

Published in final edited form as:

Best Pract Res Clin Endocrinol Metab. 2012 October ; 26(5): 701–708. doi:10.1016/j.beem.2012.03.012.

Developmental Origins of Diabetes: The Role of Oxidative Stress

Rebecca A. Simmons

Department of Pediatrics Children's Hospital Philadelphia and University of Pennsylvania,
Philadelphia, Pennsylvania 19104

Abstract

The 'thrifty phenotype' hypothesis proposes that the fetus adapts to an adverse intrauterine milieu by optimizing the use of a reduced nutrient supply to ensure survival, but by favoring the development of certain organs over that of others, this leads to persistent alterations in the growth and function of developing tissues. This concept has been somewhat controversial, however recent epidemiological, clinical, and animal studies provide support for the developmental origins of disease hypothesis. Underlying mechanisms include reprogramming of the hypothalamic-pituitary-adrenal axis, islet development, and insulin signaling pathways. Emerging data suggests that oxidative stress and mitochondrial dysfunction may also play a critical role in the pathogenesis of type 2 diabetes in individuals who were growth retarded at birth.

Keywords

intrauterine growth retardation; type 2 diabetes; Barker hypothesis; fetal origins of adult disease

Introduction

It is becoming increasingly apparent that the in utero environment in which a fetus grows and develops may have long-term effects on subsequent health and survival (1, 2). The landmark cohort study of 300,000 men by Ravelli and colleagues showed that exposure to the Dutch famine of 1944–45 during the first half of pregnancy resulted in significantly higher obesity rates at age 19 (3). Subsequent studies demonstrated a relationship between low birth weight and the later development of cardiovascular disease (4) and impaired glucose tolerance (5–7) in men in England. Those men who were smallest at birth (2.5 kg) were nearly seven times more likely to have impaired glucose tolerance or type 2 diabetes than were those who were heaviest at birth. In addition, the investigators found a similar relationship between lower birth weight and higher systolic blood pressure and triglyceride levels (8). Subsequent studies in diverse populations through the world have demonstrated a significant correlation between low birth weight and the later development of type 2 diabetes (9–18). More recent studies controlling for the confounding factors of socioeconomic status and lifestyle factors have further strengthened the association with low birth weight and increased risk of coronary heart disease, stroke, and type 2 diabetes (11, 18). In 1976 the Nurses' Health Study was initiated and a large cohort of U.S. women born from 1921 to 1946 was established. The associations with low birth weight and increased risk of coronary

© 2012 Elsevier Ltd. All rights reserved.

Correspondence, Proofs, and Reprint requests: Rebecca Simmons M.D., University Pennsylvania, BRB II/III, Rm 1308, 421 Curie Blvd, Philadelphia, PA 19104, Tele: 215-746-5139, Fax: 215-573-7627, rsimmons@mail.med.upenn.edu.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

heart disease, stroke, and type 2 diabetes remain strong even after adjusting for lifestyle factors such as smoking, physical activity, occupation, income, dietary habits, and childhood socio-economic status and occur independently of the current level of obesity or exercise (18). In a study of 22,000 American men, those born lighter than 5.5 lb had a significantly higher incidence of adult hypertension and type 2 diabetes compared with average birth weight adults (11). Similar to the Nurses Health Study, the association between birth weight and later disease is largely independent of the lifestyle risk factors (11).

Recent observations have shown that impaired growth in infancy and rapid childhood weight gain exacerbate the effects of impaired prenatal growth. The highest risk for the development of type 2 diabetes is among adults who were born small and become overweight during childhood (17, 19–21).

The mechanisms underlying the association between size at birth and impaired glucose tolerance or type 2 diabetes are unclear. A number of studies in children and adults have shown that non or pre-diabetic subjects with low birth weight are insulin resistant and thus predisposed to development of type 2 diabetes (9, 12–15, 20–27). Intrauterine growth retardation (IUGR) is known to alter the fetal development of adipose tissue, which is closely linked to the development of insulin resistance (15, 28, 29). Other studies have shown that the adverse effect of intrauterine growth retardation on glucose homeostasis was mediated through programming of the fetal endocrine pancreas (1, 30, 31). Jensen and colleagues (31) measured insulin secretion and insulin sensitivity in a well-matched Caucasian population of 19-year-old glucose tolerant men with birth weights of either below the 10th percentile (small for gestational age-SGA) or between the 50th–75th percentile (controls). To eliminate the major confounders such as "diabetes genes", none of the participants had a family history of diabetes, hypertension, or ischemic heart disease. There was no difference between the groups with regard to current weight, body mass index (BMI), body composition, and lipid profile. When controlled for insulin sensitivity, insulin secretion was reduced by 30%. However insulin sensitivity was normal in the SGA subjects. The investigators hypothesized that defects in insulin secretion may precede defects in insulin action and that once SGA individuals accumulate body fat, they will develop insulin resistance (31).

What Animal Models Can Tell Us

Animal models have a normal genetic background upon which environmental effects during gestation or early postnatal life can be tested for their role in inducing diabetes. For a comprehensive survey of the numerous animal models of fetal growth retardation, the reader is referred to two excellent reviews (32,33). The most commonly used animal models are caloric or protein restriction, glucocorticoid administration, or induction of uteroplacental insufficiency in the pregnant rodent. In the rat, maternal dietary protein restriction (approximately 40–50% of normal intake) throughout gestation and lactation has been reported to alter glucose homeostasis and hypertension in the adult offspring (34–39). Offspring are significantly growth retarded, remain growth retarded throughout life, and in some cases develop mild β -cell secretory abnormalities (34–38) and in others insulin resistance (36,39). Aged rats develop hyperglycemia characterized by defects in insulin signaling in muscle, adipocytes, and liver (39–43).

Fetal overexposure to glucocorticoids either via maternal administration or by inhibition of placental 11 β -hydroxysteroid Dehydrogenase Type 2 (11 β HSD2) in the rat induces hypertension, glucose intolerance and abnormalities in hypothalamic-pituitary-adrenal (HPA) function after birth (44–47).

To extend these experimental studies of growth retardation, we developed a model of uteroplacental insufficiency (IUGR) induced by bilateral uterine artery ligation at day 18 of gestation (term is 22 days) in the rat that restricts fetal growth (48,49). Growth retarded fetal rats have critical features of a metabolic profile characteristic of growth retarded human fetuses: decreased levels of glucose, insulin, insulin-like-growth factor 1 (IGF-I), amino acids, and oxygen (50–52). By 6 months of age, IUGR rats develop diabetes with a phenotype remarkably similar to that observed in the human with type 2 diabetes: progressive dysfunction in insulin secretion and insulin action. Thus, the studies in various animal models support the hypothesis that an abnormal intrauterine milieu can induce permanent changes in glucose homeostasis after birth and lead to type 2 diabetes in adulthood.

Cellular Mechanisms: Mitochondrial dysfunction and oxidative stress

The intrauterine environment influences development of the fetus by modifying gene expression in both pluripotential cells or terminally differentiated, poorly replicating cells. The long-range effects on the offspring (into adulthood) depend upon the cells undergoing differentiation, proliferation, and/or functional maturation at the time of the disturbance in maternal fuel economy. The fetus also adapts to an inadequate supply of substrates (such as glucose, amino acids, fatty acids, and oxygen) by metabolic changes, redistribution of blood flow, and changes in the production of fetal and placental hormones which control growth.

The fetus' immediate metabolic response to placental insufficiency is catabolism: it consumes its own substrates to provide energy. A more prolonged reduction in availability of substrates leads to a slowing in growth. This enhances the fetus' ability to survive by reducing the use of substrates and lowering the metabolic rate. Slowing of growth in late gestation leads to disproportion in organ size, since organs and tissues that are growing rapidly at the time are affected the most.

Uteroplacental insufficiency, caused by such disorders as preeclampsia, maternal smoking and abnormalities of uteroplacental development, is one of the most common cause of fetal growth retardation. The resultant abnormal intrauterine milieu restricts the supply of crucial nutrients to the fetus thereby limiting fetal growth. Multiple studies have now shown that intrauterine growth retardation is associated with increased oxidative stress in the human fetus (53–59). A major consequence of limited nutrient availability is an alteration in the redox state in susceptible fetal tissues leading to oxidative stress. In particular, low levels of oxygen, evident in growth-retarded fetuses, will decrease the activity of complexes of the electron transport chain, which will generate increased levels of reactive oxygen species (ROS) (60–62). Overproduction of ROS initiates many oxidative reactions that lead to oxidative damage not only in the mitochondria but also in cellular proteins, lipids, and nucleic acids. Increased ROS levels inactivate the iron-sulfur centers of the electron transport chain complexes, and tricarboxylic acid cycle aconitase, resulting in shutdown of mitochondrial energy production.

A key adaptation enabling the fetus to survive in a limited energy environment may be the reprogramming of mitochondrial function (62–64). However, these alterations in mitochondrial function can have deleterious effects, especially in cells that have a high energy requirement, such as the β -cell. The β -cell depends upon the normal production of ATP for nutrient-induced insulin secretion (65–72) and proliferation (73). Thus, an interruption of mitochondrial function can have profound consequences for the β -cell.

Mitochondrial dysfunction can also lead to increased production of reactive oxygen species (ROS), which will lead to oxidative stress if the defense mechanisms of the cell are overwhelmed. β -cells are especially vulnerable to attacks by ROS because expression of

antioxidant enzymes in pancreatic islets is very low (74,75), and β -cells have a high oxidative energy requirement. Increased ROS impair glucose stimulated insulin secretion (73,76,77), decrease gene expression of key β -cell genes (78–84), and induce cell death (85–87).

We have found that uteroplacental insufficiency induces oxidative stress and marked mitochondrial dysfunction in the fetal β -cell (88). ATP production is impaired and continues to deteriorate with age. The activities of complexes I and III of the electron transport chain progressively decline in IUGR islets.

Mitochondrial DNA point mutations accumulate with age and are associated with decreased mtDNA content and reduced expression of mitochondrial-encoded genes in IUGR islets. Mitochondrial dysfunction results in impaired insulin secretion. These results demonstrate that IUGR induces mitochondrial dysfunction in the fetal β -cell leading to increased production of ROS, which in turn damage mtDNA (88). A self-reinforcing cycle of progressive deterioration in mitochondrial function leads to a corresponding decline in β -cell function. Finally, a threshold in mitochondrial dysfunction and ROS production is reached and diabetes ensues.

Mitochondrial dysfunction is not limited to the β -cell in the IUGR animal. IUGR animals exhibit marked insulin resistance early in life (prior to the onset of hyperglycemia), characterized by blunted whole body glucose disposal in response to insulin and impaired insulin suppression of hepatic glucose output (89). Basal hepatic glucose production is also increased (89). Oxidation rates of pyruvate, glutamate, succinate, and α -ketoglutarate are significantly blunted in isolated hepatic mitochondria from IUGR pups (prior to the onset of diabetes) (63). Rotenone-sensitive NADH- O_2 oxidoreductase activity is similar in control and IUGR mitochondria, showing that the defect responsible for decreased pyruvate, glutamate and α -ketoglutarate oxidation in IUGR liver precedes the electron transport chain and involves pyruvate and α -ketoglutarate dehydrogenases. Increased levels of manganese superoxide dismutase (MnSOD) suggest that an antioxidant response has been mounted and 4-hydroxynonenal (HNE) modification of pyruvate dehydrogenase E2 catalytic and E3 binding protein subunits suggests that HNE-induced inactivation of this key enzyme may play a role in the mechanism of injury. These results indicate that uteroplacental insufficiency impairs mitochondrial oxidative phosphorylation in the liver and this derangement predisposes the IUGR rat to increased hepatic glucose production by suppressing pyruvate oxidation and increasing gluconeogenesis (63).

Mitochondria in muscle of IUGR young adult rats, prior to the onset of hyperglycemia, exhibit significantly decreased rates of state 3 oxygen consumption with pyruvate, glutamate, α -ketoglutarate and succinate (64). Decreased pyruvate oxidation in IUGR mitochondria is associated with decreased ATP production, decreased pyruvate dehydrogenase activity and increased expression of pyruvate dehydrogenase kinase 4 (PDK4). Such a defect in IUGR mitochondria leads to a chronic reduction in the supply of ATP available from oxidative phosphorylation. Impaired ATP synthesis in muscle compromises energy-dependent GLUT4 recruitment to the cell surface, glucose transport and glycogen synthesis, which contributes to insulin resistance and hyperglycemia of type 2 diabetes (64).

Other animal models of fetal growth retardation also show mitochondrial abnormalities. Mitochondrial DNA content is reduced in liver, pancreas and skeletal muscle of male offspring of dams fed a low-protein diet during pregnancy and lactation (90,91). This was associated with reduced expression of mitochondrial DNA-encoded genes (91).

A number of recent studies in humans further suggest that mitochondrial dysfunction may contribute to type 2 diabetes. Studies using ^{13}C and ^{31}P magnetic resonance spectroscopy (MRS) have shown decreases in mitochondrial activity and increases in intramyocellular fat content in young insulin-resistant offspring of parents with type 2 diabetes, a group that has a strong tendency to develop diabetes later in life (92). Expression of genes involved in oxidative phosphorylation is reduced among patients with type 2 diabetes mellitus and insulin resistance (93), although this may be an effect rather than a cause of diabetes.

Conclusions

The combined epidemiological, clinical, and animal studies clearly demonstrate that the intrauterine environment influences both growth and development of the fetus and the subsequent development of adult diseases. There are critical specific windows during development, often coincident with periods of rapid cell division, during which a stimulus or insult may have long-lasting consequences on tissue or organ function postnatally. Birthweight is only one marker of an adverse fetal environment and confining studies to this population only may lead to erroneous conclusions regarding etiology. Studies using animal models of uteroplacental insufficiency suggest that mitochondrial dysfunction and oxidative stress play an important role in the pathogenesis of the fetal origins of adult disease.

References

1. Hales CN, Barker DJP. Type 2 diabetes mellitus: the thrifty phenotype hypothesis. *Diabetologia*. 1992; 35:595–601. [PubMed: 1644236]
2. Kermack WO. Death rates in Great Britain and Sweden. *Lancet*. 1934; 1 :698–703.
3. Ravelli GP, Stein ZA, Susser MW. Obesity in young men after famine exposure in utero and early infancy. *NEJM*. 1976; 295:349–353. [PubMed: 934222]
4. Barker DJP, Winter PD, Osmond C, Margetts B, Simmons SJ. Weight in infancy and death from ischaemic heart disease. *Lancet*. 1989; 2 :577–580. [PubMed: 2570282]
5. Hales CN, Barker DJP, Clark PMS, Cox LJ, Fall C, Osmond C, Winter PD. Fetal and infant growth and impaired glucose tolerance at age 64. *BMJ*. 1991; 303:1019–1022. [PubMed: 1954451]
6. Phipps K, Barker DJ, Hales CN, Fall CH, Osmond C, Clark PM. Fetal growth and impaired glucose tolerance in men and women. *Diabetologia*. 1993; 36:225–228. [PubMed: 8462770]
7. Fall CHD, Osmond C, Barker DJP, Clark PMS, Hales CN, Stirling Y, Meade TW. Fetal and infant growth and cardiovascular risk factors in women. *BMJ*. 1995; 310:428–432. [PubMed: 7873947]
8. Barker DJP, Hales CN, Fall CHD, Osmond C, Phipps K, Clark PMS. Type 2 diabetes mellitus, hypertension, and hyperlipidemia (syndrome X): relation to reduced fetal growth. *Diabetologia*. 1993; 36:62–67. [PubMed: 8436255]
9. Hales CN, Barker DJ. The Thrifty phenotype hypothesis. *Br Med Bull*. 2001; 60:5–20. [PubMed: 11809615]
10. Valdez R, Athens MA, Thompson GH, Bradshaw BS, Stern MP. Birthweight and adult health outcomes in a biethnic population in the USA. *Diabetologia*. 1994; 37:624–531. [PubMed: 7926349]
11. Curhan GC, Willett WC, Rimm EB, Spiegelman D, Ascherio AL, Stampfer MJ. Birthweight and adult hypertension, diabetes mellitus and obesity in US men. *Circulation*. 1996; 94:3246–3250. [PubMed: 8989136]
12. Lithell HO, McKeigue PM, Berglund L, Mohsen R, Lithell UBN, Leon DA. Relation of size at birth to non-insulin dependent diabetes and insulin concentrations in men aged 50–60 years. *BMJ*. 1996; 312:406–410. [PubMed: 8601111]
13. McKeigue PM, Lithell HO, Leon DA. Glucose tolerance and resistance to insulin-stimulated glucose uptake in men aged 70 years in relation to size at birth. *Diabetologia*. 1998; 41:1133–1138. [PubMed: 9794098]

14. Leger J, Levy-Marchal C, Bloch J, Pinet A, Chevenne D, Porquet D, Collin D, Czernichow P. Reduced final height and indications for insulin resistance in 20 year olds born small for gestational age: regional cohort study. *BMJ*. 1997; 315:341–347. [PubMed: 9270455]
15. Jaquet D, Gaboriau A, Czernichow P, Levy-Marchal C. Insulin resistance early in adulthood in subjects born with intrauterine growth retardation. *J Clin Endocrin Metab*. 2000; 85:1401–1406.
16. Egeland GM, Skjaerven R, Irgrens LM. Birth characteristics of women who develop gestational diabetes: population based study. *BMJ*. 2000; 321:546–547. [PubMed: 10968815]
17. Forsen T, Eriksson J, Tuomilehto J, Reunanen A, Osmond C, Barker D. The fetal and childhood growth of persons who develop type 2 diabetes. *Ann Int Med*. 2000; 133:176–182. [PubMed: 10906831]
18. Rich-Edwards JW, Colditz GA, Stampfer MJ, Willett WC, Gillman MW, Hennekens CH, Speizer FE, Manson JE. Birthweight and the risk for type 2 diabetes mellitus in adult women. *Ann Intern Med*. 1999; 130:278–284. [PubMed: 10068385]
19. Eriksson J, Forsen T, Tuomilehto J, Osmond C, Barker D. Fetal and childhood growth and hypertension in adult life. *Hypertension*. 2000; 36 :790–94. [PubMed: 11082144]
20. Bavdekar A, Sachdev HS, Fall CHD, Osmond C, Lakshmy R, Barker DJP, Biswas SKD, Ramji S, Prabhakaran D, Reddy KS. Relation of Serial Changes in Childhood Body-Mass Index to Impaired Glucose Tolerance in Young Adulthood. *NEJM*. 2004; 350:865–875. [PubMed: 14985484]
21. Bavdekar A, Yajnik CS, Fall CH, Bapat S, Pandit AN, Deshpande V, Bhawe S, Kellingray SD, Joglekar C. Insulin resistance syndrome in 8-year-old Indian children: small at birth, big at 8 years, or both? *Diabetes*. 1999; 48:2422–2429. [PubMed: 10580432]
22. Hoffman PL, Cutfield WS, Robinson EM, Bergman RN, Menon RK, Sperling MA, Gluckman PD. Insulin resistance in short children with intrauterine growth retardation. *J Clin Endocrinol Metab*. 1997; 82 :402–406. [PubMed: 9024226]
23. Li C, Johnson MS, Goran MI. effects of low birth weight on insulin resistance syndrome in Caucasian and African-American children. *Diabetes Care*. 2001; 24:2035–2042. [PubMed: 11723079]
24. Yajnik CS, Fall CH, Vaidya U, Pandit AN, Bavdekar A, Bhat DS, Osmond C, Hales CN, Barker DJ. Fetal growth and glucose and insulin metabolism in four-year-old Indian children. *Diabet Med*. 1995; 12 :330–336. [PubMed: 7600749]
25. Clausen JO, Borch-Johnsen K, Pedersen O. Relation between birth weight and the insulin sensitivity index in a population sample of 331 young, healthy Caucasians. *Am J Epidemiol*. 1997; 146:23–31. [PubMed: 9215220]
26. Flanagan DE, Moore VM, Godsland IF, Cockington RA, Robinson JS, Phillips DI. Fetal growth and the physiological control of glucose tolerance in adults: a minimal model analysis. *Am J Physiol Endocrinol Metab*. 2000; 278:E700–E706. [PubMed: 10751205]
27. Phillips DI, Barker DJ, Hales CN, Hirst S, Osmond C. Thinness at birth and insulin resistance in adult life. *Diabetologia*. 1994; 37:150–154. [PubMed: 8163048]
28. Widdowson, EM.; Southgate, DAT.; Hey, EN. Nutrition and metabolism of the fetus and infant. Visser, HKA., editor. The Hague: Martinus Nijhoff; 1979. p. 169-177.
29. Lapillonne A, Braillon P, Chatelain PG, Delmas PD, Salle BD. Body composition in appropriate and small for gestational age infants. *Acta Paediatr*. 1997; 86:196–200. [PubMed: 9055893]
30. Van Assche FA, De Prins F, Aerts L, Verjans F. The endocrine pancreas in small-for dates infants. *Br j Obstet Gynaecol*. 1977; 84:751–753. [PubMed: 336076]
31. Jensen CB, Storgaard H, Dela F, Holst JJ, Madsbad S, Vaag AA. Early differential defects of insulin secretion and action in 19-year-old Caucasian men who had low birth weight. *Diabetes*. 2002; 51:1271–1280. [PubMed: 11916955]
32. Fowden AL, Forhead AJ. Endocrine mechanisms of intrauterine programming. *Reproduction*. 2004; 127:515–526. [PubMed: 15129007]
33. McMillen C, Robinson JS. Developmental origins of the metabolic syndrome: Prediction, plasticity, and programming. *Physiol Rev*. 2005; 85:571–633. [PubMed: 15788706]
34. Dahri S, Snoeck A, Reusens-Billen B, Remacle C, Hoet JJ. Islet function in off-spring of mothers on low-protein diet during gestation. *Diabetes*. 1991; 40:115–20. [PubMed: 1748239]

35. Snoeck A, Remacle C, Reusens B, Hoet JJ. Effect of a low protein diet during pregnancy on the fetal rat endocrine pancreas. *Biol Neonate*. 1990; 57:107–118. [PubMed: 2178691]
36. Ozanne SE, Wang CL, Coleman N, Smith GD. Altered muscle insulin sensitivity in the male offspring of protein-malnourished rats. *Amer Journ Physiol*. 1996; 271:E1128–E1134.
37. Berney DM, Desai M, Palmer DJ, Greenwald S, Brown A, Hales CN, Berry CL. The effects of maternal protein deprivation on the fetal rat pancreas: major structural changes and their recuperation. *J Pathol*. 1997; 183:109–115. [PubMed: 9370956]
38. Wilson MR, Hughes SJ. The effect of maternal protein deficiency during pregnancy and lactation on glucose tolerance and pancreatic islet function in adult rat offspring. *J Endocrinology*. 1997; 154:177–185. [PubMed: 9246952]
39. Burns SP, Desai M, Cohen RD, Hales CN, Iles RA, Germain JP, Going TCH, Bailey RA. Gluconeogenesis, glucose handling, and structural changes in livers of the adult offspring of rats partially deprived of protein during pregnancy and lactation. *J Clin Inves*. 1997; 100:1768–1774.
40. Ozanne SE, Jensen CB, Tingey KJ, Storgaard H, Madsbad S, Vaag AA. Low birthweight is associated with specific changes in muscle insulin-signalling protein expression. *Diabetologia*. 2005; 48:547–52. [PubMed: 15729577]
41. Ozanne SE, Olsen GS, Hansen LL, Tingey KJ, Nave BT, Wang CL, Hartil K, Petry CJ, Buckley AJ, Mosthaf-Seedorf L. Early growth restriction leads to down regulation of protein kinase C zeta and insulin resistance in skeletal muscle. *J Endocrinol*. 2003; 177:235–41. [PubMed: 12740011]
42. Petry CJ, Dorling MW, Pawlak DB, Ozanne SE, Hales CN. Diabetes in old male offspring of rat dams fed a reduced protein diet. *Int J Exp Diabetes Res*. 2001; 2:139–43. [PubMed: 12369717]
43. Fernandez-Twinn DS, Wayman A, Ekizoglou S, Martin MS, Hales CN, Ozanne SE. Maternal protein restriction leads to hyperinsulinemia and reduced insulin-signaling protein expression in 21-mo-old female rat offspring. *Am J Physiol Regul Integr Comp Physiol*. 2005; 288:R368–73. [PubMed: 15514105]
44. Benediktsson R, Lindsay R, Noble J, Seckl JR, Edwards CRW. Glucocorticoid exposure *in utero*, a new model for adult hypertension. *Lancet*. 1993; 341:339–341. [PubMed: 8094115]
45. Lindsay RS, Lindsay RM, Edwards CRW, Seckl JR. Inhibition of 11 β -hydroxysteroid dehydrogenase in pregnant rats and the programming of blood pressure in the offspring. *Hypertension*. 1996; 27:1200–1204. [PubMed: 8641724]
46. Lindsay RS, Lindsay RM, Waddell B, Seckl JR. Programming of glucose tolerance in the rat: role of placental 11 β -hydroxysteroid dehydrogenase. *Diabetologia*. 1996; 39:1299–1305. [PubMed: 8932995]
47. Niyirenda MJ, Seckl JR. Intrauterine events and the programming of adulthood disease: the role of fetal glucocorticoid exposure. *Int J Mol Med*. 1998; 2:607–614. [PubMed: 9858661]
48. Simmons RA, Templeton L, Gertz S, Niu H. Intrauterine Growth Retardation Leads to Type II Diabetes in Adulthood in the Rat. *Diabetes*. 2001; 50:2279–2286. [PubMed: 11574409]
49. Boloker J, Gertz S, Simmons RA. Offspring of Diabetic Rats Develop Obesity and Type II Diabetes in Adulthood. *Diabetes*. 2002; 51:1499–1506. [PubMed: 11978648]
50. Ogata ES, Bussey M, Finley S. Altered gas exchange, limited glucose, branched chain amino acids, and hypoinsulinism retard fetal growth in the rat. *Metabolism*. 1986; 35:950–977. [PubMed: 3020345]
51. Simmons RA, Gounis AS, Bangalore SA, Ogata ES. Intrauterine growth retardation: Fetal glucose transport is diminished in lung but spared in brain. *Pediatr Res*. 1991; 31:59–63. [PubMed: 1594332]
52. Unterman T, Lascon R, Gotway M, Oehler D, Gounis A, Simmons RA, Ogata ES. Circulating levels of insulin-like growth factor binding protein-1 (IGFBP-1) and hepatic mRNA are increased in the small for gestational age fetal rat. *Endocrinology*. 1990; 127:2035–2037. [PubMed: 1698152]
53. Myatt L, Eis ALW, Brockman DE, Kossenjans W, Greer IA, Lyall F. Differential localization of superoxide dismutase isoforms in placental villous tissue of normotensive, pre-eclamptic, and intrauterine growth-restricted pregnancies. *J Histochem Cytochem*. 1997; 45:1433–1438. [PubMed: 9313805]

54. Karowicz-Bilinska A, Suzin J, Sieroszewski P. Evaluation of oxidative stress indices during treatment in pregnant women with intrauterine growth retardation. *Med Sci Monit.* 2002; 8:CR211–216. [PubMed: 11889459]
55. Ejima K, Nanri H, Toki N, Kashimura M, Ikeda M. Localization of thioredoxin reductase and thioredoxin in normal human placenta and their protective effect against oxidative stress. *Placenta.* 1999; 20:95–101. [PubMed: 9950150]
56. Kato H, Yoneyama Y, Araki T. Fetal plasma lipid peroxide levels in pregnancies complicated by preeclampsia. *Gynecol Obstet Invest.* 1997; 43 :158–61. [PubMed: 9127127]
57. Bowen RS, Moodley J, Dutton MF, Theron AJ. Oxidative stress in pre-eclampsia. *Acta Obstet Gynecol Scand.* 2001; 80:719–25. [PubMed: 11531614]
58. Wang Y, Walsh SW. Increased superoxide generation is associated with decreased superoxide dismutase activity and mRNA expression in placental trophoblast cells in pre-eclampsia. *Placenta.* 2001; 22 :206–212. [PubMed: 11170825]
59. Wang Y, Walsh SW. Placental mitochondria as a source of oxidative stress in pre-eclampsia. *Placenta.* 1998; 19:581–6. [PubMed: 9859861]
60. Esposti MD, McLennan H. Mitochondria and cells produce reactive oxygen species in virtual anaerobiosis: relevance to ceramide-induced apoptosis. *FEBS Letters.* 1998; 430:338–342. [PubMed: 9688567]
61. Chandel NS, Budinger GRS, Schumacker PT. Molecular oxygen modulates cytochrome c oxidase function. *J Biol Chem.* 1996; 271:8672–18677.
62. Gorgias N, Maidatsi P, Tsolaki M, Alvanou A, Kiriazis G, Kaidoglou K, Giala M. Hypoxic pretreatment protects against neuronal damage of the rat hippocampus induced by severe hypoxia. *Brain Res.* 1996; 714:215–25. [PubMed: 8861628]
63. Peterside IE, Selak MA, Simmons RA. Impaired oxidative phosphorylation in hepatic mitochondria of growth retarded rats alters glucose metabolism. *American Journal of Physiology.* 2003; 285:E1258–1264. [PubMed: 14607783]
64. Selak MA, Storey BT, Peterside IE, Simmons RA. Impaired Oxidative Phosphorylation in Skeletal Muscle Contributes to Insulin Resistance and Hyperglycemia. *American Journal of Physiology.* 2003; 285:E130–E137. [PubMed: 12637257]
65. Panten U, Zielman S, Langer J, Zunkler BJ, Lenzen S. Regulation of insulin secretion by energy metabolism in pancreatic β -cell mitochondria. *Biochem J.* 1984; 219:189–196. [PubMed: 6372787]
66. Newgard CB, McGarry JD. Metabolic coupling factors in pancreatic β -cell signal transduction. *Annu Rev Biochem.* 1995; 64:689–719. [PubMed: 7574498]
67. Schuit F. Metabolic fate of glucose in purified islet cells. Glucose regulated anaplerosis in β -cells. *J Biol Chem.* 1997; 272:18572–18579. [PubMed: 9228023]
68. Mertz RJ, Worley JF III, Spencer BHJJ, Dukes ID. Activation of stimulus-secretion coupling in pancreatic β -cells by specific products of glucose metabolism. *J Biol Chem.* 1996; 271:4838–3845. [PubMed: 8617753]
69. Ortsater H, Liss P, Akerman KEO. Contribution of glycolytic and mitochondrial pathways in glucose-induced changes in islet respiration and insulin secretion. *Pflugers Arch Eur J Physiol.* 2002; 444:506–512. [PubMed: 12136270]
70. Antinozzi PA, Ishihara H, Newgard CB, Wollheim CB. Mitochondrial metabolism sets the maximal limit of fuel-stimulated insulin secretion in a model pancreatic beta cell. A survey of four fuel secretagogues. *J Biol Chem.* 2002; 277:11746–11755. [PubMed: 11821387]
71. Malaisse WJ, Hutton JC, Carpinelli AR, Herchuelz A, Senner A. The stimulus-secretion coupling of amino acid-induced insulin release. Metabolism and cationic effects of leucine. *Diabetes.* 1980; 29:431–437. [PubMed: 6769728]
72. Lenzen S, Schmidt W, Rustenbeck I, Panten U. 2-Ketoglutarate generation in pancreatic β -cell mitochondria regulates insulin secretory action of amino acids and 2-keto acids. *Biosci Rep.* 1986; 6 :163–169. [PubMed: 3521757]
73. Noda M, Yamashita S, Takahashi N, Eto K, Shen LM, Izumi K, Daniel S, Tsubamoto Y, Nemoto T, Lino M, Kasai H, Sharp GW, Kadowaki T. Switch to anaerobic glucose metabolism with NADH accumulation in the beta-cell model of mitochondrial diabetes. Characteristics of betaHC9

- cells deficient in mitochondrial DNA transcription. *J Biol Chem.* 2002; 277:41817–26. [PubMed: 12169697]
74. Lenzen S, Drinkgern J, Tiedge M. Low antioxidant enzyme gene expression in pancreatic islets compared with various other mouse tissues. *Free Radic Biol Med.* 1996; 20:463–366. [PubMed: 8720919]
 75. Tiedge M, Lortz S, Drinkgern J, Lenzen S. Relationship between antioxidant enzyme gene expression and antioxidant defense status of insulin-producing cells. *Diabetes.* 1997; 46:1733–1742. [PubMed: 9356019]
 76. Maechler P, Jornot L, Wollheim CB. Hydrogen peroxide alters mitochondrial activation and insulin secretion in pancreatic beta cells. *J Biol Chem.* 1999; 274:27905–27913. [PubMed: 10488138]
 77. Sakai K, Matsumoto K, Nishikawa T, Suefuji M, Nakamaru K, Hirashima Y, Kawashima J, Shirotani T, Ichinose I, Brownlee M, Araki E. Mitochondrial reactive oxygen species reduce insulin secretion by pancreatic β -cells. *Biochem Biophys Res Comm.* 2003; 300:216–222. [PubMed: 12480546]
 78. Kaneto H, Xu G, Fujii N, Kim S, Bonner-Weir S, Weir GC. Involvement of c-Jun N-terminal kinase in oxidative stress-mediated suppression of insulin gene expression. *J Biol Chem.* 2002; 277:30010–30018. [PubMed: 12011047]
 79. Kaneto HH, Xu G, Fujii N, Kim S, Bonner-Weir S, Weir GC. Involvement of protein kinase C beta 2 in c-myc induction by high glucose in pancreatic beta-cells. *J Biol Chem.* 2002; 277:3680–3685. [PubMed: 11714718]
 80. Kaneto H, Xu G, Song KH, Suzuma K, Bonner-Weir S, Sharma A, Weir GC. Activation of the hexosamine pathway leads to deterioration of pancreatic beta-cell function through the induction of oxidative stress. *J Biol Chem.* 2001; 276:31099–31104. [PubMed: 11390407]
 81. Kaneto H, Kajimoto Y, Fujitani Y, Matsuoaka T, Sakamoto K, Matsuhisa M, Yamasaki Y, Hori M. Oxidative stress induces p21 expression in pancreatic islet cells: possible implication in beta-cell dysfunction. *Diabetologia.* 1999; 42:1093–1097. [PubMed: 10447521]
 82. Jonas JC, Laybutt DR, Steil GM, Trivedi N, Pertusa JG, Van de Castele M, Weir GC, Henquin JC. High glucose stimulates early response gene c-Myc expression in rat pancreatic beta cells. *J Biol Chem.* 2001; 276:35375–35381. [PubMed: 11457846]
 83. Jonas JC, Sharma A, Hasenkamp W, Ilkova H, Patane G, Laybutt R, Bonner-Weir S, Weir GC. Chronic hyperglycemia triggers loss of pancreatic beta cell differentiation in an animal model of diabetes. *J Biol Chem.* 1999; 274:14112–14121. [PubMed: 10318828]
 84. Efanova IB, Zaitsev SV, Zhivotovsky B, Kohler M, Efendic S, Orrenius S, Berggren PO. Glucose and tolbutamide induce apoptosis in pancreatic β -cells. *J Biol Chem.* 1998; 273:22501–22507.
 85. Moran A, Zhang HJ, Olsonm LK, Harmon JS, Poitoust V, Robertson RP. Differentiation of glucose toxicity from β -cell exhaustion during the evolution of defective insulin gene expression in the pancreatic islet cell line, HIT-T15. *J Clin Invest.* 2000; 99:534–539. [PubMed: 9022089]
 86. Donath MY, Gross DJ, Cerasi E, Kaiser N. Hyperglycemia-induced β -cell apoptosis in pancreatic islets of *Psammomys obesus* during development of diabetes. *Diabetes.* 1999; 48:738–744. [PubMed: 10102689]
 87. Silva JP, Kohler M, Graff C, Oldfors A, Magnuson MA, Berggren PO, Larsson NG. Impaired insulin secretion and β -cell loss in tissue specific knockout mice with mitochondrial diabetes. *Nature Genetics.* 2000; 26:336–340. [PubMed: 11062475]
 88. Simmons RA, Suponitsky-Kroyter I, Selak MA. Progressive accumulation of mitochondrial DNA mutations and decline in mitochondrial function lead to beta-cell failure. *J Biol Chem.* 2005; 280:28785–28791. [PubMed: 15946949]
 89. Vuguin P, Raab E, Liu B, Barzilai N, Simmons RA. Hepatic Insulin Resistance Precedes the Development of Diabetes in a Model of Intrauterine Growth Retardation. *Diabetes.* 2004; 53:2617–2622. [PubMed: 15448092]
 90. Park HK, Jin CJ, Cho YM, Park doJ, Shin CS, Park KS, Kim SY, Cho BY, Lee HK. Changes of mitochondrial DNA content in the male offspring of protein-malnourished rats. *Ann N Y Acad Sci.* 2004; 1011:205–216. [PubMed: 15126298]

91. Park KS, Kim SK, Kim MS, Cho EY, Lee JH, Lee KU, Pak YK, Lee HK. Fetal and early postnatal protein malnutrition cause long-term changes in rat liver and muscle mitochondria. *J Nutr.* 2003; 133:3085–3090. [PubMed: 14519789]
92. Petersen KF, Dufour S, Befroy D, Garcia R, Shulman GI. Impaired Mitochondrial Activity in the Insulin-Resistant Offspring of Patients with Type 2 Diabetes. *N Engl J Med.* 2004; 350:664–667. [PubMed: 14960743]
93. Mootha VK, Lindgren CM, Eriksson KF, Subramanian A, Sihag S, Lehar J, Puigserver P, Carlsson E, Ridderstrale M, Laurila E, Houstis N, Daly MJ, Patterson N, Mesirov JP, Golub TR, Tamayo P, Spiegelman B, Lander ES, Hirschhorn JN, Altshuler D, Groop LC. PGC-1alpha-responsive genes involved in oxidative phosphorylation are coordinately downregulated in human diabetes. *Nature Genet.* 2003; 34:267–273. [PubMed: 12808457]

Practice Points

- Uteroplacental insufficiency, caused by such disorders as preeclampsia, maternal smoking and abnormalities of uteroplacental development, is one of the most common cause of fetal growth retardation.
- Intrauterine growth retardation is linked to the later development of type 2 diabetes.
- A key adaptation enabling the fetus to survive in a limited energy environment may be the reprogramming of mitochondrial function
- An abnormal intrauterine milieu leads to the development of mitochondrial dysfunction which in turn impairs function of key cells such as the β -cell.

Research Agenda

- The effect of antioxidants on the long-term complications of intrauterine growth retardation should be investigated.
- The underlying molecular mechanisms linking oxidative stress and the development of diabetes in individuals who were growth retarded at birth needs to be further elucidated.
- Nutritional strategies optimizing postnatal growth of IUGR babies needs further evaluation.

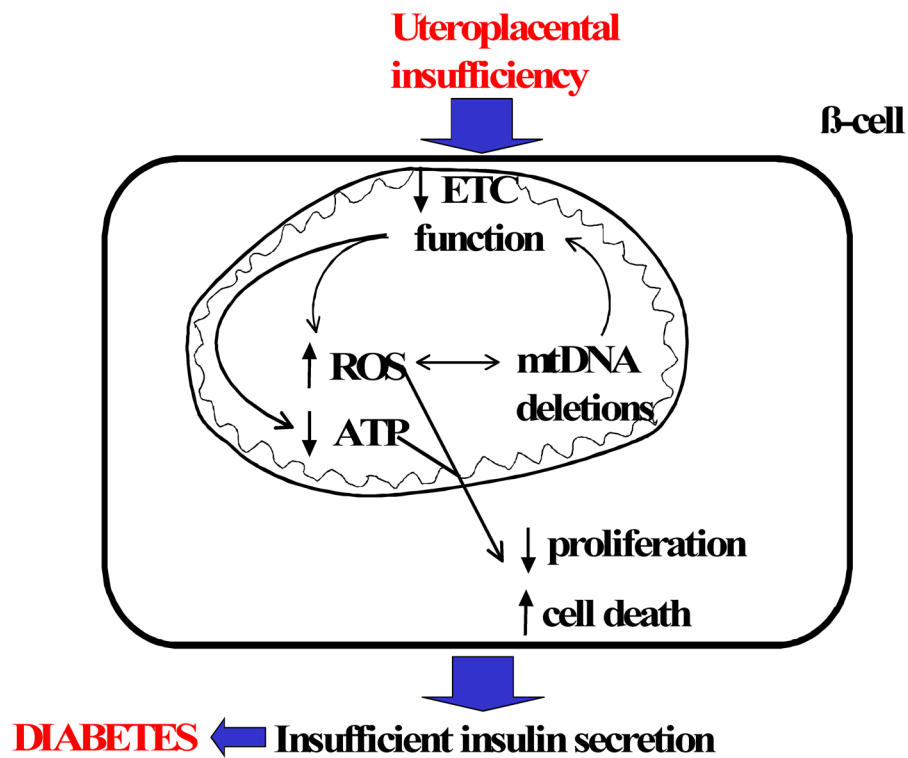


Figure 1.

Decreased nutrients and oxygen availability to the fetus result in mitochondrial dysfunction which in turn results in increased production of reactive oxygen species (ROS) and decreased production of ATP. Mitochondrial dysfunction in key cells such as the beta-cell in the pancreas decreases cell proliferation and increases cell death culminating in the development of type 2 diabetes.