38.08 mmol) and Et₃N (10.6 mL, 76.2 mmol) in anhyd DCM (131 mL) was added freshly prepared acryloyl chloride⁴¹ (4.62 mL, 57.1 mmol) dropwise at 0 °C and under N₂. At this time color change from light yellow to dark yellow was observed. The reaction was stirred for 1 h at the same temperature and then it was quenched by the addition of brine (50 mL) and saturated Rochelle's salt solution (50 mL). The emulsion was stirred until complete layer separation (~ 3 h). The layers were separated and the aqueous phase was extracted with Et₂O (2 × 100 mL). The combined organic layers were dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude product was purified by flash column chromatography (hexanes/Et₂O 7:3) to give (*S*)-1-((*tert*-butyldiphenylsilyl)oxy)pent-4-en-2-yl acrylate (12.92 g, 32.74 mmol, 86%) as a yellow oil. [α]D²⁰ – 9 (c 1.05, CHCl₃). Lit.⁴² (antipode) [α]D²⁶ + 8.3 (c 1, CHCl₃). Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature for its enantiomer.⁴²

To a solution of (*S*)-1-((*tert*-butyldiphenylsilyl)oxy)pent-4-en-2-yl acrylate (4.00 g, 10.1 mmol) in anhyd DCM (1 L) at reflux was added Grubbs catalyst 1st generation (0.823 g, 1.03 mmol, 10 mol%) in three portions (275 mg, each hour) dissolved in anhyd DCM (10 mL). The reaction mixture was refluxed for 20 h. The temperature was cooled to rt and the solvent was evaporated under reduced pressure. The crude product was purified by flash column chromatography (hexanes/AcOEt 6:4) to provide lactone (*S*)-15 (3.08 g, 8.40 mmol, 83%) as a brown oil. $[\alpha]_D^{20} - 48$ (*c* 1.03, CHCl₃). Lit. ⁴² (antipode) $[\alpha]_D^{26} + 47.8$ (*c* 1, CHCl₃). Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature for its enantiomer. ⁴²

tert-Butyl(((2S)-6-isopropoxy-3,6-dihydro-2H-pyran-2-yl)methoxy)diphenylsilane, (5S)-16

To a solution of lactone (S)-15 (1.2 g, 3.27 mmol) in anhyd DCM (40 mL) and under N_2 was added a solution of DIBALH (4 mL, 1.2 M in toluene) at $-78\,^{\circ}\text{C}$. The reaction was stirred for 1 h at the same temperature and then it was quenched by the addition of saturated NaHCO3 solution (40 mL) and Rochelle's salt solution (60 mL). The emulsion formed was stirred until complete phase separation (~ 3 h). The layers were separated and the aqueous phase was extracted with DCM (3 \times 100 mL). The combined organic layers were dried over MgSO4, filtered and concentrated under reduced pressure. The product was used in the next step without purification.

The crude product was taken up in toluene (16 mL) and then isopropyl alcohol (8.00 mL, 105 mmol) and PPTS (0.025 g, 0.098 mmol, 3 mol%) were added at rt. The reaction mixture was stirred for 12 h at the same temperature and it was quenched by the addition of saturated

NaHCO $_3$ solution (20 mL). The mixture was stirred for 30 min. The layers were separated and the aqueous phase was extracted with DCM (3 × 40 mL). The combined organic layers were dried over Na $_2$ SO $_4$, filtered and concentrated under reduced pressure. The product was purified by flash column chromatography (hexanes/AcOEt 6:4) to give (5S)-16 (1.16 g, 2.82 mmol, 86%, 2 steps) as a yellow oil and as a single isomer. [α] $_D^{20}$ – 28 (c 0.66, DCM). Lit. 20 (antipode) [α] $_D^{24}$ + 28.2 (c 0.68, DCM). Spectral data (1 H NMR and 1 3C NMR) are in accordance with those reported in the literature for its enantiomer. 20 ,23

(R)-1-((tert-Butyldiphenylsilyl)oxy)pent-4-en-2-ol, (R)-14

(*R*)-*tert*-butyl(oxiran-2-ylmethoxy)diphenylsilane was prepared from (*S*)-**10** (3.66 g, 49.5 mmol) by similar procedure as described above for its enantiomer. The crude product was purified by flash column chromatography (gradient elution, 2 to 4% of AcOEt in hexanes) to provide the expected product (13.6 g, 43.5 mmol, 88%) as a colorless oil. [α]_D²⁰ + 2.5 (c 2, CHCl₃). Lit.⁴³ [α]_D²⁰ + 2.3 (c 2, CHCl₃). Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature.⁴³

Alcohol (*R*)-**14** was obtained from (*R*)-*tert*-butyl(oxiran-2-ylmethoxy)diphenylsilane (13.4 g, 42.9 mmol) as described above for (*S*)-**14**. The crude product was purified by flash column chromatography (hexanes/AcOEt 75:25) to give (*R*)-**14** (12.8 g, 37.6 mmol, 87%) as a colorless oil. $[\alpha]_D^{20} + 3$ (*c* 1, CHCl₃). Lit.⁴³ $[\alpha]_D^{20} + 2.9$ (*c* 1, CHCl₃). Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature.⁴³

(R)-6-(((tert-Butyldiphenylsilyl)oxy)methyl)-5,6-dihydro-2H-pyran-2-one, (R)-15

(*R*)-1-((*tert*-butyldiphenylsilyl)oxy)pent-4-en-2-yl acrylate was prepared from (*R*)-14 (12.7 g, 37.3 mmol) using a similar procedure to that reported above for its enantiomer. The crude product was purified by flash column chromatography (hexanes/Et₂O 8:2) to furnish the desired product (13.9 g, 35.2 mmol, 94%) as a yellow oil. [α]_D²⁰ + 11 (c 1, CHCl₃). Lit.⁴² [α]_D²⁶ + 8.3 (c 1, CHCl₃). Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature.⁴²

Lactone (*R*)-**15** was obtained from (*R*)-1-((*tert*-butyldiphenylsilyl)oxy)pent-4-en-2-yl acrylate (5.0 g, 13 mmol) by similar procedure described for (*S*)-**15**. The crude product was purified by flash column chromatography (hexanes/AcOEt 7:3) to provide (*R*)-**15** (3.81 g, 10.4 mmol, 82%) as a brown oil. $[\alpha]_D^{20} + 41$ (*c* 1, CHCl₃). Lit.⁴² $[\alpha]_D^{26} + 47.8$ (*c* 1, CHCl₃). Lit.⁴³ $[\alpha]_D^{23} + 34.2$ (*c* 1.5, CHCl₃). Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature. ^{42,43}

tert-Butyl(((2R)-6-isopropoxy-3,6-dihydro-2H-pyran-2-yl)methoxy)diphenylsilane, (5R)-16

Compound (5*R*)-16 was prepared from (*R*)-15 (3.0 g, 8.4 mmol) as described for (5*S*)-16. The product was purified by flash column chromatography (hexanes/AcOEt 6:4) giving (5*R*)-16 (3.05 g, 7.42 mmol, 88%, 2 steps) as a yellow oil and as a single isomer. $[\alpha]_D^{20}$ + 30 (*c* 0.65, DCM). Lit.²⁰ $[\alpha]_D^{24}$ + 28.2 (*c* 0.68, DCM). Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature.^{20,23}

Ethyl (E)-6-hydroxyhex-2-enoate, 17

To a solution of 1,4-butanediol (11) (3.10 g, 34.3 mmol) in DCM (1.2 L) was added respectively (ethoxycarbonylmethylene)triphenylphosphorane (14.3 g, 41.2 mmol) and MnO₂ (10.0 g, 115 mmol) and the mixture was stirred at rt. After 24 h and 48 h, respectively, other two additional portions of MnO₂ (2 × 10.0 g) were added. The mixture was stirred for a further 5 d and filtered through Celite® using DCM for washing. The solvent was evaporated under reduced pressure and the residue was treated with Et₂O to

allow precipitation of Ph₃PO. After filtration and solvent evaporation, the crude product was purified by flash chromatography (hexanes/AcOEt 1:1), affording the E isomer **17** (4.595 g, 25.92 mmol, 75%) as the major product, together with its Z isomer (Z)-6-hydroxyhex-2-enoate (622 mg, 3.93 mmol, 11%) and the diester **18** (316 mg, 1.40 mmol, 4%) as byproducts. All compounds were obtained as colorless oils and their spectral data (IR, 1 H NMR and 13 C NMR) are in accordance with those reported in the literature. 21

Diethyl (2E,6E)-octa-2,6-dienedioate, 18

A mixture of **17** (2.35 g, 14.9 mmol) and PCC (6.5 g, 30.1 mmol, ground with 12.9 g of silica) in DCM (0.6 L) was stirred at rt for 5 h. Imidazole was added (2.02 g, 29.7 mmol) and the reaction mixture was stirred for an additional 1 h. After addition of (ethoxycarbonylmethylene)triphenylphosphorane (12.4 g, 35.6 mmol), stirring was continued for 24 h. The mixture was filtered through Celite[®] using DCM for washing. The solvent was removed under reduced pressure and the crude material was treated with Et₂O to promote precipitation of Ph₃PO. After filtration, the solvent was evaporated and the residue was purified by flash chromatography (hexanes/AcOEt 8:2) to give E:E diester **18** (2.598 g, 11.48 mmol, 77%) and its Z:E isomer diethyl (2Z,6E)-octa-2,6-dienedioate (228 mg, 1.00 mmol, 7%) as colorless oils. Spectral data (IR, ¹H NMR and ¹³C NMR) of **18** and its geometric isomer are in accordance with those reported in the literature.²¹

(2E,6E)-8-((tert-Butyldimethylsilyl)oxy)octa-2,6-dien-1-ol, 8

To a solution of **18** (2.598 g, 11.48 mmol) in anhyd DCM (50 mL) was slowly added via cannula a solution of DIBALH (9 mL, 50.5 mmol) in DCM (15 mL) at -78 °C and under N₂. The reaction mixture was stirred under reduced temperature for 2 h. After that, were added Et₂O (150 mL) and saturated solution of Rochelle's salt (50 mL), maintaining vigorous stirring for 1 h. After phase separation, the aqueous one was extracted with Et₂O (2 × 50 mL). The combined organic phases were washed with brine (50 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The crude product was purified by flash chromatography (AcOEt) to afford the corresponding diol (2*E*,6*E*)-octa-2,6-diene-1,8-diol (1.539 g, 10.82 mmol, 94%) as a light yellow oil. Spectral data (IR, ¹H NMR and ¹³C NMR) are in accordance with those reported in the literature.⁴⁴

To a solution of (2E,6E)-octa-2,6-diene-1,8-diol (1.27 mg, 8.93 mmol) in anhyd DMF (18 mL) was added TBSCl (897 mg, 5.95 mmol) and imidazole (608 mg, 8.93 mmol). The resulting mixture was stirred at rt for 24 h and the reaction was quenched by addition of H_2O (50 mL). After extraction with AcOEt $(3 \times 100 \text{ mL})$, the combined organic phases were dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by flash chromatography (gradient elution, 10 to 100% of AcOEt in hexanes) affording the mono-protected compound **8** (894 mg, 3.49 mmol, 59%), the bis-silyl ether (6E,10E)-2,2,3,3,14,14,15,15-octamethyl-4,13-dioxa-3,14-disilahexadeca-6,10-diene (251 mg, 0.677 mmol, 11%) as a byproduct, and recovery of starting material (532 mg, 3.74 mmol, 42%). Spectral data $(IR, {}^1H \text{ NMR} \text{ and } {}^{13}C \text{ NMR})$ of the isolated compounds are in agreement with the literature data.

(((2E,6E)-8-Bromoocta-2,6-dien-1-yl)oxy)(tert-butyl)dimethylsilane, 19

To a solution of **8** (611 mg, 2.38 mmol) in anhyd MeCN (14 mL) was added Ph₃P (1.25 mg, 4.77 mmol), 2,6-lutidine (0.55 mL, 4.75 mmol) and CBr₄ (1.58 g, 4.76 mmol), respectively, at 0 °C. The resulting solution was stirred at rt for 10 min. The reaction was quenched with H₂O (50 mL) and extracted with Et₂O (2 × 100 and 1 × 50 mL). The combined organic phases were washed with brine (50 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The crude product was purified by flash chromatography (hexanes/AcOEt

95:5) to give **19** (744 mg, 2.3 mmol, 98%) as a colorless oil. R_f : 0.64 (hexanes/AcOEt 95:5). IR (ATR) v_{max}/cm^{-1} : 2928, 2856, 1472, 1254, 966, 835, 775. 1 H NMR (500 MHz, CDCl₃) δ : 0.06 (s, 6H), 0.90 (s, 9H), 2.14 (s,br, 4H), 3.93 (d, J = 5.0 Hz, 2H), 4.11 (d, J = 2.5 Hz, 2H), 5.49–5.73 (m, 4H). 13 C NMR (125 MHz, CDCl₃) δ : - 5.1 (2 C), 18.4, 26.0 (3 C), 31.4, 31.6, 33.3, 63.8, 126.8, 129.7, 130.3, 135.6. HRMS (ESI-TOF) m/z: [M + Na]⁺ Calcd for C₁₄H₂₇BrOSiNa 341.09067; Found 341.09049.

2-((((1S,2S)-2-Methylcyclopropyl)methyl)sulfonyl)benzo[d]thiazole, (1S,2S)-22

To a mixture of DME (2 mL, 19 mmol) and anhyd DCM (18 mL) was added a solution of diethylzinc (1 M in hexanes, 19 mL) at -10 °C and under N₂. Diiodomethane (3 mL, 37 mmol) was added dropwise and the solution was stirred at -10 °C for 20 min. This fresh Zn(CH₂I)₂·DME solution was added by cannula over a solution of *trans*-crotyl alcohol (12) (540 mg, 7.50 mmol) and butylboronic acid *N*,*N*,*N'*,*N'*-tetramethyl-L-tartaric acid diamide ester, (4*R*,5*R*)-13 (2.2 g, 8.1 mmol), in DCM (37 mL) at -10 °C and the mixture was stirred at 0 °C for 4 h. Saturated aqueous NH₄Cl (50 mL) was added and the mixture was extracted with Et₂O (4 × 50 mL). The combined organic phases were stirred with NaOH 2 M (100 mL) overnight. After phase separation, the organic one was washed with aqueous solution of HCl 1 M (50 mL), saturated solution of NaHCO₃ (50 mL) and brine (50 mL). The organic phase was dried over MgSO₄, filtered, and concentrated under reduced pressure (450 mbar). The crude product ((1*S*,2*S*)-2-methylcyclopropyl)methanol, (1*S*,2*S*)-9, was obtained as a light yellow oil and used in the next step without purification to avoid losses due to its volatility. The enantiomeric ratio was determined to be 94:6 after analysis of the corresponding Mosher's ester by ¹H NMR (500 MHz).

To a solution of (1S,2S)-9 obtained above in THF (15 mL) was successively added 2-mercaptobenzothiazole (1.0 g, 6.0 mmol), Ph_3P (1.57 g, 5.97 mmol) and DIAD (1.3 mL, 6.6 mmol). The mixture was stirred at rt for 24 h and the solvent excess was removed under reduced pressure. Et_2O was added to the residue and the precipitate was removed by filtration. After solvent evaporation, the crude material was partially purified by flash chromatography (hexanes/AcOEt 95:5) to give 2-((((1S,2S)-2-methylcyclopropyl)methyl)thio)benzo[d]thiazole, (1S,2S)-21 (1.033 g), as a light yellow oil. This intermediate was not characterized because it was still impure.

To a solution of (1S,2S)-**21** (1.033 g) in EtOH (40 mL) was added a solution of ammonium molybdate tetrahydrate (540 mg, 0.437 mmol) in H_2O_2 30% v/v (2 mL) at 0 °C and under N_2 . The resulting mixture was stirred at rt for 18 h and the solvent was removed under reduced pressure. The residue was diluted with H_2O (50 mL) and extracted with DCM $(5 \times 50 \text{ mL})$. The combined organic phases were washed with brine (50 mL), dried with MgSO₄, evaporated, and the residue was purified by flash chromatography (hexanes/AcOEt 8:2) to afford (1S,2S)-**22** (890 mg, 3.33 mmol, 39% for 3 steps, er 94:6) as a white solid. mp: 77–80 °C. Lit.²⁵ mp: 94–96 °C. [α]_D²⁰ + 8 (c 1.13, CHCl₃). Lit.²⁵ [α]_D²⁰ + 7 (c 1.14, CHCl₃). The enantiomeric ratio of (1S,2S)-**22** was determined by chiral HPLC analysis²⁶ and its spectral data $(\text{IR}, {}^1\text{H} \text{ NMR})$ and ${}^{13}\text{C} \text{ NMR})$ are in agreement with the literature data.²⁵

2-((((1R,2R)-2-Methylcyclopropyl)methyl)sulfonyl)benzo[d]thiazole, (1R,2R)-22

Sulfone (1R,2R)-22 was prepared from 12 in 31% overall yield and er 94:6 employing a similar procedure as described above for (1S,2S)-22, but using butylboronic acid N,N,N',N'-tetramethyl-D-tartaric acid diamide ester, (4S,5S)-13. mp: 79–82 °C. [α]_D²⁰ – 8 (c 1.11, CHCl₃). Lit.²⁵ [α]_D²⁰ (antipode) + 7 (c 1.14, CHCl₃). The enantiomeric ratio of (1R,2R)-22 was determined by chiral HPLC analysis²⁶ and its spectroscopic data (IR, ¹H NMR and ¹³C NMR) are in accordance with those reported for its enantiomer by Charette.²⁵

2-((((trans)-2-Methylcyclopropyl)methyl)sulfonyl)benzo[d]thiazole (22)

Racemic sulfone **22** was produced from **12** in 83% overall yield using similar procedure as described for (1*S*,2*S*)-**22**, but employing Simmons–Smith cyclopropanation in place of asymmetric Charette's reaction. mp: 84–86 °C. The spectra data of racemic **22** (IR, ¹H NMR and ¹³C NMR) are in accordance with those reported for (1*S*,2*S*)-**22** by Charette.²⁵

tert-Butyl(((2E,6E,8E)-9-((2S)-6-isopropoxy-3,6-dihydro-2H-pyran-2-yl)nona-2,6,8-trien-1-yl)oxy)dimethylsilane, (5S)-23

To a solution of **19** (664 mg, 2.08 mmol) in anhyd MeCN (9.5 mL) was added *n*-Bu₃P (0.77 mL, 3.1 mmol) and the mixture was stirred at rt for 12 h. After that, the excess of solvent was removed under reduced pressure and the crude product was maintained under high vacuum (0.2 mmHg) for 8 h. The tri-*n*-butylphosphonium salt **20** was used in the Wittig olefination without purification.

To a solution of dihydropyran (5*S*)-**16** (1.41 g, 3.43 mmol) in anhyd THF (35 mL) at 0 °C and under N_2 was added a solution of TBAF (3.6 mL, 1 M in THF). The solution was allowed to warm to rt and stirred for 3 h. The reaction was quenched by the addition of H_2O (35 mL). The layers were separated and the aqueous layer was extracted with AcOEt (5 × 40 mL). The combined organic layers were dried over Na_2SO_4 , filtered and concentrated under reduced pressure. The crude product was purified by column chromatography (hexanes/AcOEt 6:4) to give the corresponding alcohol ((2*S*)-6-isopropoxy-3,6-dihydro-2*H*-pyran-2-yl)methanol (0.492 g, 2.85 mmol, 83%) as a white solid. mp: 43–45 °C. $[\alpha]_D^{20}$ – 47 (c 1, DCM). Lit.²⁰ (antipode) $[\alpha]_D^{24}$ + 40.4 (c 0.47, DCM). Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature for its enantiomer.²⁰

To a solution of oxalyl chloride (0.192 mL, 2.23 mmol) in anhyd DCM (10 mL) at -78 °C and under N₂, was added dropwise DMSO (0.227 mL, 3.12 mmol). After stirring for 15 min, a solution of the alcohol obtained above (0.240 g, 1.39 mmol) in anhyd DCM (4 mL) was added dropwise by cannula. The resultant white emulsion was stirred for more 15 min and then Et₃N (1.0 mL, 6.9 mmol) was added at the reaction. After 5 min the mixture was allowed to warm to rt. The reaction was quenched by the addition of saturated NH₄Cl solution (15 mL). The layers were separated and the aqueous phase was extracted with Et₂O (3 × 25 mL). The organic layers were combined, washed with brine (20 mL), dried over Na₂SO₄, filtered and carefully concentrated (bath temperature: 30 °C, pressure 400–420 mbar) to prevent loss of the volatile aldehyde. The crude product (2*S*)-6-isopropoxy-3,6-dihydro-2*H*-pyran-2-carbaldehyde, (5*S*)-7, was obtained as a colorless oil which was used in the next step without purification because its instability. R_E 0.57 (hexanes/AcOEt 7:3).

Wittig Olefination—To a solution of freshly prepared aldehyde (5*S*)-7 (236 mg, 1.39 mmol) and tri-*n*-butylphosphonium salt **20** (2.08 mmol) in anhyd PhMe (25 mL) was added dropwise a solution of NHMDS (2.8 mL, 2.8 mmol, 1 M in THF) at -78 °C and under N₂. The mixture was warmed slowly and stirred over 6 h. The reaction was quenched with H₂O (50 mL) and extracted with AcOEt (3 × 100 mL). The combined organic phases were washed with brine (50 mL), dried over MgSO₄ and the solvent was removed under reduced pressure. The crude product was purified by flash chromatography (hexanes/AcOEt 95:5) to furnish (5*S*)-**23** (383 mg, 0.975 mmol, 70%) as a light yellow oil and as a 6:1 mixture of the two isomers at C-6/C-7 double bond. R_f : 0.50 (hexanes/AcOEt 95:5). IR (ATR) v_{max}/cm^{-1} : 2958, 2926, 2895, 2851, 1254, 1125, 1099, 1028, 988, 836, 776. ¹H NMR (500 MHz, CDCl₃) &: (major isomer) 0.06 (s, 6H, Si(CH₃)₂), 0.90 (s, 9H, C(CH₃)₃), 1.16 (d, J = 5.0 Hz, 3H, C(CH₃)₂), 1.22 (d, J = 5.0 Hz, 3H, C(CH₃)₂), 1.97–2.26 (m, 6H, H-4, H-10 and H-11), 4.00 (sept, J = 5.0 Hz, 1H, CH(CH₃)₂), 4.11 (d, J = 4.7 Hz, 2H, H-14), 4.42–4.46 (m, 1H, H-5), 5.10 (s,br, 1H, H-1), 5.52–5.72 (m, 5H, H-13, H-6, H-12, H-9 and H-2), 5.97–6.08 (m,

2H, H-3 and H-8), 6.22 (dd, J = 15.3 and 10.5 Hz, 1H, H-7). 13 C NMR (125 MHz, CDCl₃) δ : (major isomer) – 5.1 (2 CH₃, Si(CH₃)₂), 18.4 (C, Si-C), 22.0 (CH₃, C(CH₃)₂), 23.8 (CH₃, C(CH₃)₂), 26.0 (3 CH₃, C(CH₃)₃), 30.7 (CH₂, C-4), 31.8 (CH₂, C-11), 32.2 (CH₂, C-10), 63.9 (CH₂, C-14), 66.4 (CH, C-5), 69.4 (CH, C(CH₃)₂), 93.1 (CH, C-1), 126.1 (CH, C-2), 128.4 (CH, C-3), 129.7 (CH, C-13), 130.0 (CH, C-8), 130.2 (CH, C-12*), 130.8 (CH, C-6*), 131.1 (CH, C-7), 134.5 (CH, C-9); *indicates that assignments may be interchanged. HRMS (ESI-TOF) m/z: [M – OCH(CH₃)₂]⁺ Calcd for C₂₀H₃₃O₂Si 333.22443; Found 333.22458.

(2E,6E,8E)-9-((2S)-6-Isopropoxy-3,6-dihydro-2H-pyran-2-yl)nona-2,6,8-trienal, (5S)-24

To a solution of (5S)-23 (0.375 g, 0.955 mmol) in anhyd THF (48 mL) was added a solution of TBAF (1 mL, 1 mmol, 1 M in THF) at 0 °C and under N2. The yellow solution was allowed to warm to rt and stirred for 2 h. The reaction was quenched by the addition of H₂O (30 mL) and AcOEt (60 mL). The layers were separated and the aqueous phase was extracted with AcOEt (4 × 50 mL). The combined organic phases were dried over MgSO₄, filtered and concentrated under reduced pressure. The product was used in the next step without purification. To a solution of the alcohol obtained above in anhyd DCM (95 mL) was added MnO₂ (5 g, 57.3 mmol, 85% activated, Sigma-Aldrich) in 3 portions (every 24 hour). The reaction was allowed to stir for additional 24 h. The crude reaction mixture was directly filtered through a plug of silica gel, eluted with a mixture of hexanes/AcOEt 1:1, and concentrated under reduced pressure. The product was purified by flash chromatography (hexanes/AcOEt 6:4) to give (5S)-24 (227 mg, 0.821 mmol, 86 %, 2 steps) as a light yellow oil and as a 6:1 mixture of the two isomers at C-6/C-7 double bond. R_f: 0.54 (hexanes/ AcOEt 1:1). IR (ATR) $v_{\text{max}}/\text{cm}^{-1}$: 2959, 2922, 2850, 1691, 1124, 1099, 1026, 993, 668, 651. ¹H NMR (500 MHz, CDCl₃) δ : (major isomer) 1.19 (d, J = 6.1 Hz, 3H, C(CH₃)₂), 1.25 $(d, J = 6.1 \text{ Hz}, 3H, C(CH_3)_2), 2.01-2.14 \text{ (m, 2H, H-4)}, 2.32-2.36 \text{ (m, 2H, H-10)}, 2.45-2.49$ $(m, 2H, H-11), 4.02 \text{ (sept, } J = 6.1 \text{ Hz}, 1H, CH(CH_3)_2), 4.46-4.49 \text{ (m, 1H, H-5)}, 5.13 \text{ (s,br,}$ 1H, H-1), 5.66–5.79 (m, 3H, H-6, H-9 and H-2), 6.00–6.03 (m, 1H, H-3), 6.09–6.17 (m, 2H, H-8 and H-13), 6.25 (dd, J = 15.4 and 10.5 Hz, 1H, H-7), 6.85 (dd, J = 15.6 and 6.7 Hz, 1H, H-12), 9.52 (d, J = 7.9 Hz, 1H, H-14). ¹³C NMR (125 MHz, CDCl₃) δ : (major isomer) 22.0 (CH₃, C(CH₃)₂), 23.8 (CH₃, C(CH₃)₂), 30.6 (CH₂, C-4), 30.7 (CH₂, C-10), 32.2 (CH₂, C-11), 66.2 (CH₂, C-5), 69.5 (CH, C(CH₃)₂), 93.1 (CH, C-1), 126.1 (CH, C-2), 128.3 (CH, C-3), 130.5 (CH, C-7), 131.1 (CH, C-8), 131.9 (CH, C-9*), 132.5 (CH, C-6*), 133.3 (CH, C-13), 157.4 (CH, C-12), 193.9 (CHO, C-14); *indicates that assignments may be interchanged. HRMS (ESI-TOF) m/z: [M + Na]⁺ Calcd for C₁₇H₂₄O₃Na 299.1623; Found 299.1637.

(2S)-6-Isopropoxy-2-((1E,3E,7E,9E)-10-((1S,2S)-2-methylcyclopropyl)deca-1,3,7,9-tetraen-1-yl)-3,6-dihydro-2*H*-pyran, (5S,16S,18S)-25

Julia Olefination with THF. To a solution of aldehyde (5*S*)-**24** (16 mg, 0.058 mmol) and sulfone (1*S*,2*S*)-**22** (23 mg, 0.086 mmol) in anhyd THF (2.5 mL) was added dropwise a solution of NHMDS (120 μ L, 0.12 mmol, 1 M in THF) at – 78 °C and under N₂. The light yellow resulting mixture was stirred for 10 min and the reaction was quenched with H₂O (6 mL). The mixture was extracted with AcOEt (30 + 10 mL) and the organic layer was washed with brine (10 mL), dried over MgSO₄, and concentrated under reduced pressure. The crude product was purified by flash chromatography (hexanes/AcOEt 95:5) to afford (5*S*,16*S*,18*S*)-**25** as 1.7:1 *E:Z* mixture at C-14 double bond (12.4 mg, 0.0377 mmol, 65%), contaminated with minor amounts of the 6*Z* double bond isomers as a light yellow oil. R_f : 0.67 (hexanes/AcOEt 9:1). IR (ATR) v_{max}/cm^{-1} : 2969, 2925, 1654, 1447, 1380, 1316, 1181, 1099, 1027, 985, 718. ¹H NMR (500 MHz, CDCl₃) δ : (major isomer) 0.47–0.50 (m, 1H), 0.53–0.57 (m, 1H), 0.72–0.79 (m, 1H), 1.04–1.10 (m, 1H), 1.06 (d, J = 6.0 Hz, 3H), 1.17 (d, J = 6.2 Hz, 3H), 1.24 (d, J = 6.2 Hz, 3H), 2.03–2.25 (m, 6H), 4.01 (sept, J = 6.2 Hz, 1H), 4.42–4.47 (m, 1H), 5.10 (s,br, 1H), 5.15 (dd, J = 14.8 and 8.9 Hz, 1H), 5.48–5.56 (m, 1H),

5.62 (d, J = 15.3 and 6.1 Hz, 1H), 5.68–5.76 (m, 2H), 5.94–6.09 (m, 4H), 6.22 (dd, J = 15.3 and 10.2 Hz, 1H), (second largest isomer) 0.53–0.57 (m, 2H), 0.72–0.79 (m, 1H), 1.04–1.10 (m, 1H), 1.09 (d, J = 5.8 Hz, 3H), 1.17 (d, J = 6.1 Hz, 3H), 1.24 (d, J = 6.2 Hz, 3H), 2.03–2.25 (m, 6H), 4.01 (sept, J = 6.2 Hz, 1H), 4.42–4.47 (m, 1H), 4.72 (t, J = 10.4 Hz, 1H), 5.10 (s,br, 1H), 5.48–5.56 (m, 1H), 5.68–5.76 (m, 2H), 5.87 (t, J = 10.9 Hz, 1H), 5.94–6.09 (m, 4H), 6.47 (dd, J = 14.8 and 11.2 Hz, 1H). 13 C NMR (63 MHz, CDCl₃) δ : (major isomer) 15.6 (2 C), 18.5, 22.0, 22.9, 23.9, 30.7, 32.3, 32.6, 66.4, 69.4, 93.1, 126.1, 127.5, 128.5, 130.0, 130.1, 130.7, 130.9, 131.2, 134.6, 136.1; (second largest isomer) 15.8, 16.0, 18.5, 19.3, 21.9, 23.9, 30.8, 32.2, 32.7, 62.7, 69.3, 92.8, 126.3, 126.5, 128.6, 130.0, 130.9, 131.2, 132.8, 134.2, 134.6, 136.2. HRMS (ESI-TOF) m/z: [M + Na]⁺ Calcd for C₂₂H₃₂O₂Na 351.2300; Found 351.2288.

(S)-6-((1E,3E,7E,9E)-10-((1S,2S)-2-Methylcyclopropyl)deca-1,3,7,9-tetraen-1-yl)-5,6-dihydro-2*H*-pyran-2-one, (5S,16S,18S)-1

A mixture of PCC (323 mg, 1.50 mmol) and CaCO₃ (600 mg, 6 mmol) in anhyd DCM (6 mL) was stirred at rt for 30 min. Then a solution of (5S,16S,18S)-25 (100 mg, 0.304 mmol) in DCM (13 mL) was added by cannula and the reaction mixture was stirred at rt for 1 h. After filtration over silica gel and solvent evaporation under reduced pressure, the crude product was purified by flash chromatography (gradient elution, 20 to 30% of AcOEt in hexanes) to give (5S,16S,18S)-1' (19 mg, 0.067 mmol, 22%) as a mixture of E:Z isomers at C-14/C-15 double bond and contaminated with minor amounts of the 6Z isomers. This mixture was separated using preparative HPLC with SunFire[™] Prep C₁₈ OBD[™] column (particle size 5 μm, dimensions 19 mm ×100 mm, 60% MeCN/40% H₂O, 17 mL/min) to yield the major isomer with E-configured double bonds (5S,16S,18S)-1 (6.7 mg, t_R 18.4 min) as a colorless oil. R_f : 0.41 (hexanes/AcOEt 7:3). [α]_D²⁰ – 10 (c 0.1, CHCl₃). IR (ATR) $v_{\text{max}}/\text{cm}^{-1}$: 2995, 2918, 1720, 1658, 1381, 1244, 986, 816. ¹H NMR (500 MHz, CDCl₃) δ : 0.47-0.51 (m, 1H, H-17), 0.54-0.58 (m, 1H, H-17), 0.73-0.80 (m, 1H, H-18), 1.05 (d, J = 0.47-0.51 (m, 1H, H-17), 0.54-0.58 (m, 1H, H-17), 0.73-0.80 (m, 1H, H-18), 0.54-0.58 (m, 1H, H-18), 0.54-0.58 (m, 1H, H-17), 0.73-0.80 (m, 1H, H-18), 0.54-0.58 (m, 1H, H-17), 0.73-0.80 (m, 1H, H-18), 0.54-0.58 (m, 1H, H-17), 0.73-0.80 (m, 1H, H-18), 0.54-0.58 (m, 1H, H-18), 0.54-0.58 (m, 1H, H-18), 0.54-0.58 (m, 1H, H-18), 0.73-0.80 (m, 1H, H-18), 0.54-0.58 (m, 1H, H-18), 0.546.0 Hz, 3H, H-19), 1.05–1.10 (m, 1H, H-16), 2.14–2.18 (m, 4H, H-10 and H-11), 2.43–2.45 (m, 2H, H-4), 4.92-4.96 (m, 1H, H-5), 5.15 (dd, J = 14.7 and 9.0 Hz, 1H, H-15), 5.50 (dt, J = 14.7), 5.50 = 14.1 and 6.6 Hz, 1H, H-12), 5.64 (dd, J = 15.3 and 6.6 Hz, 1H, H-6), 5.77 (dt, J = 15.2 and 6.6 Hz, 1H, 1H-9), 5.94–6.06 (m, 4H, H-2, H-8, H-13 and H-14), $6.30 \text{ (dd, } J = 15.3 \text{ and } 10.4 \text{ (d$ Hz, 1H, H-7), 6.85–6.89 (m, 1H, H-3). ¹³C NMR (125 MHz, CDCl₃) δ: 15.7 (CH₂, C-17; CH, C-18), 18.5 (CH₃, C-19), 22.8 (CH, C-16), 29.8 (CH₂, C-4), 32.1 (CH₂, C-11), 32.6 (CH₂, C-10), 77.9 (CH, C-5), 121.6 (CH, C-2), 126.7 (CH, C-6), 127.4 (CH, C-14), 129.1 (CH, C-8), 129.9 (CH, C-12), 130.8 (CH, C-13), 133.7 (CH, C-7), 136.3 (CH, C-15), 136.8 (CH, C-9), 144.6 (CH, C-3), 164.0 (CO, C-1). HRMS (ESI-TOF) m/z: [M + Na]+ Calcd for C₁₉H₂₄ONa 307.1674; Found 307.1663.

(2S)-6-Isopropoxy-2-((1E,3E,7E,9E)-10-((1R,2R)-2-methylcyclopropyl)deca-1,3,7,9-tetraen-1-yl)-3,6-dihydro-2*H*-pyran, (5S,16R,18R)-25

Julia Olefination with DME. To a solution of aldehyde (5S)-24 (103 mg, 0.373 mmol) and sulfone (1R,2R)-22 (150 mg, 0.561 mmol) in anhyd DME (17 mL) was added dropwise a solution of NHMDS (0.78 mL, 0.78 mmol, 1 M in THF) at -78 °C and under N₂. The light yellow mixture was stirred for 10 min and the reaction was quenched with brine (50 mL). The mixture was extracted with AcOEt (3 × 50 mL) and the organic layer was dried over MgSO₄ and concentrated under reduced pressure. The crude product was purified by flash chromatography (hexanes/AcOEt 95:5) to afford (5S,16R,18R)-25 as a 2.7:1 E:Z mixture at C-14 double bond (116 mg, 0.353 mmol, 95%), contaminated with minor amounts of the 6Z double bonds isomers as a light yellow oil. R_f : 0.67 (hexanes/AcOEt 9:1). The IR and NMR data of (5S,16R,18R)-25 are identical to those described for (5S,16S,18S)-25, despite their diastereoisomeric relationship. HRMS (ESI-TOF) m/z: [M + K]⁺ Calcd for C₂₂H₃₂O₂K 367.2039; Found 367.2030.

(S)-6-((1E,3E,7E,9E)-10-((1R,2R)-2-Methylcyclopropyl)deca-1,3,7,9-tetraen-1-yl)-5,6-dihydro-2*H*-pyran-2-one, (5S,16R,18*R*)-1

A mixture of PCC (361 mg, 1.67 mmol) and CaCO₃ (670 mg, 6.70 mmol) in anhyd DCM (7 mL) was stirred at rt for 2 h. Then a solution of (5*S*,16*R*,18*R*)-**25** (111 mg, 0.338 mmol) in DCM (12 mL) was added by cannula and the reaction mixture was stirred at rt for 2 h. After filtration over silica gel and solvent evaporation under reduced pressure, the crude product was purified by flash chromatography (gradient elution, 20 to 30% of AcOEt in hexanes) to afford (5*S*,16*R*,18*R*)-**1'** (28 mg, 0.098 mmol, 29%) as a mixture of *E:Z* isomers at C-14/C-15 double bond and contaminated with minor amounts of the 6*Z* isomers. This mixture was separated using preparative HPLC with SunFireTM Prep C₁₈ OBDTM column (particle size 5 μ m, dimensions 19 mm ×100 mm, 60% MeCN/40% H₂O, 17 mL/min) to give the major isomer with *E*-configured double bonds (5*S*,16*R*,18*R*)-**1** (12.9 mg, t_R 18.4 min) as a colorless oil. R_f : 0.41 (hexanes/AcOEt 7:3). [α]_D²⁰ – 100 (c 0.1, CHCl₃). The IR and NMR data of (5*S*,16*R*,18*R*)-**1** are identical to those described for (5*S*,16*S*,18*S*)-**1** despite their diastereoisomeric relationship. HRMS (ESI-TOF) m/z: [M + H]⁺ Calcd for C₁₉H₂₅O₂ 285.1855; Found 285.1866.

tert-Butyl(((2E,6E,8E)-9-((2R)-6-isopropoxy-3,6-dihydro-2H-pyran-2-yl)nona-2,6,8-trien-1-yl)oxy)dimethylsilane, (5R)-23

To a solution of **19** (568 mg, 1.78 mmol) in anhyd MeCN (8 mL) was added *n*-Bu₃P (0.70 mL, 2.8 mmol) and the mixture was stirred at rt for 12 h. After that, the excess of solvent was removed under reduced pressure and the crude product was maintained under high vacuum (0.2 mmHg) for 8 h. The tri-*n*-butylphosphonium salt **20** was used without purification.

Deprotection of (5R)-**16** (3.05 g, 7.31 mmol) employing a similar procedure described above for its enantiomer provided the corresponding alcohol. The crude product was purified by column chromatography (hexanes/AcOEt 6:4) to give the desired product ((2R)-6-isopropoxy-3,6-dihydro-2*H*-pyran-2-yl)methanol (1.14 g, 6.62 mmol, 90%) as a white solid. mp: 42- $44 ^{\circ}$ C. [α]_D²⁰ + 46 (c 1, DCM). Lit.²⁰ [α]_D²⁴ + 40.4 (c 0.47, DCM). Spectral data (1 H NMR and 13 C NMR) are in accordance with those reported in the literature.²⁰

Aldehyde (2R)-6-isopropoxy-3,6-dihydro-2H-pyran-2-carbaldehyde, (5R)-7, was prepared from the alcohol prepared above (0.205 g, 1.19 mmol) as described for (5S)-7. The crude product was obtained as a colorless oil that was used in the next step without purification because its instability. R_f : 0.57 (hexanes/AcOEt 7:3).

Wittig Olefination—Compound (5*R*)-23 was obtained from freshly prepared aldehyde (5*R*)-7 (202 mg, 1.19 mmol) and tri-*n*-butylphosphonium salt 20 (1.78 mmol) following the procedure described above for (5*S*)-23. The crude product was purified by flash chromatography (gradient elution, 0 to 10% of AcOEt in hexanes) to furnish (5*R*)-23 (368 mg, 0.937 mmol, 79%) as a light yellow oil and as a 5:1 mixture of the two isomers at C-6/C-7 double bond. R_f : 0.50 (hexanes/AcOEt 95:5). The IR and NMR data of (5*R*)-23 are identical to those described for its enantiomer (5*S*)-23. HRMS (ESI-TOF) m/z: [M + Na]⁺ Calcd for $C_{23}H_{40}O_3SiNa$ 415.2644; Found 415.2633.

(2E,6E,8E)-9-((2R)-6-Isopropoxy-3,6-dihydro-2H-pyran-2-yl)nona-2,6,8-trienal, (5R)-24

Aldehyde (5R)-24 was obtained from (5R)-23 (0.699 g, 1.78 mmol) using a similar procedure as described for (5S)-24 except for the reaction time of the MnO₂ oxidation which was reduced from 96 h to 72 h by using of another batch of MnO₂ (6.19 g, 71.2 mmol, 90% activated, Fluka Analytical). The product was purified by flash chromatography

(hexanes/AcOEt 6:4) to give (5R)-**24** (371 mg, 1.34 mmol, 75 %, 2 steps) as a light yellow oil and as a 5:1 mixture of the two isomers at C-6/C-7 double bond. R_f : 0.54 (hexanes/AcOEt 1:1). The IR and NMR data of (5R)-**24** are identical to those described for its enantiomer (5S)-**24**. HRMS (ESI-TOF) m/z: [M + Na]⁺ Calcd for C₁₇H₂₄O₃Na 299.1623; Found 299.1618.

(2R)-6-Isopropoxy-2-((1E,3E,7E,9E)-10-((1S,2S)-2-methylcyclopropyl)deca-1,3,7,9-tetraen-1-yl)-3,6-dihydro-2H-pyran, (5R,16S,18S)-25

Intermediate (5R,16S,18S)-**25** was prepared from aldehyde (5R)-**24** (105 mg, 0.380 mmol) and sulfone (1S,2S)-**22** (150 mg, 0.561 mmol) as described for (5S,16R,18R)-**25**. The crude product was purified by flash chromatography (hexanes/AcOEt 95:5) to afford (5R,16S,18S)-**25** as a 2.7:1 *E:Z* mixture at C-14 double bond (117 mg, 0.356 mmol, 94%), contaminated with minor amounts of the 6Z double bonds isomers as a light yellow oil. R_f : 0.67 (hexanes/AcOEt 9:1). The IR and NMR data of (5R,16S,18S)-**25** are identical to those described for (5S,16S,18S)-**25** despite their diastereoisomeric relationship. HRMS (ESITOF) m/z: $[M + Na]^+$ Calcd for $C_{22}H_{32}O_2Na$ 351.2300; Found 351.2306.

(*R*)-6-((1*E*,3*E*,7*E*,9*E*)-10-((1*S*,2*S*)-2-Methylcyclopropyl)deca-1,3,7,9-tetraen-1-yl)-5,6-dihydro-2*H*-pyran-2-one, (5*R*,16*S*,18*S*)-1

Compound (5R,16S,18S)-1 was synthesized from (5R,16S,18S)-25 (111 mg, 0.338 mmol) using a similar procedure to that described for (5S,16R,18R)-1. The crude product was purified by flash chromatography (gradient elution, 20 to 30% of AcOEt in hexanes) to give (5R,16S,18S)-1' (31 mg, 0.11 mmol, 33%) as a mixture of E:Z isomers at C-14/C-15 double bond and contaminated with minor amounts of the 6Z isomers. After separation in preparative HPLC with SunFire TM Prep C_{18} OBD Column (particle size 5 μ m, dimensions 19 mm ×100 mm, 60% MeCN/40% H₂O, 17 mL/min) the major isomer with *E*-configured double bonds (5R,16S,18S)-1 (11.9 mg, t_R 18.4 min) was obtained as a colorless oil. R_f : 0.41 (hexanes/AcOEt 7:3). [α]_D²⁰ + 100 (c 0.1, CHCl₃). IR (ATR) ν _{max}/cm⁻¹: 2995, 2918, 1720, 1658, 1381, 1244, 986, 816. ¹H NMR (500 MHz, CDCl₃) δ: 0.47–0.51 (m, 1H, H-17), 0.54– 0.58 (m, 1H, H-17), 0.73-0.80 (m, 1H, H-18), 1.05 (d, J = 6.0 Hz, 3H, H-19), 1.05-1.10 (m, 1H, H-16), 2.14–2.18 (m, 4H, H-10 and H-11), 2.43–2.45 (m, 2H, H-4), 4.92–4.96 (m, 1H, H-5), 5.15 (dd, J = 14.7 and 9.0 Hz, 1H, H-15), 5.50 (dt, J = 14.1 and 6.6 Hz, 1H, H-12), 5.64 (dd, J = 15.3 and 6.6 Hz, 1H, H-6), 5.77 (dt, J = 15.2 and 6.6 Hz, 1H, H-9), 5.94–6.06 (m, 4H, H-2, H-8, H-13 and H-14), 6.30 (dd, J = 15.3 and 10.4 Hz, 1H, H-7), 6.85-6.89 (m, 10.4 Hz, 1.0 Hz1H, H-3). ¹³C NMR (125 MHz, CDCl₃) δ: 15.7 (CH₂, C-17; CH, C-18), 18.5 (CH₃, C-19), 22.8 (CH, C-16), 29.8 (CH₂, C-4), 32.1 (CH₂, C-11), 32.6 (CH₂, C-10), 77.9 (CH, C-5), 121.6 (CH, C-2), 126.7 (CH, C-6), 127.4 (CH, C-14), 129.1 (CH, C-8), 129.9 (CH, C-12), 130.8 (CH, C-13), 133.7 (CH, C-7), 136.3 (CH, C-15), 136.8 (CH, C-9), 144.6 (CH, C-3), 164.0 (CO, C-1). HRMS (ESI-TOF) m/z: [M + Na]⁺ Calcd for C₁₉H₂₄O₂Na 307.1674; Found 307.1678.

(2R)-6-Isopropoxy-2-((1E,3E,7E,9E)-10-((1R,2R)-2-methylcyclopropyl)deca-1,3,7,9-tetraen-1-yl)-3,6-dihydro-2*H*-pyran, (5*R*,16*R*,18*R*)-25

Intermediate (5R,16R,18R)-**25** was synthesized from aldehyde (5R)-**24** (105 mg, 0.380 mmol) and sulfone (1R,2R)-**22** (145 mg, 0.543 mmol) as described for (5S,16R,18R)-**25**. The crude product was purified by flash chromatography (hexanes/AcOEt 95:5) to furnish (5R, 16R,18R)-**25** as a 2.7:1 *E:Z* mixture at C-14 double bond (118 mg, 0.359 mmol, 95%), contaminated with minor amounts of the 6Z double bonds isomers as a light yellow oil. R_f : 0.67 (hexanes/AcOEt 9:1). The IR and NMR data of (5R,16R,18R)-**25** are identical to those described for (5S,16S,18S)-**25**. HRMS (ESI-TOF) m/z: $[M + Na]^+$ Calcd for $C_{22}H_{32}O_2Na$ 351.2300; Found 351.2294.

(R)-6-((1E,3E,7E,9E)-10-((1R,2R)-2-Methylcyclopropyl)deca-1,3,7,9-tetraen-1-yl)-5,6-dihydro-2*H*-pyran-2-one, (5R,16R,18R)-1

Compound (5R,16R,18R)-1 was synthesized from (5R,16R,18R)-25 (110 mg, 0.335 mmol) using a similar procedure to that described above for (5S,16R,18R)-1. The crude product was purified by flash chromatography (gradient elution, 20 to 30% of AcOEt in hexanes) to furnish (5R,16R,18R)-1′ (23 mg, 0.081 mmol, 24%) as a mixture of E:Z isomers at C-14′ C-15 double bond and contaminated with minor amounts of the 6Z isomers. After separation in preparative HPLC with SunFireTM Prep C₁₈ OBDTM column (particle size 5 μ m, dimensions 19 mm ×100 mm, 60% MeCN/40% H₂O, 17 mL/min) the major isomer with E-configured double bonds (5R,16R,18R)-1 (6.2 mg, t_R 18.4 min) was obtained as a colorless oil. R_f : 0.41 (hexanes/AcOEt 7:3). [α]_D²⁰ + 10 (c 0.1, CHCl₃). The IR and NMR data of (5R, 16R,18R)-1 are identical to those described for (5S,16S,18S)-1. HRMS (ESI-TOF) m/z: [M + Na]⁺ Calcd for C₁₉H₂₄O₂Na 307.1674; Found 307.1663.

Ethyl (E)-6-hydroxyhex-2-enoate, 26

To a solution of **17** (1.54 g, 9.74 mmol) in anhyd DCM (40 mL) was added Et₃N (3.3 mL, 23.4 mmol), DMAP (120 mg, 1.15 mmol) and TBSCl (1.76 g, 11.7 mmol), respectively, under N_2 . The resulting mixture was stirred at rt for 4.5 h and the solvent was removed under reduced pressure. The residue was purified by flash chromatography (hexanes/AcOEt 95:5) affording ethyl (*E*)-6-((*tert*-butyldimethylsilyl)oxy)hex-2-enoate (2.54 g, 9.32 mmol, 96%) as a colorless oil. Spectral data (1 H NMR and 1 3C NMR) are in accordance with those reported in the literature. 45

To a solution of ethyl (*E*)-6-((*tert*-butyldimethylsilyl)oxy)hex-2-enoate (2.35 g, 8.63 mmol) in anhyd DCM (40 mL) was slowly added via cannula a solution of DIBALH (3.9 mL, 21.6 mmol) in DCM (10 mL), at -78 °C and under N₂. The reaction mixture was stirred under reduced temperature for 2 h. After that, were added Et₂O (100 mL) and saturated solution of Rochelle's salt (50 mL), maintaining vigorous stirring for 30 min at 0 °C and for 1 h at rt. After phase separation, the aqueous one was extracted with Et₂O (2 × 100 mL). The combined organic phases were dried over MgSO₄, filtered, and concentrated under reduced pressure. The crude product was purified by flash chromatography (hexanes/AcOEt 7:3) to afford **26** (1.95 g, 8.46 mmol, 98%) as a colorless oil. Spectral data (¹H NMR and ¹³C NMR) are in accordance with those reported in the literature. ⁴⁶

tert-Butyl(((4E,6E)-7-((2R)-6-isopropoxy-3,6-dihydro-2H-pyran-2-yl)hepta-4,6-dien-1-yl)oxy)dimethylsilane, (5R)-29

To a solution of **26** (1.00 g, 4.34 mmol) in anhyd MeCN (14 mL) was added 2,6-lutidine (1.0 mL, 8.9 mmol), Ph₃P (2.28 mg, 8.69 mmol) and CBr₄ (2.88 g, 8.68 mmol), respectively, at 0 °C. The resulting solution was stirred for 10 min and the reaction was quenched with H₂O (50 mL) and extracted with Et₂O (3 × 100 mL). The combined organic phases were washed with brine (50 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The crude product was purified by flash chromatography (gradient elution, 0 to 5% of AcOEt in hexanes) to give (E)-((6-bromohex-4-en-1-yl)oxy)(tert-butyl)dimethylsilane (**27**) (1.16 g, 3.97 mmol, 91%) as a colorless oil. R_f : 0.64 (hexanes/AcOEt 95:5). IR (ATR) v_{max}/cm^{-1} : 2929, 2857, 1472, 1255, 1104, 835, 775. ¹H NMR (250 MHz, CDCl₃) δ : 0.04 (s, 6H), 0.89 (s, 9H), 1.54–1.65 (m, 2H), 2.04–2.17 (m, 2H), 3.60 (t, J = 6.3 Hz, 2H), 3.94 (d, J = 6.6 Hz, 2H), 5.63–5.85 (m, 2H). ¹³C NMR (63 MHz, CDCl₃) δ : -5.3 (2 C), 18.3, 25.9 (3 C), 28.4, 31.9, 33.5, 62.3, 126.6, 136.1. We were unable to obtain HRMS data of **27** using ESI-TOF and APCI mass spectrometry techniques.

To a solution of **27** (912 mg, 3.12 mmol) in anhyd MeCN (17.2 mL) was added n-Bu₃P (1.15 mL, 4.66 mmol) and the mixture was stirred at rt for 12 h. After that, the excess of

solvent was removed under reduced pressure and the crude product was maintained under high vacuum (0.2 mmHg) for 8 h. The tri-*n*-butylphosphonium salt **28** was employed in the Wittig reaction without purification.

Wittig Olefination—To a solution of freshly prepared aldehyde (5R)-7 (354 mg, 2.08 mmol) and tri-n-butylphosphonium salt 28 (3.12 mmol) in anhyd PhMe (38 mL) was added dropwise a solution of NHMDS (4.2 mL, 4.2 mmol, 1 M in THF) at - 78 °C and under N₂. The mixture was stirred for 13 h at -78 °C and for 1 h at rt. The reaction was quenched with H_2O (50 mL) and extracted with AcOEt (100 mL and 2 × 50 mL). The combined organic phases were washed with brine (50 mL), dried over MgSO₄ and the solvent was removed under reduced pressure. The crude product was purified by flash chromatography (hexanes/ AcOEt 95:5) to furnish (5R)-29 (526 mg, 1.43 mmol, 69%) as a colorless oil and as a 6:1 mixture of the two isomers at C-6/C-7 double bond. R_f: 0.50 (hexanes/AcOEt 95:5). IR (ATR) v_{max}/cm^{-1} : 2955, 2929, 2893, 2857, 1255, 1181, 1100, 1028, 988, 835, 775. ¹H NMR (600 MHz, CDCl₃) δ: (major isomer) 0.04 (s, 6H, Si(CH₃)₂), 0.89 (s, 9H, C(CH₃)₃), H-11), 1.99–2.04 (m, 1H, H-4), 2.07–2.10 (m, 1H, H-4), 2.12–2.16 (m, 2H, H-10), 3.61 (t, J $= 6.4 \text{ Hz}, 1\text{H}, \text{H}-12), 4.01 \text{ (sept, } J = 6.2 \text{ Hz}, 1\text{H}, \text{CH(CH}_3)_2), 4.43-4.47 \text{ (m, 1H, H}-5), 5.11$ (s,br, 1H, H-1), 5.62 (dd, J = 15.4 and 6.3 Hz, 1H, H-6), 5.70–5.75 (m, 2H, H-9 and H-2), 5.99-6.01 (m, 1H, H-3), 6.05 (dd, J = 15.4 and 10.6 Hz, 1H, H-8), 6.23 (dd, J = 15.4 and 10.6 Hz, 1H, H-7). ¹³C NMR (150 MHz, CDCl₃) δ: (major isomer) – 5.3 (2 CH₃, Si(CH₃)₂), 18.3 (C, Si-C), 22.0 (CH₃, C(CH₃)₂), 23.9 (CH₃, C(CH₃)₂), 26.0 (3 CH₃, C(CH₃)₃), 28.9 (CH₂, C-10), 30.7 (CH₂, C-4), 32.3 (CH₂, C-11), 62.5 (CH₂, C-12), 66.4 (CH, C-5), 69.5 (CH, C(CH₃)₂), 93.1 (CH, C-1), 126.1 (CH, C-2), 128.5 (CH, C-3), 129.9 (CH, C-8), 130.7 (CH, C-6), 131.3 (CH, C-7), 135.0 (CH, C-9). HRMS (ESI-TOF) m/z: [M – C(CH₃)₂ + Na]⁺ Calcd for C₁₈H₃₂O₃SiNa 347.20129; Found 347.20134.

(4E,6E)-7-((2R)-6-Isopropoxy-3,6-dihydro-2H-pyran-2-yl)hepta-4,6-dien-1-ol, (5R)-30

To a solution of (5R)-29 (520 mg, 1.42 mmol) in anhyd THF (71 mL) was added a solution of TBAF (1.5 mL, 1.5 mmol, 1 M in THF) at 0 °C and under N₂. The yellow solution was allowed to warm to rt and stirred for 5.5 h. The reaction was quenched by the addition of H_2O (50 mL) and AcOEt (3 × 100 mL). The combined organic layers were washed with brine (50 mL), dried over MgSO₄, filtered and concentrated under reduced pressure. The crude product was purified by flash chromatography (hexanes/AcOEt 7:3) to give (5R)-30 (334.5 mg, 1.326 mmol, 94%) as a colorless oil and as a 6:1 mixture of the two isomers at C-6/C-7 double bond. R_f : 0.30 (hexanes/AcOEt 7:3). IR (ATR) v_{max}/cm^{-1} : 3409, 2971, 2930, 2887, 1657, 1181, 1099, 1027, 989. ¹H NMR (500 MHz, CDCl₃) δ: (major isomer) 1.17 (d, J = 6.2 Hz, 3H, C(CH₃)₂), 1.23 (d, J = 6.2 Hz, 3H, C(CH₃)₂), 1.64–1.70 (m, 2H, 2H, 2H)H-11), 1.98–2.04 (m, 1H, H-4), 2.06–2.13 (m, 1H, H-4), 2.16–2.20 (m, 2H, H-10), 3.66 (t, J = 6.4 Hz, 1H, H-12), 4.00 (sept, J = 6.2 Hz, 1H, CH(CH₃)₂), 4.43–4.47 (m, 1H, H-5), 5.10 (s,br, 1H, H-1), 5.63 (dd, J = 15.4 and 6.3 Hz, 1H, H-6), 5.69–5.75 (m, 2H, H-9 and H-2), 5.99-6.01 (m, 1H, H-3), 6.07 (dd, J = 14.8 and 10.5 Hz, 1H, H-8), 6.23 (dd, J = 15.4 and 10.5 Hz, 1H, H-7). ¹³C NMR (125 MHz, CDCl₃) δ: (major isomer) 22.0 (CH₃, C(<u>C</u>H₃)₂), 23.8 (CH₃, C(<u>C</u>H₃)₂), 28.9 (CH₂, C-10), 30.7 (CH₂, C-4), 32.1 (CH₂, C-11), 62.4 (CH₂, C-12), 66.4 (CH, C-5), 69.5 (CH, C(CH₃)₂), 93.1 (CH, C-1), 126.1 (CH, C-2), 128.4 (CH, C-3), 130.2 (CH, C-8), 131.0 (CH, C-6)*, 131.1 (CH, C-7)*, 134.4 (CH, C-9). *indicates that assignments may be interchanged. HRMS (ESI-TOF) m/z: $[M - CH_2(CH_3)_2 + H]^+$ Calcd for C₁₂H₁₇O₃ 209.11722; Found 209.11709.

(2R)-6-Isopropoxy-2-((1E,3E,7E)-8-((1S,2S)-2-methylcyclopropyl)octa-1,3,7-trien-1-yl)-3,6-dihydro-2H-pyran, (5R,14S,16S)-31

To a solution of oxalyl chloride (133 μ L, 1.54 mmol) in DCM (16 mL) was added DMSO (160 μ L, 2.19 mmol) at – 78 °C and under N₂. After 15 min, was added dropwise (5*R*)-**30** (240 mg, 0.951 mmol) in DCM (7 mL) and the mixture was stirred for a further 15 min. Et₃N (0.7 mL, 4.8 mmol) was added and after 5 min the mixture was allowed to warm to rt. The reaction was quenched by the addition of saturated NH₄Cl solution (20 mL). The layers were separated and the aqueous phase was extracted with Et₂O (3 × 50 mL). The organic layers were combined, washed with brine (20 mL), dried over MgSO₄, filtered and concentrated under reduced pressure. The crude product was obtained as a yellow oil, which was used in the next step without purification. $R_{\dot{F}}$: 0.67 (hexanes/AcOEt 7:3).

To a solution of crude aldehyde obtained in the previous step and sulfone (1S,2S)-22 (390 mg, 1.46 mmol) in anhyd DME (40 mL) was added dropwise a solution of potassium hexamethyldisilazide (1.9 mL, 1.9 mmol, 1 M in THF) at - 78 °C and under N₂. The dark brown resulting mixture was stirred for 30 min and the reaction was quenched with brine (100 mL). The mixture was extracted with AcOEt (3×100 mL) and the organic layer was washed with brine (50 mL), dried over MgSO₄, and concentrated under reduced pressure. The crude product was purified by flash chromatography (hexanes/AcOEt 9:1) to afford (5R, 14S,16S)-31 as 2.2:1 E:Z mixture at C-12 double bond (119.3 mg, 0.394 mmol, 41%), contaminated with minor amounts of the 6Z double bond isomers as a light yellow oil. Rf. 0.58 (hexanes/AcOEt 9:1). IR (ATR) $v_{\text{max}}/\text{cm}^{-1}$: 2969, 2924, 1659, 1400, 1380, 1316, 1181, 1099, 1027, 988. ¹H NMR (600 MHz, CDCl₃) δ: (major isomer) 0.39–0.41 (m, 1H), 0.46– $0.49 \text{ (m, 1H)}, 0.66 - 0.72 \text{ (m, 1H)}, 1.00 - 1.10 \text{ (m, 1H)}, 1.05 \text{ (d, } J = 6.0 \text{ Hz, 3H)}, 1.17 \text{ (d, } J = 0.049 \text{ (m, 1H)}, 0.66 - 0.72 \text{ (m, 1H)}, 1.00 - 1.10 \text{ (m, 1H)}, 1.05 \text{ (d, } J = 0.049 \text{ (m, 1H)}, 0.66 - 0.72 \text{ (m, 1H)}, 1.00 - 1.10 \text{ (m, 1H)}, 1.05 \text{ (d, } J = 0.049 \text{ (m, 1H)}, 1.04 - 0.049 \text{ (m, 1H)}, 1.00 - 0.049 \text$ 6.2 Hz, 3H), 1.23 (d, J = 6.2 Hz, 3H), 1.99–2.28 (m, 6H), 4.01 (sept, J = 6.2 Hz, 1H), 4.43– 4.47 (m, 1H), 5.01 (dd, J = 15.3 and 8.6 Hz, 1H), 5.11 (s,br, 1H), 5.44 (dt J = 15.3 and 6.7 and 6Hz, 1H), 5.62 (d, J = 15.3 and 6.2 Hz, 1H), 5.68 - 5.76 (m, 2H), 5.99 - 6.11 (m, 2H), 6.23 (dd, J = 15.3 and 10.2 Hz, 1H); (second largest isomer) 0.46–0.49 (m, 2H), 0.76–0.72 (m, 1H), 1.00-1.10 (m, 1H), 1.07 (d, J = 5.9 Hz, 3H), 1.17 (d, J = 6.1 Hz, 3H), 1.23 (d, J = 6.2 Hz, 3H), 1.99–2.28 (m, 6H), 4.01 (sept, J = 6.2 Hz, 1H), 4.43–4.47 (m, 1H), 4.79 (t, J = 10.3 Hz, 1H), 5.11 (s,br, 1H), 5.26 (dt, J = 10.3 and 7.4 Hz, 1H), 5.60–5.65 (m, 1H), 5.68–5.76 (m, 2H), 5.99–6.11 (m, 2H), 6.21–6.27 (m, 1H). ¹³C NMR (150 MHz, CDCl₃) δ: (major isomer) 15.1, 15.5, 18.5, 22.0, 22.4, 23.9, 30.7, 32.2, 32.8, 66.4, 69.5, 93.1, 126.1, 126.6, 128.5, 129.8, 130.8, 131.3, 134.1, 134.9; (second largest isomer) 15.1, 15.5, 18.6, 18.7, 22.0, 23.9, 27.3, 30.7, 32.8, 66.5, 69.5, 93.1, 126.1, 126.5, 128.5, 129.9, 130.8, 131.3, 134.2, 135.0. HRMS (ESI-TOF) m/z: $[M + Na]^+$ Calcd for $C_{20}H_{30}O_2Na$ 325.21380; Found 325.21354.

(R)-6-((1E,3E,7E)-8-((1S,2S)-2-Methylcyclopropyl)octa-1,3,7-trien-1-yl)-5,6-dihydro-2*H*-pyran-2-one, (5*R*,14*S*,16*S*)-2

A mixture of PCC (395 mg, 1.82 mmol) and CaCO₃ (730 mg, 7.28 mmol) in anhyd DCM (10 mL) was stirred at rt for 2 h. Then a solution of (5R,14S,16S)-31 (110 mg, 0.364 mmol) in DCM (10 mL) was added by cannula and the reaction mixture was stirred at rt for 2 h. After filtration over silica gel and solvent evaporation under reduced pressure, the crude product was purified by flash chromatography (hexanes/AcOEt 7:3) to afford (5R,14S, 16S)-2′ (43 mg, 0.166 mmol, 46%) as a mixture of E:Z isomers at C-12/C-13 double bond and contaminated with minor amounts of the 6Z isomers. This mixture was separated using preparative HPLC with SunFireTM Prep C₁₈ OBDTM column (particle size 5 μ m, dimensions 19 mm × 100 mm, 55% MeCN/45% H₂O, 17 mL/min) to yield the major isomer with E-configured double bonds (5R,14S,16S)-2 (14.5 mg, t_R 18.6 min) as a colorless oil. R_f : 0.64 (hexanes/AcOEt 7:3). [α]_D²⁰ + 95 (c 0.1, CHCl₃). IR (ATR) ν _{max}/cm⁻¹: 2996, 2925, 1720, 1382, 1244, 991, 961, 816. ¹H NMR (500 MHz, CDCl₃) δ : 0.38–0.42 (m, 1H, H-15), 0.46–0.49 (m, 1H, H-15), 0.66–0.71 (m, 1H, H-16), 0.99–1.05 (m, 1H, H-14), 1.04 (d, J = 6.2 Hz,

3H, H-17), 2.04–2.08 (m, 2H, H-11), 2.10–2.16 (m, 2H, H-10), 2.43–2.45 (m, 2H, H-4), 4.92–4.96 (m, 1H, H-5), 5.00 (dd, J = 15.2 and 8.6 Hz, 1H, H-13), 5.43 (dt, J = 15.3 and 6.9 Hz, 1H, H-12), 5.64 (dd, J = 15.3 and 6.6 Hz, 1H, H-6), 5.77 (dt, J = 15.2 and 6.6 Hz, 1H, H-9), 6.01–6.05 (m, 2H, H-2, H-8), 6.31 (dd, J = 15.4 and 10.5 Hz, 1H, H-7), 6.85–6.89 (m, 1H, H-3). ¹³C NMR (125 MHz, CDCl₃) δ : 14.7 (CH, C-16), 14.8 (CH₂, C-15), 18.5 (CH₃, C-17), 22.4 (CH, C-14), 29.8 (CH₂, C-4), 32.0 (CH₂, C-11), 32.8 (CH₂, C-10), 77.9 (CH, C-5), 121.7 (CH, C-2), 126.3 (CH, C-12), 126.5 (CH, C-6), 128.9 (CH, C-8), 133.7 (CH, C-7), 134.3 (CH, C-13), 137.1 (CH, C-9), 144.6 (CH, C-3), 164.0 (CO, C-1). HRMS (ESITOF) m/z: [M + Na]⁺ Calcd for C₁₇H₂₂O₂Na 281.15120; Found 281.15065.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

- 1. Gerwick WH, Moore BS. Chem Biol. 2012; 19:85. [PubMed: 22284357]
- 2. Lewy DS, Gauss CM, Soenen DR, Boger DL. Curr Med Chem. 2002; 9:2005. [PubMed: 12369868]
- 3. Stampwala SS, Bunge RH, Hurley TR, Willmer NE, Brankiewicz AJ, Steinman CE, Smitka TA, French JC. J Antibiot. 1983; 36:1601. [PubMed: 6689324]
- 4. Tunac JB, Graham BD, Dobson WE. J Antibiot. 1983; 36:1595. [PubMed: 6689323]
- 5. Kalesse M, Christmann M. Synthesis. 2002:981.
- 6. Kobayashi M, Higuchi K, Murakami N, Tajima H, Aoki S. Tetrahedron Lett. 1997; 38:2859.
- 7. Murakami N, Wang W, Aoki M, Tsutsui Y, Higuchi K, Aoki S, Kobayashi M. Tetrahedron Lett. 1997; 38:5533.
- 8. Maier ME. Nat Prod Rep. 2009; 26:1105. [PubMed: 19693411]
- 9. Nicolaou KC, Snyder SA. Angew Chem Int Ed. 2005; 44:1012.
- 10. Balunas MJ, Grosso MF, Villa FA, Engene N, McPhail KL, Tidgewell K, Pineda LM, Gerwick L, Spadafora G, Kyle DE, Gerwick WH. Org Lett. 2012; 14:3878. [PubMed: 22794317]
- 11. Gerwick WH, Proteau PJ, Nagle DG, Hamel E, Blokhin A, Slate DL. J Org Chem. 1994; 59:1243.
- Edwards DJ, Marquez BL, Nogle LM, McPhail K, Goeger DE, Roberts MA, Gerwick WH. Chem Biol. 2004; 11:817. [PubMed: 15217615]
- Green LC, Wagner DA, Glogowski J, Skipper PL, Wishnok JS, Tannenbaum SR. Anal Biochem. 1982; 126:131. [PubMed: 7181105]
- Barcelos RC, Pastre JC, Caixeta V, Vendramini-Costa DB, De Carvalho JE, Pilli RA. Bioorg Med Chem. 2012; 20:3635. [PubMed: 22537680]
- Bruder M, Vendramini-Costa DB, De Carvalho JE, Pilli RA. Bioorg Med Chem. 2013; 21:5107.
 [PubMed: 23876338]
- De Fátima A, Kohn LK, Antonio MA, De Carvalho JE, Pilli RA. Bioorg Med Chem. 2005;
 13:2927. [PubMed: 15781402]
- 17. De Fátima A, Kohn LK, De Carvalho JE, Pilli RA. Bioorg Med Chem. 2006; 14:622. [PubMed: 16202605]

18. Vendramini-Costa DB, De Castro IDB, Ruiz AL, Marquissolo C, Pilli RA, De Carvalho JE. Bioorg Med Chem. 2010; 18:6742. [PubMed: 20729093]

- 19. Boger DL, Ichikawa S, Zhong W. J Am Chem Soc. 2001; 123:4161. [PubMed: 11457179]
- 20. Crimmins MT, King BW. J Am Chem Soc. 1998; 120:9084.
- 21. Phillips DJ, Pillinger KS, Li W, Taylor AE, Graham AE. Chem Commun. 2006:2280.
- 22. Charette AB, Juteau H. J Am Chem Soc. 1994; 116:2651.
- 23. Although axial orientation of the isopropyl substituent can be assumed based on anomeric effect, we refrained from depict the stereochemistry at C-1 due to the lack of unambiguous evidence as the NOESY spectrum does show a weak correlation between H-1 and H-5.
- 24. Corrêa IR Jr, Pilli RA. Angew Chem Int Ed. 2003; 42:3017.
- 25. Charette AB, Lebel H. J Am Chem Soc. 1996; 118:10327.
- 26. Chiralpak® IA column (particle size 5 μm, dimensions 4.6 mm × 250 mm, 98% hexanes/2% *i*-PrOH, 1 mL/min). For chromatograms see supporting information.
- 27. Tamura R, Saegusa K, Kakihana M, Oda D. J Org Chem. 1988:53.
- 28. Aissa C. Eur J Org Chem. 2009:1831.
- 29. Liu P, Jacobsen EN. J Am Chem Soc. 2001; 123:10772. [PubMed: 11674024]
- 30. Smith AB, Brandt BM. Org Lett. 2001; 3:1685. [PubMed: 11405686]
- 31. The yield and diastereoisomeric ratio for preparation of (5*S*,16*S*,18*S*)-25 were not optimized because they were obtained before the studies shown in Table 1.
- 32. Li D, Zhao Y, Ye L, Chen C, Zhang J. Synthesis. 2010:3325.
- 33. SunFireTM Prep C18 OBDTM column (particle size 5 μ m, dimensions 19 mm \times 100 mm, 60% MeCN/40% H₂O, 17 mL/min).
- 34. Chiralpak® IA (particle size 5 μ m, dimensions 4.6 mm \times 250 mm, 90% MeCN/10% MeOH, 0.7 mL/min). For chromatograms see supporting information.
- 35. Beecham AF. Tetrahedron. 1972; 28:5543.
- 36. Nagle DG, Geralds RS, Yoo HD, Gerwick WH, Kim TS, Nambu M, White JD. Tetrahedron Lett. 1995; 36:1189.
- 37. Gu L, Wang B, Kulkarni A, Geders TW, Grindberg RV, Gerwick L, Hakansson K, Wipf P, Smith JL, Gerwick WH, Sherman DH. Nature. 2009; 459:731. [PubMed: 19494914]
- 38. SunFireTM Prep C18 OBDTM column (particle size 5 μ m, dimensions 19 mm \times 100 mm, 55% MeCN/45% H₂O, 17 mL/min).
- 39. Guivisdalsky PN, Bittman R. J Org Chem. 1989; 54:4637.
- 40. Huckins JR, De Vicente J, Rychnovsky SD. Org Lett. 2007; 9:4757. [PubMed: 17944478]
- 41. Stempel GH, Cross RP, Mariella RP. J Am Chem Soc. 1950; 72:2299.
- 42. Kamal A, Balakrishna M, Reddy PV, Faazil S. Tetrahedron: Asymmetry. 2010; 21:2517.
- 43. Hansen TV. Tetrahedron: Asymmetry. 2002; 13:547.
- 44. Fox RJ, Lalic G, Bergman RG. J Am Chem Soc. 2007; 129:14144. [PubMed: 17973391]
- 45. Kelly BD, Lambert TH. Org Lett. 2011; 13:740. [PubMed: 21247136]
- 46. Bahde RJ, Rychnovsky SD. Org Lett. 2008; 10:4017. [PubMed: 18700774]

Figure 1. Structures of marine cyanobacterial metabolites: coibacins A–D (1–4), curacin A (5) and jamaicamide A (6).

Figure 2. Specific optical rotation of the four isomers of coibacin A (1).

Scheme 1. Retrosynthetic plan for coibacin A (1) isomers.

Scheme 2. Preparation of aldehyde (5*S*)-7.

$$\begin{array}{c} \text{HO} \\ \text{OH} \\ \text{II} \\ \end{array} \\ \begin{array}{c} \text{Ph}_3 \text{PCHCO}_2 \text{Et} \\ \text{MnO}_2, \text{ DCM} \\ \hline \\ 75\% \\ \end{array} \\ \text{II} \\ \end{array} \\ \begin{array}{c} \text{HO} \\ \hline \\ 75\% \\ \end{array} \\ \begin{array}{c} \text{II} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \text{II} \\ \end{array} \\ \begin{array}{c} \text{Ph}_3 \text{PCHCO}_2 \text{Et} \\ \hline \\ 17 \\ \end{array} \\ \begin{array}{c} \text{II} \\ \end{array} \\ \begin{array}{c} \text{II} \\ \end{array} \\ \begin{array}{c} \text{II} \\ \end{array} \\ \begin{array}{c} \text{DIBALH} \\ \hline \\ 2) \text{ TBSCI, Imid.} \\ \hline \\ \text{HO} \\ \hline \\ \end{array} \\ \begin{array}{c} \text{Ph}_3 \text{P, CBr}_4 \\ \hline \\ 2,6 \text{-lutidine} \\ \hline \\ 98\% \\ \end{array} \\ \\ \text{PG = TBS)} \\ \\ \\ \text{Br} \\ \\ \end{array} \\ \begin{array}{c} \text{Ph}_3 \text{P, CBr}_4 \\ \hline \\ 2,6 \text{-lutidine} \\ \hline \\ 98\% \\ \end{array} \\ \\ \text{OTBS} \\ \begin{array}{c} \text{Ph}_3 \text{P, CBr}_4 \\ \hline \\ \text{OTBS} \\ \end{array} \\ \begin{array}{c} \text{Ph}_3 \text{P, MeCN} \\ \hline \\ \text{quantitative} \\ \end{array} \\ \begin{array}{c} \text{Ph}_3 \text{P, CBr}_4 \\ \hline \\ \text{OTBS} \\ \end{array} \\ \begin{array}{c} \text{OTBS} \\ \end{array} \\ \begin{array}{c}$$

Scheme 3. Synthesis of tri-*n*-butyl phosphonium salt **20**.

Me₂NOC CONMe₂

$$O_{B}O$$
Bu
$$OH$$

$$OH$$

$$(4R,5R)-13$$

$$Zn(CH2I)2·DME$$

$$OH$$

$$OH$$

$$(1S,2S)-9$$

$$er 94:6$$

Scheme 4. Synthesis of sulfone (1*S*,2*S*)-22.

Scheme 5. Synthesis of aldehyde (5*S*)-24.

Scheme 6. Synthesis of enantiomerically pure isomers (5*S*,16*S*,18*S*)- and (5*S*,16*R*,18*R*)-1

Scheme 7.
Retrosynthetic plan for correct isomer of coibacin B (2).

Scheme 8. Synthesis of tri-*n*-butyl phosphonium salt **28**.

Scheme 9. Synthesis of (5*R*,14*S*,16*S*)-**2** (coibacin B).

Table 1

Modified Julia olefination involving aldehyde (5S)-24 and racemic sulfone 22.

H	
H (ESA) H. A MEHADS (2 eq) - 78 - 7	

Entry ^a	Solvent	MHMDS	Time	Ratio E :Z at C-14 b	Yield
1	DMF	NHMDS	NHMDS 1.5 h (–78 °C), 1.5 h (rt)	1.7:1	4%c
2	THF	NHMDS	1.5 h (-78 °C), 1.5 h (rt)	1.7:1	28%c
ю	THF	NHMDS	10 min	1.7:1	65% de
4	DME	NHMDS	20 min	2.7:1	<i>p</i> %56
S	THF/HMPA 4:1	NHMDS	1.5 h	1.7:1	p%99
9	DME/HMPA 4:1	NHMDS	1.5 h	2:1	23%d
7	DME	LHMDS	10 min	1:10	54%d
∞	DME	KHMDS	10 min	2.4:1	p%56

 $^{^{\}it a}$ Scale: 0.06 mmol of (5*S*)-24; Volume of solvent: 2.5 mL.

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 $^{^{}b}E$:Z ratio of the double bond at C-14 of the isomers with E geometry on the C-6 double bond.

 $^{^{\}mathcal{C}}$ Addition order: sulfone, NHMDS and aldehyde.

 $d_{\mbox{Addition}}$ order: sulfone, aldehyde and NHMDS.

 $[^]e$ When the scale was duplicated the yield was 70%.