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Additional Information

1 **Understanding the relevance of in-mouth food processing. A review of *in vitro* techniques**

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13
14
15 **Abstract**

16 Oral processing of food is the first step in the eating process. Although the food undergoes a
17 number of changes during mastication that influence the subsequent steps, this stage has very
18 often been neglected in studies of digestion, bioavailability, flavor release, satiety potential,
19 glycaemic index determination, etc. The present review draws on different sources such as
20 nutrition, medicine, phoniatry and dentistry to explain some *in vitro* oral processing methods
21 and techniques that could be transferred to food technology studies to mimic *in vivo*
22 comminution, insalivation, and bolus formation, describing, as a necessary reference, the
23 respective *in vivo* physiological processes they attempt to imitate.

24 Developing a deeper understanding of all the aspects of in-mouth process will help food
25 technologists to give this crucial step the necessary attention its due importance and to
26 consider better ways to incorporate it into their studies.

27

28 **Introduction**

29 Food is a mixture of proteins, carbohydrates and lipids that interact physically and chemically
30 in an aqueous environment to create a food-specific native or processed structure. Differences
31 in the chemical composition of foods are therefore associated with differences in their
32 macrostructure and texture which affect various food characteristics, including resistance to
33 hydrolysis or to breakdown during oral food processing and simultaneous (oral) or subsequent
34 (gastric, intestinal digestion. In-mouth actions results from a dynamic process in which the
35 textural characteristics of food are continuously analyzed by the oral sensory systems (Pineau,
36 *et al.*, 2009). Chen (2009) reviewed the physiology as well as the rheological principles of food
37 texture and sensory perception, since food texture is the main factor that determines the
38 different processes for transforming food into a material that is ready to be swallowed.

39 In a pioneering work, Hutchings and Lillford (1988) stated that texture perception in the mouth
40 is a dynamic sensory monitor of changes made to a food. They proposed a groundbreaking
41 general model, defining the breakdown path of the food during oral processing through three
42 aspects or dimensions: the mechanical and rheological behavior of the food (degree of
43 structure), the oral experience via saliva participation (degree of lubrication), and the
44 sequences of oral processing (time). Involving the oral experience and time in texture studies
45 was a significant development which turned texture appreciation from a static process into a
46 dynamic one. Several years later Prinz and Lucas (1997) proposed the optimum swallow
47 model, in which swallowing was defined as the moment when the food bolus reaches a peak
48 cohesive force, driven by the interaction between the food particles (degree of structure) and
49 saliva (degree of lubrication). In this way the duality of separating thresholds for food particle
50 size and for particle lubrication is eliminated: swallowing is initiated when it is sensed that a
51 batch of food particles is binding together under viscous forces so as to form a bolus.

52 In plain words, digestion is the process of breaking food down into simpler substances that can
53 be absorbed by the body. Food digestion in humans depends on both the chemical and
54 physical characteristics of the food and on how it changes as it passes through the different
55 areas of the digestive tract. Within this framework, the relevance of oral processing up to the
56 instant of swallowing is evident.

57 Inside the mouth, food undergoes a number of changes. Some of them, such as comminution,
58 are not strictly speaking digestive processes but are undoubtedly necessary before these can
59 take place, and could be considered a “pre-digestive step”.

60 During in-mouth food processing, food is subjected to several major mechanical and chemical
61 modifications. The solid food is fractured by the teeth and diluted and broken down by saliva.
62 These joint actions induce its progressive comminution and adherence of the resulting smaller

63 particles through saliva impregnation, formed into a cohesive bolus and finally swallowed (Van
64 der Bilt, Mojet, Tekamp, & Abbink, 2010). It would appear that saliva is involved at every step,
65 not only as a digestive medium but as a lubricant, providing surface smoothness and weak
66 inter-particle adhesive forces (Lillford, 2011). Although mastication seems a simple process, it
67 involves many factors: the physiological characteristics of the individual performing the
68 chewing action, such as facial anatomy, gender, age, personality type, time of day, or dentition
69 status, as well as the properties of the food being chewed, such as hardness, moisture content,
70 fat content, food portion size, or food structure, all have an effect on the formation of the food
71 bolus (Bornhorst & Singh, 2012). The bolus is eventually swallowed when its structural
72 characteristics have become suitable for safe swallowing.

73 Over the years, researchers from different disciplines such as nutrition, pharmacy, medicine or
74 dentistry have been working on this subject. However, it is in the last decade that food
75 technology research has fully approached oral processing, with enormous interest, as the
76 bridge between food texture, microstructure and sensory perception (Stieger & Van de Velde,
77 2013). As it constitutes a short step (about 20–30 seconds) in the overall ingestion process
78 compared with the length of the gastric and intestinal stages (1–10 hours), it has often been
79 neglected in studies such as those dealing with food digestion.

80 *In vitro* studies covering bioavailability, determination of the carbohydrate glycaemic index,
81 transportation and absorption of nutrients, flavor release, evaluation of the satiety potential of
82 ingredients or whole food systems, etc. are only a few examples of current interests in the
83 area of food science and technology research where the release of some food components
84 from their physicochemical dietary matrix is necessary. This release begins in the mouth.
85 Depending on the scope of each specific study, the selection of methods for mimicking oral
86 actions in *in vitro* studies has to consider a number of factors.

87 The principal aim of the present work is to give an overview of the main strategies that could
88 be used in Food Technology research for *in vitro* studies in which oral processing plays a role.
89 For this purpose, it offers a review of the main equipment and techniques that have been
90 designed to reproduce human mouth processing, emphasizing the newest of these. The
91 physiological actions they attempt to imitate are necessary references and are also described.
92 This paper will help Food Technology researchers to choose the proper tool for their *in vitro*
93 studies.

94

95 **Oral comminution**

96 *In vivo* scenario

97 The oral breakdown or disruption of food during mastication is highly variable, depending on
98 the food itself (texture, dryness, hardness, size) and on the characteristics of each person
99 (dental health, degree of hunger, particular habits). Many authors have pointed out that the
100 pre-swallow bolus is characterized by a specific particle size distribution that is similar across
101 individuals for the same food (Jalabert-Malbos, Mishellany-Dutour, Woda, & Peyron, 2007;
102 Mishellany, Woda, Labas, & Peyron, 2006). Nevertheless, some studies have also revealed
103 important inter-individual differences in food bolus formation and in chewing behavior (Loret,
104 *et al.*, 2011; Tárrega, Yven, Sémon, & Salles, 2011; Tournier, Grass, Zope, Salles, & Bertrand,
105 2012).

106 A recent study by Hwang *et al.* (2012), with banana, tofu, cooked rice, and biscuits eaten by
107 healthy subjects, showed that the particle size distribution of the ready-to-swallow bolus
108 depended essentially on food type and on mechanical properties of the food such as hardness,
109 cohesiveness, and adhesiveness, and not on individual differences. Mishellany *et al.* (2006),
110 working with three nuts and three vegetables, showed that the sizes of the bolus particles just
111 before swallowing were comparable in all subjects, whereas the number of cycles and duration
112 of sequences varied widely between individuals. They stated that fracture and fragmentation
113 of food (ingestion involving fracture of particles by the incisors) were closely correlated with
114 the ratio of toughness to Young's modulus in foods with approximate linear stress-strain
115 relationships (the stress-strain gradient provides the Young's modulus value of the food and
116 toughness is the work required to fracture it). Since the stress-strain relations of a number of
117 food products are distinctly nonlinear, more complex fracture models have to be introduced in
118 these cases (Lucas, Prinz, Agrawal, & Bruce, 2004). Of course, other factors such as water
119 content, the ability to absorb saliva (Hutchings & Lillford, 1988) and the fibrous structure of the
120 food also influence the way in which they are broken down (Mishellany, Woda, Labas, &
121 Peyron, 2006).

122 In a study by Jalabert-Malbos *et al.* (2007), foods that were swallowed rapidly (14–20
123 masticatory cycles) were soft and had a high water content, like egg white, pickled cucumbers,
124 mushrooms or olives. The boluses obtained from these foods contained many large particles.
125 Harder foods such as coconuts and carrots needed more cycles and longer mastication before
126 swallowing, probably because more time was needed to process the food and to disrupt the
127 fibers. They also needed more complete insalivation to produce a lubricated bolus that was
128 safe to swallow. To be swallowed easily, particles must be smaller than 2 mm, with the
129 exception of soft particles that are not liable to injure the upper digestive mucosae. Jalabert-
130 Malbos, Mishellany-Dutour, Woda, and Peyron (2007) showed that for a range of foods, sizes
131 from 0.4 to 4 mm with a median of around 2 mm were found in boluses when ready for

132 swallowing. Mastication reduced bread to an increasing number of small particles. Le Bleis,
133 Chaunier, Della Valle, Panouillé, and Réguerre (2013) found that mastication reduced two
134 types of bread of different textures into an increasing number of small particles. However, the
135 number of small particles did not always increase with the number of masticatory cycles,
136 probably because many small particles are lost during intermediary swallows that are not
137 generally analyzed (Jalabert-Malbos, Mishellany-Dutour, Woda, & Peyron, 2007).

138 One important parameter that describes the bolus just before swallowing is its median particle
139 size (d_{50}), defined as the theoretical sieve size through which 50% of its mass can pass
140 (Jalabert-Malbos, Mishellany-Dutour, Woda, & Peyron, 2007; Ngom, Diagne, Aïdara-Tamba, &
141 Sene, 2007). The d_{50} value is a useful way to classify foods used in masticatory evaluation
142 (Veyrune, Opé, Nicolas, Woda, & Hennequin, 2013) according to how easily they are processed
143 in the mouth to form a suitable bolus.

144 The *in vivo* results highlight two characteristics of mastication in humans. Firstly, the intra-
145 individual variability of food bolus particle size distribution is very narrow. Secondly, there is a
146 contrast between the narrow inter-individual variability of the food bolus d_{50} and the much
147 broader variability of the physiological variables among individuals, such as duration of the
148 sequence, number of strokes, and electromyographic activity (Jalabert-Malbos, Mishellany-
149 Dutour, Woda, & Peyron, 2007; Mishellany, Woda, Labas, & Peyron, 2006; Peyron, Mishellany,
150 & Woda, 2004).

151 Quantitative electromyography (EMG) has been used to explain the physiological process of
152 mastication, to assess muscle function, and also to diagnose temporomandibular disorders
153 (González, Montoya, & Cárcel, 2001). EMG emerged timidly in the late '80s (Boyar & Kilcast,
154 1986) as a new tool in texture evaluation. It is a non-invasive technique that does not interfere
155 with the mastication process and gives a detailed account of the activity of the masticatory
156 muscles. EMG offers the possibility of monitoring muscle activity during mastication (González,
157 Montoya, Benedito, & Rey, 2004; González, Montoya, & Cárcel, 2001). The results obtained
158 provide time-dependent information to characterize food texture. By monitoring the activities
159 of the facial muscles, this technique makes it possible to correlate food physics with the
160 physiology of oral processing and the sensory perception of food (González, Montoya,
161 Benedito, & Rey, 2004).

162 Electrognathography, also known as jaw tracking (JT), is a three-dimensional method for
163 tracking mandibular movements that provides information on mandibular velocity and
164 direction as well as the extent of the jaw movements.

165 EMG and JT are the methods most commonly used to study the relationships between oral
166 processing and food texture. Together with mechanical and sensory analyses, these two

167 techniques constitute a powerful combination for characterizing the complex nature of food
168 texture (Chen, 2009). A number of EMG and JT parameters are used to understand changes in
169 chewing behavior in relation to different textural properties. The typical measurements are
170 number of chews, chewing time, chewing frequency, total or mean muscle activity, peak
171 muscle activity, jaw movement amplitudes, and jaw-opening and -closing velocities, as well as
172 opening, closing, and occlusal phase durations. These parameters can be examined over the
173 complete chewing sequence or over different parts of it (Koç, Vinyard, Essick, & Foegeding,
174 2013). A new intraoral bite force recorder which would allow the study of natural mastication
175 without an increase in the occlusal vertical dimension was recently proposed by Shimada,
176 Yamabe, Torisu, Baad-Hansen, Murata, and Svensson (2012) for subsequent analysis of the
177 relation between electromyographic (EMG) activity of jaw-closing muscles, jaw movements
178 and bite force during mastication of five different types of food.

179 Oral physiology also exerts an important influence on chewing (Van der Bilt, Engelen, Pereira,
180 Van der Glas, & Abbink, 2006), as do characteristics such as bite force, chewing performance
181 and salivary flow rate. Chewing performance can be determined by quantifying the degree of
182 fragmentation through sieving artificial (for example, silicon rubber cubes) or real food. Other
183 methods involve evaluating the ability to mix and knead a food bolus using two-colored
184 chewing gum or paraffin wax (Van der Bilt, Mojet, Tekamp, & Abbink, 2010).

185 Besides teeth, masticatory muscles, and the temporomandibular joint, the tongue plays an
186 important role in orofacial motor behavior such as mastication and swallowing. As Kakizaki,
187 Uchida, Yamamura, and Yamada (2002) stated, the neuronal network plays a major role in
188 triggering and sequencing the neuromuscular events associated with movements, and the
189 tongue and masticatory muscles have been shown to be active in a well-coordinated manner
190 during semiautomatic movements. It is believed that the tongue senses the size and
191 lubrication status of food particles. Chewed food particles of the right size are pushed by the
192 elevated tongue to the back of the oral cavity (Mioche, Bourdiol, Monier, & Martin, 2002;
193 Okada, Honma, Nomura, & Yamada, 2007), while large particles are selected for further size
194 reduction. From a physiological point of view, it is the combined action of pushing, pulling, and
195 twisting by the tongue that transports the food particle, either to push it back to the molar
196 teeth for further size reduction or to pull it to the back of oral cavity for bolus formation. The
197 structural characteristics of the tongue, which is made up of 17 muscles, allow it to perform a
198 wide range of movements to seal the bolus content anteriorly and laterally and generate
199 pressure for its posterior propulsion. The videofluorography technique has made it possible to
200 track and analyze the tongue movement during mastication by gluing small lead markers to
201 the teeth and tongue surface (Taniguchi, *et al.*, 2013).

202 Nevertheless, there are other food bolus characteristics that could influence the exact
203 conditions for starting to swallow. Data involving not only granularity but also the rheological
204 properties of the food bolus need to be collected in order to gain a better understanding of the
205 link between physiological properties and the final d_{50} values observed just before swallowing.
206 It could be hypothesized that the moderate correlation seen between the number of cycles
207 and pre-swallow d_{50} reflects a need to attain certain rheological states that are partially
208 independent of particle size. Mishellany-Dutour *et al.* (2007) reported that subjects who
209 display long masticatory sequences, with many cycles, probably masticate less efficiently but
210 still need to achieve certain rheological conditions in terms of the viscosity, cohesiveness or
211 stickiness of the final bolus.

212 Recently, some devices have been developed to measure tongue function objectively during
213 swallowing. Some of these methods have limitations for measuring tongue-palate contact
214 function quantitatively. For example, dynamic palatography can be effective in showing
215 temporary changes in tongue contact position but cannot measure the amplitude of tongue
216 pressure (Taniguchi, Tsukada, Ootaki, Yamada, & Inoue, 2008). Developed for dysphagia
217 rehabilitation and often used by phoniatricians, this method consists of instrumentation which
218 records linguopalatal contacts during continuous speech and is used to evaluate areas of the
219 palate contacted by the tongue.

220 A technique reported by Kieser *et al.* (2008) allowed accurate measurement of tongue
221 pressure during swallowing, using an intraoral appliance with multichannel pressure sensors.
222 These sensors are capable of measuring absolute pressures to a chrome-cobalt palatal
223 appliance with a labial bow. However, the details of the movement of the tongue surface
224 during different functions remain unclear. Sugita, Inoue, Taniguchi, Ootaki, Igarashi, and
225 Yamada (2006) recorded tongue pressures at two sites on the palate during swallowing of
226 model gels with different consistencies, and demonstrated that bolus consistency affected the
227 tongue pressure of the anterior and posterior portions against the hard palate in different
228 ways. The results suggested that a basic pattern of tongue pressure is maintained during
229 swallowing but is modulated differently, by sensory feedback between the anterior and
230 posterior portions of the tongue, to complete the propulsion of the bolus in the oral cavity.

231

232 *In vitro* scenario

233 A few artificial mouths that simulate mastication have been developed in recent years. One of
234 these, called the chewing simulator (Salles, *et al.*, 2007), makes it possible to set and control
235 some of the masticatory variables, such as the number of masticatory cycles, the amplitude of
236 the mechanical movements or the bite force. Another, the BITE Master II, can measure

237 variables to be replicated such as fractal force and energy to fracture, but in this case only for
238 the first bite (Meullenet & Gandhapuneni, 2006).

239 Most of the existing prototypes have been developed for dental or orthodontic research and
240 use compressive forces with teeth that have anatomical shapes. However, the complex shapes
241 of natural teeth are operative because of the action of the central nervous system and it is
242 very difficult to mimic this. In most machines only one functional variable (e.g. speed,
243 deformation or piston movement) can be controlled at a time (Woda, *et al.*, 2010).

244 Other machines are oriented towards the mechanical properties of the mouth and make no
245 attempt to reproduce the conditions in which foods are processed within a closed mouth
246 (Hoebler, *et al.*, 2002). Conserva *et al.* (2008) developed a machine for *in vitro* study of the
247 stress transmitted to a bone-implant in dentistry. Daumas, Xu and Bronlund (2005) developed
248 another, called the mechatronic chewing device, to evaluate the dynamic changes in the
249 texture of foods quantitatively, reproducing human chewing behavior. In this device, the jaw
250 mechanism design first needs to be modelled and analyzed through simulations with the
251 corresponding mathematical model.

252 Arvisenet, Billy, Poinot, Vigneau, Bertrand, and Prost (2008) also developed an artificial mouth.
253 Their aim was not to reproduce the human mouth exactly but to determine whether
254 mastication conditions have an effect on the release of volatile compounds.

255 Comprehension of the physiology of taste perception is a key to preparing some food
256 products. Using a newly patented mastication simulator called AMADEUS (Automated
257 Mastication for Artificial Destructuration and Extensive Understanding of Sensoriality),
258 Guilloux, *et al.* (2013) obtained salt release kinetics and compared the results with sensory
259 data.

260 Researchers from the University of Auvergne developed a mastication simulator called the
261 Artificial Masticatory Advanced Machine (AM2) (Figure 1 and Figure 2) (Monique, *et al.*, 2007).
262 It simulates the mastication function, producing a bolus, while allowing permanent control of
263 the process and collection of the whole bolus at any time. This simulator produces a food bolus
264 with physical properties similar to those of the food bolus produced after natural mastication
265 just before deglutition of the same food. In the AM2, a number of mastication variables are
266 replicated and controlled. The experimenter can select the type of constraints exerted on the
267 food, the number of masticatory cycles, the cycle duration and the duration of the mastication
268 sequence, the force range applied to the food, the mastication chamber temperature and the
269 quantity of artificial saliva. As pre-swallow food particle size distribution is a good indicator of
270 food bolus characteristics, Mishellany-Dutour *et al.* (2011) used d_{50} to check the efficiency of
271 AM2. They compared the d_{50} particle size values obtained in healthy human subjects with

272 those obtained using the AM2. The results showed that the AM2 was able to simulate the d_{50}
273 food bolus particle size of peanuts and carrots produced by humans. Food bolus d_{50} values
274 obtained *in vitro* and *in vivo* at different times during the mastication process were also
275 similar.

276 In simulating mastication with mechanical devices, the intention has been to break down solid
277 foods into particles of a similar average size to those achieved by chewing.

278 If equipment to simulate the masticatory process is not available, the sample can simply be
279 minced. Experiments with rice, spaghetti and sweetcorn have shown that mincing is an
280 appropriate means of mimicking mastication, giving similar starch content values to the mean
281 values obtained by chewing. Hoebler, Devaux, Karinthi, Belleville, & Barry (2000) compared the
282 particle sizes of food after human mastication and *in vitro* mincing. The particles obtained after
283 human mastication were described as heterogeneous in size and shape, moist, limp, and not
284 easily wet-sieved. The results showed that mincing gave an acceptable reproduction of the
285 particle size distribution of bread, pasta and tortiglioni after *in vivo* mastication. The variability
286 in size and distribution of the minced bread particles was high, but satisfactory for the purpose
287 of *in vitro* simulation of mastication. Applied to foods of differing sizes (spaghetti and
288 tortiglioni) and physical textures (bread and pasta), mincing allowed large amounts of food to
289 be broken down, and thus seems to be a suitable means of mimicking chewing in a wide range
290 of foods. This method of breaking down food is simple, suitable for routine analysis and easy
291 to use in an *in vitro* procedure.

292 As discussed above, some devices have been developed to measure *in vivo* tongue function. A
293 technique reported by Ishihara *et al.* (2013) has established an *in vitro* evaluation system for
294 determining the deformation of both the tongue and the food, particularly tongue-palate
295 compression, using an artificial tongue made of silicone rubber and an aluminum plate that
296 mimics the hard palate in a conventional uniaxial compression apparatus. They used this
297 method to determine the fracture profiles of gels prepared from different agar sources.

298 Consequently, existing *in vitro* models can be improved by including an *in vitro* oral phase that
299 mimics chewing behavior. When exact imitation is not feasible, at least a particle size
300 characterization of the sample (prior to subsequent steps) should be carried out (Van
301 Buggenhout, *et al.*, 2010).

302

303 Quantifying the bolus particle size distribution

304 To quantify the particle size distribution of chewed foods, the method most commonly used
305 has been sieving. Image analysis (IA) is another frequently used method to characterize the
306 size and shape of the bolus particles (Hoebler, Devaux, Karinthi, Belleville, & Barry, 2000). This

307 method has been used to determine whether the size and shape properties of a ready-to-
308 swallow food bolus were independent of the subjects (Peyron, Mishellany, & Woda, 2004).
309 Chen, Khandelwal, Liu and Funami (2013) used image analysis to study the correlation
310 between the particle size distribution of food bolus and the hardness of the food. Le Bleis,
311 Chaunier, Della Valle, Panouillé, and Réguerre (2013) also used IA to characterize the degree of
312 fragmentation and heterogeneity of boluses from two types of bread. Mishellany, Woda, Labas
313 and Peyron (2006) listed a number of additional methods that have been used to quantify
314 particle size during *in vitro* digestion studies, such as laser diffraction, microscopy,
315 sedimentation analysis and diffusion of light.

316 Six natural foods using sieving and laser diffraction methods were compared by Peyron,
317 Mishellany and Woda (2004); after *in vivo* mastication, they noted that each of these two
318 methods analyzed only one interval of the full range of particle sizes. Particles smaller than the
319 aperture of the finest sieve were lost by sieving and laser diffraction lost large particles
320 because of its technical limits. Therefore, food boluses of raw vegetables consisting of larger
321 particles are better characterized by sieving but laser diffraction is the best method for
322 measuring the granularity of dry and brittle foods such as nuts, because these contain a high
323 percentage of small particles.

324 The use of IA to ascertain the particle size of food has been described as rapid, accurate and
325 reliable, providing precise particle enumeration over a wide range of sizes with detailed two-
326 dimensional data and obviating the unpleasant and time-consuming sieving and laser
327 diffraction processes. However, the IA technique has the same limitation as the sieving
328 method with respect to the range of values: the smallest particles in the food boluses are
329 missed because they are eliminated during preparation, which involves diluting, washing and
330 arranging the samples, so distribution curves obtained with IA are similar to those obtained by
331 sieving. Importantly, however, the IA technique offers an additional insight, as the particle
332 shape can be observed and quantified by the particle shape index.

333 Arvisenet, Billy, Poinot, Vigneau, Bertrand, and Prost (2008) studied food boluses with low
334 levels of distinguishable particles by using an image texture analysis technique, the grey level
335 co-occurrence matrix method (GLCM). They showed that this method can provide reliable
336 differentiation using images of apple crunched in an artificial mouth under different
337 compression movement frequency conditions and with different rotation speeds. Hoebler,
338 Devaux, Karinthi, Belleville, and Barry (2000) showed that GLCM can be used to investigate
339 food bolus formation during mastication of different breads and different types of pasta. The
340 use of GLCM textural features for image classification enabled an average of 67% of images to
341 be classified correctly into their respective chewing cycles. Tournier, Grass, Zope, Salles, and

342 Bertrand (2012) used GLCM in four different breads and identified contrast as the best marker
343 of food degradation.

344 Hence, the choice of one method rather than another will depend on both the goal of the
345 proposed study and the nature of the food. It should also be considered that whatever
346 technique is used, not all the particles will be spat out even when the material obtained by
347 rinsing out the oral cavity is added to the sample (Mishellany, Woda, Labas, & Peyron, 2006).

348

349 **Insalivation**

350 *In vivo* scenario

351 The oral food stage is short but it also plays another important role: hydrating and lubricating
352 the food by mixing it with saliva. The saliva interacts with the food components, leading to
353 structure formation or structure breakdown (Chen, 2009).

354 Human saliva is a complex biological fluid, consisting mainly of water (99.5% w/w), various
355 proteins (0.3% w/w), small organic compounds and inorganic salts. It has a pH of around 6.8,
356 rising to around 7-8 after food ingestion. Saliva is typically secreted at a rate of about 0.2 to 4
357 ml per minute, with a total saliva output of 500 to 1500 mL per day (McClements & Li, 2010).
358 Resting or unstimulated salivary flow is the result of low-level autonomic stimulation by the
359 higher brain centers. Salivary secretion is upregulated above the resting rate by taste and
360 chewing and to a lesser degree by smell stimulation (Carpenter, 2013).

361 The major protein component of human saliva is mucin. Other proteins in saliva include
362 various enzymes such as α -amylase, immunoglobulins, antibacterial proteins, proline-rich
363 proteins (up to 45 % of the total weight of protein) and peptides such as histatins and cystatins
364 (Sarkar, Goh, & Singh, 2009). The parotid gland contributes the greatest flow (as much as 60%
365 of the total) to stimulated saliva but less to resting salivary flow. It secretes a serous substance
366 that contains no mucins but is rich in amylase and in proline-rich proteins. The submandibular
367 and sublingual glands contribute more to the resting salivary flow rate and their saliva is rich in
368 mucins. Mucins are high-molecular-weight glycoproteins with an elongated structure that
369 contribute significantly to the viscoelastic behavior of saliva.

370 Amylase is the single most abundant protein in saliva and is involved in the initial digestion of
371 starch-containing foods. Because of this, when the food under study is rich in starch the oral
372 digestion step has been taken into consideration, as in studies with potatoes (Parada &
373 Aguilera, 2009), pasta (Petitot, *et al.*, 2009) or a starch-based custard dessert (Engelen, *et al.*,
374 2003). During insalivation, which is particularly important for starchy semi-fluid foods, the
375 rapid action of salivary amylase reduces the viscosity (Hoebler, *et al.*, 2002).

376 Since the activity of salivary amylase is greatly reduced as soon as it reaches the acidic
377 environment of the stomach, pancreatic amylase is much more likely to be involved in the
378 digestion of starch in foods, in the opinion of Carpenter (2013). Also, in studies on pancreatic
379 digestion pancreatic activity has been found to overwhelm salivary amylase activity, so
380 Woolnough, Bird, Monro, & Brennan (2010) considered that oral digestion can be neglected.
381 Structural variability among foods can give rise to different rates of starch hydrolysis as a
382 consequence of their different degree of accessibility to enzymes. Hoebler, Devaux, Karinthi,
383 Belleville, and Barry (2000) found that in cereal-based products, about 50% of bread starch and
384 25% of pasta starch were hydrolyzed during the short period of oral processing. Butterworth,
385 Warre, and Ellis (2011) stated that some uncertainty still remains with regard to the
386 physiological significance of salivary amylase. According to Nantanga, Chan, Suleman, Bertoft,
387 and Seetharaman (2013), who worked with cooked starch treated with saliva from six
388 participants at equal activity under conditions mimicking oral digestion, further research is
389 needed to understand whether the hydrolyzate structure obtained, rather than the level of
390 amylase activity, is the determinant of oral digestion of starch.

391 Lingual lipase is another salivary digestive enzyme. This enzyme breaks down a small fraction
392 of dietary triglycerides in the oral cavity and stomach. However, lingual lipase is considered to
393 be of limited significance in lipolysis for healthy individuals (Pedersen, Bardow, Jensen, &
394 Nauntofte, 2002).

395 Many factors such as the flow rate, time of day, type and size of the salivary glands, duration
396 and type of the stimulus, diet, drugs, age, sex and blood type affect the amount and
397 composition of saliva secreted in humans (Vingerhoeds, Blijdenstein, Zoet, & Van Aken, 2005).
398 When subjects display marked differences in their saliva composition their potential for oral
399 interaction with food may differ, as in the subsequent release and perception of taste
400 compounds (Neyraud, Palicki, Schwartz, Nicklaus, & Feron, 2012). The role of saliva in the
401 perception of the taste, flavor and texture of foods has been also taken into account. During
402 consumption, food mixes with saliva, so it is not the food itself but the products of its
403 interactions with saliva which we perceive. Consequently, the role of saliva in perception
404 appears to be essential (Neyraud, Palicki, Schwartz, Nicklaus, & Feron, 2012). For example, the
405 action of the enzyme α -amylase, initiating the digestion of starch, can result in a drop in the
406 perceived thickness of certain food products, as commented above. In addition, the large
407 salivary proteins influence lubrication and hence, possibly, the perception of attributes such as
408 smoothness and astringency (Engelen, *et al.*, 2003). Saliva also plays a major role in the
409 detection and perception of fat, as it is directly involved in the orosensory detection of
410 triglycerides and their hydrolysis products (Feron & Poette, 2013).

411 The perception of texture attributes is strongly related to the way the food is processed during
412 food intake, mastication, and swallowing and during the cleaning of the mouth after
413 swallowing. It is also modulated by the interaction with other basic properties, such as taste
414 and aroma attributes. The most important dynamic feature of an eating process in association
415 with texture perception is the change of length scale. Understanding the in-mouth processes
416 at the colloidal scale turned out to be essential to grasping the interplay between perception,
417 oral physiology and food properties. In this regard, two aspects have to be taken into account:
418 first, food particles are chewed and reduced in size from centimeter scale initially to sub-
419 millimeter scale at the point of swallowing, and second, a thick film of food-saliva mixture
420 between oral surfaces (i.e. tongue and hard palate) is gradually reduced to a final thin film of a
421 few micrometers (Van Vliet, Van Aken, de Jongh, & Hamer, 2009). These changes have
422 important implications for the perceived texture and, more importantly, for the underpinning
423 mechanisms applied for texture perception (Chen & Stokes, 2012).

424 Saliva acts as a buffering system (De Almeida, Grégio, Machado, de Lima, & Azevedo, 2008),
425 affecting the degree to which sourness is perceived. Significant decreases in perception with
426 increasing salivary flow rates were observed for citric acid and sodium chloride. Although this
427 can partially be explained by a dilution effect, bitterness and sweetness remained unaffected
428 by the salivary flow conditions (Heinzerling, Stieger, Bult, & Smit, 2011).

429

430 *In vitro* scenario

431 The important role of saliva in the oral processing of foods makes it clear that saliva needs to
432 be used in *in vitro* studies. Exact reproduction of human saliva is especially difficult because of
433 its complexity, unstable character and inter-individual variability, as well as its dependence on
434 the type of saliva stimulation (Roger-Leroi, Mishellany-Dutour, Woda, Marchand, & Peyron,
435 2012). In addition, its complex composition varies over the day. It is thus only possible to
436 imitate an average saliva composition (Gal, Fovet, & Adib-Yadzi, 2001).

437 The compositional complexity of simulated saliva fluids (SSF) used in the literature varies
438 widely depending on the objectives of the research. Some researchers use a simple buffer
439 solution without any additional component to simulate oral conditions. Others use simulated
440 saliva fluids that contain many of the components found in human saliva, such as acids,
441 buffers, minerals, mucins and enzymes (McClements & Li, 2010). In the food technology field,
442 in studies where digestion processes are to be emulated, the SSF should be as similar as
443 possible to naturally occurring saliva. For example, Van Ruth, Grossmann, Geary, and
444 Delahunty (2001) found that significant differences in the volatility of compounds when

445 artificial saliva or water was added indicated that saliva replacement was inadequate in aroma
446 release studies.

447 Some recipes for preparing simulated saliva solutions can be found in the literature (Björklund,
448 Ouwehand, & Forssten, 2011; Gal, Fovet, & Adib-Yadzi, 2001; Leung & Darvell, 1997;
449 Mishellany-Dutour, *et al.*, 2011; Sarkar, Goh, & Singh, 2009).

450 As mentioned above, during oral processing the effect of saliva on the food can lead to
451 impressive changes in rheological and other related properties. Saliva acts as a glue, holding
452 the fragmented solid particles together. The lubrication or tribological qualities of saliva are
453 central to many of its food processing roles, such as facilitating the swallowing of the food
454 bolus and its transport through the body. Surprisingly, according to Bongaerts, Rossetti and
455 Stokes (2007) there are few studies on the lubricating properties of whole human saliva in
456 terms of how it is influenced by surface roughness or surface compliance.

457 The results from the *in vitro* study carried out by Engelen *et al.* (2003) suggested that for a
458 semi-solid food like custard, breakdown by α -amylase in the mouth is limited because the time
459 it spends in the mouth (about 4-5 seconds) is too short for the saliva and custard to become
460 properly mixed, so the effects of breakdown are undoubtedly present but not extensive. In
461 contrast, during mastication of solids the mixing is more vigorous, and probably more efficient,
462 enabling the enzyme to come into contact with more starch particles rather than being
463 confined to the initial surface. Therefore, enzyme activity is more valuable for breaking down
464 solid foods that remain in the mouth for a longer time, such as bread and other cereal
465 products. Using a mixing simulator, Prinz, Janssen and de Wijk (2007) demonstrated with video
466 images of the recovered samples that saliva-induced structure breakdown exerts a dramatic
467 effect on the viscosity of starch-based custards despite the incomplete mixing of custard and
468 saliva that occurs *in vivo*. Several authors (Ferry, Hort, Mitchell, Lagarrigue, & Pamies, 2004;
469 Sorba & Sopade, 2013) used the Rapid Visco Analyser (Newport Scientific, Warriewood,
470 Australia) to measure the decrease in viscosity over time on adding amylase to starch pastes.

471 To quantify the susceptibility of starch-based semisolid foods to salivary α -amylase and the
472 rate of enzyme-induced structure breakdown, Janssen, Terpstra, de Wijk, and Prinz (2007)
473 developed a measuring system, the Structure Breakdown Cell (SBC), consisting of a helical
474 rotating vane. This system aims to achieve near-perfect mixing with saliva while monitoring
475 the resulting change in the torque required to rotate the vane through the food sample. The
476 use of complex geometries in rotational rheometry offers numerous benefits for the
477 mechanical characterization of saliva-induced breakdown, compared with the conventional
478 geometries used in rotational rheometry, as it is more effective in simulating the mixing
479 process in the mouth and tracking the evolution of the structure.

480 “Melting”, defined by Engelen *et al.* (2003) as the rate of decrease in thickness and spreading
481 of the product in the mouth, is a sensory attribute that could be affected considerably by the
482 presence of salivary enzymes. Since starch is broken down by the salivary enzyme α -amylase,
483 sensory melting could be affected more by saliva than by water. However, why does saliva
484 affect melting more than an α -amylase solution? A possible reason is that the α -amylase in the
485 water solution is less active than in saliva. Early work by Erickson (1992) has provided support
486 for this explanation by showing that the presence of chloride ions is essential for α -amylase to
487 reach full activity. The molecular basis for this effect was further studied by Qian, Ajandouz,
488 Payan, and Nahoum (2005). Studies performed with mice have indicated that α -amylase is
489 more active in saliva than in the gland. It can therefore be speculated that other components
490 of saliva (e.g. hydrolyzing enzymes) or products originating in microorganisms can also
491 influence the activity of salivary α -amylase. The choice of kinetic models for studying starch
492 amylolysis *in vitro* is also a subject of some controversy (Butterworth, Warren, & Ellis, 2011).
493 As described above, several masticatory apparatuses have been employed to date to produce
494 a food bolus with the closest possible resemblance to that resulting from *in vivo* chewing. To
495 achieve the goal of producing the expected food bolus, Roger-Leroi, Mishellany-Dutour, Woda,
496 Marchand, and Peyron (2012) stated that it is mandatory to develop artificial saliva with
497 chemical and rheological characteristics that are close to those of human saliva and proposed
498 a formulation that satisfies the major requirement of viscosity.

499

500 **Bolus formation**

501 Bolus characterization

502 Understanding the dynamic changes in food structure that take place during oral processing is
503 a key factor for texture design. A knowledge of bolus rheology is one of the more important
504 approaches to such understanding. From a rheological point of view, the bolus should behave
505 as a weak gel for ease of mastication and swallowing. A homogeneous and cohesive state
506 allows the mass flow of bolus through the pharyngeal phase, increasing swallowing comfort
507 (Ishihara, Nakauma, Funami, Odake, & Nishinari, 2011).

508 Prinz and Lucas (1997) stated that the decisive factor for swallowing should be the combined
509 effect of particle size and oral lubrication with the participation of saliva. According to these
510 authors the optimum moment for swallowing is defined in terms of a peak cohesive force
511 between food particles: a swallow should be triggered when it is sensed that a batch of food
512 particles is binding together under viscous forces so as to form a bolus. As Chen and Lolivret
513 (2011) commented, experimental evidence suggests that rather than maximum consistency,
514 appropriate flow-ability is a likely trigger point for swallowing. They proved this with different

515 food boluses expectorated by volunteers and simulated boluses made with SSF, using a tensile
516 method in which the boluses were stretched vertically and the force at separation was
517 recorded as a function of stretching distance. Some other experimental evidence in the
518 literature supports this premise. With the help of magnetic resonance imaging (MRI) and
519 videofluorescence techniques, for example, Buettner, Beer, Hannig, and Settles (2001)
520 observed that a food bolus became highly stretched or extensionally deformed during
521 swallowing. This was further confirmed by Kumagai, Tashiro, Hasegawa, Kohyama, and
522 Kumagai (2009), who observed the velocity profile of various bolus flows in the pharynx by the
523 Ultrasonic Pulse Doppler method. Pereira, Gavião, Engelen, and van der Bilt (2007)
524 demonstrated that the addition of fluid could significantly reduce the number of chewing
525 cycles for some dry foods because of enhanced bolus flowability in the presence of extra fluid.
526 The importance of bolus stretchability was also confirmed by Seo, Hwang, Han, and Kim (2007)
527 on investigating sensory and instrumental slipperiness and compliance of foods during
528 swallowing by human subjects using non-invasive techniques. All this experimental evidence
529 suggests that maximum consistency is not a criterion for the point of swallowing and that the
530 key criterion in swallowing is stretchability (Chen and Lolivret (2011).

531 Peyron *et al.* (2011) were also of the opinion that particle size and bolus hardness are not the
532 only decisive factors in the swallowing threshold, since d_{50} and hardness values barely change
533 after the middle of the masticatory sequence. Particle size (Peyron, Mishellany, & Woda,
534 2004), lubrication by saliva and bolus wetting (Gavião, Engelen, & Van der Bilt, 2004) are initial
535 contributing factors to the final rheological values obtained for the swallowing threshold.

536 On the other hand, the several critical thresholds for swallowing may not be reached
537 simultaneously in a bolus: the swallowing threshold is probably an integrative process that
538 combines the perceptions of the various bolus properties enabling swallowing (Peyron, *et al.*,
539 2011). Evidently, the swallowing threshold comprises many components. As formation of a
540 swallowable bolus is assumed to be a key driving constraint, to avoid dangerous aspiration of
541 small particles, each individual uses his or her physiological resources to chew a given food
542 until a safe bolus is made and the swallowing threshold is reached.

543

544 Current techniques for studying bolus rheology

545 Ishihara, Nakauma, Funami, Odake, and Nishinari (2011) listed a number of techniques for
546 inspecting the physiology of swallowing, such as videoendoscopy, the ultrasonic (ultrasound)
547 method and acoustic analysis, not only for clinical studies but also for texture studies
548 (Kumagai, Tashiro, Hasegawa, Kohyama, & Kumagai, 2009; Saitoh, *et al.*, 2007). Other
549 techniques such as Doppler velocimetry might allow direct information concerning bolus

550 velocity to be obtained without the need to track the boundaries of a bolus (e.g. in
551 videofluoroscopy) (Engmann & Burbidge, 2013).

552 Videofluorography (VI) (Okada, Honma, Nomura, & Yamada, 2007; Ono, Hori, Masuda, &
553 Hayashi, 2009) and the real-time MRI technique (Buettner, Beer, Hannig, & Settles, 2001;
554 Kulinna-Cosentini, Schima, & Cosentini, 2007), both developed for medical applications, have
555 been used successfully to provide insight into the visual evidence of food transformation and
556 transportation at different stages of oral processing (Figure 3). It is foreseeable that the use of
557 such imaging techniques, together with the classic mechanical and sensory methods, will be a
558 powerful combination in characterizing food texture (Chen, 2009).

559 VI is currently one of the best ways of evaluating the swallowing function because it enables
560 visualization of the movement of all the anatomical components related to chewing and
561 swallowing (Ono, Hori, Masuda, & Hayashi, 2009). These components include the lips, cheeks,
562 jaw, tongue, hyoid bone, pharynx, larynx, and esophagus. This technique also makes it possible
563 to visualize the passage of a food or drink containing a contrast medium (typically barium
564 sulfate powder or soluble iodine complexes) in two dimensions (sagittal and frontal). However,
565 its application involves radiation exposure and is therefore limited to patients with severe
566 dysfunction in chewing and swallowing.

567 Kulinna-Cosentini *et al.* (2007) have proved that MRI is a feasible, non-invasive method for
568 swallowing evaluations because it has excellent potential for providing fully three-dimensional
569 static images of the gastroesophageal junction and its anatomical structures involved in
570 swallowing, and their degree of variation. In comparison to VI, MRI offers several advantages:
571 it provides a better evaluation of soft tissues, the ability to acquire various series of images
572 with excellent time resolution, and – if adequately processed, which is no trivial challenge
573 (Engmann & Burbidge, 2013) – the possibility of resolving three-dimensional details from
574 different angles without changing the patient's position, but its main advantage is the lack of
575 ionizing radiation to the patient.

576 Currently, these physiological measurements suffer from limitations. For instance,
577 videoendoscopy presents low quantitative performance because of the 2D projection
578 character of the technique. The ultrasonic method is applicable preferably to females, as they
579 lack the thyroid cartilage which could interfere with the transit of the ultrasonic pulse. Acoustic
580 analysis is an alternative approach for recording swallowing profiles that has been utilized for
581 diagnostic purpose as a non-invasive method in both healthy and dysphagic individuals
582 (Lazareck & Moussavi, 2004), but has been used less in the field of food technology.

583 Despite the aid of the above techniques, difficulties in measuring the rheological properties of
584 boluses still remain owing to personal physiological differences, including mastication ability

585 and saliva secretion, which sometimes lead to poor reproducibility of experiments. This could
586 be one of the reasons why more research on bolus rheology has been conducted from a
587 physiological perspective, in medical research, than by food scientists from the food
588 technology point of view. Different stages of the swallowing mechanism, which involve
589 different fluid mechanics regimes (from creeping flow to turbulent flow conditions) depending
590 on the boundary conditions and bolus rheology, need to be studied (Engmann & Burbidge,
591 2013). It is important for food scientists to establish experimental procedures to prepare a
592 bolus *in vitro* with high reproducibility (Ishihara, Nakauma, Funami, Otake, & Nishinari, 2011).

593

594 *In silico* scenario

595 The last few decades have been witnessing the rise of alternative research models, the so-
596 called *in silico* approaches, using computational environments. The expression *in silico*,
597 imitating the common biological Latin expressions *in vivo* and *in vitro*, refers to performing
598 experiments using computers (Noori & Spanagel, 2013).

599 *In silico* models are gaining importance in the food science and technology field. The
600 development and validation of such models require more and more in-depth knowledge of the
601 physiological mechanisms of mastication. Mathematical models of oral processing are
602 proposed, generally based on geometrical considerations, to emulate certain physiological
603 features during mastication. *In vitro*, *in vivo* and *in silico* approaches have been compared
604 when studying the dynamics of the perception of saltiness and solute release from model dairy
605 products of varying composition and rheological behavior (Panouillé, *et al.*, 2010). In another
606 study, the mechanical human mastication of commercial breakfast cereals was modelled by
607 using X-ray tomography data to quantify crack propagation in brittle airy products (Hedjazi,
608 Guessasma, Martin, Della Valle, & Dendievel, 2012). Le Révérend, Loret and Hartmann (2012)
609 studied how force is distributed along the mandibular arch and how force distribution is
610 related to the space available to fit foods between the teeth.

611 *In silico* models have found a number of applications in characterizing mastication. Of special
612 interest are the studies on aroma release and its particularities, some of which are more
613 closely related to oral processing. Tréléa *et al.* (2008) described a mechanistic mathematical
614 model for aroma release in the oropharynx reaching the nasal cavity during consumption of
615 flavored yogurt. The model was based on the physiology of the swallowing process and was
616 validated via mass spectrometry measurements of aroma concentration. According to the
617 authors, this work constitutes a first step towards computer-aided product formulation. An
618 elastohydrodynamic model of swallowing was developed by De Loubens, Magnin, Doyennette,
619 Tréléa, and Souchon (2011) to quantify physical mechanisms that explain pharyngeal mucosa

620 coating. Considering complex physiological conditions, the results were applied to predicting
621 aroma release kinetics. Using a coupled biomechanical-SPH (Smoothed Particle
622 Hydrodynamics) model, Harrison *et al.* (2012) studied food breakdown and flavor release
623 during mastication. SPH is a numerical method that allows complexities such as fluid free
624 surfaces or solid fracture and interactions with complicated deforming boundaries and
625 chemical dynamics to be modelled. De Loubens, Magnin, Doyennette, Tréléa, and Souchon
626 (2010) developed an experimental device in order to gain insight into the biomechanics of the
627 pharyngeal peristalsis; the results demonstrated the influence of food bolus viscosity on flavor
628 release. Délérís *et al.* (2012) developed a mathematical model of mass transfer in the mouth
629 during eating that made it possible to identify the parameters and properties associated with
630 the product, or with the subject eating the product, that explain stimuli release in the mouth.
631 To examine the effect of various oral and gastric factors, the disintegration profiles obtained
632 by measuring the mass retention of different artificially masticated boluses were fitted to a
633 linear-exponential model, demonstrating that the bread structure and moisture content were
634 key features controlling the process (Bornhorst & Singh, 2012).
635 Model predictions have generally been in good agreement with the experimental data, so, *in*
636 *silico* approaches could be a promising tool in food oral processing studies.

637

638 **Conclusions**

639 While we are eating, a whole series of transformations take place in the mouth before
640 swallowing. Thanks to research in a number of very different disciplines we are gradually but
641 constantly learning more about these processes, and in greater detail.

642 Physically, the food is broken down in the mouth into smaller particles in preparation for the
643 following stages: gastric and intestinal digestion. Physiologically, the processes that take place
644 in the mouth must be viewed from three different angles. The first is the beginning of starch
645 digestion, thanks to the α -amylase in the saliva, the second is the chewing process (number of
646 chews, chewing time, chewing frequency, bite force, fracture energy, oral – or simulation
647 chamber – temperature, quantity and type of saliva) in relation to the food involved (size,
648 shape, viscosity, cohesiveness, hardness, stickiness) and the third is that the particles obtained
649 have to be formed into a cohesive, hydrated bolus that can be swallowed safely and
650 comfortably.

651 While it is practically impossible to reproduce such a complicated mechanism as in-mouth
652 processing, there are tools that can achieve similar results.

653 Researchers should ask themselves which steps, in relation to the food in question and the
654 parameters to be analyzed, necessarily precede the procedures they wish to apply in their
655 study.

656 The choice of one method or another will depend on the physical state of the food (liquid or
657 solid), and its initial mechanical and structural properties. For example, a researcher who
658 wishes to study how a food's texture affects its consumer acceptability needs to consider the
659 in-mouth sensations aroused by all the chewing and insalivation mechanisms involved through
660 to formation of the bolus to be swallowed, and not merely measure some single mechanical
661 property as an indicator of texture, while the researcher who wants to know how the lipids
662 contained in a given food could be digested by pancreatic lipases needs to consider which of
663 the structural breakdowns the food undergoes is responsible for releasing the fat from the
664 matrix. In addition, a cohesive, consistent bolus has many different properties to those of a
665 food that is simply minced and diluted in water or in artificial saliva. The question is: do all
666 these differences affect the results of my study?

667 The path of research related to the oral processing of food is very broad and many crossroads
668 and shortcuts may be encountered along the way. Only a profound knowledge of the
669 processes and a clear vision of the aims of the study will make it possible to take the right
670 course.

671

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676

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948 **Figure captions**

949 Figure 1. General view of the Artificial Masticatory Advanced Machine (AM2) masticator
950 simulator.

951 Figure 2. The AM2 masticatory chamber. It is a cylindrical cavity whose two ends are formed by
952 the stationary “maxillary disk” and the moving “mandibular disk”; this can move back and
953 forth along and rotate around the central axis of the cylinder. Both AM2 disks are shown in the
954 different positions during operation.

955 Figure 3. Oral and pharyngeal segments of a subject. Dynamic sequence in the sagittal view
956 shows a normal peristaltic wave with propagation of the bolus. Upper: during rest; middle: at
957 the beginning of swallowing; bellow: complete swallowing (velopharyngeal closure prevents
958 nasal penetration). Left: videofluorography images; right: magnetic resonance images.

959