ORIGINAL ARTICLE





A survey study on gastrointestinal parasites of stray cats in Azarshahr, (East Azerbaijan province, Iran)

Nasser Hajipour · Aabbas Imani Baran · Mohammad Yakhchali · Seyed Mehdi Banan Khojasteh · Farzam Sheikhzade Hesari · Bijan Esmaeilnejad · Jafar Arjmand

Received: 14 January 2015/Accepted: 9 February 2015/Published online: 28 February 2015 © Indian Society for Parasitology 2015

Abstract Gastrointestinal parasites are among the most common parasitic infections found in stray cats, which might act potential helminthic parasites to domestic cats. The objective of this study was to determine the prevalence of gastrointestinal parasites in stray cats in the city of Azarshahr, which is located in East Azerbaijan province, Iran. A cross—sectional study was conducted on 50 necropsied stray cats, trapped and collected from different geographic regions of Azarshahr. From a total 50 stray cats examined, 15 (30 %) were female and 35 (70 %) were male. Overall 47 cats (94 %) were identified as infected with at least one of the endoparasites. The prevalence of parasites found were: Taenia taeniaeformis (60 %), Dipylidium caninum (58 %), Taenia hydatigera (24 %), Mesocestoides lineatus (78 %), Ancylostoma tubaeforme (14 %), Toxascaris leonina (30 %), Toxocara cati (78 %), Physaloptera praeputialis (10 %), and Syphacia obvelata (10 %). Contamination rate for zoonotic parasites of cat was greater than expected in AzarShahr region. Therefore, appropriate control measures should be taken and preventive methods should be applied.

N. Hajipour (\boxtimes) · M. Yakhchali · B. Esmaeilnejad · J. Arjmand Department of Pathobiology, Faculty of Veterinary Medicine, Urmia University, Urmia, Iran e-mail: n.hajipour@yahoo.com

A. Imani Baran

Department of Pathobiology, Parasitology Division, Faculty of Veterinary Medicine, Tabriz University, Tabriz, Iran

S. M. Banan Khojasteh · F. Sheikhzade Hesari Department of Animal Biology, Faculty of Natural Sciences, University of Tabriz, Tabriz, Iran **Keywords** Gastrointestinal · Parasite · Stray cat · AzarShahr

Introduction

Stray cats play a main role in the epidemiology of gastrointestinal helminthic parasites. In as much as they, harbor a wide variety of parasites which can be detrimental to their and human health (Calvete et al. 1998; Hille et al. 2014; Krecek et al. 2010; Ramos et al. 2013; Scorza et al. 2011; Silaghi et al. 2014), and also represent potential reservoirs of helminthic parasites to domestic cats and human, especially in urban areas (Hille et al. 2014; Polak et al. 2014; Youssef and Uga 2014). Besides, due to their contact with both domestic cats and humans, stray cat may play an important role in the transmission of several pathogenic agents; such as Toxoplasma gondii (Can et al. 2014; Lappin 2010; Vilares et al. 2014) and Cryptosporidium (Bush et al. 2011; Scorza et al. 2011; Spada et al. 2013; Spain et al. 2001) that have the ability to cause life-threatening infections in individuals (Davoust et al. 2014; Johnson et al. 2013), or Leishmania infantum (Chatzis et al. 2014; Hatam et al. 2010; Pennisi et al. 2013; Richter et al. 2014) and Toxocara spp. which have been associated with two main clinical syndromes in humans (ocular larva migrans and visceral larva migrans) (Bowman et al. 2010; Fisher 2003; Petithory 2007; Prokopowicz and Sosnowska 1990; Rubinsky-Elefant et al. 2010). Various factors likes influence on the parasitic burden in stray cats; geographical region, season of the year, administration of antiparasitic treatment and factors related to population parameters (age, sex, breed) (Mircean et al. 2010). Previous studies have usually recorded a high prevalence of parasites in stray and feral cats (Adams et al. 2008; Becker



et al. 2012; Calvete et al. 1998; Canto et al. 2013; Capari et al. 2013; Coman 1972; Duarte et al. 2010; Engbaek et al. 1984; Jittapalapong et al. 2007; Khalafalla 2011; Krecek et al. 2010; Labarthe et al. 2004; Lefkaditis et al. 2014; Millan and Casanova 2009; Ramos et al. 2013; Riggio et al. 2013; Stoichev et al. 1982). In Iran, little is known about the prevalence of gastrointestinal parasites in stray cats. Thus, the main aim the present investigation was to determine the prevalence of gastrointestinal helminthic parasites in stray cats in this part of the country.

Materials and methods

Study areas

The study was conducted from March 2013 to October 2013 in Azarshahr (East Azerbaijan province-Iran). This city lies in the northwest of Iran and is located 1468 m above mean sea level between 37°46′ North Latitude and 45°85′ East Longitude. The studied region has a moderate climate in winters and mild in summers, with an annual rainfall of about 388 mm.

Sample collection

This study was performed based on cross sectional design. The sample cats including fifty, 15 (30 %) female and 35 (70 %) male stray cats, trapped by baited cage-traps with chicken meat or beef and collected from different areas after permission from the Environment Agency and Bureau veterinary of Azarshahr city, were sent to the Department of Animal Biology, Faculty of Natural Science, Tabriz University. For each cat, some characteristics such as breed, age (based on their teeth condition) sex, weight were registered (Eldredge et al. 2008), they were divided into three age groups (Table 1). The cats were anesthetized by diethylether and euthanized by intra muscular injection of

high doses of the Ketamine 10 % according to the protocol described by Arabali and Hooshyar with some modifications (Arbabi and Hooshyar 2009). Immediately their carcasses were autopsied and examined for the presence of protozoa, helminthes. For this purpose, after opening the abdominal cavity and removing the internal organs including stomach, intestine, kidney, liver, heart and lungs. The small intestine of each cats was detach from the mesenterium, then was cut longitudinally, immersed in phosphate buffered saline (pH 7.2) for 10 min and the mucosa was scraped with the blades of scalpel, thereafter, both the contents of the gut and scraping of the mucosa were washed with tap water in sieve NO. 60 and examined with the naked eyes as well as under a stereomicroscope (Borji et al. 2011; Zibaei et al. 2007). The examination of livers and lungs were done in two steps. First, the samples were sliced in small pieces and examined carefully under stereomicroscope in order to find the mature and immature helminthes or nymph of linguatula serrata. In the second step, the minced tissues were put in 50 ml of digestion solution containing 5 g pepsin enzyme (7178, Merck) and 25 ml hydrochloric acid (374, Merck) in 1 liter water (pH=3.5), incubating at 37 °C for 24 h. After that, the suspensions were examined (Haddadzadeh et al. 2010).

Parasitological procedures

All helminthes parasites recovered were relaxed in water, then fixed in 70 % alcohol and 5 % glycerin (Ash and Orihel 1987), nematodes were mounted and cleared in lactophenol and tapeworms were mounted and stained in acid alum carmine for identification (Ash and Orihel 1987). The identification was carried out following specific keys described by Yamaguti (1961) and Soulsby (1977). The fecal samples were examined for presence of cyst and trophozoite of protozoa, egg and larva of helminthes by direct and flotation methods using saturated sugar solution as described by Soulsby (1977).

Table 1 Distribution of the helminthic infections among age groups of the studied cats

Parasite (intensity of infection)	Age group (year)			Total n (%)	P value
	$\leq 1 \ (n = 14)$	1-2 (n = 17)	>2 (n = 19)		
Taenia taeniaeformis (1.60)	4 (28.6)	9 (52.9)	17 (89.5)	30 (60)	0.07
Mesocestoides lineatus (0.74)	4 (28.6)	16 (94.1)	19 (100)	39 (78)	0.001
Dipylidium caninum (0.60)	3 (21.4)	14 (82.3)	12 (63.1)	29 (58)	0.003
Taenia hydatigera (0.40)	2 (14.2)	5 (29.4)	5 (26.3)	12 (24)	0.48
Ancylostoma tubaeforme (0.14)	0	0	7 (36.8)	7 (14)	0.001
Toxascaris leonine (0.30)	0	2 (11.8)	13 (68.4)	15 (30)	0.001
Toxocara cati (0.82)	6 (42.8)	16 (94.1)	17 (89.5)	39 (78)	0.059
Physaloptera praeputialis (0.2)	0	1 (5.8)	0	1 (5.8)	0.58
Syphacia abvelata (0.2)	0	1 (5.8)	0	1 (5.8)	0.58



Statistical analysis

To run statistical analysis, the prevalence with 95 % confidence interval was calculated for each parasitic species. Associations between host factors (age, gender, and weight and parasitism) was evaluated using the Chi square test with SPSS software version 16.2.

Results

Of the fifty stray cats examined, forty seven (94 %) were positive for at least one of the parasite, 41 (82 %) indicated nematodes and 41 (82 %) cestodes. Mesocestoides lineatus (78 %) and Toxocara cati (78 %) were the most common parasites. Prevalence and quantity for the rest of parasites were: Taenia taeniaeformis (60 %, 80 worms), Dipylidium caninum (58 %, 30 worms), Taenia hydatigera (24 %, 19 worms), Ancylostoma tubaeforme (14 %, 7 worms), Toxascaris leonina (30 %, 15 worms), Physaloptera praeputialis (10 %, one) and Syphacia obvelata (10 %, one) (Table 1). No parasites were found in the respiratory tract, heart, liver and kidney. This study showed that there was no significant difference both in infection rate between male and female cats (P > 0.05). However, females had the higher rates of infection than males did. The association between age and prevalence of parasite, with exception of T. taeniaeformis, T. hydatigera, Ph. praeputialis, S. obvelata was significant (P < 0.05). The frequency of individual parasites in more than 2 year old cats was higher than 1-2 and $1 \ge (Table 2)$.

Discussion

Some of the parasites found in our study are zoonotic and play an important action as reservoir and these helminths represent important public health problems (Hille et al.

2014: Krecek et al. 2010: Polak et al. 2014: Youssef and Uga 2014). Therefore these, parasites must be considered as an alert to veterinaries and public health agencies. In the present study, the overall prevalence of parasitic infection (at least with one of the intestinal helminth species in stray cats of 94 % was similar to that reported in previous studies, conducted in other Iranian cities such as Isfahan (98.5 %) (Jamshidi et al. 2002), Shiraz (92.9 %) (Zibaei et al. 2007), Kashan (97.3 %) (Arbabi and Hooshyar 2009), North of Iran (90 %) (Changizi et al. 2007), Mashhad (88.46 %) (Borji et al. 2011), Kashan (95.6 %) (Arbabi and Hooshyar 2009) and in other countries, i.e. Spain (89.7, 100 %) (Calvete et al. 1998; Millan and Casanova 2009), Brazil (98. 6 %) (Labarthe et al. 2004), Egypt (91 %) (Khalafalla 2011), Nigeria (80.77 %) (Raji et al. 2013). However, the identified prevalence rate in this study was more than that of the earlier researches, conducted in different contexts such as Italy (35 %) (Riggio et al. 2013), northern Italy (50.4 %) (Spada et al. 2013) Hungary (39.6 %) (Capari et al. 2013), Germany (33.6 %) (Becker et al. 2012), Mexico (53 %) (Canto et al. 2013), Greece (55.8 %) (Lefkaditis et al. 2014), Thailand (11.9 %) (Jittapalapong et al. 2007), Brazil (67.12 %) (Ramos et al. 2013). Moreover, the most common helminth parasites, detected in our study, were M. lineatus (78 %) and T. cati (78 %). The prevalence of *T. cati* infection was higher than the estimated prevalence previously found in Christmas Island (53.57 %) (Adams et al. 2008), Northern Germany (27.1 %) (Becker et al. 2012), Spain (55.2 %) (Calvete et al. 1998), Mexico (3 %) (Canto et al. 2013), Hungary (17.4 %) (Capari et al. 2013), Thailand (3.5 %) (Jittapalapong et al. 2007), Egypt (9 %) (Khalafalla 2011), India (4 %) (Krecek et al. 2010), Brazil (25.2 %) (Labarthe et al. 2004), Greece (18.14 %) (Lefkaditis et al. 2014), Spain (35 %) (Millan and Casanova 2009), Romania (20.3 %) (Mircean et al. 2010), Australia (3.2 %), (Palmer et al. 2008), Nigeria (16.67 %) (Raji et al. 2013), England (34.8 %) (Nichol et al. 1981), Brazil (4.11 %) (Ramos

Table 2 Distribution of helminthic infections among sexes in the studied cats

Parasite (intensity of infection)	Sex n (%)	P value	
	Male $(n = 35)$	Female $(n = 15)$	
Taenia taeniaeformis	20 (57.1)	10 (66.6)	0.78
Mesocestoides lineatus	22 (77.1)	12 (80)	0.83
Dipylidium caninum	21 (60)	8 (53.3)	0.73
Taeniahydatigera	10 (28.6)	2 (13.3)	0.14
Ancylostoma tubaeforme	4 (11.4)	3 (20)	0.42
Toxascaris leonine	9 (52.7)	6 (40)	0.31
Toxocara cati	25 (71.4)	14 (93.3)	0.15
Physaloptera praeputialis	1 (2.8)	0	0.51
Syphacia abvelata	1 (2.8)	0	0.51



et al. 2013), Italy (22. 2 %) (Riggio et al. 2013), Argentina (61.2 %) (Sommerfelt et al. 2006), Northern Italy (33.1 %) (Spada et al. 2013), as well as in Iranian contexts including Shiraz (42.6 %) (Zibaei et al. 2007), Northern Iran (44 %) (Sharif et al. 2007), Mashhad (28.84 %) (Borji et al. 2011), North of Iran (8 %) (Changizi et al. 2007), Isfahan (13 %) (Jamshidi et al. 2002), Tehran (42.31 %) (Mirzayans 1973), Kashan (13.3) (Arbabi and Hooshyar 2009), Ahvaz (8.3 %), Tehran (23 %) (Bahadori et al. 2004), However, similar prevalence of T. cati infection has been recorded in Denmark (79 %) by Engback et al. (1984). The most likely reasons for the increased prevalence of T. cati in our studies were the poor hygiene, lack of anthelmintics drug used in stray cats, the presence of high humidity, and moderate temperatures (Arbabi and Hooshyar 2009). All theses factors were clearly observed in our research context. However, another invisible factor or reason or the common occurrence of this parasite can be attributed to the role of some intermediary, such as rodents and invertebrates, in the life cycle of T cati and in the diet of stray cats (Adams et al. 2008). M. lineatus was the dominant tapeworm reported in our study with a prevalence rate of 79 % which was higher than that reported in Spain (13.8 %) (Calvete et al. 1998), Italy (1.2 %) (Riggio et al. 2013), Iran (13.4 and 8 %) (Arbabi and Hooshyar 2009; Borji et al. 2011). The T. taeniaeformis prevalence (60 %) was relatively similar to that found by Coman (1972) in Australia (78 %), Stoichev et al. (1982) in Bulgaria (75.5 %), Abu-Madi et al. (2008) in Qatar (74 %) and Borthakur and Mukharjee (2011) in India (70.4 %) but the prevalence rate of this cestode in our study was higher than that recorded in Spain (8.6 and 22 %) (Calvete et al. 1998; Millan and Casanova 2009), Nile Delta of Egypt (22 %) (Khalafalla 2011), Christmas Island (7.14 %) (Adams et al. 2008), Greece (8.37 %) (Lefkaditis et al. 2014), Mexico (4 %) (Canto et al. 2013), Nigeria (6.67 %) (Raji et al. 2013), Brazil (0.68 %) (Ramos et al. 2013) and Iran (2, 9.6, 15, 12.3 and 9.2 %) (Arbabi and Hooshyar 2009; Borji et al. 2011; Changizi et al. 2007; Jamshidi et al. 2012; Zibaei et al. 2007). Coman attribute the prevalence of this cestode with the importance of rodent in the diet of these cats (Coman 1972). In our study, the prevalence of the D. caninum was 58 %, which is compatible with the findings of the previous studies done in Brazil (52.6) (Labarthe et al. 2004) and Iran (68.1 and 49.5 %) (Arbabi and Hooshyar 2009; Zibaei et al. 2007), However, the prevalence of this parasite was higher than the previously recorded rates by other surveys from various countries (Borji et al. 2011; Calvete et al. 1998; Canto et al. 2013; Dalimi and Mobedi 1992; Jittapalapong et al. 2007; Khalafalla 2011; Lefkaditis et al. 2014; Raji et al. 2013; Ramos et al. 2013). High prevalence of this parasite in resent survey could be due to the fact that D. caninum requires fleas as intermediary hosts and cats infected after ingesting infected fleas (Labarthe et al. 2004). T. hydatigera prevalence of 24 % found in the current study was higher than that reported by Borji et al. (2011) Furthermore, compared with some previous studies, such as; 5 % in Egypt (Khalafalla 2011), 7.69 and 12.9 % in Iran (Borji et al. 2011; Zibaei et al. 2007), 11.9 % in Brazil (Labarthe et al. 2004), 1 % in Mexico (Canto et al. 2013), 3.33 % in Nigeria (Raji et al. 2013), the prevalence of T. leonine was higher (30 %). But Lefkaditis et al. (2014) detected a rate of 93 % in Greece (Lefkaditis et al. 2014). The definitive hosts for *T. leonina* are both feline and canine species (Okulewicz et al. 2012). Cats become infected by ingesting either the eggs or rodents that contain the larvae of *T. leonine* which all can impact the prevalence of T. leonina. Ph. praeputialis (10 %) and S. abvelata (10 %) were the other nematode species found in the present survey. More interestingly, for the first time in the relevant research literature S. abvelata was detected in the sample stray cats of our study.

Conclusion

Findings of this study indicated a high prevalence of endoparasite infections in stay cats in this particular Iranian context. Some of the identified parasites, known as agents of zoonotic diseases, most probably deteriorate the cats' health, and the zoonotic character of some parasites found in this study must serve as an alert to public health agencies, veterinarians and pet owners (Becker et al. 2012).

Acknowledgments The authors would like to thank Mortaza Amini and Armen Badali for their laboratorial support.

References

Abu-Madi MA, Pal P, Al-Thani A, Lewis JW (2008) Descriptive epidemiology of intestinal helminth parasites from stray cat populations in Qatar. J Helminthol 82(1):59–68. doi:10.1017/s0022149x07870830

Adams PJ, Elliot AD, Algar D, Brazell RI (2008) Gastrointestinal parasites of feral cats from Christmas Island. Aust Vet J 86(1–2):60–63. doi:10.1111/j.1751-0813.2007.00246.x

Arbabi M, Hooshyar H (2009) Gastrointestinal parasites of stray cats in Kashan, Iran. Trop Biomed 26(1):16–22

Ash LR, Orihel TC (1987) Parasites: a guide to laboratory procedures and identification. ASCP Press, Chicago

Bahadori ShR, Eslami A, Meshgi B, Poor Hoseini S (2004) Study on stray cats infested with parasitic helminthes in Tehran, Iran. J Fac Vet Med Univ Tehran 59(2):171–174

Becker AC, Rohen M, Epe C, Schnieder T (2012) Prevalence of endoparasites in stray and fostered dogs and cats in Northern Germany. Parasitol Res 111(2):849–857. doi:10.1007/s00436-012-2909-7

Borji H, Razmi G, Ahmadi A, Karami H, Yaghfoori S, Abedi V (2011) A survey on endoparasites and ectoparasites of stray cats



- from Mashhad (Iran) and association with risk factors. J Parasit Dis 35(2):202–206. doi:10.1007/s12639-011-0057-0
- Borthakur SK, Mukharjee SN (2011) Gastrointestinal helminthes in stray cats (*Felis catus*) from Aizawl, Mizoram, India. Southeast Asian J Trop Med Public Health 42(2):255–258
- Bowman DD, Montgomery SP, Zajac AM, Eberhard ML, Kazacos KR (2010) Hookworms of dogs and cats as agents of cutaneous larva migrans. Trends Parasitol 26(4):162–167. doi:10.1016/j.pt.2010.01.005
- Bush JM, Speer B, Opitz N (2011) Disease transmission from companion parrots to dogs and cats: what is the real risk? Vet Clin North Am Small Anim Pract 41(6):1261–1272. doi: 10.1016/j.cvsm.2011.07.004
- Calvete C, Lucientes J, Castillo JA et al (1998) Gastrointestinal helminth parasites in stray cats from the mid-Ebro Valley, Spain. Vet Parasitol 75(2–3):235–240
- Can H, Doskaya M, Ajzenberg D et al (2014) Genetic characterization of Toxoplasma gondii isolates and toxoplasmosis seroprevalence in stray cats of Izmir, Turkey. PLoS One 9(8):e104930. doi:10.1371/journal.pone.0104930
- Canto GJ, Guerrero RI, Olvera-Ramirez AM, Milian F, Mosqueda J, Aguilar-Tipacamu G (2013) Prevalence of fleas and gastro-intestinal parasites in free-roaming cats in central Mexico. PLoS One 8(4):e60744. doi:10.1371/journal.pone.0060744
- Capari B, Hamel D, Visser M, Winter R, Pfister K, Rehbein S (2013) Parasitic infections of domestic cats, *Felis catus*, in western Hungary. Vet Parasitol 192(1–3):33–42. doi:10.1016/j.vetpar. 2012.11.011
- Changizi E, Mobedi I, Salimi-Bejestani M, Rezaei-Doust A (2007) Gastrointestinal helminthic parasites in stray cats (*Felis catus*) from North of Iran. Iran J Parasitol 2(4):25–29
- Chatzis MK, Andreadou M, Leontides L et al (2014) Cytological and molecular detection of Leishmania infantum in different tissues of clinically normal and sick cats. Vet Parasitol 202(3–4): 217–225. doi:10.1016/j.vetpar.2014.02.044
- Coman BJ (1972) A survey of the gastro-intestinal parasites of the feral cat in victoria. Aust Vet J 48(4):133–136. doi:10.1111/j.1751-0813.1972.tb09260.x
- Dalimi A, Mobedi I (1992) Helminth parasites of carnivores in the northern part of Iran. Ann Trop Med Parasitol 86(4):395–397
- Davoust B, Mediannikov O, Sambou M et al (2014) [Serological survey of animal toxoplasmosis in Senegal.]. Bull Soc Pathol Exot. doi:10.1007/s13149-014-0403-4
- Duarte A, Castro I, Pereira da Fonseca IM et al (2010) Survey of infectious and parasitic diseases in stray cats at the Lisbon Metropolitan Area, Portugal. J Feline Med Surg 12(6):441–446. doi:10.1016/j.jfms.2009.11.003
- Eldredge DM, Carlson DG, Carlson LD, Griffin JM (2008) Cats owner's home veterinary handbook, 3rd edn. John Wiley & Sons, Hoboken
- Engbaek K, Madsen H, Larsen SO (1984) A survey of helminths in stray cats from Copenhagen with ecological aspects. Z Parasitenkd 70(1):87–94
- Fisher M (2003) *Toxocara cati*: an underestimated zoonotic agent. Trends Parasitol 19(4):167–170
- Haddadzadeh H, Athari S, Abedini R et al (2010) One-humped camel (*Camelus dromedarius*) infestation with Linguatula serrata in Tabriz, Iran. J Arthropod-Borne Dis 4(1):54–59
- Hatam GR, Adnani SJ, Asgari Q et al (2010) First report of natural infection in cats with Leishmania infantum in Iran. Vector Borne Zoonotic Dis 10(3):313–316. doi:10.1089/vbz.2009.0023
- Hille K, Mobius N, Akmatov MK et al (2014) Zoonoses research in the German National Cohort: feasibility of parallel sampling of pets and owners. Bundesgesundheitsblatt Gesundheitsforschung

- Gesundheitsschutz 57(11):1277–1282. doi:10.1007/s00103-014-2047-8
- Jamshidi S, Meshgi B, Toghani M (2002) A study of helminthic infection of gastrointestinal tract in stray cats at urban, area in Isfahan. J Fac Vet Med Univ Tehran 57(2):25–27
- Jamshidi S, Maazi N, Ranjbar-Bahadori S, Rezaei M, Morakabsaz P, Hosseininejad M (2012) A survey of ectoparasite infestation in dogs in Tehran, Iran. Rev Bras Parasitol Vet 21(3):326–329
- Jittapalapong S, Inparnkaew T, Pinyopanuwat N, Kengradomkij C, Sangvaranond A, Wongnakphet S (2007) Gastrointestinal parasites of stray cats in Bangkok Metropolitan areas, Thailand. Katsetsart J Nat Sci 41:69–73
- Johnson CR, Gorla SK, Kavitha M et al (2013) Phthalazinone inhibitors of inosine-5'-monophosphate dehydrogenase from *Cryptosporidium parvum*. Bioorg Med Chem Lett 23(4): 1004–1007. doi:10.1016/j.bmcl.2012.12.037
- Khalafalla RE (2011) A survey study on gastrointestinal parasites of stray cats in northern region of Nile delta, Egypt. PLoS One 6(7):e20283. doi:10.1371/journal.pone.0020283
- Krecek RC, Moura L, Lucas H, Kelly P (2010) Parasites of stray cats (Felis domesticus L., 1758) on St. Kitts, West Indies. Vet Parasitol 172(1–2):147–149. doi:10.1016/j.vetpar.2010.04.033
- Labarthe N, Serrao ML, Ferreira AM, Almeida NK, Guerrero J (2004) A survey of gastrointestinal helminths in cats of the metropolitan region of Rio de Janeiro, Brazil. Vet Parasitol 123(1–2):133–139
- Lappin MR (2010) Update on the diagnosis and management of Toxoplasma gondii infection in cats. Top Companion Anim Med 25(3):136–141. doi:10.1053/j.tcam.2010.07.002
- Lefkaditis MA, Paştiu AI, Rodi-Buriel A et al (2014) Helminth burden in stray cats from Thessaloniki, Greece. Helminthologia 51(1):73–76. doi:10.2478/s11687-014-0211-1
- Millan J, Casanova JC (2009) High prevalence of helminth parasites in feral cats in Majorca Island (Spain). Parasitol Res 106(1):183–188. doi:10.1007/s00436-009-1647-y
- Mircean V, Titilincu A, Vasile C (2010) Prevalence of endoparasites in household cat (*Felis catus*) populations from Transylvania (Romania) and association with risk factors. Vet Parasitol 171(1–2):163–166. doi:10.1016/j.vetpar.2010.03.005
- Mirzayans A (1973) *Toxocara cati* in a new mammalian host. Vet Rec 92(10):262
- Nichol S, Ball SJ, Snow KR (1981) Prevalence of intestinal parasites in feral cats in some urban areas of England. Vet Parasitol 9(2):107–110
- Okulewicz A, Perec-Matysiak A, Buńkowska K, Hildebrand J (2012) Toxocara canis, Toxocara cati and Toxascaris leonina in wild and domestic carnivores. Helminthologia 49(1):3–10. doi: 10.2478/s11687-012-0001-6
- Palmer CS, Thompson RC, Traub RJ, Rees R, Robertson ID (2008) National study of the gastrointestinal parasites of dogs and cats in Australia. Vet Parasitol 151(2–4):181–190. doi:10.1016/j.vetpar.2007.10.015
- Pennisi MG, Hartmann K, Lloret A et al (2013) Leishmaniosis in cats: ABCD guidelines on prevention and management. J Feline Med Surg 15(7):638–642. doi:10.1177/1098612x13489229
- Petithory JC (2007) Visceral and cutaneous larva migrans. Rev Prat 57(18):1977–1983
- Polak KC, Levy JK, Crawford PC, Leutenegger CM, Moriello KA (2014) Infectious diseases in large-scale cat hoarding investigations. Vet J 201(2):189–195. doi:10.1016/j.tvjl.2014.05.020
- Prokopowicz D, Sosnowska D (1990) Toxocariasis. Przegl Epidemiol 44(3):193–198
- Raji A, Magaji A, Bello M, Lawal M, Mamuda A (2013) Prevalence of gastrointestinal parasites of stray cats: a case study of two hospitals in Sokoto Metropolis, Sokoto, Nigeria. J Bacteriol Parasitol 4(175):2



- Ramos DG, Scheremeta RG, Oliveira AC, Sinkoc AL, Pacheco Rde C (2013) Survey of helminth parasites of cats from the metropolitan area of Cuiaba, Mato Grosso, Brazil. Rev Bras Parasitol Vet 22(2):201–206. doi:10.1590/s1984-29612013000200040
- Richter, Schaarschmidt K, Krudewig (2014) Ocular signs, diagnosis and long-term treatment with allopurinol in a cat with leishmaniasis. Schweiz Arch Tierheilkd 156(6):289–294. doi:10.1024/0036-7281/a000593
- Riggio F, Mannella R, Ariti G, Perrucci S (2013) Intestinal and lung parasites in owned dogs and cats from central Italy. Vet Parasitol 193(1–3):78–84. doi:10.1016/j.vetpar.2012.11.026
- Rubinsky-Elefant G, Hirata CE, Yamamoto JH, Ferreira MU (2010) Human toxocariasis: diagnosis, worldwide seroprevalences and clinical expression of the systemic and ocular forms. Ann Trop Med Parasitol 104(1):3–23. doi:10.1179/136485910x1260 7012373957
- Scorza AV, Duncan C, Miles L, Lappin MR (2011) Prevalence of selected zoonotic and vector-borne agents in dogs and cats in Costa Rica. Vet Parasitol 183(1–2):178–183. doi:10.1016/j.vetpar. 2011.06.025
- Sharif M, Nasrolahei M, Ziapour SP et al (2007) *Toxocara cati* infections in stray cats in northern Iran. J Helminthol 81(1): 63–66. doi:10.1017/s0022149x07214117
- Silaghi C, Knaus M, Rapti D et al (2014) Survey of Toxoplasma gondii and Neospora caninum, hemotropic mycoplasmas and other arthropod-borne pathogens in cats from Albania. Parasit Vectors 7:62. doi:10.1186/1756-3305-7-62

- Sommerfelt IE, Cardillo N, Lopez C, Ribicich M, Gallo C, Franco A (2006) Prevalence of *Toxocara cati* and other parasites in cats' faeces collected from the open spaces of public institutions: Buenos Aires, Argentina. Vet Parasitol 140(3–4):296–301
- Soulsby EJL (1977) Helminths, arthropods and protozoa of domesticated animals, vol 6. Lea and Febiger, Philadelphia
- Spada E, Proverbio D, Della Pepa A et al (2013) Prevalence of faecalborne parasites in colony stray cats in northern Italy. J Feline Med Surg 15(8):672–677. doi:10.1177/1098612x12473467
- Spain CV, Scarlett JM, Wade SE, McDonough P (2001) Prevalence of enteric zoonotic agents in cats less than 1 year old in central New York State. J Vet Intern Med 15(1):33–38
- Stoichev I, Janchev J, Svilenov D (1982) Helminths and pathomorphological lesions in cats from villages of Bulgaria with human endemic nephropathy. Zentralbl Vet B 29(4):292–302
- Vilares A, Gargate MJ, Ferreira I et al (2014) Isolation and molecular characterization of Toxoplasma gondii isolated from pigeons and stray cats in Lisbon, Portugal. Vet Parasitol. doi:10.1016/j.vetpar.2014.08.006
- Yamaguti N (1961) Nematodes of vertebrates Systema helminthum, vol III. Inter Science, New York
- Youssef AI, Uga S (2014) Review of parasitic zoonoses in egypt. Trop Med Health 42(1):3–14. doi:10.2149/tmh.2013-23
- Zibaei M, Sadjjadi SM, Sarkari B (2007) Prevalence of *Toxocara cati* and other intestinal helminths in stray cats in Shiraz, Iran. Trop Biomed 24(2):39–43

