Characterizing Nitrogen Sites in Nitrogen-Doped Reduced Graphene Oxide: A Combined Solid-State ¹⁵N NMR, XPS and DFT Approach

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Abstract: Despite the potential applications in energy storage and conversion systems such as Lioxygen batteries and fuel cells, the nature and distribution of doped nitrogen sites in reduced graphene oxides are still not well understood. In this work, we report a combined approach of ¹⁵N solid-state nuclear magnetic resonance (NMR) spectroscopy alongside the predominantly used X-ray photoelectron spectroscopy (XPS) to characterize the nitrogen environments in reduced graphene oxides. Application of ¹H-¹⁵N low-power double quantum cross polarization under fast magic angle spinning with Carr-Purcell-Meiboom-Gill scheme shows selective detection of protonated sites with low-power radiofrequency irradiation. NMR shift calculations of a series of N-containing molecules and a graphene nanoflake model were performed to help interpret the experimental data. This work demonstrates a powerful approach to identify and quantify the different nitrogen environments in doped graphene materials and can also be widely applied to similar graphitic carbon-based materials with other dopants.

KEYWORDS Doped graphenes, NMR, XPS, ab initio, cross polarization

Introduction

Research on graphene-based materials possessing two-dimensional (2D) carbon structures has gained significant attention due to the versatile functionalities of these materials, originating from the flexibility of tuning pore size/distribution, surface area, and especially, modifying electronic structure in the basal plane by substitution of carbon by other heteroatoms such as boron, nitrogen, sulfur, phosphorus, and halogens.¹⁻³ There are strong demands for non-metallic catalysts for reactions of significant importance in energy storage and conversion such as hydrogen evolution reaction in water splitting, oxygen reduction reaction in fuel cells and metal-oxygen batteries, and oxygen evolution reactions in metal-oxygen batteries and water splitting.^{2,4-7} Current industrial applications heavily rely on the use of metal-based catalysts but many suffer from known drawbacks such as cost, toxicity, and loss of catalytic activity due to poisoning.² To tackle the aforementioned problems, the use of various doped-graphene/carbon-based materials has been evaluated in energy-related applications; for example, recent studies demonstrated that introducing heteroatom-dopants such as nitrogen in graphene facilitates oxygen reduction reaction (ORR) in fuel cells operated in aqueous alkaline media.^{8,9} Similarly, reduced graphene oxide (rGO) with nitrogen dopants is a promising electrode material with potential use in Li-air batteries.^{4,10,11} these batteries possessing a higher theoretical specific energy density than the conventional Li-ion batteries.^{12–14} As nitrogen-containing functional groups are expected to play an important role in the ORR process,¹⁵ fully characterizing the nature of those nitrogen sites is essential to further improve the oxygen electrocatalysis.

Many previous studies reported N-doped graphenes synthesized by reaction of graphene oxide, which is prepared by a modified Hummer's method, with several sources of nitrogen such as urea, ammonia, melamine or hydrazine.^{3,16,17} The doping level of nitrogen varies from 1-16 atomic % based on the synthetic method, precursors and reaction temperatures.^{3,18} Due to the low concentration of dopant nitrogen and lack of crystallinity, previous characterizations of rGOs, especially their nitrogen sites, reported in the literature have predominantly relied on X-ray Photoelectron Spectroscopy (XPS).^{19,20} Four nitrogen environments are commonly reported: pyridinic (398.1-399.3 eV), pyrrolic (399.8-401.2 eV), graphitic (401.1-402.7 eV), and N-oxide (pyridinic N at ~402.8 eV).^{3,17,21} Recent studies have suggested that among these N sites, the major catalytic activity likely stems from pyridinic N sites rather than from all of the others.²² Full characterization of the functional groups present in N-doped rGO is difficult due to the challenges associated with XPS fitting, often showing the lack of sufficient resolution and peak overlap.^{17,21} Thus, other quantitative spectroscopic methods are required to complement the XPS.

Solid-state NMR, being a local characterization technique, offers a promising route to identify and potentially quantify the functional groups. In particular, ¹⁵N NMR is particularly promising

since ¹⁵N is a spin-1/2 nucleus with a wide chemical shift range. However, its measurement suffers from its low natural abundance (0.4 vs. 99.6 % for ¹⁵N and ¹⁴N, respectively). Solid-state ¹⁵N NMR spectroscopy has been employed to characterize the nitrogen environments in graphitic carbonbased materials, revealing useful chemical shift information on diverse nitrogen sites; ¹H-¹⁵N cross polarization (CP) and ¹⁵N direct polarization (DP) magic angle spinning (MAS) NMR measurements of nitrogen–containing carbonaceous solids reported shifts for pyrrolic, graphitic, and pyridinic nitrogen at around -245, -180, and -80 ppm (with respect to CH₃NO₂ (*I*)), respectively.²³ Similarly, more recent ¹⁵N MAS NMR measurements of a N-containing pyrolized carbon material identified a combination of pyridinic, graphitic, and edge nitrogen features with the aid of a CP pulse sequence; however, poor resolution was observed due to the relatively slow sample spinning (MAS rate = 20 kHz) and moderate field (11.7 T).²⁴ In a different study, two distinct graphitic N shifts of -180 (N bonded to three carbons) and -223 ppm (N bonded to two carbons) in a melem unit have been reported alongside *ab initio* calculations of ¹⁵N and ¹³Cenriched graphitic carbon nitride (g-C₃N₄).²⁵ Another NMR study of hydrazine-treated graphite oxide reported that pyrazole formation at edges gives rise to a broad ¹⁵N resonance at -200 ppm.¹⁶

To enhance the site resolution, fast spinning (>40 kHz) would be desirable; the normal disadvantages of such fast MAS, however, are: (a) smaller available rotor sample volume and poor absolute signal sensitivity, and (b) an increase in the CP radiofrequency (rf) power delivered to the sample to satisfy the optimum Hartmann-Hahn (HH) match conditions,²⁶ which is likely to result in sample degradation due to stronger rf heating in these conductive graphenes.^{27,28} To some extent, the use of small rotors is often appropriate, given the small amount of sample available (typically <10 mg from one synthesis batch), and as the sensitivity per amount of material is improved for small rotors (due to the higher Q-factors); however, even with ¹⁵N enrichment of the dopant precursor, the dilute nature of ¹⁵N in these samples makes the acquisition time-consuming. Hence, we have employed Carr-Purcell-Meiboom-Gill (CPMG) detection of multiple echoes to drastically improve the sensitivity.²⁹ (b) was solved by employing a low-power *Double Quantum* (DQ) HH condition in the CP sequence.³⁰ In brief, this modifies the normal Zero Quantum HH condition of $\omega_H = \omega_C \pm n\omega_r$ (where $\omega_H, \omega_C, \omega_r$ are the proton rf, carbon rf, and MAS frequencies, respectively) to $\omega_H + \omega_C = n\omega_r$, thereby allowing polarization transfer under low-power irradiation and consequently less sample heating. As we will show, a combination of DP-CPMG and DQ-CP-CPMG sequences under fast MAS yields high resolution ¹⁵N spectra in N-rGO in moderate measurement time with site-specific observation, which would not otherwise be possible with simple echoes under slow MAS.

Ab initio calculations of NMR shifts are almost routinely performed for molecular systems,³¹ but only a few systematic studies on the ¹⁵N isotope are reported in the literature.^{32–35} Care should also be taken when comparing chemical shifts as ¹⁵N shifts are often referenced to different compounds (NH₃ (*l*), CH₃NO₂ (*l*), NH₄Cl (*s*), or NH₄NO₃ (*s*)).³⁶ Furthermore, as nitrogen in N-rGO is in a heterocyclic *sp*² environment, a different shielding-shift relationship to other *sp*³-nitrogen environments may be expected. Hence, we have performed a systematic *ab initio* study of ¹⁵N shifts in aromatic N-heterocycles to aid our interpretation of the N-rGO NMR spectra.

Experimental Methods

Synthesis and characterization of N-rGO. Aqueous graphene oxide (GO) solution was prepared by a modified Hummer's method, as described previously³⁷ and its concentration is nominally 2 mg/ml. For any further reduction process, the GO solution was sonicated for an hour prior to use. To synthesize nitrogen-doped reduced graphene oxide (N-rGO) powder, the obtained graphene oxide solution was first concentrated by heating and stirring it in a vial at ~100 °C, resulting in graphene oxide concentration of ~4 mg/ml. The concentrated GO solution was cooled down to room temperature and ¹⁵N-enriched urea (98 atom% ¹⁵N, 99 % purity, Sigma Aldrich) was added into the GO solution with 1 : 4 wt. ratio (GO : urea) and stirred for 10 mins. The mixture was hydrothermally reduced at 180 °C for 8 hours and then rinsed with deionized water and freeze dried in vacuo. The collected solid was subject to further pyrolysis in a tube furnace under Ar flow (10 cm³/min) at 600 °C for 2 hours, to obtain the final N-rGO powder. The N-rGO was kept and handled in an inert Ar atmosphere to avoid humidity. Pristine rGO was also prepared under similar conditions except adding urea for comparison.

Powder X-ray diffraction (PXRD) measurements with a Panalytical Empyrean diffractometer were performed in a reflection mode with Cu K α radiation, λ =1.5406 Å in a low background Si sample holder. Scanning electron microscopy (SEM) images were recorded with a Hitachi S-5500 in lens field emission electron microscope. Transmission electron microscopy (TEM) images were taken using FEI Philips Tecnai 20 at 200 keV. The rGO aqueous solution was drop cast on a TEM grid and then dried in air before transferring into the TEM chamber. Raman spectra (Horiba Scientific, Labram HR Evoltuion) were measured by focusing the laser onto the sample and a 633 nm laser (1.4 mW) was used with 20 s acquisition time and 10 co-added scans are used for each spectrum. XPS (Thermo Fisher Scientific, ESCALAB 250Xi) spectra were measured by an Al K α monochromated X-ray source at an energy level of 1486.68 eV. X-ray spot size was 200 µm and the XPS survey was measured three times at three different spots to confirm uniform distribution of dopants and to estimate errors in N content. For the high resolution scans, pass energy of 50 eV, step size of 0.1 eV, and dwell time of 50 ms were used. The obtained XPS data were analyzed (and deconvoluted) using CasaXPS software.

Solid-state NMR. All solid-state NMR spectra were acquired on a 16.4 T Bruker Avance III spectrometer using a 1.3 mm HX probehead. A rotor synchronized Hahn-echo pulse (for ¹H) and Carr-Purcell-Meiboom-Gill (CPMG) pulse sequence (for ¹³C and ¹⁵N) were used to acquire ¹H, ¹³C and ¹⁵N magic angle spinning (MAS) spectra with a spinning speed of 50 kHz, with recycle delays of 20, 40 and 5 s for ¹H, ¹³C and ¹⁵N, respectively, and radio frequency (RF) field strength of 150, 125 and 83 kHz, respectively. ¹H-¹⁵N cross polarization with CPMG detection at 50 kHz MAS was performed using a double-quantum cross polarization (DQCP) as detailed in a previous paper.³⁰ DQCP conditions were optimized on a ¹³C-¹⁵N-labelled glycine sample. Low-power ¹⁵N and ¹H rf amplitudes of 40 kHz and 10 kHz were employed. No ¹H decoupling was applied during the acquisition due to long acquisition time. ¹H, ¹³C, and ¹⁵N shifts were externally referenced to

solid adamantane at 1.87 and 38.6 (of CH group) and solid NH₄Cl at -338.2 ppm (with respect to CH₃NO₂ at 0 ppm), respectively. Despite of IUPAC recommends using CH₃NO₂ as a primary reference, using liquid NH₃ as an external reference is practically common so, for ¹⁵N, chemical shift values referenced by either CH₃NO₂ or NH₃ can be interconverted by: δ_{iso} (NH₃) = δ_{iso} (CH₃NO₂) + 380.5 ppm.³⁶ A summed echo was generated from the experimental CPMG spectrum using a Python script embedded in Bruker Topspin 3.5 and then Fourier transformed to generate a 1D spectrum. Spectral deconvolutions of the obtained 1D spectra were performed using Bruker Topspin 3.5.

First principles calculation. First principles calculations were employed to estimate the chemical shifts of various doped nitrogen moieties present in the (planar) graphitic structure. The following functional groups were considered: pyrrolic, pyridinic (neutral), pyridinium (+1 charged), edge, N-oxide (N⁺-O⁻), and graphitic. A graphene monolayer with a distribution of functional groups was used to simulate a more realistic condition of doping.

Gaussian16 was used for all calculations.³⁸ All structures were initially relaxed under a B3LYP/6-31G(d) level of theory, followed by a shift calculation using a Gauge Independent Atomic Orbital (GIAO) approach under a B3LYP/6-31+G(d,p) level of theory using a more extended basis set. To convert the calculated chemical shielding σ to chemical shift δ , a linear relationship of $\delta_{iso} = a \cdot \sigma_{iso} + b$ was used. Values of a and b were obtained from a linear regression of the shift-shielding data of 71 heterocyclic molecules (120 nitrogen environments) containing various types of nitrogen functional groups (see Table S1 in SI), where the shielding was evaluated using the same approach as above for consistency. The calculated shifts were also referenced to a primary reference of nitromethane (CH₃NO₂) at 0 ppm.³⁶

Results and Discussion

The N-rGO was synthesized by a two-step process reported previously³⁷ involving hydrothermal reaction of a mixture of urea and graphite oxide (GO), dispersed in aqueous solution and the subsequent thermal reduction of the resulting solid mixture at 600 °C in an argon flow. The structural integrity of the prepared N-rGO was characterized by powder X-ray diffraction (PXRD), Raman, scanning electron microscopy (SEM), and transmission electron microscopy (TEM). Figure 1a shows the XRD patterns of GO, N-rGO, and pristine graphite for comparison. The (002) reflection of GO and N-rGO corresponds to the broad reflections at 10.0 and 26.4° 20 respectively. The peak shift of the (002) reflection from 10° for GO to 26.4°, 20 for N-rGO indicates a significant decrease in the interlayer spacing from 8.84 Å to 3.37 Å. As previously reported, this change is due to removal of various oxygen functional groups (hydroxyl, epoxy, carbonyl, *etc.*) upon thermal reduction.³⁹ The broadening of the (002) reflection in graphite-like materials is commonly associated with the turbostratic nature, fluctuation in interlayer spacing, and/or variation in the local curvature of the graphene layers.⁴⁰ Raman spectra of both rGO and N-rGO (Figure 1b) show two dominant and characteristic peaks known as D and G bands at ~1330 and ~1590 cm⁻¹, respectively. Unlike graphenes, the Raman spectrum of the pristine graphite reference displays a

very narrow and intense G band along with very weak D and noticeably intense 2D bands. The G band originates from the Raman-active in-plane E_{2g} mode. The D band stems from the in-plane breathing modes (A_{1g}) of six-atom rings, which are only Raman-active in the presence of defects in the rings.⁴¹ Therefore, the intensity ratio of the D and G bands (I_D/I_G) is also a sensitive measure of disorder in graphitic structures. The slightly increased I_D/I_G ratio of 1.3 is obtained from N-rGO with respect to 1.2 from rGO (see also Figure S1), the increase being tentatively ascribed to the defects induced by nitrogen dopants in the N-rGO. The graphitic morphologies of doped rGO also have been confirmed by SEM and TEM. SEM images of pristine and doped rGO (Figure 1c and S2 in SI) show very similar, hierarchical macroporous and uniform structures. No segregation is observed from addition of urea (for doping), which was mostly removed by the thermal reduction process. A TEM image of doped rGO in Figure 1d exhibits lattice fringes of graphene layers.



Figure 1. (a) XRD patterns of N-rGO and GO, (b) Raman spectra of N-rGO and rGO, and (c-d) SEM and TEM images of N-rGO, respectively. In both the XRD and Raman data, measurements of pristine graphite are also shown for comparison.



Figure 2. (a) XPS survey spectrum of N-rGO and GO, and (b) high resolution N 1s XPS spectrum (black solid line). The experimental and a total fit are shown as black solid and red dashed lines; the peak areas are filled with various colors denoting individual fits to different N species.

XPS has been employed to confirm the extent of nitrogen doping into the graphitic structure and to assign the various nitrogen sites. XPS survey spectra of N-rGO and GO are shown in Figure 2a. The N-rGO survey exhibts a major C 1s peak with two minor O 1s and N 1s peaks, indicating the nitrogen dopant implanted in the structure; a doping level of ~7 atomic % nitrogen is estimated. A high resolution N 1s XPS spectrum (Figure 2b) shows two overlapped peaks with a broad tail at a higher binding energy region and its spectral deconvolution further reveals at least four nitrogen environments corresponding to pyridinic (398.3 eV with 33.6 %), pyrrolic (399.4 eV with 20.4 %), graphitic (400.1 eV with 27.4 %), and N-oxide (401.8 eV with 8.70 %) with $\pi - \pi^*$ satellite at 404.2 eV (9.96%), making use of assignments from previous XPS data of N-doped carbon structures.^{18,21} The relative composition of the various species differs however, with higher pyrrolic and pyridinic compositions alongside a lower graphitic nitrogen content being observed due to the lower reduction temperature applied (600 °C) compared to the temperature used to prepare the previously reported sample.²¹

A solid-state ¹³C NMR spectrum of pristine rGO and N-rGO are shown in Figure S3. The two spectra are only marginally different, with the N-rGO showing a slightly higher peak intensity around the 150 ppm region (where *sp*² C-N bonded carbons are expected to resonate); but in general the resolution is insufficient to distinguish between different carbon environments. Therefore, a solid-state ¹⁵N CPMG MAS NMR measurement of N-rGO has been used to directly observe various nitrogen environments in the graphitic structure. The ¹⁵N direct polarization (DP) CPMG spectrum (Figure 3a) contains two overlapping peaks with small shoulders. The lineshape is much broader than the reported ¹⁵N spectrum of graphitic carbon nitride (g-C₃N₄), this being ascribed to the distribution of nitrogen dopants and conductive nature of graphene. The ¹⁵N spectrum is deconvoluted into four peaks at -70, -116, -183, and -227 ppm, which we assign as pyridinic, N-oxide, graphitic, and pyrrolic/pyridinium nitrogen sites, respectively (*vide infra*).



Figure 3. Solid-state (a) ¹⁵N DP-CPMG and (b) ¹H-¹⁵N CP-CPMG MAS NMR spectra of N-rGO, acquired at a 16.4 T with a MAS frequency of 50 kHz. Experimental and total fits are shown as black solid and red dashed lines, respectively; peak areas are filled with different colors to identify and assign the individual peaks. The range of shifts obtained from singly-doped nanoflakes (model A) are shown as colored bars, with black line denoting the shifts obtained from the multiple-doped nanoflake (model B; also shown as an inset on the right hand side). In the model structure, gray, white, and red balls denote carbon, hydrogen, and oxygen atoms, respectively. CP contact times are shown in the figure.

To aid the identification of protonated species through spectral editing, a high resolution $^{1}H^{-15}N$ CP CPMG NMR measurents under fast MAS (50 kHz) of N-rGO was also performed (Figure 3b). With a short CP contact time of 200 µs, the signal at -227 ppm is selectively excited, indicating that this site is directly bonded to a proton. Thus, this peak must be assigned to either pyrrolic (N-H) or pyridinium (N⁺-H) environments, consistent with the previous report on N-containing pyrolized carbon.²⁴ The calculated proton shieldings of pyrrolic and pyridinium species in the nanoflake model (*vide infra*) are similar (20.8 and 21.8 ppm, respectively). With a longer CP contact time of 4000 µs, other N sites more distant from the protons also start to build up in intensity, resembling the DP CPMG spectrum. Of note, the pyridinic and N-oxide overlapping resonance is now seen (-50 to -150 ppm), consistent with the location of these environments on the edge of the graphene flakes, close to the terminating C-H species. The graphitic N environment (-190 ppm) is not observed on this CP spectrum, consistent with its location in the center of the flakes.

Despite the spectral editing with CP technique, the overlapping high-frequency signals are still not resolved clearly. Hence, ab initio NMR chemical shift calculations were performed on Ncontaining model systems to aid the assignment of different nitrogen species. The investigated models include (i) 71 N-containing small heterocyclic molecules and (ii) graphene flakes doped with various nitrogen functional groups. Figure 4 shows the relationship of the ¹⁵N chemical shielding (σ_{iso}) in N-heterocycles *versus* their ¹⁵N isotropic chemical shifts (δ_{iso}), together with approximate shift ranges for each functional group. A linear relation is found between σ_{iso} and δ_{iso} , with a functional form of $\delta_{iso} = -0.925\sigma_{iso} - 125.05$ relative to liquid nitromethane (CH₃NO₂). The shifts can be grouped into four regions according to the functional groups: (i) diazine N=N (50 to -50 ppm), (ii) pyridinic N, N-oxide, and pyrryl N (-50 to -150 ppm), (iii) graphitic/tertiary N (-150 to -200 ppm), and (iv) pyrrolic N (-200 to -300 ppm). Considering the different shift ranges, it is observed that (i) diazine N=N group is highly deshielded, possibly due to the more electronegative nature of the nitrogen relative to carbon; (ii) pyridinic, pyrryl, and N-oxides share similar ranges of shifts, although N-oxides have narrower, less shielded shift range (-50 to -100 ppm; due to oxygen coordination) compared to the other the two (-50 to -150 ppm); and (iii) graphitic and pyrrolic shifts are well separated from the others, so these should be easily identified in a spectrum.



Figure 4. (a) Calculated ¹⁵N isotropic chemical shielding (σ_{iso}) *versus* experimental isotropic chemical shift (δ_{iso}) in a series of small N-heterocyclic molecules. The linear regression shows the trend between δ_{iso} and σ_{iso} . (b) Calculated ¹⁵N chemical shift ranges for the collection of heterocyclic molecules presented in (a), referenced to NH₃ (*l*) and CH₃NO₂ (*l*).

The calculated range of shifts is in agreement with previous reports by Solum *et al.* on nitrogen heterocycles, where the shifts were referenced by simply subtracting the σ_{iso} from the nitromethane reference, effectively setting a = -1 in the regression as expected from the shielding-shift relationship.³⁴ However, the value of a = -0.925 clearly deviates from -1, which is generally ascribed to deficiencies in the functionals used in the DFT calculations (the so-called 'self-interaction error');⁴² the added degree of freedom by performing a two-variable linear regression (slope and intercept) is expected to yield a more accurate shielding-shift relationship. A more recent study by Xin *et al.* on ¹⁵N shifts of general nitrogen functional groups found the relationship $\delta_{iso} = -0.978\sigma_{iso} - 126.77$, which is close to the current result.³⁵ However, it needs to be stressed that a more extensive cc-pDVZ basis set was used in their study, compared to our 6-31+G(d,p); good agreement between the two regression data shows that a dual basis scheme with cheaper 6-31G(d)/6-31+G(d,p) basis sets could be used to calculate the shielding, which becomes important for evaluating the shielding in the larger graphene nanoflake models used here (*vide infra*).

To simulate more realistic doped nitrogen environments, two classes of N-doped graphite models were then constructed based on an undoped graphene nanoflake, composed of 77 carbon atoms: (i) 22 model structures simulating single-nitrogen doping by five different functional groups (N-oxide, graphitic, pyridine, pyridinium, and pyrrolic), with each nitrogen placed in different positions relative to the flake edge, denoted model A and (ii) one structure of multiple-nitrogen doping (one of each functional group) to simulate a more realistic scenario at 6.5 atomic %, denoted model B. The range of shifts obtained from singly-doped model A are shown below the experimental spectra in Figure 3a, alongside the values of shifts obtained from the multiple-doped graphene flake of model B (shown on inset). The calculated ranges of the shifts correspond well to the experimental spectrum, shown with the same color scheme on both the deconvoluted peaks and the model structure of graphene nanoflake.

These DFT calculations, therefore, further corroborate the assignment of -227 ppm peak to pyrrolic/pyridinium species from the CP spectra; furthermore, the partially overlapping resonances at -70, -116, and -183 ppm are now unambiguously assigned to pyridinic, N-oxide, and graphitic species, respectively. Our assignments are also in good agreement with the previous NMR study by MacIntosh *et al.*, although the resolution and sensitivity of their ¹⁵N spectrum is poorer due to the slower MAS rate used (15 kHz *versus* 50 kHz used in this work).²⁴ We also note that the nature of our sample, a reduced graphene oxide prepared with nitrogen dopants, is significantly different to their work, in which a N-containing carbon was prepared from pyrolyzing a polymerized aniline precursor. The peak at -183 ppm is also in a good agreement with the shift from the reported graphitic carbon nitride²⁵ and it is assigned to graphitic N. Of note, the intensity of the shift corresponding to graphitic N (a blue peak in Figure 3) is noticeably lower than what expected from the XPS data. This could arise from a very different spin-lattice relaxation behavior of graphitic N (the reported T₁ is >1000 s and thus the signal being saturated by the use of a shorter recycle delay of 5 s).²⁵ A distribution of graphitic nitrogen resonances over a broad range is not expected, as

evidenced from the narrow range of graphitic ¹⁵N shifts in a series of graphite nanoflake models doped with graphitic nitrogen placed at different positions (model A; Figure 3a, blue bar).

Conclusion

Our work demonstrated that various nitrogen sites in nitrogen-doped reduced graphene oxide can be identified and assigned by a combined approach of XPS, NMR, and *ab initio* calculations. Along with ¹⁵N enrichment, ¹⁵N CPMG and CP pulse sequences have been employed for NMR signal enhancement to determine the nature of doping in reduced graphene oxide. Fast sample spinning, combined with CP under fast MAS conditions, enables high resolution spectra to be acquired to allow clear assignments of various nitrogen sites. At least four nitrogen sites are characterized and a good agreement between our experimental and computational data is obtained. This study establishes a baseline for characterizing various nitrogen environments in graphenes and heterocyclics in general and can also be extended to study similar graphitic carbon-based materials with other dopants.

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Notes

The authors declare no competing financial interests.

Supporting Information

Supporting information including Raman and SEM images, and ¹³C CPMG NMR spectra, and library of calculated ¹⁵N chemical shielding versus chemical shifts is available.

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