*Highlights (for review)

Highlights

- > Genotyping for four polymorphic sites in the VDR gene in patients with Parkinson's disease.
- Association was detected between FokI C polymorphism and Parkinson's disease.
- ➤ No association could be detected between ApaI, BsmI, TaqI polymorphisms and Parkinson's disease.

Association of vitamin D receptor gene polymorphisms and Parkinson's

disease in Hungarians

Rita Torok^a, Nora Torok^a, Levente Szalardy^a, Imola Plangar^a, Zoltan Szolnoki^b, Ferenc

Somogyvari^c, Laszlo Vecsei^{a,d}, Peter Klivenyi^a

^a Department of Neurology, University of Szeged, H-6725 Szeged, Semmelweis u. 6, Hungary

^b Department of Neurology and Cerebrovascular Diseases, Pándy Kálmán County Hospital,

H-5700 Gyula, Semmelweis u. 1., Hungary

^c Department of Medical Microbiology and Immunology, Faculty of Medicine, University of

Szeged, H-6725 Szeged, Dóm tér 10., Hungary

^d Neuroscience Research Group of the Hungarian Academy of Sciences and University of

Szeged, H-6725 Szeged, Semmelweis u. 6, Hungary

Corresponding author:

Peter Klivenyi, MD, PhD

Department of Neurology, University of Szeged, H-6725 Szeged, Semmelweis u. 6, Hungary

Phone: +36 62 545348; Fax: +36 62 545597

E-mail: klivenyi.peter@med.u-szeged.hu

1

Running Title:

Vitamin D receptor polymorphisms in Parkinson's disease

Abstract

Vitamin D receptor (VDR) gene encodes a transcription factor that influences calcium

homeostasis and immunoregulation, and may play a role in neurological disorders including

Parkinson's disease (PD). The investigations of the association between VDR and PD in

different populations revealed various results. In a present study 100 PD patients and 109

healthy controls from the Hungarian population were genotyped for four polymorphic sites

(BsmI, ApaI, FokI and TaqI) in the VDR gene. The polymorphisms were determined by

polymerase chain reaction and restriction fragment length polymorphism (PCR-RFLP). Our

results demonstrate an association between the FokI C allele and PD; the frequency of the C

allele was significantly higher in PD patients than in controls, suggesting that this

polymorphism may have a role in the development of PD in these patients.

Keywords: FokI, Parkinson's disease, polymorphism, vitamin D receptor

3

1. Introduction

Parkinson's disease (PD) is one of the most common neurodegenerative disorders. The characteristic neuropathological features are the presence of Lewy bodies and the loss of dopaminergic cells in the substantia nigra pars compacta, but the precise pathomechanism is still not fully understood. The most common concepts are related to the genetic background and environmental effects [5]. Indeed, a number of genetic risk factors and gene-environment interactions have been implicated in the pathogenesis of PD [13, 14, 19-21, 24, 25, 29, 37]. Vitamin D, as an environmental factor, has been the subject of various studies on different neurological disorders, from which it has emerged that a vitamin D deficiency is associated with an increased risk of many diseases, including schizophrenia, autism, multiple sclerosis, Alzheimer's disease and PD [1, 3, 4, 7, 9, 22, 26-28, 30, 33, 35].

In humans, the majority of vitamin D is synthesized via the cleavage of a cholesterol metabolite in the epidermis by UVB, with further metabolism to the primary circulating form of vitamin D, 25-hydroxyvitamin D (250HD), in the liver. This compound circulates in the blood in a form bound to vitamin D binding protein and in the kidneys 250HD is metabolized by 1- α -hydroxylase to its active form, 1,25-dihydroxyvitamin D (1,250HD). 1,250HD binds to vitamin D receptors (VDRs) and influences calcium homeostasis, neurotrophic signalling, immunoregulation, cell growth and differentiation [8, 16, 17]. Both 1- α -hydroxylase and VDRs are expressed in numerous body tissues, including the brain [10].

The VDR gene encodes a nuclear transcription factor. The human gene is localized to 12q12 and various polymorphisms have been reported in it [38]. The level of vitamin D in PD has been analysed in number of studies. A Japanese population exhibited a higher incidence of hip fractures and lower serum levels of 25OHD in PD patients as compared with the healthy controls [34, 35], observations that were confirmed in a Caucasian population [9]. The

serum 250HD level correlated negatively with the severity of PD, measured in terms of the Hoehn &Yahr stage and the Unified Parkinson's Disease Rating Scale (UPDRS) [34-36]. Whereas Sato et al. [34] reported a negative correlation between the 1,250HD level and the UPDRS score, Suzuki et al. [36] could not confirm this finding.

However, only limited data are available regarding the association between VDR polymorphisms and PD. Suzuki et al. [36] found that FokI CC genotype was associated with milder forms of PD in a Japanese population. Furthermore, an association between the BsmI bb genotype and PD was demonstrated in a Korean population [18]. A recent genome-wide association study revealed the association of VDR polymorphisms with the risk of PD and the age at onset in a Caucasian population [6]. In a Chinese study it was described that FokI C allele associates with an increased risk of PD as well as early-onset PD [12].

The study we report here related to whether the known VDR gene polymorphisms ApaI, FokI, TaqI and BsmI are associated with PD in the Hungarian population, which belongs to the Caucasian race.

2. Material and methods

2.1 Subjects

100 idiopathic PD patients (mean age: 66.4±9.3 yr; 44 men and 56 women) were enrolled. The patients were evaluated by movement disorders specialists, who confirmed the diagnosis of idiopathic PD. Secondary forms of parkinsonism were excluded. The age at onset was determined from the medical records and the cases were categorized as early-onset (diagnosed ≤60 yr) or late-onset (diagnosed >60 yr) PD (Table 1). The Park2 and LRRK2 mutations were not present.

The control group comprised 109 healthy individuals (mean age: 64.0±8.2 yr; 54 men and 55 women) who had no history of neurological or psychiatric disorders (Table 1). The patients and healthy controls, all of Hungarian origin, were selected from the Department of Neurology and Department of Medical Microbiology and Immunobiology at the University of Szeged. The study was approved by the Ethics Committee of the Faculty of Medicine, University of Szeged. All study participants gave their written informed consent, in accordance with the Declaration of Helsinki.

Table 1. Characteristics of PD patients and healthy controls

	PD group (%)	Control group (%)	
No.	100	109	
Age (mean±SEM)	66.4±9.3	64.0 ± 8.2	
Age at onset ≤ 60	52 (52)		
Age at onset > 60	48 (48)		
Gender			
Male	44 (44)	54 (49.5)	
Female	56 (56)	55 (50.5)	

2.2 DNA isolation

Genomic DNA was isolated from the peripheral blood by a standard desalting method [23], and stored at -20° C until further use.

The VDR polymorphisms were determined by polymerase chain reaction (PCR) techniques in a thermal cycler (Applied Biosystems 2720 Thermal Cycler, Applied Biosystems, Foster City, CA, USA) and restriction fragment-length polymorphism (RFLP).

2.3 Genotyping

For amplification of the FokI C/T polymorphism (rs10735810) the following primers were used [15]: forward primer 5'-AGC TGG CCC TGG CAC TGA CTC TGC TCT-3' and reverse primer 5'-ATG GAA ACA CCT TGC TTC TTC TCC CTC-3'. The PCR amplification was carried out with the following cycling parameters: 95°C for 5 min, and then 30 cycles of 95°C for 30 s, 60°C for 30 s, and 72°C for 30 s, and finally 72°C for 7 min. The PCR products were digested with the FokI restriction enzyme (Fermentas, Vilnius, Lithuania) at 55°C for 3 h. The digested products were separated by agarose gel electrophoresis. The genotypes were defined as CC (265 bp), TT (169 and 96 bp) or CT (265, 169 and 96 bp).

The BsmI A/G polymorphic site of intron 8 (rs1544410) was amplified with previously described primers [18]: forward primer 5'-CAA CCA AGA CTA CAA GTA CCG CGT CAG TGA-3' and reverse primer 5'-AAC CAG CGG GAA GAG GTC AAG GG-3'. The PCR conditions were as follows: 95°C for 10 min, 95°C for 30 s, 59°C for 30 s and 72°C for 50 s for 35 cycles, and finally 72°C for 10 min. The PCR products were digested with the restriction enzyme Mva12691 (Fermentas, Vilnius, Lithuania) at 37°C overnight. Fragments were separated by electrophoresis in 2% stained agarose gels and visualized in UV light. The genotypes were defined as AA (825 bp), GG (650 and 175 bp) or AG (825, 650 and 175 bp).

PCR amplification of the polymorphic TaqI T/C site (rs731236) was performed with the following primers [31]: forward 5'-CAG AGC ATG GAC AGG GAG CAA-3' and reverse 5'-CAC TTC GAG CAC AAG GGG CGT TAG C-3'. The PCR conditions were identical to those for the BsmI polymorphism. The PCR products were digested with the TaqI restriction enzyme (Fermentas, Vilnius, Lithuania) at 65°C for 3 h and fragments were analysed by electrophoresis in 2% agarose gel. The absence of the TaqI restriction site on both alleles (TT) led to the 501 bp fragment, whereas the presence of the restriction site on both alleles (CC) yielded bands of 295 and 206 bp. The presence of the 501, 295 and 206 bp fragments reflected the TC heterozygotes.

The ApaI G/T polymorphic site (rs7976091) was amplified with previously described primers [32]: forward primer 5'-CAA CCA AGA CTA CAA GTA CCG CGT CAG TGA-3' and reverse primer 5'-CAC TTC GAG CAC AAG GGG CGT TAG C-3' The PCR conditions were: 95°C for 10 min, followed by 35 cycles of 95°C for 30 s, 59°C for 30 s and 72°C for 2 min, and finally 72°C for 7 min. The PCR products were incubated with the ApaI restriction enzyme (Fermentas, Vilnius, Lithuania) at 37°C overnight. The genotypes were defined as TT (absence of restriction site, one band at 2000 bp), TG (heterozygote, three bands at 2000, 1700 and 300 bp) and GG (presence of the restriction site, two bands at 1700 and 300 bp).

2.4 Statistical analysis

All statistical analyses were performed with the SPSS Statistics 17.0 software (SPSS Inc., Chicago, IL, USA). The difference in genotype frequencies was analysed by using the Fisher's exact test or the χ^2 test. The associations between the genotypes and the PD were estimated via the odds ratio (OR), with a 95% confidence interval (CI) (95% CI). A p value of less than 0.05 was considered statistically significant. The observed FokI, BsmI, ApaI and

TaqI genotype frequencies were in accordance with the Hardy–Weinberg equilibrium in both the patients and the controls.

3. Results

3.1 VDR FokI polymorphism

The distributions of FokI restriction site genotypes in the PD patients and the controls are shown in Table 2. There was a significant difference in genotypes between the PD patients and the healthy controls ($\chi^2 = 6.7$; p = 0.035). The frequency of genotype with C (CC+CT) was significantly higher among the patients with PD relative to the controls: OR = 2.677 and 95% CI = 1.214-5.91, p = 0.015 for CC+CT vs. TT. Moreover, the C allele showed a significant association with PD group (OR = 1.615, 95% CI = 1.087-2.399, p = 0.017). There was no difference between the FokI polymorphism and gender in PD group, and no significant association was found between this polymorphism and the age at onset (Table 3).

Table 2. VDR FokI genotypes and allele frequencies in the PD patients and the controls

Genotype					Allele frequency		
	CC (%)	CT (%)	TT (%)	p	C (%)	T (%)	p
PD patients	42 (42)	48 (48)	10 (10)	0.025	132 (66)	68 (34)	0.017
Control	35 (32.1)	49 (45)	25(22.9)	0.035	119 (54.6)	99 (45.4)	0.017

Table 3.

Relationship between age at onset and gender in the PD patients as_a function of the VDR FokI genotyping

	CC (%)	CT (%)	TT (%)	p	
Age at onset					
≤60 yr	22 (42.3)	24 (46.2)	6 (11.5)	0.041	
>60 yr	20 (41.7)	24 (50)	4 (8.3)	0.841	
Gender					
Male	18 (40.9)	22 (50)	4 (9.1)	0.958	
Female	24 (42.9)	26 (46.4)	6 (10.7)	0.938	

3.2 VDR BsmI polymorphism

There was no significant difference in the BsmI genotypic distribution (OR = 0.890, 95% CI = 0.478-1.654, p = 0.753 for GG vs. AA+AG) and allele frequency (OR = 0.977, 95% CI = 0.665-1.434, p = 0.905) between the PD patients and the healthy controls. The BsmI genotypic distribution, the allele frequency, the male to female ratio and the age at onset of the PD patients are presented in Table 4.

Table 4. VDR BsmI genotypes and allele distributions in patients with PD and controls

Genotype				Allele f	requency		
	AA (%)	AG (%)	GG (%)	p	A (%)	G (%)	p
PD patients	24 (24)	49 (49)	27 (27)	0.902	97 (48.5)	103 (51.5)	0.905
Controls	25 (22.9)	57 (52.3)	27 (24.8)	0.902	107 (49)	111 (51)	0.905
Age at onset							
≤60 yr	9 (17.3)	27 (51.9)	16 (30.8)	0.261			
>60 yr	15 (31.3)	22 (45.8)	11 (22.9)	0.201			
Gender							
Male	7 (15.9)	22 (50)	15 (34.1)	0.161			
Female	17 (30.4)	27 (48.2)	12 (21.4)	0.161			

3.3 VDR TaqI polymorphism

The frequencies of the TaqI genotype in the PD group and the controls were similar (OR = 0.840, 95% CI = 0.399-1.767, p = 0.646 for TT+TC vs. CC) and we did not find differences in allele distribution (OR = 0.802, 95% CI = 0.540-1.190, p = 0.273). There was no significant difference in the male to female ratio and the age at onset in the various TaqI polymorphism subgroups (Table 5).

Table 5. VDR TaqI genotypes and allele frequencies in patients with PD and controls

	Genotype			All	ele frequency			
	TT (%)	TC (%)	CC (%)	p	T (%)	C (%)	p	
PD patients	35 (35)	48 (48)	17 (17)	0.405	0.485	118 (59)	82 (41)	0.273
Controls	47 (43.1)	46 (42.2)	16 (14.7)	0.463	140 (64.2)	78 (35.8)	0.273	
Age at onset								
≤60 yr	22 (42.3)	23 (44.2)	7 (13.5)	0.265				
>60 yr	13 (27.1)	25 (52.1)	10 (20.8)	0.203				
Gender								
Male	18 (40.9)	20 (45.5)	6 (13.6)	0.528				
Female	17 (30.4)	28 (50)	11 (19.6)	0.328				

3.4 VDR ApaI polymorphism

The ApaI genotype frequencies (OR = 1.352, 95% CI = 0.654-2.796, p = 0.466 for GG vs. TT+TG) and the allele distribution (OR = 1.177, 95% CI = 0.793-1.748, p = 0.417) were similar in the healthy controls and the patients with PD (Table 6). There was no statistically significant association between the ApaI polymorphism and the age at onset in PD patients, and no significant difference was found between this polymorphism and gender in the PD group.

Table 6. VDR ApaI genotypes and allele frequencies in the PD patients and the controls

Genotype					All	ele frequency	
	TT (%)	TG (%)	GG (%)	p	T (%)	G (%)	p
PD patients Controls	42 (42) 42 (38.5)	43 (43) 46 (42.2)	15 (15) 21 (19.3)	0.691	127 (63.5) 130 (59.6)	73 (36.5) 88 (40.4)	0.417
Age at onset ≤60 yr >60 yr Gender	18 (34.6) 24 (50)	23 (44.2) 20 (41.7)	11 (21.2) 4 (8.3)	0.130			
Male Female	13 (29.5) 29 (51.8)	22 (50) 21 (37.5)	9 (20.5) 6 (10.7)	0.07			

4. Discussion

This work involved a study of VDR polymorphism in PD patients among Hungarians. Earlier reports revealed that 25OHD levels were decreased in PD patients in Japanese and Caucasian populations [9, 34, 35] and demonstrated a higher incidence of osteoporosis in PD patients of both genders, with a decreased bone mass index and a reduced bone mineral density [11, 34]. Overall, it has been clearly shown that the vitamin D metabolism is affected in PD patients.

Differences have also been demonstrated in the VDR polymorphisms in PD in Japanese, Korean, Chinese and Caucasian populations [6, 12, 18, 36].

Our results have indicated a significant difference in the FokI genotype distribution between PD and controls in Hungarian population; the frequency of the C allele was significantly higher in PD patients than in the healthy control group, suggesting that the C allele may have a role in the development of PD.

Previously, a Japanese and a Chinese study detected difference in this polymorphism between healthy subjects and PD patients. In Japan, FokI CC genotype was associated with milder forms of PD [36]. Han et al. [12] suggested that FokI C allele might be a risk factor for sporadic PD development.

FokI polymorphism is located in exon 2 at the 5' coding region of the gene. This polymorphism results in different translation initiation sites: if the VDR gene contains C allele, the protein will be three amino acids shorter. Difference in length may result in altered VDR function [2, 38].

BsmI, ApaI and TaqI polymorphisms are located in the 3'-end region of the VDR gene, which do not result in changes in the amino acid sequence of the VDR [38]. We did not identify significant associations with these VDR polymorphisms. A Korean study detected difference in the genotype frequency of BsmI polymorphism between healthy subjects and PD patients; the bb genotype in that study_was more common in Korean PD patients than in

controls [18]. No difference in BsmI polymorphism was identified in Japanese patients [36], where the FokI CC genotype displayed a strong association with the milder forms of PD.

Our data demonstrated no significant associations between VDR ApaI polymorphism and PD, whereas the ApaI polymorphism was associated with the early-onset form of PD in American Caucasians. Interestingly, there was no association between FokI polymorphism and PD patients among American Caucasians, reflecting differences in ethnicity among Caucasians [6].

These diverse data suggest that the Caucasian population is not homogeneous in this respect. We did not detect an association between the age at onset, the male-female ratio and the VDR polymorphisms in the PD group. The differences between the results of the various authors must be interpreted with regard to the facts that the study populations and sample sizes differed, with the additional possibility of certain ethnic variations.

As far as we are aware, this is the first report on the potential correlation between a VDR polymorphism and PD from a European country. We conclude overall that the C allele of the VDR FokI polymorphism may be associated with PD in a Caucasian population.

5. Acknowledgements

This work was supported by grants ETT 026-04 and TÁMOP-4.2.1/B-09/1/KONV-2010-0005. The project "TÁMOP-4.2.1/B-09/1/KONV-2010-0005 – Creating the Centre of Excellence at the University of Szeged" is supported by the European Union and co-financed by the European Regional Development Fund.

6. Conflict of interest

The authors declare there is no conflict of interest.

7. Author contributions

Conceived and designed the experiments: RT, PK. Performed the experiments: RT, NT.

Collected the samples: NT, FS, ZSZ, PK. Analyzed the data: LSZ, RT. Wrote the paper: RT,

LSZ, IP, PK. Study supervision or coordination: LV, PK.

8. References

- [1] L. Amezcua, R.H. Chung, D.V. Conti, A.M. Langer-Gould, Vitamin D levels in Hispanics with multiple sclerosis, J Neurol 259 (2012) 2565-2570.
- [2] H. Arai, K. Miyamoto, Y. Taketani, H. Yamamoto, Y. Iemori, K. Morita, T. Tonai, T. Nishisho, S. Mori, E. Takeda, A vitamin D receptor gene polymorphism in the translation initiation codon: effect on protein activity and relation to bone mineral density in Japanese women, J Bone Miner Res 12 (1997) 915-921.
- [3] A. Ascherio, K.L. Munger, K.C. Simon, Vitamin D and multiple sclerosis, Lancet Neurol 9 (2010) 599-612.
- [4] J.S. Buell, B. Dawson-Hughes, T.M. Scott, D.E. Weiner, G.E. Dallal, W.Q. Qui, P. Bergethon, I.H. Rosenberg, M.F. Folstein, S. Patz, R.A. Bhadelia, K.L. Tucker, 25-Hydroxyvitamin D, dementia, and cerebrovascular pathology in elders receiving home services, Neurology 74 (2010) 18-26.
- [5] L.F. Burbulla, R. Kruger, Converging environmental and genetic pathways in the pathogenesis of Parkinson's disease, J Neurol Sci 306 (2011) 1-8.
- [6] M.W. Butler, A. Burt, T.L. Edwards, S. Zuchner, W.K. Scott, E.R. Martin, J.M. Vance, L. Wang, Vitamin D receptor gene as a candidate gene for Parkinson disease, Ann Hum Genet 75 (2011) 201-210.
- [7] J.J. Cannell, Autism and vitamin D, Med Hypotheses 70 (2008) 750-759.
- [8] A.S. Dusso, A.J. Brown, E. Slatopolsky, Vitamin D, Am J Physiol Renal Physiol 289 (2005) F8-28.
- [9] M.L. Evatt, M.R. Delong, N. Khazai, A. Rosen, S. Triche, V. Tangpricha, Prevalence of vitamin d insufficiency in patients with Parkinson disease and Alzheimer disease, Arch Neurol 65 (2008) 1348-1352.
- [10] D.W. Eyles, S. Smith, R. Kinobe, M. Hewison, J.J. McGrath, Distribution of the vitamin D receptor and 1 alpha-hydroxylase in human brain, J Chem Neuroanat 29 (2005) 21-30.
- [11] H.A. Fink, M.A. Kuskowski, E.S. Orwoll, J.A. Cauley, K.E. Ensrud, Association between Parkinson's disease and low bone density and falls in older men: the osteoporotic fractures in men study, J Am Geriatr Soc 53 (2005) 1559-1564.
- [12] X. Han, L. Xue, Y. Li, B. Chen, A. Xie, Vitamin D receptor gene polymorphism and its association with Parkinson's disease in Chinese Han population, Neurosci Lett 525 (2012) 29-33.

- [13] D.B. Hancock, E.R. Martin, K. Fujiwara, M.A. Stacy, B.L. Scott, J.M. Stajich, R. Jewett, Y.J. Li, M.A. Hauser, J.M. Vance, W.K. Scott, NOS2A and the modulating effect of cigarette smoking in Parkinson's disease, Ann Neurol 60 (2006) 366-373.
- [14] D.B. Hancock, E.R. Martin, J.M. Vance, W.K. Scott, Nitric oxide synthase genes and their interactions with environmental factors in Parkinson's disease, Neurogenetics 9 (2008) 249-262.
- [15] S.S. Harris, T.R. Eccleshall, C. Gross, B. Dawson-Hughes, D. Feldman, The vitamin D receptor start codon polymorphism (FokI) and bone mineral density in premenopausal American black and white women, J Bone Miner Res 12 (1997) 1043-1048.
- [16] S. Kato, The function of vitamin D receptor in vitamin D action, J Biochem 127 (2000) 717-722.
- [17] J.P. Kesby, D.W. Eyles, T.H. Burne, J.J. McGrath, The effects of vitamin D on brain development and adult brain function, Mol Cell Endocrinol 347 (2011) 121-127.
- [18] J.S. Kim, Y.I. Kim, C. Song, I. Yoon, J.W. Park, Y.B. Choi, H.T. Kim, K.S. Lee, Association of vitamin D receptor gene polymorphism and Parkinson's disease in Koreans, J Korean Med Sci 20 (2005) 495-498.
- [19] Y.J. Li, S.A. Oliveira, P. Xu, E.R. Martin, J.E. Stenger, C.R. Scherzer, M.A. Hauser, W.K. Scott, G.W. Small, M.A. Nance, R.L. Watts, J.P. Hubble, W.C. Koller, R. Pahwa, M.B. Stern, B.C. Hiner, J. Jankovic, C.G. Goetz, F. Mastaglia, L.T. Middleton, A.D. Roses, A.M. Saunders, D.E. Schmechel, S.R. Gullans, J.L. Haines, J.R. Gilbert, J.M. Vance, M.A. Pericak-Vance, C. Hulette, K.A. Welsh-Bohmer, Glutathione S-transferase omega-1 modifies age-at-onset of Alzheimer disease and Parkinson disease, Hum Mol Genet 12 (2003) 3259-3267.
- [20] D.M. Maraganore, M. de Andrade, A. Elbaz, M.J. Farrer, J.P. Ioannidis, R. Kruger, W.A. Rocca, N.K. Schneider, T.G. Lesnick, S.J. Lincoln, M.M. Hulihan, J.O. Aasly, T. Ashizawa, M.C. Chartier-Harlin, H. Checkoway, C. Ferrarese, G. Hadjigeorgiou, N. Hattori, H. Kawakami, J.C. Lambert, T. Lynch, G.D. Mellick, S. Papapetropoulos, A. Parsian, A. Quattrone, O. Riess, E.K. Tan, C. Van Broeckhoven, Collaborative analysis of alpha-synuclein gene promoter variability and Parkinson disease, JAMA 296 (2006) 661-670.
- [21] C.C. McCulloch, D.M. Kay, S.A. Factor, A. Samii, J.G. Nutt, D.S. Higgins, A. Griffith, J.W. Roberts, B.C. Leis, J.S. Montimurro, C.P. Zabetian, H. Payami, Exploring gene-environment interactions in Parkinson's disease, Hum Genet 123 (2008) 257-265.
- [22] J. McGrath, Hypothesis: is low prenatal vitamin D a risk-modifying factor for schizophrenia?, Schizophr Res 40 (1999) 173-177.
- [23] S.A. Miller, D.D. Dykes, H.F. Polesky, A simple salting out procedure for extracting DNA from human nucleated cells, Nucleic Acids Res 16 (1988) 1215.
- [24] I. Mizuta, W. Satake, Y. Nakabayashi, C. Ito, S. Suzuki, Y. Momose, Y. Nagai, A. Oka, H. Inoko, J. Fukae, Y. Saito, M. Sawabe, S. Murayama, M. Yamamoto, N. Hattori, M. Murata, T. Toda, Multiple candidate gene analysis identifies alphasynuclein as a susceptibility gene for sporadic Parkinson's disease, Hum Mol Genet 15 (2006) 1151-1158.

- [25] I. Mizuta, T. Tsunoda, W. Satake, Y. Nakabayashi, M. Watanabe, A. Takeda, K. Hasegawa, K. Nakashima, M. Yamamoto, N. Hattori, M. Murata, T. Toda, Calbindin 1, fibroblast growth factor 20, and alpha-synuclein in sporadic Parkinson's disease, Hum Genet 124 (2008) 89-94.
- [26] E.M. Mowry, Vitamin D: evidence for its role as a prognostic factor in multiple sclerosis, J Neurol Sci 311 (2011) 19-22.
- [27] K.L. Munger, L.I. Levin, B.W. Hollis, N.S. Howard, A. Ascherio, Serum 25-hydroxyvitamin D levels and risk of multiple sclerosis, JAMA 296 (2006) 2832-2838.
- [28] H.L. Newmark, J. Newmark, Vitamin D and Parkinson's disease--a hypothesis, Mov Disord 22 (2007) 461-468.
- [29] S.A. Oliveira, Y.J. Li, M.A. Noureddine, S. Zuchner, X. Qin, M.A. Pericak-Vance, J.M. Vance, Identification of risk and age-at-onset genes on chromosome 1p in Parkinson disease, Am J Hum Genet 77 (2005) 252-264.
- [30] S.M. Orton, S.V. Ramagopalan, A.E. Para, M.R. Lincoln, L. Handunnetthi, M.J. Chao, J. Morahan, K.M. Morrison, A.D. Sadovnick, G.C. Ebers, Vitamin D metabolic pathway genes and risk of multiple sclerosis in Canadians, J Neurol Sci 305 (2011) 116-120.
- [31] B.L. Riggs, T.V. Nguyen, L.J. Melton, 3rd, N.A. Morrison, W.M. O'Fallon, P.J. Kelly, K.S. Egan, P.N. Sambrook, J.M. Muhs, J.A. Eisman, The contribution of vitamin D receptor gene alleles to the determination of bone mineral density in normal and osteoporotic women, J Bone Miner Res 10 (1995) 991-996.
- [32] J. Sainz, J.M. Van Tornout, M.L. Loro, J. Sayre, T.F. Roe, V. Gilsanz, Vitamin Dreceptor gene polymorphisms and bone density in prepubertal American girls of Mexican descent, N Engl J Med 337 (1997) 77-82.
- [33] Y. Sato, T. Asoh, K. Oizumi, High prevalence of vitamin D deficiency and reduced bone mass in elderly women with Alzheimer's disease, Bone 23 (1998) 555-557.
- [34] Y. Sato, Y. Honda, J. Iwamoto, T. Kanoko, K. Satoh, Abnormal bone and calcium metabolism in immobilized Parkinson's disease patients, Mov Disord 20 (2005) 1598-1603.
- [35] Y. Sato, M. Kikuyama, K. Oizumi, High prevalence of vitamin D deficiency and reduced bone mass in Parkinson's disease, Neurology 49 (1997) 1273-1278.
- [36] M. Suzuki, M. Yoshioka, M. Hashimoto, M. Murakami, K. Kawasaki, M. Noya, D. Takahashi, M. Urashima, 25-hydroxyvitamin D, vitamin D receptor gene polymorphisms, and severity of Parkinson's disease, Mov Disord 27 (2012) 264-271.
- [37] J.M. van der Walt, M.A. Noureddine, R. Kittappa, M.A. Hauser, W.K. Scott, R. McKay, F. Zhang, J.M. Stajich, K. Fujiwara, B.L. Scott, M.A. Pericak-Vance, J.M. Vance, E.R. Martin, Fibroblast growth factor 20 polymorphisms and haplotypes strongly influence risk of Parkinson disease, Am J Hum Genet 74 (2004) 1121-1127.
- [38] J.M. Zmuda, J.A. Cauley, R.E. Ferrell, Molecular epidemiology of vitamin D receptor gene variants, Epidemiol Rev 22 (2000) 203-217.